

Role of coagulopathy in glucocorticoid-induced osteonecrosis of the femoral head

Qiankun Zhang^{1,*}, Jin Lv^{2,*} and Lie Jin¹

Abstract

The two major theories of glucocorticoid (GC)-induced osteonecrosis of the femoral head (ONFH) are apoptosis and ischaemia. The traditional theory implicates ischaemia as the main aetiological factor because the final common pathway of ONFH is interruption of blood supply to the bone. The most common causes of interruption of blood supply include fat embolism and coagulation disorders. GCs can directly or indirectly lead to coagulation disorders, producing a hypercoagulable state, followed by poor blood flow, ischaemia, and eventually ONFH. This review summarizes the existing knowledge on coagulation disorders in the context of GC-induced ONFH, including hypofibrinolysis and thrombophilia, endothelial cell dysfunction and damage, endothelial cell apoptosis, lipid metabolism, platelet activation, and the effect of anticoagulant treatment.

Keywords

Glucocorticoid, osteonecrosis, femoral head, ischaemia, coagulopathy

Date received: 22 January 2017; accepted: 27 February 2017

Introduction

Increased glucocorticoid (GC) levels are the most common nontraumatic cause of osteonecrosis of the femoral head (ONFH).^{1,2} GC-induced ONFH in young adults usually requires hip replacement^{3,4} However, several studies have shown poor prosthetic durability in patients with ONFH.^{5–7} A previous study showed that the mean daily GC dose was strongly associated with osteonecrosis (ON).⁸ Most cross-study analyses demonstrate that a sustained large dose of GC can induce symptomatic ON.^{9,10}

There is no widely held consensus on the pathogenesis of GC-induced ON. Several mechanisms of GC-induced ON have been proposed (Figure 1). A novel mechanism of

¹Department of Nephrology, Lishui Central Hospital, Lishui, Zhejiang, China

²Department of Neurology, Lishui People's Hospital, Lishui, Zhejiang, 323000, China

*These authors contributed equally to this work.

Corresponding author:

Lie Jin, Department of Nephrology, Lishui Central Hospital, Lishui, Zhejiang, China.

Email: 296765973@qq.com



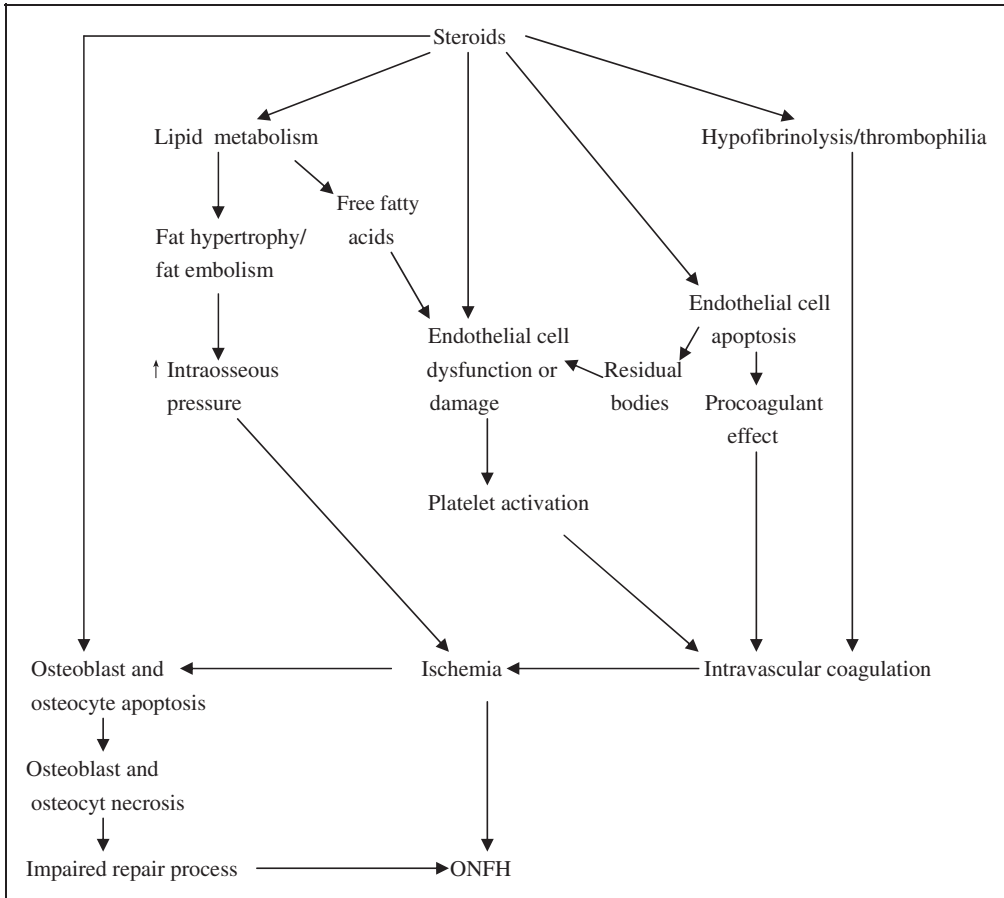


Figure 1. Plausible mechanisms for steroid-induced development of ONFH

GC-induced ON is apoptosis in osteoblasts and osteocytes, thus compromising bone formation and integrity.^{11–14} However, the traditional concept of GC-induced ON implicates ischaemia as the main aetiological factor. GCs are thought to interrupt blood supply to the bone and eventually cause ONFH in a variety of ways.^{15–18} The most common causes of interruption of the blood supply include fat embolism and coagulation disorders.^{19–22} This article summarizes existing knowledge on coagulation disorders in the context of GC-induced ON. We review the literature and highlight controversies, with emphasis on the questions of

how GC-induced coagulation disorders, directly or indirectly, relate to ischaemia in GC-induced osteonecrosis.

Hypofibrinolysis and thrombophilia

Previous studies showed that high doses of dexamethasone administered to rats inhibited fibrinolytic activity by decreasing tissue plasminogen activator (t-PA) activity and increasing plasma plasminogen activator inhibitor-1 (PAI-1) antigen levels.^{23–25} PAI-1 plays a role in fibrinolysis by forming complexes with t-PA. The t-PA/PAI-1

complexes do not have the ability to activate plasminogen to plasmin. GCs increase the activity of PAI-1, leading to hypofibrinolysis and a relatively hypercoagulable state.²⁶ Subsequent research showed decreased fibrinolytic activity, as a consequence of increased PAI-1, and decreased t-PA, by GCs in animals and patients with ON.^{18,27–29} Furthermore, as important factors of hypofibrinolysis, plasma fibrinogen and lipoprotein (a) (Lp(a)) are also abnormalities found in GC-induced or idiopathic ON.^{30–34} In an ON animal model, Drescher et al.³⁰ showed that plasma fibrinogen was significantly elevated in ON following mega-dose GC treatment, which suggested a hypercoagulable condition in GC-induced ON. In a clinical study, Pósan et al.³⁴ found that Lp(a) levels were elevated in patients with primary and secondary ONFH. Other studies have investigated the association between thrombophilia and development of ON following GC treatment.^{17,32,35–37} Guan et al.³⁵ showed that, at 24 hours after prednisolone injection, abnormal hypercoagulability occurred in a rabbit model. Glueck et al.³² compared 36 patients with primary and secondary ONFH with healthy volunteers. They found that these patients were more likely to have thrombophilic disorders, heterozygosity or homozygosity for platelet glycoprotein IIIa P1A1/A2 polymorphism, anticardiolipin antibodies, lupus anticoagulant, or both, and deficiency in proteins C and S, or antithrombin III.

However, the association between hypofibrinolysis or thrombophilia with primary or secondary ON is unclear. Seguin et al.³⁸ showed that there was no association between thrombophilia with ON and considered that GC-induced regional endothelial dysfunction was a more likely reason. Asano et al.³⁹ found that genotypes of PAI-1 4G/5G and MTHFR C677T or plasma concentrations of PAI-1 Ag and tHcy had no effect on the incidence of ONFH in

Japanese subjects, and suspected that this may differ according to race.

Endothelial cell dysfunction and damage

Endothelial dysfunction may present early in GC-induced ONFH. Yu et al.⁴⁰ found that GC significantly affected the transcriptome of vascular endothelial cells of the human femoral head. Chen et al.⁴¹ showed circulating endothelial progenitor cell damage in patients with GC-induced ONFH. In a histopathological study, Nishimura et al.⁴² found endothelial cell damage by electron microscopy in steroid-treated rabbits. Li et al.²⁷ also showed endothelial cell damage with a high coagulant and low fibrinolytic milieu in an experimental study on GC-induced ON. In patients with dysbaric osteonecrosis, Slichter et al.⁴³ found platelet thrombus formation, which was secondary to endothelial cell damage in the femoral head.

The pathogenesis of GC-induced endothelial cell dysfunction and damage is multiple, and oxidative stress may play an important role.^{44–47} After initial damage of endothelial cells triggered by GCs or other factors, a hypercoagulable state is produced. This is followed by vascular problems (thrombosis, poor blood flow, and ischaemia), and this in turn results in endothelial cell damage, which may be cyclic.^{48–51}

Endothelial cell apoptosis

GCs can induce endothelial cell apoptosis by a different signalling pathway.^{52–55} Endothelial cell apoptosis consequently promotes thrombus formation and ON by two major mechanisms. First, apoptotic bodies can indirectly lead to coagulopathic changes by endothelial dysfunction. Second, apoptotic endothelial cells can stimulate adhesion of platelets to endothelial cells and activate platelets, eventually leading to thrombus formation.⁵⁰

However, GCs can induce endothelial cell apoptosis and lead to a hypercoagulable state. Cessation or a reduction in blood flow along capillaries could also play an aetiological role in endothelial cell apoptosis.⁵⁹⁻⁶¹

Lipid metabolism

There is abundant evidence that excessive GCs can induce hyperlipidaemia, fat hypertrophy, fat deposition within the femoral head intramedullary tissue, and fat embolism. These factors may cause ischaemia by elevating intraosseous pressure and decreasing blood flow, eventually leading to ONFH.⁶²⁻⁷⁰

However, beyond the above-mentioned changes, dyslipidaemia can also lead to a hypercoagulable state and aggravate ischaemia.^{20-22,50,71} Jones et al.²² found intraosseous fibrin thromboses after induction of experimental fat emboli and speculated that fat emboli could trigger intravascular coagulation. Additionally, some vasoactive substances that are released from injured marrow adipocytes can affect endothelial cells that line blood vessels and produce a hypercoagulable state.⁵⁰

Platelet activation

High doses of GCs induce platelet aggregation.^{72,73} There is evidence that platelet activation is involved in GC-induced ON. Masuhara et al.⁷⁴ found that platelet activation may play an important role in experimental ON in rabbits. In patients with ONFH, Pósan showed that platelet activation (measured by beta triglyceride) was significantly higher compared with that in healthy controls.³⁴ Similarly, in some animal studies on GC-induced femoral head necrosis, blood platelet levels were decreased in the early stage.^{35,75} This finding indicates the occurrence of consumption coagulopathy caused by activation not only of endothelial cells, but also of platelets. Additionally, platelet thrombus formation has been

detected in arterioles adjacent to the necrotic area by histopathological observation.^{43,71,75}

In summary, platelet activation is involved in progression of GC-induced ON and the effect may be secondary to endothelial cell damage by GC.

Anticoagulant treatment

Hypofibrinolysis (decreased ability to lyse clots) and thrombophilia (increased likelihood of forming thrombi) appear to play important roles in ON. If coagulation abnormalities cause ON, then anticoagulation therapy might ameliorate it. Wada et al.⁷⁶ found that warfarin decreased the incidence of ON in spontaneously hypertensive rats. Glueck et al.⁷⁷ studied patients whose ON was caused by heritable thrombophilia or hypofibrinolysis. They showed that 12 weeks of therapy with enoxaparin before femoral head collapse may slow progression or stabilize ON, as determined by X-ray and MRI, while providing pain relief. Motomura et al.⁷⁸ demonstrated that the combined use of warfarin and probucol helps prevent steroid-induced ON in rabbits. Kang et al.⁷⁹ also found that combination treatment with enoxaparin and lovastatin reduced the incidence of GC-induced ON in the rabbit.

In summary, coagulation abnormalities may play an important role in GC-induced ON. Additionally, anticoagulation therapy can significantly decrease the incidence of ON in GC-treated rabbits.

Conclusion

This article provides an overview of the role of coagulopathy in GC-induced ON. GCs can directly lead to hypofibrinolysis and thrombophilia or indirectly lead to endothelial cell dysfunction and damage. Endothelial cell apoptosis, lipid metabolism, and platelet activation lead to a hypercoagulable state, followed by poor blood

flow, ischaemia, and eventually ONFH. Experimental studies have shown that use of an anticoagulant alone or combined with a lipid-lowering agent is beneficial in preventing GC-induced ON. Better understanding of the pathogenesis of GC-induced ON can generate better treatment options.

Declaration of conflicting interest

The Authors declare that there are no conflicts of interest.

Funding

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

References

1. Koo KH, Kim R, Kim YS, et al. Risk period for developing osteonecrosis of the femoral head in patients on steroid treatment. *Clin Rheumatol* 2002; 21: 299–303.
2. Assouline-Dayana Y, Chang C, Greenspan A, et al. Pathogenesis and natural history of osteonecrosis. *Semin Arthritis Rheum* 2002; 2: 94–124.
3. Sakaguchi M, Tanaka T, Fukushima W, et al. Impact of oral corticosteroid use for idiopathic osteonecrosis of the femoral head: a nationwide multicenter case control study in Japan. *J Orthop Sci* 2010; 15: 185–191.
4. Kubo T, Ueshima K, Saito M, et al. Clinical and basic research on steroid-induced osteonecrosis of the femoral head in Japan. *J Orthop Sci* 2016; 21: 407–413.
5. Adili A and Trousdale RT. Femoral head resurfacing for the treatment of osteonecrosis in the young patient. *Clin Orthop Relat Res* 2003; 417: 93–101.
6. Brinker MR, Rosenberg AG, Kull L, et al. Primary total hip arthroplasty using noncemented porous-coated femoral components in patients with osteonecrosis of the femoral head. *J Arthroplasty* 1994; 9: 457–468.
7. Chiu KH, Shen WY, Ko CK, et al. Osteonecrosis of the femoral head treated with cementless total hip arthroplasty: a comparison with other diagnoses. *J Arthroplasty* 1997; 12: 683–688.
8. Felson DT and Anderson JJ. Across-study evaluation of association between steroid dose and bolus steroids and avascular necrosis of bone. *Lancet* 1987; 1: 902–906.
9. Lafforgue P. Pathophysiology and natural history of avascular necrosis of bone. *Joint Bone Spine* 2006; 73: 500–507.
10. Powell C, Chang C, Naguwa SM, et al. Steroid induced osteonecrosis: an analysis of steroid dosing risk. *Autoimmun Rev* 2010; 9: 721–743.
11. Kabata T, Kubo T, Matsumoto T, et al. Apoptotic cell death in steroid induced osteonecrosis: an experimental study in rabbits. *J Rheumatol* 2000; 27: 2166–2171.
12. Manolagas SC. Birth and death of bone cells: basic regulatory mechanisms and implications for the pathogenesis and treatment of osteoporosis. *Endocr Rev* 2000; 21: 115–137.
13. Weinstein RS, Chen JR, Powers CC, et al. Promotion of osteoclast survival and antagonism of bisphosphonate-induced osteoclast apoptosis by glucocorticoids. *J Clin Invest* 2012; 109: 1041–1048.
14. Zalavras C, Shah S, Birnbaum MJ, et al. Role of apoptosis in glucocorticoid induced osteoporosis and osteonecrosis. *Crit Rev Eukaryot Gene Expr* 2003; 13: 221–235.
15. Jones JP Jr. Coagulopathies and osteonecrosis. *Acta Orthop Belg* 1999; 65(Suppl 1): 5–8.
16. Kerachian MA, Harvey EJ, Cournoyer D, et al. Avascular necrosis of the femoral head: vascular hypotheses. *Endothelium* 2006; 13: 237–244.
17. Oinuma K, Harada Y, Nawata Y, et al. Sustained hemostatic abnormality in patients with steroid-induced osteonecrosis in the early period after high-dose corticosteroid therapy. *J Orthop Sci* 2000; 5: 374–379.
18. Glueck CJ, Fontaine RN, Gruppo R, et al. The plasminogen activator inhibitor-1 gene, hypofibrinolysis, and osteonecrosis. *Clin Orthop Relat Res* 1999; 366: 133–146.
19. Fukui K, Kominami R, Shinohara H, et al. Glucocorticoid induces micro-fat embolism in the rabbit: a scanning electron

- microscopic study. *J Orthop Res* 2006; 24: 675–683.
20. Nagasawa K, Tada Y, Koarada S, et al. Very early development of steroid-associated osteonecrosis of femoral head in systemic lupus erythematosus: prospective study by MRI. *Lupus* 2005; 14: 385–390.
 21. Jones JP Jr. Intravascular coagulation and osteonecrosis. *Clin Orthop Relat Res* 1992; 277: 41–53.
 22. Jones JP Jr. Fat embolism, intravascular coagulation, and osteonecrosis. *Clin Orthop Relat Res* 1993; 292: 294–308.
 23. van Giezen JJ and Jansen JW. Correlation of in vitro and in vivo decreased fibrinolytic activity caused by dexamethasone. *Ann N Y Acad Sci* 1992; 667: 199–201.
 24. van Giezen JJ and Jansen JW. Inhibition of fibrinolytic activity in-vivo by dexamethasone is counterbalanced by an inhibition of platelet aggregation. *Thromb Haemost* 1992; 68: 69–73.
 25. van Giezen JJ, Brakkee JG, Dreteler GH, et al. Dexamethasone affects platelet aggregation and fibrinolytic activity in rats at different doses which is reflected by their effect on arterial thrombosis. *Blood Coagul Fibrinolysis* 1994; 5: 249–255.
 26. Kerachian MA, Séguin C and Harvey EJ. Glucocorticoids in osteonecrosis of the femoral head: a new understanding of mechanisms of action. *J Steroid Biochem Mol Biol* 2009; 114: 121–128.
 27. Li Y, Chen J, Zhang Z, et al. The experimental study on treatment of glucocorticoid-induced ischemic necrosis of femoral head by gu fu sheng capsule. *J Tradit Chin Med* 2004; 24: 303–307.
 28. Yamamoto Y, Ishizu A, Ikeda H, et al. Dexamethasone increased plasminogen activator inhibitor-1 expression on human umbilical vein endothelial cells: an additive effect to tumor necrosis factor- α . *Pathobiology* 2004; 71: 295–301.
 29. Ferrari P, Schroeder V, Anderson S, et al. Association of plasminogen activator inhibitor-1 genotype with avascular osteonecrosis in steroid-treated renal allograft recipients. *Transplantation* 2002; 74: 1147–1152.
 30. Drescher W, Weigert KP, Bünger MH, et al. Femoral head blood flow reduction and hypercoagulability under 24 h mega dose steroid treatment in pigs. *J Orthop Res* 2004; 22: 501–508.
 31. Hirata T, Fujioka M, Takahashi KA, et al. Low molecular weight phenotype of Apo(a) is a risk factor of corticosteroid-induced osteonecrosis of the femoral head after renal transplant. *J Rheumatol* 2007; 34: 516–522.
 32. Glueck CJ, Freiberg RA, Fontaine RN, et al. Hypofibrinolysis, thrombophilia, osteonecrosis. *Clin Orthop Relat Res* 2001; 386: 19–33.
 33. Glueck CJ, Freiberg RA and Wang P. Heritable thrombophilia-hypofibrinolysis and osteonecrosis of the femoral head. *Clin Orthop Relat Res* 2008; 466: 1034–1040.
 34. Pószán E, Hársfalvi J, Szepesi K, et al. Increased platelet activation and decreased fibrinolysis in the pathogenesis of aseptic-necrosis of the femoral head. *Platelets* 1998; 9: 233–235.
 35. Guan XY and Han D. Role of hypercoagulability in steroid-induced femoral head necrosis in Rabbits. *J Orthop Sci* 2010; 15: 365–370.
 36. Chotanaphuti T, Heebthamai D, Chuwong M, et al. The prevalence of thrombophilia in idiopathic osteonecrosis of the hip. *J Med Assoc Thai* 2009; 92(Suppl 6): S141–S146.
 37. Winkel ML, Appel IM, Pieters R, et al. Impaired dexamethasone-related increase of anticoagulants is associated with the development of osteonecrosis in childhood acute lymphoblastic leukemia. *Haematologica* 2008; 93: 1570–1574.
 38. Seguin C, Kassis J, Busque L, et al. Non-traumatic necrosis of bone (osteonecrosis) is associated with endothelial cell activation but not thrombophilia. *Rheumatology (Oxford)* 2008; 47: 1151–1155.
 39. Asano T, Takahashi KA, Fujioka M, et al. Relationship between postrenal transplant osteonecrosis of the femoral head and gene polymorphisms related to the coagulation and fibrinolytic systems in Japanese subjects. *Transplantation* 2004; 77: 220–225.
 40. Yu QS, Guo WS, Cheng LM, et al. Glucocorticoids significantly influence the transcriptome of bone microvascular

- endothelial cells of human femoral head. *Chin Med J (Engl)* 2015; 128: 1956–1963.
41. Chen C, Yang S, Feng Y, et al. Impairment of two types of circulating endothelial progenitor cells in patients with glucocorticoid-induced avascular osteonecrosis of the femoral head. *Joint Bone Spine* 2013; 80: 70–76.
 42. Nishimura T, Matsumoto T, Nishino M, et al. Histopathologic study of veins in steroid treated rabbits. *Clin Orthop Relat Res* 1997; 334: 37–42.
 43. Slichter SJ, Stegall P, Smith K, et al. Dysbaric osteonecrosis: a consequence of intravascular bubble formation, endothelial damage, and platelet thrombosis. *J Lab Clin Med* 1981; 98: 568–590.
 44. Ichiseki T, Matsumoto T, Nishino M, et al. Oxidative stress and vascular permeability in steroid-induced osteonecrosis model. *J Orthop Sci* 2004; 9: 509–515.
 45. Ichiseki T, Kaneuji A, Katsuda S, et al. DNA oxidation injury in bone early after steroid administration is involved in the pathogenesis of steroid-induced osteonecrosis. *Rheumatol (Oxford)* 2005; 44: 456–460.
 46. Kuribayashi M, Fujioka M, Takahashi KA, et al. Vitamin E prevents steroid-induced osteonecrosis in rabbits. *Acta Orthop* 2010; 81: 154–160.
 47. Iuchi T, Akaike M, Mitsui T, et al. Glucocorticoid excess induces superoxide production in vascular endothelial cells and elicits vascular endothelial dysfunction. *Circ Res* 2003; 92: 81–87.
 48. Redlich M, Maly A, Aframian D, et al. Histopathologic changes in dental and oral soft tissues in 2-butoxyethanol-induced hemolysis and thrombosis in rats*. *J Oral Pathol Med* 2004; 33: 424–429.
 49. Jones JP Jr, Ramirez S and Doty SB. The pathophysiologic role of fat in dysbaric osteonecrosis. *Clin Orthop Rel Res* 1993; 296: 256–264.
 50. Boss JH and Misselevich I. Osteonecrosis of the femoral head of laboratory animals: The lessons learned from a comparative study of osteonecrosis in man and experimental animals. *Vet Pathol* 2003; 40: 345–354.
 51. He W, Xu C, Fan Y, et al. Effects of the Chinese drugs for activating blood circulation on plasma TXB2 and 6-keto-PGF1alpha contents in rabbits with glucocorticoid-induced femoral head necrosis. *J Tradit Chin Med* 2004; 24: 233–237.
 52. Okada Y, Tanikawa T, Iida T, et al. Vascular injury by glucocorticoid: involvement of apoptosis of endothelial cells. *Clin Calcium* 2007; 17: 872–877. [in Japanese, English Abstract].
 53. Yan J, Liu Q, Dou Y, et al. Activating glucocorticoid receptor-ERK signaling pathway contributes to ginsenoside Rg1 protection against β -amyloid peptide-induced human endothelial cells apoptosis. *J Ethnopharmacol* 2013; 147: 456–466.
 54. Gaytán F, Morales C, Bellido C, et al. Selective apoptosis of luteal endothelial cells in dexamethasone-treated rats leads to ischemic necrosis of luteal tissue. *Biol Reprod* 2002; 66: 232–240.
 55. Vogt CJ and Schmid-Schönbein GW. Microvascular endothelial cell death and rarefaction in the glucocorticoid-induced hypertensive rat. *Microcirculation* 2001; 8: 129–139.
 56. Dimmeler S, Haendeler J and Zeiher AM. Regulation of endothelial cell apoptosis in athero thrombosis. *Curr Opin Lipidol* 2002; 13: 531–536.
 57. Kirwan CC, McCollum CN, McDowell G, et al. Investigation of proposed mechanisms of chemotherapy-induced venous thromboembolism: endothelial cell activation and procoagulant release due to apoptosis. *Clin Appl Thromb Hemost* 2015; 21: 420–427.
 58. Leroyer AS, Tedgui A and Boulanger CM. Role of microparticles in atherothrombosis. *J Intern Med* 2008; 263: 528–537.
 59. Shabat S, Nyska A, Long PH, et al. Osteonecrosis in a chemically induced rat model of human hemolytic disorders associated with thrombosis—a new model for avascular necrosis of bone. *Calcif Tissue Int* 2004; 74: 220–228.
 60. Langille BL, Bendeck MP and Keeley FW. Adaptations of carotid arteries of young and mature rabbits to reduced carotid blood flow. *Am J Physiol* 1989; 256(4 Pt 2): H931–H939.
 61. Azmi TI and O’Shea JD. Mechanism of deletion of endothelial cells during regression

- of the corpus luteum. *Lab Invest* 1984; 51: 206–217.
62. Jaffe WL, Epstein M, Heyman N, et al. The effect of cortisone on femoral and humeral heads in rabbits: An experimental study. *Clin Orthop Relat Res* 1972; 82: 221–228.
 63. Cui Q, Wang GJ and Balian G. Steroid-induced adipogenesis in a pluripotential cell line from bone marrow. *J Bone Joint Surg Am* 1997; 79: 1054–1063.
 64. Chen XC, Weng J, Chen XQ, et al. Relationships among magnetic resonance imaging, histological findings, and IGF-I in steroid-induced osteonecrosis of the femoral head in rabbits. *J Zhejiang Univ Sci B* 2008; 9: 739–746.
 65. Cui Q, Wang GJ, Su CC, et al. The Otto Aufranc Award. Lovastatin prevents steroid induced adipogenesis and osteonecrosis. *Clin Orthop Relat Res* 1997; 344: 8–19.
 66. Jones JP Jr. Fat embolism and osteonecrosis. *Orthop Clin North Am* 1985; 16: 595–633.
 67. Zhou Q, Li Q, Yang L, et al. Changes of blood vessels in glucocorticoid-induced avascular necrosis of femoral head in rabbits. *Zhonghua Wai Ke Za Zhi* 2000; 38: 212–215. [in Chinese, English Abstract].
 68. Cui Q, Wang GJ and Balian G. Pluripotential marrow cells produce adipocytes when transplanted into steroid-treated mice. *Connect Tissue Res* 2000; 41: 45–56.
 69. Yin L, Li YB and Wang YS. Dexamethasone-induced adipogenesis in primary marrow stromal cell cultures: mechanism of steroid-induced osteonecrosis. *Chin Med J (Engl)* 2006; 119: 581–588.
 70. Kitajima M, Shigematsu M, Ogawa K, et al. Effects of glucocorticoid on adipocyte size in human bone marrow. *Med Mol Morphol* 2007; 40: 150–156.
 71. Boss JH and Misselevich I. Osteonecrosis of the femoral head of laboratory animals: the lessons learned from a comparative study of osteonecrosis in man and experimental animals. *Vet Pathol* 2003; 40: 345–354.
 72. Liverani E, Banerjee S, Roberts W, et al. Prednisolone exerts exquisite inhibitory properties on platelet functions. *Biochem Pharmacol* 2012; 83: 1364–1373.
 73. Moraes LA, Paul-Clark MJ, Rickman A, et al. Ligand-specific glucocorticoid receptor activation in human platelets. *Blood* 2005; 106: 4167–4175.
 74. Masuhara K, Nakata K, Yamasaki S, et al. Involvement of platelet activation in experimental osteonecrosis in rabbits. *Int J Exp Pathol* 2001; 82: 303–308.
 75. Yamamoto T, Irisa T, Sugioka Y, et al. Effects of pulse methylprednisolone on bone and marrow tissues: corticosteroid-induced osteonecrosis in rabbits. *Arthritis Rheum* 1997; 40: 2055–2064.
 76. Wada M, Kumagai K, Murata M, et al. Warfarin reduces the incidence of osteonecrosis of the femoral head in spontaneously hypertensive rats. *J Orthop Sci* 2004; 9: 585–590.
 77. Glueck CJ, Freiberg RA, Fontaine RN, et al. Anticoagulant therapy for osteonecrosis associated with heritable hypofibrinolysis and thrombophilia. *Expert Opin Investig Drugs* 2001; 10: 1309–1316.
 78. Motomura G, Yamamoto T, Miyanishi K, et al. Combined effects of an anticoagulant and a lipid-lowering agent on the prevention of steroid-induced osteonecrosis in rabbits. *Arthritis Rheum* 2004; 50: 3387–3391.
 79. Kang P, Gao H, Pei F, et al. Effects of an anticoagulant and a lipid-lowering agent on the prevention of steroid-induced osteonecrosis in rabbits. *Int J Exp Pathol* 2010; 91: 35–43.