

The Pathobiology of Myocardial Recovery and Remission: From Animal Models to Clinical Observations in Heart Failure Patients

REVIEW

ARICK C. PARK, MD, PHD DOUGLAS L. MANN, MD

[*Author affiliations can be found in the back matter of this article](#page-10-0)

ABSTRACT

Heart failure with reduced left ventricular (LV) ejection fraction (HFrEF) is a morbid and lifethreatening disease, arising secondary to abnormalities of cardiac structure and function that lead to adverse LV remodeling. Implementation of medical and device therapies results in significant improvements in patient outcomes that are associated with reverse LV remodeling and improved LV ejection fraction. This review provides an overview of the pathobiology of reverse LV remodeling in animal models and in HFrEF patients. We emphasize the differences between myocardial recovery and remission as well as the fragile nature of maintaining a state of myocardial remission.

CORRESPONDING AUTHOR: Douglas L. Mann, MD

Center for Cardiovascular Research, Washington University School of Medicine, St. Louis, Missouri, US dmann@wustl.edu

KEYWORDS:

heart failure; LV remodeling; reverse LV remodeling; myocardial remission; myocardial recovery

TO CITE THIS ARTICLE:

Park AC, Mann DL. The Pathobiology of Myocardial Recovery and Remission: From Animal Models to Clinical Observations in Heart Failure Patients. Methodist DeBakey Cardiovasc J. 2024;20(4):16-30. doi: [10.14797/mdcvj.1389](https://doi.org/10.14797/mdcvj.1389)

INTRODUCTION

Reverse left ventricular (LV) remodeling represents a coordinated multi-level process that allows the failing heart to adopt a new, less pathologic steady state associated with improved pump function and improved clinical prognosis. For some, etiologies of heart failure (HF) normalization of LV structure and function and recovery from HF symptomatology may occur spontaneously once the inciting event is resolved (eg, myocarditis). For other patients with chronic HF with reduced LV ejection fraction (HFrEF), improvements in LV structure and function may occur after initiating guideline-directed medical therapy (GDMT).[1](#page-10-1) Although utilization of GDMT may reverse LV remodeling with improved LV ejection fraction (LVEF) and improved HF symptoms, for the majority of HFrEF patients treated with GDMT, LVEF and LV structural abnormalities do not fully normalize. These patients remain at increased risk of redeveloping clinical HF despite maintenance of GDMT[.1](#page-10-1) Indeed, HF patients whose ventricles resume a more normal LV size and shape will experience one of two different clinical outcomes: freedom from HF events or recurrence of HF events. Previously the authors suggested that the term "myocardial remission" should refer to the normalization of the molecular, cellular, myocardial, and LV geometric changes that provoke cardiac remodeling but that are insufficient to prevent the recurrence of HF in the face of normal and/or perturbed hemodynamic loading conditions, whereas the term "myocardial recovery" should be used to describe the normalization of the molecular, cellular, myocardial, and LV geometric heart changes that provoke cardiac remodeling, allowing the heart to maintain preserved LV structure/function in the face of normal and/ or perturbed hemodynamic loading conditions (Figure 1). [2](#page-10-2) Here we review the components of reverse LV remodeling in both experimental models and human studies. We also discuss the biological differences between myocardial recovery and remission with an emphasis on the need to maintain clinical remission from HF.

REVERSE LEFT VENTRICULAR REMODELING

Failing hearts undergo progressive LV dilation, increased sphericity, and LV wall thinning leading to ventricularafterload mismatch that contributes to decreased stroke volume. Compounding this, the dilated spherical LV geometry results in restrictive tethering of the papillary muscles, which often leads to significant functional mitral regurgitation (MR), a product of leaflet malcoaptation during ventricular systole. MR further causes enlargement and remodeling of the left atrium, resulting in mitral annular dilation, propagating worse functional MR. Aside from reducing systemic output, significant MR causes further LV volume/pressure overload and remodeling.

Figure 1 Reverse remodeling and myocardial recovery. Cardiac remodeling occurs secondary to abnormalities that arise in the biology of the cardiac myocyte (C), the myocardium (cardiocytes and extracellular matrix [M]), as well as LV geometry (LV), which have collectively been referred to as the HF phenotype. C: cardiac myocyte; M: myocardium; LV: left ventricular; HF: heart failure. Reproduced with permission from Mann DL et al.²

Collectively, these changes result in a rightward shift in the LV pressure-volume relationship, referred to as "adverse LV remodeling." In post-infarction patients, remodeling changes are associated with worse clinical outcomes compared to other seemingly more obvious factors, including LVEF or severity of coronary arterial lesions.^{3[,4](#page-10-4)} Additional longitudinal studies of patients with LV dilation but without myocardial infarctions, LV dimension, and sphericity—remain potent predictors of survival.^{5[,6](#page-10-6)}

The term "reverse LV remodeling" arose from studies that observed a leftward shift in the LV end-diastolic pressure-volume relationship towards normal values in patients who had been supported with LV assist devices (LVAD) or with a cardiomyoplasty (a novel technique developed to improve LV function by wrapping the myocardium with latissimus dorsi muscle and then pacing the skeletal muscle).^{[7-](#page-10-7)9} Importantly, while implemented, these devices allowed the heart to assume a more normal LV end-diastolic pressure-volume relationship and favorable LV elliptical geometry, which was preserved even when the LVAD was stopped/removed or pacing of the latissimus dorsi muscle was stopped. In the context

of the present discussion, it is important to recognize that the calculation of LVEF incorporates LV end-diastolic volume in the denominator of the equation. Accordingly, in the clinical setting, improvements in LVEF are generally associated with a reciprocal decrease in LV end-diastolic volume rather than improved LV contractile function per se.

BIOLOGICAL MECHANISMS OF REVERSE LV REMODELING

Remodeling events observed on the macroscopic scale, including distortion of LV geometry, deterioration of contractile function, and hemodynamic perturbations, are reflective of a dysfunctional and failing cardiac myocyte and its surrounding myocardium. Although first described as the normalization of cardiac function (LVEF $>$ 50%) and left shift of the LV end-diastolic pressure-volume relationship, reverse LV remodeling represents a return to a less pathologic state rather than a complete normalization of HF biology (Figure 2). [10](#page-10-9)

Figure 2 Proposed hypothetical model indicating that reverse left ventricular (LV) remodeling represents the summation of the complex interactions between multiple biological networks that adopt a novel nonpathological configuration, which only partly overlaps with the configuration present in normal hearts. Reproduced with permission from Weinheimer CJ et al.¹⁰

REMODELING AND RECOVERY OF CARDIAC MYOCYTE FUNCTION

Histologic evaluation of failing hearts consistently shows that cardiac myocytes become hypertrophied in response to chronic ventricular overloading and increased diastolic wall stress. In the setting of volume overload, these changes in hemodynamic load lead to elongation of the cardiac myocyte and lengthwise addition of sarcomeres, resulting in eccentric LV remodeling, which is so named

because of the eccentric position of the LV in the chest in patients with volume overload. A number of changes occur in cardiac myocyte biology in response to myocardial stretch and hemodynamic overload, neurohormonal activation (eg, norepinephrine, angiotensin II), and release of inflammatory mediators and oxidative stress. These changes in cardiac myocyte biology lead to this myocyte contractile dysfunction and increased myocyte cell death (Figure 3).

Figure 3 Summary schematic of key changes to cardiac myocyte biology from adverse remodeling (left) to reverse remodeling (right) after implementation of medical and device therapies. **(A)** Reverse remodeling leads to increased β-AR1 density and restoration of calcium handling and excitation-contraction coupling by way of decreased NCX density, decreased LTCC density albeit with increased activity (increased phosphorylation), increased SERCA density, and normalization of hyperphosphorylated RyR. **(B)** Reverse remodeling results in a shift in α-MHC and β-MHC ratios. α-MHC: alpha myosin heavy chain; β-AR1: beta adrenergic receptor 1; β-MHC: beta-myosin heavy chain; LTCC: L-type calcium channel; NCX: sodium calcium exchanger; SERCA: sarcoendoplasmic reticulum calcium ATPase; RyR: ryanodine receptor

Beta-Adrenergic Signaling

Ventricles obtained from HF patients demonstrate a marked reduction in beta-adrenergic receptor (β-AR) density, isoproterenol-mediated adenyl cyclase stimulation, and the contractile response to β-adrenergic agonists. The downregulation of β-ARs is likely mediated by increased levels of norepinephrine in the vicinity of the receptor. In patients with dilated cardiomyopathy, this reduction in receptor density involves primarily the β-AR1-receptor protein and mRNA and is proportional to the severity of HF. In contrast, the level of β-AR2-adrenergic receptor protein and mRNA are unchanged or increased.¹¹ These changes are reversed in patients treated with medical and device therapies (Figure 3A). Patients treated with metoprolol compared to placebo had a significant upregulation in total β-receptor tissue density. Interestingly, there was no receptor upregulation in patients treated with carvedilol compared to control despite the fact that it was associated with a greater improvement in LVEF and stroke volume.^{[12](#page-10-11)} It is possible that this is due to a difference in treatment length (6 months for metoprolol compared to 4 months for carvedilol). Changes in β-AR density may reflect a late effect of reverse LV remodeling effect.

In a more recent study of dilated cardiomyopathy, patients receiving carvedilol for 20 months had a significant increase in β-AR density, albeit measured indirectly and noninvasively by ¹¹C-CGP12177 PET, which also predicted improved reverse remodeling events[.13](#page-10-12) Similarly, angiotensin-converting enzyme (ACE) inhibitors increased myocardial β-AR density compared to placebo.14 The application of device therapies, LVAD and cardiac resynchronization therapy (CRT) , also has shown to enhance isoproterenol responsiveness as an indirect measure of β-AR desensitization[.15](#page-10-13)[-18](#page-10-14) In addition to changes in β-AR density, alterations in the topographic landscape of β-AR2-receptor protein is also likely to affect cardiomyocyte function. β-AR2-adrenergic receptors reside in transverse tubules (T-tubules) to produce localized cAMP signaling. However, in rat models of ischemic cardiomyopathy, β-AR2 receptors redistribute to the cell surface membranes and diffusely activate protein kinase A signaling.[19,](#page-10-15)[20](#page-10-16) Whether or not these topographic changes revert to the norm in reverse LV remodeling has yet to be determined.

Cardiac Myocyte Contractile and Structural Proteins

Myosin is a sarcomeric protein that plays a critical role in myocardial contractile function. Normal nonfailing ventricles predominantly consisted of β-myosin heavy chain (MHC) and up to 30% α -MHC (adult isoform of MHC), while failing hearts consistently downregulated α -MHC expression.^{21,22} In a transgenic rabbit model where β-MHC is similarly the dominant ventricular isoform, α-MHC overexpression was found to be protective against tachycardia-induced cardiomyopathy and post-infarction remodeling.[23](#page-11-1) Additional rodent models of hypertensive and ischemic cardiomyopathy have shown that medical treatment with an ACE inhibitor or angiotensin receptor blocker conferred higher α -MHC levels.^{[24,](#page-11-2)[25](#page-11-3)} In a trial of patients with idiopathic dilated cardiomyopathy, those that were treated with β-blocker therapy had restored transcript levels of α-MHC and sarcoplasmic-reticulum calcium ATPase (SERCA) and a decrease in β-MHC gene expression (Figure 3B). [26](#page-11-4)

Similarly, patients undergoing unloading and reverse remodeling following LVAD support had a downregulation in β-MHC expression.²⁷ Furthermore, patients who were treated with and responded to CRT consistently showed an increase in α-MHC and SERCA mRNA transcript levels.[28,](#page-11-6)[29](#page-11-7) These findings are consistent with those found in experimental dog models of ischemic cardiomyopathy and device unloading[.30](#page-11-8) Additionally biologic changes include an increase in other major sarcomeric proteins. This is evidenced by patients supported with LVAD who had increased expression of actin, myosin, tropomyosin, troponin C and T, and titin when comparing histologic tissue before and after LVAD treatment.³¹

Perhaps not unexpectedly, transduction of mechanical forces from a failing and dilated heart also leads to abnormalities in the cytoskeleton. Biopsies taken from patients with failing hearts had lower protein levels of vinculin and desmin and increased β-tubulin polymerization. These findings were reversed in patients mechanically supported with LVAD therapy.³² In a separate study, cytoskeleton transcript genes were evaluated in LVAD patients who underwent explant to transplant compared to those who underwent explant due to LV recovery. Those in the recovery group had significant changes in gene expression profiles of non-sarcomeric (lamin A/C, spectrin, and vinculin) and sarcomeric (β-actin, α-tropomyosin, a1-actinin, α-filamin A, and troponin T3) cytoskeleton markers.[33](#page-11-11) Other markers of cardiac remodeling, including natriuretic peptides, syndecan-4, vascular endothelial growth factor, and biomarkers of fibrosis (discussed below), are also dysregulated in HF[.34](#page-11-12) Specifically, a decrease in natriuretic peptides has been associated with reverse remodeling outcomes[.35](#page-11-13)

Calcium Handling and Excitation-Contraction Coupling

Failing cardiac myocytes have impaired calcium handling, which leads to myocyte contractile dysfunction. Ryanodine receptors (RyR) regulate the release of calcium from the sarcoplasmic reticulum (SR) into the cytoplasm, which is required for muscle contraction. Interestingly, patients

with HF have hyperphosphorylated levels of RyR via protein kinase A, resulting in calcium leak secondary to increased sensitivity of calcium-induced activation[.36](#page-11-14) RyR hyperphosphorylation was abrogated in failing hearts from patients treated with β-blocker therapy³⁷ and in patients following LVAD support.[36](#page-11-14) However, this is controversial and likely an oversimplification as prior studies using mutant nonphosphorylatable RyR had no effect on cardiac function or sympathetic stimulation.³⁸ On the other hand, SERCA, responsible for transporting cytosolic calcium back into the SR, is transcriptionally dysregulated in failing myocytes and also contributes to dysfunctional contraction. As described above, β-blocker and CRT therapy result in normalization of depressed SERCA transcript levels.^{[26,](#page-11-4)[28](#page-11-6)[,29](#page-11-7)}

More recent data demonstrated that despite decreased transcripts, SERCA protein levels remained unchanged in dilated and ischemic cardiomyopathies.³⁹ What remains to be determined are SERCA protein levels in reverse remodeling. Lastly, failing cardiac myocytes have impaired excitation. L-type calcium channels (LTCC) exist as voltage sensors on the cell membrane, coupling membrane depolarization to SR calcium release through RyR. NCX is a sodium calcium exchanger responsible for extruding intracellular calcium to restore resting calcium levels. Together, LTCC and NCX are major regulators in membrane potential activation and are both dysregulated in failing cardiac myocytes. In a tropomodulin-overexpressing transgenic mouse model of HF, treatment with propranolol normalized protein levels and the density of SERCA, NCX, and LTCC.[40](#page-11-18) Similarly, mechanical LV unloading resulted in restoration of LTCC density,⁴¹ increased mRNA transcripts encoding SERCA, RyR, and NCX, and restored cardiac myocyte contractile force.^{[42](#page-11-20)} Together, these data suggest that the dysregulation of excitation-contraction coupling found in failing cardiac myocytes is reversible after the implementation of medical and device therapies (Figure 3).

Myocyte Cell Death

Basic studies have suggested that progressive myocyte loss, through necrotic, apoptotic, or cell death pathways linked to autophagy, may contribute to progressive cardiac dysfunction and LV remodeling. Apoptosis, or programmed cell death, is an evolutionarily conserved process that allows multicellular organisms to selectively remove cells through a highly regulated program of cell suicide. Apoptosis is mediated by two pathways. The extrinsic pathway utilizes cell surface receptors, whereas the intrinsic pathway involves the mitochondria and endoplasmic reticulum (ER). Each of these pathways leads to caspase activation. The intrinsic pathway is responsible for transducing most apoptotic stimuli, including those caused by inadequate nutrients or survival factors, hypoxia, oxidative stress, nutrient stress, proteotoxic stress, DNA damage, and chemical and physical toxins. These stimuli ultimately converge on the mitochondria to trigger the release of apoptogenic proteins, such as cytochrome c, which leads to activation of proteolytic caspases, with subsequent DNA fragmentation.^{43,44} In rat models of ischemic HF, inhibition of apoptosis partially protected against cardiac myocyte drop-out, replacement fibrosis, and myocardial dysfunction.^{45,[46](#page-12-2)}

In healthy donor hearts, cytochrome c was localized in mitochondria, as expected under normal nonapoptotic conditions. In contrast, hearts obtained from patients with dilated cardiomyopathies had cytochrome c released into the sarcoplasmic space, a finding that was reversed after LVAD support. 47 A separate study demonstrated that LVAD unloading significantly reduces TUNEL staining, a marker of cell death, although the overall incidence of apoptosis and its downstream mediators was quite low[.48](#page-12-4) However, when cardiac myocytes are closely scrutinized by electron microscopy, TUNEL-positive myocytes from dilated cardiomyopathy were not apoptotic but, rather, had increased DNA repair activity.⁴⁹ This, in part, is likely due to the complex nature of cell death.

The exact mechanism of apoptosis in LV remodeling and volume overload is incompletely understood. Some insight can be gained from a study in cultured cardiac myocytes, which demonstrated activation of apoptotic pathways through mechanical stretch.^{[50](#page-12-6)} Furthermore, common pathways associated with apoptosis (eg, NF-kβ and MAPK) are activated in failing hearts and are normalized with LVAD support.^{51[-53](#page-12-8)} In addition to normalization of apoptotic factors, LVAD support also resulted in increased myocyte proliferation as measured by markers of cardiomyocyte mitosis and cytokinesis[.54](#page-12-9) In addition to LVAD support, CRT device implementation also has been shown to reduce apoptosis. Specifically, in a study where endomyocardial biopsies were obtained before and after CRT device implantation, patients receiving resynchronization had improved metrics LV remodeling and decreased histologic evidence of apoptosis.⁵⁵ Viewed together, these data suggest that cell death pathways are activated in failing cardiac myocytes and that device therapies can attenuate this effect, but future studies are needed to further elucidate the mechanisms and biologic relevance of myocyte death.

REMODELING AND CHANGES IN THE EXTRACELLULAR MATRIX

In addition to the biological changes that occur in cardiac myocyte during reverse LV remodeling, the extracellular matrix also undergoes important changes in organization and chemical composition.

Extracellular structure and function

One salient feature of the extracellular matrix is its network of fibrillar proteins that provide mechanical support to surrounding cardiac myocytes and translating myocyte contractile forces into LV ejection. Under normal homeostatic circumstances, type I and III collagens are the major fibrillar protein serving as a scaffold to transduce mechanical forces and maintain LV geometry. In transverse aortic constriction animal models of pressure overload, TGF-β, Smad 2/3 and Smad 1/5 pathways activate cardiac fibroblast to deposit collagens and other matrix proteins.^{56,[57](#page-12-11)} These changes result in myocardial stiffness akin to what is observed in patients with hypertensive heart disease.

In contrast, studies in patients with various cardiomyopathies are less conclusive and the data are mixed. This, in part, is likely due to the diverse etiologies and various stages of HF. For example, in ischemic cardiomyopathy, cardiac myocyte cell death is replaced by fibrinous scar and collagen deposition between capillaries and in the interstices surrounding cardiac myocytes. Correctively, patients with ischemic cardiomyopathy have a total increase in collagen deposition and decrease in the collagen I:III ratio, both of which were normalized with ACE inhibitor therapy.⁵⁸ In a separate study, patients with dilated cardiomyopathy similarly had an increase in total collagen, but with an increase in the collage I:III ratio.⁵⁹

Similar to ACE inhibitors, treatment with metoprolol also reduced myocardial fibrosis both in dog models of HF and in patients with dilated cardiomyopathies.^{[60,](#page-12-14)[61](#page-12-15)} Further evidence of reverse remodeling was demonstrated in a substudy of the RALES (Randomized Aldactone Evaluation Study) trial, showing spironolactone therapy reduced procollagen type III amino-terminal peptide (PIIINP), a serum marker of collagen production and cardiac fibrosis.^{[62](#page-12-16)} Additional studies show that patients receiving CRT device therapy have increased reverse LV remodeling, decreased fibrosis, and reduced circulating MMP-9 concentrations.^{55[,63](#page-12-17)} The data for LVAD support in reverse LV remodeling collagen deposition are conflicting and less clear. These discrepancies likely reflect the diverse and dynamic nature of cardiomyopathies and their medical treatment.

Collagen content is a highly regulated process that is not only influenced by production but also by turnover and post-translational modifications. Metalloproteases (MMPs) are collagenolytic enzymes commonly activated within a failing and dilated heart. Although the precise regulatory activators of MMPs are not known, tissue inhibitors of matrix metalloproteinases (TIMPs) can directly target and inhibit MMPs and their proteolytic function. Decreased TIMP-1 and TIMP-3 transcript and protein levels were found in failing hearts from patient hearts of both ischemic and dilated etiologies.[64](#page-13-0) In fact, TIMP3 deficient mice developed spontaneous LV dilation, cardiac myocyte hypertrophy, decreased collagen volume, and LV systolic dysfunction, mirroring remodeling events seen in human dilated cardiomyopathies[.64](#page-13-0)

Matricellular Signaling Proteins and Myofibroblasts

Once thought to be a simple and inert network of mechanical structural proteins, the extracellular matrix is a complex milieu of diverse and dynamic signaling molecules. These previously underappreciated matricellular proteins can drive both inflammation and repair through "outsidein" signaling. In addition to the collagenolytic role of metalloproteases described above, MMPs can modulate inflammatory pathways through proteolytic processing of signaling molecules. Specifically, MMPs process cell membrane-bound tumor necrosis factor (TNF) to yield mature TNF, a proinflammatory cytokine and mediator of tissue repair mechanisms consistently elevated in patients with congestive HF.[65-](#page-13-1)[67](#page-13-2) Additional studies have found that thrombospondin-1 (TSP-1), an inhibitor of angiogenesis and activator of TGFβ, is induced after tissue injury and is decreased in failing hearts.^{68,69} In a pressure overload mouse model, TSP-1 expression was increased and a loss of TSP-1 function resulted in early hypertrophy and worse LV dilation without affecting collagen content.⁷⁰ Other matricellular proteins, including tenascin-C, secreted protein acidic and rich in cysteine (SPARC), osteopontin, and periostin, have also been described in cardiac injury and remodeling[.71](#page-13-4) Cardiac fibroblasts are major effector cells responsible for secreting collagens' matricellular proteins and can be activated by injury and neurohormonal activation.

In patients with dilated cardiomyopathy, elevated serum levels of carboxy-terminal propeptide of collagen type I (PICP) are associated with worse outcomes.⁷² Treatment with losartan or eplerenone can decrease concentrations of serum PICP, further suggesting a role of cardiac fibroblast in reverse remodeling.^{73,74} Recent studies suggest a role for pirfenidone in reducing myocardial fibrosis in patients with HF with preserved ejection fraction[.75](#page-13-7) *Wisper* (Wisp2 superenhancer-associated RNA) is a cardiac fibroblast-enriched lncRNA that regulates cardiac fibrosis after myocardial injury. Antisense oligonucleotides-mediated silencing of *Wisper* in vivo attenuated MI-induced fibrosis and cardiac dysfunction in a murine model of acute coronary ligation.⁷⁶ Together, these data describe a complex network of extracellular matrix signaling molecules that drive biologic responses to cardiac stress. At the time of this writing, it is unclear how these matricellular proteins play a role in reverse LV remodeling.

Myocardial Microvasculature and Endothelial Dysfunction

Many of the aforementioned signaling molecules regulate endothelial cell survival and angiogenesis. This, in part, may explain why patients with dilated cardiomyopathies have alterations in capillary density. The concept of capillary rarefaction, or a reduced capillary density, in dilated cardiomyopathy suggests a mismatch between the oxygen supply and the demands of the myocardial tissue. This can contribute to the progression of HF due to inadequate perfusion and the resulting ischemia. In animal models of HF, medical therapy with β-blockers, ACE inhibitors, and aldosterone antagonists restored the loss of capillary density.[60,](#page-12-14)[77](#page-13-9)[-79](#page-13-10) Additionally, patients mechanically unloaded with LVAD therapy underwent significant gene expression changes in vascular organization and migration signaling pathways.⁸⁰ This is further supported by an increase in microvascular density with pulsatile LVAD unloading[.81](#page-13-12) Additional evidence of microvascular density recovery was demonstrated after 6 months of CRT.55 More recent data suggest proteins related to endothelial function are dysregulated. ANGPT2 (angiopoietin2) and VEGFR1 (vascular endothelial growth factor receptor 1) concentrations directly correlate with worse LV remodeling events, and a decrease in ANGPT2 is associated with LV reverse remodeling and outcomes.⁸² Although the data are consistent across animal and human models, further studies are needed to understand the primary triggers of microvascular density and its role in cardiomyopathies.

MYOCARDIAL RECOVERY VERSUS MYOCARDIAL REMISSION

At the beginning of this review, we noted that HF patients whose ventricles resume a more normal LV size and shape may experience freedom from or recurrence of HF events. Although we have discussed the various components of reverse LV remodeling, none of these studies directly addresses which of these changes is required to maintain clinical stability in ventricles that have undergone reverse LV remodeling.

Although the precise reasons for the lack of clinical stability in hearts that have undergone reverse LV remodeling are not known, one plausible explanation is that many of the multilevel molecular changes that occur during forward LV remodeling remain dysregulated in reverse LV remodeled hearts even though they may look phenotypically normal (Figure 2). As one example, transcriptional profiling studies of failing hearts have shown that only about 5% of genes that are dysregulated in failing hearts revert back to normal following LVAD support despite

typical morphologic and functional responses to LVAD support.⁸³ Second, although maximal calcium saturated force generation is improved in myocytes following LVAD support, force generation is still less than in myocytes from nonfailing controls despite reversal of cardiac myocyte hypertrophy.[84](#page-13-14) Third, most studies that have examined changes in the ECM following LVAD have suggested that the ECM does not revert to normal on its own and can actually be characterized by increased myocardial fibrosis. Moreover, our current understanding of changes in the ECM during LVAD support has focused on ECM content and not on the more fundamental issues of its 3D organization or the interactions between the collagen matrix and the resident cardiac myocytes, which are likely to be critically important. Fourth, although the LV end-diastolic pressurevolume relationship of LVAD-supported hearts are shifted leftward and overlap those found in nonfailing ventricles, the ratio of LV wall thickness to LV wall radius does not return to normal despite normalization of LV chamber geometry.[85](#page-13-15) This has important implications for the stability of LV function insofar as it implies that the LV wall stress elevated in reverse LV remodeled hearts.

Given that end-diastolic wall stress represents the load on the cardiac myocyte at the onset of systole, the observation that the r/h ratio is not normalized despite the normalization of LV global chamber properties suggests that the cardiac myocytes in reverse remodeled ventricles are still exposed to increased physiological stresses. Whether this represents the loss of functioning cardiac myocytes or failure of the 3D organization of the ECM to revert to normal is unknown. Thus, regression of the HF phenotype and the accompanying return towards a more normal cardiac phenotype during reverse remodeling does not, in and of itself, signify that the cellular/molecular biology and physiology of these hearts are normal, which may explain why reverse remodeling may be associated with different clinical outcomes.

Do Potential Biological Differences Explain Disparate Clinical Outcomes After Reverse Remodeling?

Although the potential biological differences between myocardial recovery and myocardial remission are not known, there are parallels in mechanical engineering science that may help to illuminate potential important differences and to frame future mechanistic discussions. In mechanics, deformation of a material refers to the change in the shape or size of an object due to an applied force. Figure 4A shows a representative one-dimensional stress versus strain diagram of a material that is exposed to an increased load[.2](#page-10-2) With increasing stress, there is an increase in the length of the material up until the point when no further changes in length are possible without the material

Figure 4 Mechanical engineering science and cardiac remodeling. **(A)** Diagram of a stress-strain curve of a ductile material illustrating the relationship between an applied force (stress) and deformation (strain). Deformation can lead to reversible changes in a material (elastic deformation) if the properties of the material are not changed, and irreversible changes in a material (plastic deformation). **(B)** Hypothetical model of reverse remodeling in a heart that has undergone irreversible damage (plastic deformation). **(C)** Hypothetical model of reverse remodeling with recovery in heart that has undergone reversible damage (elastic deformation). Reproduced with permission from Mann DL et al.^{[2](#page-10-2)}

breaking. Importantly, if the material returns to its original state when the load is removed, this is referred to as "elastic deformation." In contrast, if the mechanical properties of the material are changed irreversibly when applying stress, such that the object will return only partially to its original properties when the stress is removed, this is referred to as "plastic deformation." It is sometimes the case that elastic deformation occurs under a certain level of stress and plastic deformation occurs when that stress level is exceeded. Regardless, the important distinction is whether or not the material returns to its original state when the stress is removed. Although precise parallels between cardiac remodeling in HF and deformation of solid materials following loading are not appropriate, there could be a heuristic parallel between reverse remodeling and plastic deformation insofar as the reverse remodeled heart does not revert completely to normal after cessation of hemodynamic overloading (Figure 4B). Although speculative, it is possible that myocardial recovery is more analogous to elastic deformation in that the recovered heart reverts back to normal after hemodynamic overloading is removed (Figure 4C).

HUMAN MODELS OF REVERSE LV REMODELING

Reverse LV remodeling and normalization of LVEF have been observed in numerous clinical scenarios wherein discontinuation of the inciting event, including cardiotoxic agents, peripartum cardiomyopathy, stress-induced cardiomyopathy, and viral myocarditis, often results in significant improvement in LV size, shape, and function. However, rates of myocardial recovery have been increasing in more recent clinical studies, which is most likely attributable to advances in and widespread application of GDMT and device therapies.

MEDICAL THERAPIES

Several studies show that treatment with enalapril compared to placebo prevent progression of LV remodeling and result in fewer HF-related deaths, [86,](#page-14-0)[87](#page-14-1) and a substudy of the Val-HeFT (Valsartan Heart Failure) trial demonstrated that the addition of valsartan to an existing ACE inhibitor or β-blocker therapy further reduced LV diameter.⁸⁸ Additional therapies such as carvedilol and spironolactone have been shown to reduce LV volumes when added to ACE inhibitors alone and with β-blockers, respectively, and have been longstanding cornerstone therapies in reducing mortality.[89,](#page-14-3)[90](#page-14-4) Interestingly, patients on equivalent drug therapy (β-blocker, ACE inhibitor or ARB, and aldosterone receptor blocker) and with LVEF between 40% to 50% had significantly improved clinical outcomes if they recovered from a worse LV function (LVEF < 40%) compared to either unchanged (LVEF 40-50%) or deteriorated function (LVEF > 50%).⁹¹ Lastly, SGLT2 inhibitors are the most recent of adopted medical therapies to demonstrate a reduction primarily in HF hospitalizations when added to standard therapy and also have been shown to reduce LV volumes in some patients.^{[92](#page-14-6)}

DEVICE THERAPIES

Device therapies also have shown to reduce ventricular volumes and contribute towards reverse remodeling. Both pulsatile and continuous-flow LVADs and axial or centrifugal pumps provided equivalent LV pressure unloading, albeit pulsatile devices provided a greater unloading to LV volume due to higher pump output. Despite these differences, both pulsatile and continuous-flow therapies were equivalent in hemodynamic measures of LV reverse remodeling, although there may be serum biomarkers to suggest greater biological remodeling with the use of pulsatile pumps.[93-](#page-14-7)[95](#page-14-8) Cardiac resynchronization therapy has been widely implemented in patients with moderate to severe systolic HF and LV desynchrony as evidenced by a prolonged QRS interval, portending poorer outcomes. Patients treated with CRT had improved contractile function, smaller LV volumes, and less mitral regurgitation at both short- and long-term follow-up. 96,[97](#page-14-10)

DURABILITY OF HF WITH REVERSE LV REMODELING AND IMPROVED LVEF

Although applications of medical and device therapies have reshaped the landscape and management of HF, it remains a remarkably morbid and mortal disease with rates of death comparable to those of aggressive malignancies. Additionally, the efficacy of current HF therapeutics is highly variable and often dependent on the etiology of the cardiomyopathy as well as duration and severity. Even with reverse LV remodeling that results in normalization of LV structure and function, the vast majority of HF patients remain vulnerable to future HF events. One recent study evaluated the durability of clinical stability in patients with dilated cardiomyopathy who were started on GDMT and followed for 10 years. Of the 408 enrolled patients with dilated cardiomyopathy, only 15% normalized their LV structure and function on GDMT (defined as an LVEF > 50% and LV end-diastolic dimension index < 33 mm/m2). Remarkably, only 60% of the patient cohort with normalized LV structure and function on GDMT were able to maintain clinical stability over the ensuing decade.⁹⁸ In the TRED-HF (Therapy Withdrawal in Recovered DCM) study, patients with dilated cardiomyopathy and normal LV size and function were randomized to continued treatment with GDMT or phased withdrawal. The investigators found that 44% of patients who discontinued medical therapy had HF relapse within 6 months, which was not observed in the patients who continued GDMT and remained clinically stable.[99](#page-14-12) As noted by the authors of the TRED-HF study, their findings suggest that "for many patients, improvement in cardiac function following treatment does not reflect full and sustained recovery but rather reflects remission, which requires at least some treatment to be maintained.["99](#page-14-12) Viewed together, these data not only highlight the heterogenous response to conventional medical therapies but also the fragility of maintaining

clinical stability despite normalization of LV structure and function.

CONCLUSION

Although reverse LV remodeling occurs in the vast majority of patients who have prolonged LVAD support—and occurs in a significant proportion of HFrEF patients treated with GDMT—reverse LV remodeling with an improved LVEF does not necessarily signify that the patient is cured from HF. Indeed, many HF patients whose ventricles resume a more normal LV size and shape will experience recurrent HF events despite being maintained on state-of-theart GDMT and receiving excellent clinical care. Further, the results of the TRED-HF study suggest that GDMT is required to maintain clinical stability even in patients who have normalization of their LV structure and function. Collectively, these studies suggest that while achieving normalization of their LV structure and function should be an important goal of HF therapy, we also need to think more holistically about the totality of the HF patient experience and concentrate our future research efforts on identifying the optimal therapeutic approach(es) that will maintain patients in remission, free from future HF events.

KEY POINTS

- **•** Reverse remodeling is the process by which a failing heart undergoes efforts towards normalization of geometric, hemodynamic, and biologic changes associated with improved left ventricular ejection fraction and clinical outcomes.
- **•** Patients and animal models with heart failure can achieve reverse remodeling through the implementation of medical and device therapies.
- **•** The biological underpinnings of myocardial reverse remodeling are complex and associated with a reversion towards a more normal cardiac myocyte and extracellular biology.
- Reverse remodeling and myocardial recovery represent improved yet fragile new steady states continuously at risk for future adverse remodeling events. A more complete mechanistic understanding is required to leverage the full therapeutic potential of novel medical and device treatments.

COMPETING INTERESTS

The authors have no competing interests to declare.

AUTHOR AFFILIATIONS

Arick C. Park, MD, PhDorcid.org/0000-0001-5668-0701 Washington University School of Medicine, St. Louis, Missouri, US **Douglas L. Mann, MD**orcid.org/0000-0002-2516-0145 Washington University School of Medicine, St. Louis, Missouri, US

REFERENCES

- 1. **Wilcox JE, Fang JC, Margulies KB, Mann DL.** Heart Failure With Recovered Left Ventricular Ejection Fraction: JACC Scientific Expert Panel. J Am Coll Cardiol. 2020 Aug 11;76(6):719-734. doi: [10.1016/j.jacc.2020.05.075](https://doi.org/10.1016/j.jacc.2020.05.075)
- 2. **Mann DL, Barger PM, Burkhoff D.** Myocardial recovery: myth, magic or molecular target? J Am Coll Cardiol. 2012 Dec 18;60(24):2465-72. doi: [10.1016/j.jacc.2012.06.062](https://doi.org/10.1016/j.jacc.2012.06.062)
- 3. **St John Sutton MG, Pfeffer MA, Plappert T,** et al. Quantitative two-dimensional echocardiographic measurements are major predictors of adverse cardiovascular events after acute myocardial infarction. The protective effects of captopril. Circulation. 1994 Jan;89(1):68-75. doi: [10.1161/01.cir.89.1.68](https://doi.org/10.1161/01.CIR.89.1.68)
- 4. **White HD, Norris RM, Brown MA, Brandt PW, Whitlock RM, Wild CJ.** Left ventricular end-systolic volume as the major determinant of survival after recovery from myocardial infarction. Circulation. 1987 Jul;76(1):44-51. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.76.1.44) [cir.76.1.44](https://doi.org/10.1161/01.CIR.76.1.44)
- 5. **Douglas PS, Morrow R, Ioli A, Reicheck N.** Left ventricular shape, afterload and survival in idiopathic dilated cardiomyopathy. J Am Coll Cardiol. 1989 Feb;13(2):311-5. doi: [10.1016/0735-1097\(89\)90504-4](https://doi.org/10.1016/0735-1097(89)90504-4)
- 6. **Vasan RS, Larson MG, Benjamin EJ, Evans JC, Levy D.** Left ventricular dilatation and the risk of congestive heart failure in people without myocardial infarction. N Engl J Med. 1997 May 8;336(19):1350-5. doi: [10.1056/](https://doi.org/10.1056/NEJM199705083361903) [NEJM199705083361903](https://doi.org/10.1056/NEJM199705083361903)
- 7. **Levin HR, Oz MC, Chen JM, Packer M, Rose EA, Burkhoff D.** Reversal of chronic ventricular dilation in patients with endstage cardiomyopathy by prolonged mechanical unloading. Circulation. 1995 Jun 1;91(11):2717-20. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.91.11.2717) [cir.91.11.2717](https://doi.org/10.1161/01.CIR.91.11.2717)
- 8. **Kass DA, Baughman KL, Pak PH,** et al. Reverse remodeling from cardiomyoplasty in human heart failure. External constraint versus active assist. Circulation. 1995 May 1;91(9):2314-8. doi: [10.1161/01.cir.91.9.2314](https://doi.org/10.1161/01.CIR.91.9.2314)
- 9. **Westaby S, Jin XY, Katsumata T, Taggart DP, Coats AJ, Frazier OH.** Mechanical support in dilated cardiomyopathy: signs of early left ventricular recovery. Ann Thorac Surg. 1997 Nov;64(5):1303-8. doi: [10.1016/S0003-4975\(97\)00910-7](https://doi.org/10.1016/S0003-4975(97)00910-7)
- 10. **Weinheimer CJ, Kovacs A, Evans S,** et al. Load- Dependent Changes in Left Ventricular Structure and Function in a Pathophysiologically Relevant Murine Model of Reversible Heart Failure. Circ Heart Fail. 2018 May;11(5):e004351. doi: [10.1161/CIRCHEARTFAILURE.117.004351](https://doi.org/10.1161/CIRCHEARTFAILURE.117.004351)
- 11. **Bristow MR, Ginsburg R, Minobe W,** et al. Decreased catecholamine sensitivity and beta-adrenergic-receptor density in failing human hearts. N Engl J Med. 1982 Jul 22;307(4):205-11. doi: [10.1056/NEJM198207223070401](https://doi.org/10.1056/NEJM198207223070401)
- 12. **Gilbert EM, Abraham WT, Olsen S, et al. Comparative** hemodynamic, left ventricular functional, and antiadrenergic effects of chronic treatment with metoprolol versus carvedilol in the failing heart. Circulation. 1996 Dec 1;94(11):2817-25. doi: [10.1161/01.cir.94.11.2817](https://doi.org/10.1161/01.CIR.94.11.2817)
- 13. **Naya M, Tsukamoto T, Morita K,** et al. Myocardial betaadrenergic receptor density assessed by 11C-CGP12177 PET predicts improvement of cardiac function after carvedilol treatment in patients with idiopathic dilated cardiomyopathy. J Nucl Med. 2009 Feb;50(2):220-5. doi: [10.2967/jnumed.108.056341](https://doi.org/10.2967/jnumed.108.056341)
- 14. **Gilbert EM, Sandoval A, Larrabee P, Renlund DG, O'Connell JB, Bristow MR.** Lisinopril lowers cardiac adrenergic drive and increases beta-receptor density in the failing human heart. Circulation. 1993 Aug;88(2):472-80. doi: [10.1161/01.cir.88.2.472](https://doi.org/10.1161/01.CIR.88.2.472)
- 15. **Klotz S, Barbone A, Reiken S,** et al. Left ventricular assist device support normalizes left and right ventricular betaadrenergic pathway properties. J Am Coll Cardiol. 2005 Mar 1;45(5):668-76. doi: [10.1016/j.jacc.2004.11.042](https://doi.org/10.1016/j.jacc.2004.11.042)
- 16. **Mullens W, Bartunek J, Wilson Tang WH,** et al. Early and late effects of cardiac resynchronization therapy on force-frequency relation and contractility regulating gene expression in heart failure patients. Heart Rhythm. 2008 Jan;5(1):52-9. doi: [10.1016/j.hrthm.2007.09.009](https://doi.org/10.1016/j.hrthm.2007.09.009)
- 17. **Ogletree-Hughes ML, Stull LB, Sweet WE, Smedira NG, McCarthy PM, Moravec CS.** Mechanical unloading restores beta-adrenergic responsiveness and reverses receptor downregulation in the failing human heart. Circulation. 2001 Aug 21;104(8):881-6. doi: [10.1161/](https://doi.org/10.1161/hc3301.094911) [hc3301.094911](https://doi.org/10.1161/hc3301.094911)
- 18. **Schnee PM, Shah N, Bergheim M,** et al. Location and density of alpha- and beta-adrenoreceptor sub-types in myocardium after mechanical left ventricular unloading. J Heart Lung Transplant. 2008 Jul;27(7):710-7. doi: [10.1016/j.](https://doi.org/10.1016/j.healun.2008.03.015) [healun.2008.03.015](https://doi.org/10.1016/j.healun.2008.03.015)
- 19. **Nikolaev VO, Moshkov A, Lyon AR,** et al. Beta2-adrenergic receptor redistribution in heart failure changes cAMP compartmentation. Science. 2010 Mar 26;327(5973):1653-7. doi: [10.1126/science.1185988](https://doi.org/10.1126/science.1185988)
- 20. **Schobesberger S, Wright P, Tokar S,** et al. T-tubule remodelling disturbs localized β2-adrenergic signalling in rat ventricular myocytes during the progression of heart failure. Cardiovasc Res. 2017 Jun 1;113(7):770-782. doi: [10.1093/](https://doi.org/10.1093/cvr/cvx074) [cvr/cvx074](https://doi.org/10.1093/cvr/cvx074)
- 21. **Lowes BD, Minobe W, Abraham WT,** et al. Changes in gene expression in the intact human heart. Downregulation of alpha-myosin heavy chain in hypertrophied, failing ventricular myocardium. J Clin Invest. 1997 Nov 1;100(9):2315-24. doi: [10.1172/JCI119770](https://doi.org/10.1172/JCI119770)
- 22. **Nakao K, Minobe W, Roden R, Bristow MR, Leinwand LA.** Myosin heavy chain gene expression in human heart failure. J Clin Invest. 1997 Nov 1;100(9):2362-70. doi: [10.1172/](https://doi.org/10.1172/JCI119776) [JCI119776](https://doi.org/10.1172/JCI119776)
- 23. **James J, Martin L, Krenz M,** et al. Forced expression of alpha-myosin heavy chain in the rabbit ventricle results in cardioprotection under cardiomyopathic conditions. Circulation. 2005 May 10;111(18):2339-46. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.0000164233.09448.B1
) [CIR.0000164233.09448.B1](https://doi.org/10.1161/01.CIR.0000164233.09448.B1
)
- 24. **Brooks WW, Bing OHL, Robinson KG, Slawsky MT, Chaletsky DM, Conrad CH.** Effect of angiotensin-converting enzyme inhibition on myocardial fibrosis and function in hypertrophied and failing myocardium from the spontaneously hypertensive rat. Circulation. 1997 Dec 2;96(11):4002-10. doi: [10.1161/01.cir.96.11.4002](https://doi.org/10.1161/01.CIR.96.11.4002)
- 25. **Wang J, Guo X, Dhalla NS.** Modification of myosin protein and gene expression in failing hearts due to myocardial infarction by enalapril or losartan. Biochim Biophys Acta. 2004 Oct 14;1690(2):177-84. doi: [10.1016/j.](https://doi.org/10.1016/j.bbadis.2004.06.004) [bbadis.2004.06.004](https://doi.org/10.1016/j.bbadis.2004.06.004)
- 26. **Lowes BD, Gilbert EM, Abraham WT,** et al. Myocardial gene expression in dilated cardiomyopathy treated with betablocking agents. N Engl J Med. 2002 May 2;346(18):1357-65. doi: [10.1056/NEJMoa012630](https://doi.org/10.1056/NEJMoa012630)
- 27. **Blaxall BC, Tschannen-Moran BM, Milano CA, Koch WJ.** Differential gene expression and genomic patient stratification following left ventricular assist device support. J Am Coll Cardiol. 2003 Apr 2;41(7):1096-106. doi: [10.1016/](https://doi.org/10.1016/S0735-1097(03)00043-3) [s0735-1097\(03\)00043-3](https://doi.org/10.1016/S0735-1097(03)00043-3)
- 28. **Iyengar S, Haas G, Lamba S,** et al. Effect of cardiac resynchronization therapy on myocardial gene expression in patients with nonischemic dilated cardiomyopathy. J Card Fail. 2007 May;13(4):304-11. doi: [10.1016/j.](https://doi.org/10.1016/j.cardfail.2007.01.005) [cardfail.2007.01.005](https://doi.org/10.1016/j.cardfail.2007.01.005)
- 29. **Vanderheyden M, Mullens W, Delrue L,** et al. Myocardial gene expression in heart failure patients treated with cardiac resynchronization therapy responders versus nonresponders. J Am Coll Cardiol. 2008 Jan 15;51(2):129-36. doi: [10.1016/j.](https://doi.org/10.1016/j.jacc.2007.07.087) [jacc.2007.07.087](https://doi.org/10.1016/j.jacc.2007.07.087)
- 30. **Rastogi S, Mishra S, Gupta RC, Sabbah HN.** Reversal of maladaptive gene program in left ventricular myocardium of dogs with heart failure following long-term therapy with the Acorn Cardiac Support Device. Heart Fail Rev. 2005 Jun;10(2):157-63. doi: [10.1007/s10741-005-4643-z](https://doi.org/10.1007/s10741-005-4643-z)
- 31. **de Jonge N, van Wichen DF, Schipper ME,** et al. Left ventricular assist device in end-stage heart failure: persistence of structural myocyte damage after unloading. An immunohistochemical analysis of the contractile myofilaments. J Am Coll Cardiol. 2002 Mar 20;39(6):963-9. doi: [10.1016/s0735-1097\(02\)01713-8](https://doi.org/10.1016/S0735-1097(02)01713-8)
- 32. **Aquila LA, McCarthy PM, Smedira NG, Young JB, Moravec CS.** Cytoskeletal structure and recovery in single

human cardiac myocytes. J Heart Lung Transplant. 2004 Aug;23(8):954-63. doi: [10.1016/j.healun.2004.05.018](https://doi.org/10.1016/j.healun.2004.05.018)

- 33. **Birks EJ, Hall JL, Barton PJ,** et al. Gene profiling changes in cytoskeletal proteins during clinical recovery after left ventricular-assist device support. Circulation. 2005 Aug 30;112(9 Suppl):I57-64. doi: [10.1161/](https://doi.org/10.1161/CIRCULATIONAHA.104.526137) [CIRCULATIONAHA.104.526137](https://doi.org/10.1161/CIRCULATIONAHA.104.526137)
- 34. **Ponikowska B, Iwanek G, Zdanowicz A,** et al. Biomarkers of Myocardial Injury and Remodeling in Heart Failure. J Pers Med. 2022 May 16;12(5):799. doi: [10.3390/jpm12050799](https://doi.org/10.3390/jpm12050799)
- 35. **Daubert MA, Adams K, Yow E,** et al. NT-proBNP Goal Achievement Is Associated With Significant Reverse Remodeling and Improved Clinical Outcomes in HFrEF. JACC Heart Fail. 2019 Feb;7(2):158-168. doi: [10.1016/j.](https://doi.org/10.1016/j.jchf.2018.10.014) [jchf.2018.10.014](https://doi.org/10.1016/j.jchf.2018.10.014)
- 36. **Marx SO, Reiken S, Hisamatsu Y,** et al. PKA phosphorylation dissociates FKBP12.6 from the calcium release channel (ryanodine receptor): defective regulation in failing hearts. Cell. 2000 May 12;101(4):365-76. doi: [10.1016/s0092-](https://doi.org/10.1016/S0092-8674(00)80847-8) [8674\(00\)80847-8](https://doi.org/10.1016/S0092-8674(00)80847-8)
- 37. **Reiken S, Gaburjakova M, Guatimosim S,** et al. Protein kinase A phosphorylation of the cardiac calcium release channel (ryanodine receptor) in normal and failing hearts. Role of phosphatases and response to isoproterenol. J Biol Chem. 2003 Jan 3;278(1):444-53. doi: [10.1074/jbc.M207028200](https://doi.org/10.1074/jbc.M207028200)
- 38. **Zhang H, Makarewich CA, Kubo H, et al.** Hyperphosphorylation of the cardiac ryanodine receptor at serine 2808 is not involved in cardiac dysfunction after myocardial infarction. Circ Res. 2012 Mar 16;110(6):831-40. doi: [10.1161/CIRCRESAHA.111.255158](https://doi.org/10.1161/CIRCRESAHA.111.255158)
- 39. **Ragone I, Barallobre-Barreiro J, Takov K,** et al. SERCA2a Protein Levels Are Unaltered in Human Heart Failure. Circulation. 2023 Aug 15;148(7):613-616. doi: [10.1161/](https://doi.org/10.1161/CIRCULATIONAHA.123.064513) [CIRCULATIONAHA.123.064513](https://doi.org/10.1161/CIRCULATIONAHA.123.064513)
- 40. **Plank DM, Yatani A, Ritsu H,** et al. Calcium dynamics in the failing heart: restoration by beta-adrenergic receptor blockade. Am J Physiol Heart Circ Physiol. 2003 Jul;285(1):H305-15. doi: [10.1152/ajpheart.00425.2002](https://doi.org/10.1152/ajpheart.00425.2002)
- 41. **Chen X, Piacentino V 3rd, Furukawa S, Goldman B, Margulies KB, Houser SR.** L-type Ca2+ channel density and regulation are altered in failing human ventricular myocytes and recover after support with mechanical assist devices. Circ Res. 2002 Sep 20;91(6):517-24. doi: [10.1161/01.](https://doi.org/10.1161/01.RES.0000033988.13062.7C) [res.0000033988.13062.7c](https://doi.org/10.1161/01.RES.0000033988.13062.7C)
- 42. **Heerdt PM, Holmes JW, Cai B,** et al. Chronic unloading by left ventricular assist device reverses contractile dysfunction and alters gene expression in end-stage heart failure. Circulation. 2000 Nov 28;102(22):2713-9. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.102.22.2713) [cir.102.22.2713](https://doi.org/10.1161/01.CIR.102.22.2713)
- 43. **Narula J, Haider N, Virmani R,** et al. Apoptosis in myocytes in end-stage heart failure. N Engl J Med. 1996 Oct 17;335(16):1182-9. doi: [10.1056/NEJM199610173351603](https://doi.org/10.1056/NEJM199610173351603)
- 44. **Narula J, Pandey P, Arbustini E,** et al. Apoptosis in heart failure: release of cytochrome c from mitochondria and activation of caspase-3 in human cardiomyopathy. Proc Natl Acad Sci U S A. 1999 Jul 6;96(14):8144-9. doi: [10.1073/](https://doi.org/10.1073/pnas.96.14.8144) [pnas.96.14.8144](https://doi.org/10.1073/pnas.96.14.8144)
- 45. **Chandrashekhar Y, Sen S, Anway R, Shuros A, Anand I.** Long-term caspase inhibition ameliorates apoptosis, reduces myocardial troponin-I cleavage, protects left ventricular function, and attenuates remodeling in rats with myocardial infarction. J Am Coll Cardiol. 2004 Jan 21;43(2):295-301. doi: [10.1016/j.jacc.2003.09.026](https://doi.org/10.1016/j.jacc.2003.09.026)
- 46. **Hayakawa K, Takemura G, Kanoh M,** et al. Inhibition of granulation tissue cell apoptosis during the subacute stage of myocardial infarction improves cardiac remodeling and dysfunction at the chronic stage. Circulation. 2003 Jul 8;108(1):104-9. doi: [10.1161/01.CIR.0000074225.62168.68](https://doi.org/10.1161/01.CIR.0000074225.62168.68)
- 47. **Haider N, Narula N, Narula J.** Apoptosis in heart failure represents programmed cell survival, not death, of cardiomyocytes and likelihood of reverse remodeling. J Card Fail. 2002 Dec;8(6 Suppl):S512-7. doi: [10.1054/](https://doi.org/10.1054/jcaf.2002.130034) [jcaf.2002.130034](https://doi.org/10.1054/jcaf.2002.130034)
- 48. **de Jonge N, van Wichen DF, van Kuik J,** et al. Cardiomyocyte death in patients with end-stage heart failure before and after support with a left ventricular assist device: low incidence of apoptosis despite ubiquitous mediators. J Heart Lung Transplant. 2003 Sep;22(9):1028-36. doi: [10.1016/](https://doi.org/10.1016/S1053-2498(02)01160-9) [s1053-2498\(02\)01160-9](https://doi.org/10.1016/S1053-2498(02)01160-9)
- 49. **Kanoh M, Takemura G, Misao J,** et al. Significance of myocytes with positive DNA in situ nick end-labeling (TUNEL) in hearts with dilated cardiomyopathy: not apoptosis but DNA repair. Circulation. 1999 Jun 1;99(21):2757-64. doi: [10.1161/01.cir.99.21.2757](https://doi.org/10.1161/01.CIR.99.21.2757)
- 50. **Cheng WP, Wang BW, Lo HM, Shyu KG.** Mechanical Stretch Induces Apoptosis Regulator TRB3 in Cultured Cardiomyocytes and Volume-Overloaded Heart. PLoS One. 2015 Apr 21;10(4):e0123235. doi: [10.1371/journal.](https://doi.org/10.1371/journal.pone.0123235) [pone.0123235](https://doi.org/10.1371/journal.pone.0123235)
- 51. **Baba HA, Stypmann J, Grabellus F,** et al. Dynamic regulation of MEK/Erks and Akt/GSK-3beta in human end-stage heart failure after left ventricular mechanical support: myocardial mechanotransduction-sensitivity as a possible molecular mechanism. Cardiovasc Res. 2003 Aug 1;59(2):390-9. doi: [10.1016/s0008-6363\(03\)00393-6](https://doi.org/10.1016/S0008-6363(03)00393-6)
- 52. **Wong SC, Fukuchi M, Melnyk P, Rodger I, Giaid A.** Induction of cyclooxygenase-2 and activation of nuclear factorkappaB in myocardium of patients with congestive heart failure. Circulation. 1998 Jul 14;98(2):100-3. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.98.2.100) [cir.98.2.100](https://doi.org/10.1161/01.CIR.98.2.100)
- 53. **Flesch M, Margulies KB, Mochmann HC, Engel D, Sivasubramanian N, Mann DL.** Differential regulation of mitogen-activated protein kinases in the failing human heart

in response to mechanical unloading. Circulation. 2001 Nov 6;104(19):2273-6. doi: [10.1161/hc4401.099449](https://doi.org/10.1161/hc4401.099449)

- 54. **Canseco DC, Kimura W, Garg S,** et al. Human ventricular unloading induces cardiomyocyte proliferation. J Am Coll Cardiol. 2015 Mar 10;65(9):892-900. doi: [10.1016/j.](https://doi.org/10.1016/j.jacc.2014.12.027) [jacc.2014.12.027](https://doi.org/10.1016/j.jacc.2014.12.027)
- 55. **D'Ascia C, Cittadini A, Monti MG, Riccio G, Sacca L.** Effects of biventricular pacing on interstitial remodelling, tumor necrosis factor-alpha expression, and apoptotic death in failing human myocardium. Eur Heart J. 2006 Jan;27(2):201- 6. doi: [10.1093/eurheartj/ehi579](https://doi.org/10.1093/eurheartj/ehi579)
- 56. **Xia Y, Lee K, Li N, Corbett D, Mendoza L, Frangogiannis NG.** Characterization of the inflammatory and fibrotic response in a mouse model of cardiac pressure overload. Histochem Cell Biol. 2009 Apr;131(4):471-81. doi: [10.1007/s00418-008-](https://doi.org/10.1007/s00418-008-0541-5) [0541-5](https://doi.org/10.1007/s00418-008-0541-5)
- 57. **Herum KM, Lunde IG, Skrbic B,** et al. Syndecan-4 is a key determinant of collagen cross-linking and passive myocardial stiffness in the pressure-overloaded heart. Cardiovasc Res. 2015 May 1;106(2):217-26. doi: [10.1093/cvr/](https://doi.org/10.1093/cvr/cvv002) [cvv002](https://doi.org/10.1093/cvr/cvv002)
- 58. **Mukherjee D, Sen S.** Alteration of collagen phenotypes in ischemic cardiomyopathy. J Clin Invest. 1991 Oct;88(4):1141-6. doi: [10.1172/JCI115414](https://doi.org/10.1172/JCI115414)
- 59. **Marijianowski MM, Teeling P, Mann J, Becker AE.** Dilated cardiomyopathy is associated with an increase in the type I/type III collagen ratio: a quantitative assessment. J Am Coll Cardiol. 1995 May;25(6):1263-72. doi: [10.1016/0735-](https://doi.org/10.1016/0735-1097(94)00557-7) [1097\(94\)00557-7](https://doi.org/10.1016/0735-1097(94)00557-7)
- 60. **Morita H, Suzuki G, Mishima T,** et al. Effects of long-term monotherapy with metoprolol CR/XL on the progression of left ventricular dysfunction and remodeling in dogs with chronic heart failure. Cardiovasc Drugs Ther. 2002 Sep;16(5):443-9. doi: [10.1023/a:1022142620189](https://doi.org/10.1023/A:1022142620189)
- 61. **Shigeyama J, Yasumura Y, Sakamoto A,** et al. Increased gene expression of collagen Types I and III is inhibited by beta-receptor blockade in patients with dilated cardiomyopathy. Eur Heart J. 2005 Dec;26(24):2698-705. doi: [10.1093/eurheartj/ehi492](https://doi.org/10.1093/eurheartj/ehi492)
- 62. **Zannad F, Alla F, Dousset B, Perez A, Pitt B.** Limitation of excessive extracellular matrix turnover may contribute to survival benefit of spironolactone therapy in patients with congestive heart failure: insights from the randomized aldactone evaluation study (RALES). Rales Investigators. Circulation. 2000 Nov 28;102(22):2700-6. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.102.22.2700) [cir.102.22.2700](https://doi.org/10.1161/01.CIR.102.22.2700)
- 63. **Hessel MH, Bleeker GB, Bax JJ,** et al. Reverse ventricular remodelling after cardiac resynchronization therapy is associated with a reduction in serum tenascin-C and plasma matrix metalloproteinase-9 levels. Eur J Heart Fail. 2007 Oct;9(10):1058-63. doi: [10.1016/j.ejheart.2007.07.007](https://doi.org/10.1016/j.ejheart.2007.07.007)
- 64. **Li YY, Feldman AM, Sun Y, McTiernan CF.** Differential expression of tissue inhibitors of metalloproteinases in the failing human heart. Circulation. 1998 Oct 27;98(17):1728- 34. doi: [10.1161/01.cir.98.17.1728](https://doi.org/10.1161/01.CIR.98.17.1728)
- 65. **Torre-Amione G, Kapadia S, Benedict CR, Oral H, Young JB, Mann DL.** Proinflammatory cytokine levels in patients with depressed left ventricular ejection fraction: a report from the Studies of Left Ventricular Dysfunction (SOLVD). J Am Coll Cardiol. 1996 Apr;27(5):1201-6. doi: [10.1016/0735-](https://doi.org/10.1016/0735-1097(95)00589-7) [1097\(95\)00589-7](https://doi.org/10.1016/0735-1097(95)00589-7)
- 66. **Gearing AJH, Beckett P, Christodoulou M,** et al. Matrix metalloproteinases and processing of pro-TNF-alpha. J Leukoc Biol. 1995 May;57(5):774-7. doi: [10.1002/jlb.57.5.774](https://doi.org/10.1002/jlb.57.5.774)
- 67. **Gullestad L, Ueland T, Kjekshus J,** et al. The predictive value of galectin-3 for mortality and cardiovascular events in the Controlled Rosuvastatin Multinational Trial in Heart Failure (CORONA). Am Heart J. 2012 Dec;164(6):878-83. doi: [10.1016/j.ahj.2012.08.021](https://doi.org/10.1016/j.ahj.2012.08.021)
- 68. **Batlle M, Pérez-Villa F, Lázaro A,** et al. Decreased expression of thrombospondin-1 in failing hearts may favor ventricular remodeling. Transplant Proc. 2009 Jul-Aug;41(6):2231-3. doi: [10.1016/j.transproceed.2009.06.009](https://doi.org/10.1016/j.transproceed.2009.06.009)
- 69. **Vila V, Martínez-Sales V, Almenar L, Lazaro IS, Villa P, Reganon E.** Inflammation, endothelial dysfunction and angiogenesis markers in chronic heart failure patients. Int J Cardiol. 2008 Nov 12;130(2):276-7. doi: [10.1016/j.](https://doi.org/10.1016/j.ijcard.2007.07.010) [ijcard.2007.07.010](https://doi.org/10.1016/j.ijcard.2007.07.010)
- 70. **Xia Y, Dobaczewski M, Gonzalez-Quesada C,** et al. Endogenous thrombospondin 1 protects the pressureoverloaded myocardium by modulating fibroblast phenotype and matrix metabolism. Hypertension. 2011 Nov;58(5):902- 11. doi: [10.1161/HYPERTENSIONAHA.111.175323](https://doi.org/10.1161/HYPERTENSIONAHA.111.175323)
- 71. **Frangogiannis NG.** Regulation of the inflammatory response in cardiac repair. Circ Res. 2012 Jan 6;110(1):159-73. doi: [10.1161/CIRCRESAHA.111.243162](https://doi.org/10.1161/CIRCRESAHA.111.243162)
- 72. **Raafs AG, Verdonschot JAJ, Henkens M,** et al. The combination of carboxy-terminal propeptide of procollagen type I blood levels and late gadolinium enhancement at cardiac magnetic resonance provides additional prognostic information in idiopathic dilated cardiomyopathy - A multilevel assessment of myocardial fibrosis in dilated cardiomyopathy. Eur J Heart Fail. 2021 Jun;23(6):933-944. doi: [10.1002/ejhf.2201](https://doi.org/10.1002/ejhf.2201)
- 73. **López B, Querejeta R, Varo N, González A, Larman M, Martínez Ubago JL, Díez J.** Usefulness of serum carboxyterminal propeptide of procollagen type I in assessment of the cardioreparative ability of antihypertensive treatment in hypertensive patients. Circulation. 2001 Jul 17;104(3):286- 91. doi: [10.1161/01.cir.104.3.286](https://doi.org/10.1161/01.CIR.104.3.286)
- 74. **Stienen S, Ferreira JP, Pitt B,** et al. Eplerenone prevents an increase in serum carboxy-terminal propeptide of procollagen type I after myocardial infarction complicated

by left ventricular dysfunction and/or heart failure. Eur J Heart Fail. 2020 May;22(5):901-903. doi: [10.1002/ejhf.1812](https://doi.org/10.1002/ejhf.1812)

- 75. **Lewis GA, Dodd S, Clayton D,** et al. Pirfenidone in heart failure with preserved ejection fraction: a randomized phase 2 trial. Nat Med. 2021 Aug;27(8):1477-1482. doi: [10.1038/](https://doi.org/10.1038/s41591-021-01452-0) [s41591-021-01452-0](https://doi.org/10.1038/s41591-021-01452-0)
- 76. **Micheletti R, Plaisance I, Abraham BJ,** et al. The long noncoding RNA Wisper controls cardiac fibrosis and remodeling. Sci Transl Med. 2017 Jun 21;9(395):eaai9118. doi: [10.1126/scitranslmed.aai9118](https://doi.org/10.1126/scitranslmed.aai9118)
- 77. **Abraham ST, Benscoter HA, Schworer CM, Singer HA.** A role for Ca2+/calmodulin-dependent protein kinase II in the mitogen-activated protein kinase signaling cascade of cultured rat aortic vascular smooth muscle cells. Circ Res. 1997 Oct;81(4):575-84. doi: [10.1161/01.res.81.4.575](https://doi.org/10.1161/01.RES.81.4.575)
- 78. **Akhand AA, Du J, Liu W,** et al. Redox-linked cell surfaceoriented signaling for T-cell death. Antioxid Redox Signal. 2002 Jun;4(3):445-54. doi: [10.1089/15230860260196236](https://doi.org/10.1089/15230860260196236)
- 79. **Suzuki G, Morita H, Mishima T,** et al. Effects of longterm monotherapy with eplerenone, a novel aldosterone blocker, on progression of left ventricular dysfunction and remodeling in dogs with heart failure. Circulation. 2002 Dec 3;106(23):2967-72. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.0000039104.56479.42) [cir.0000039104.56479.42](https://doi.org/10.1161/01.CIR.0000039104.56479.42)
- 80. **Hall JL, Grindle S, Han X,** et al. Genomic profiling of the human heart before and after mechanical support with a ventricular assist device reveals alterations in vascular signaling networks. Physiol Genomics. 2004 May 19;17(3):283-91. doi: [10.1152/physiolgenomics.00004.2004](https://doi.org/10.1152/physiolgenomics.00004.2004)
- 81. **Drakos SG, Kfoury AG, Hammond EH,** et al. Impact of mechanical unloading on microvasculature and associated central remodeling features of the failing human heart. J Am Coll Cardiol. 2010 Jul 27;56(5):382-91. doi: [10.1016/j.](https://doi.org/10.1016/j.jacc.2010.04.019) [jacc.2010.04.019](https://doi.org/10.1016/j.jacc.2010.04.019)
- 82. **Harrington J, Nixon AB, Daubert MA,** et al. Circulating Angiokines Are Associated With Reverse Remodeling and Outcomes in Chronic Heart Failure. J Card Fail. 2023 Jun;29(6):896-906. doi: [10.1016/j.cardfail.2022.12.011](https://doi.org/10.1016/j.cardfail.2022.12.011)
- 83. **O'Connor CM, Whellan DJ, Lee KL,** et al. Efficacy and safety of exercise training in patients with chronic heart failure: HF-ACTION randomized controlled trial. JAMA. 2009 Apr 8;301(14):1439-50. doi: [10.1001/jama.2009.454](https://doi.org/10.1001/jama.2009.454)
- 84. **Krause A, Sillard R, Kleemeier B,** et al. Isolation and biochemical characterization of LEAP-2, a novel blood peptide expressed in the liver. Protein Sci. 2003 Jan;12(1):143-52. doi: [10.1110/ps.0213603](https://doi.org/10.1110/ps.0213603)
- 85. **Higgins SL, Hummel JD, Niazi IK,** et al. Cardiac resynchronization therapy for the treatment of heart failure in patients with intraventricular conduction delay and malignant ventricular tachyarrhythmias. J Am Coll Cardiol. 2003 Oct 15;42(8):1454-9. doi: [10.1016/s0735-](https://doi.org/10.1016/S0735-1097(03)01042-8) [1097\(03\)01042-8](https://doi.org/10.1016/S0735-1097(03)01042-8)
- 86. **Yusuf S, Pitt B, Davis CE, Hood WB, Cohn JN.**; SOLVD Investigators. Effect of enalapril on survival in patients with reduced left ventricular ejection fractions and congestive heart failure. N Engl J Med. 1991 Aug 1;325(5):293-302. doi: [10.1056/NEJM199108013250501](https://doi.org/10.1056/NEJM199108013250501)
- 87. Greenberg B, Quinones MA, Koilpillai C, et al. Effects of long-term enalapril therapy on cardiac structure and function in patients with left ventricular dysfunction. Results of the SOLVD echocardiography substudy. Circulation. 1995 May 15;91(10):2573-81. doi: [10.1161/01.cir.91.10.2573](https://doi.org/10.1161/01.CIR.91.10.2573)
- 88. Wong M, Staszewsky L, Latini R, et al. Valsartan benefits left ventricular structure and function in heart failure: Val-HeFT echocardiographic study. J Am Coll Cardiol. 2002 Sep 4;40(5):970-5. doi: [10.1016/s0735-](https://doi.org/10.1016/S0735-1097(02)02063-6) [1097\(02\)02063-6](https://doi.org/10.1016/S0735-1097(02)02063-6)
- 89. **Chan AK, Sanderson JE, Wang T,** et al. Aldosterone receptor antagonism induces reverse remodeling when added to angiotensin receptor blockade in chronic heart failure. J Am Coll Cardiol. 2007 Aug 14;50(7):591-6. doi: [10.1016/j.](https://doi.org/10.1016/j.jacc.2007.03.062) [jacc.2007.03.062](https://doi.org/10.1016/j.jacc.2007.03.062)
- 90. **Doughty RN, Whalley GA, Gamble G, MacMahon S, Sharpe N.** Left ventricular remodeling with carvedilol in patients with congestive heart failure due to ischemic heart disease. Australia-New Zealand Heart Failure Research Collaborative Group. J Am Coll Cardiol. 1997 Apr;29(5):1060-6. doi: [10.1016/s0735-1097\(97\)00012-0](https://doi.org/10.1016/S0735-1097(97)00012-0)
- 91. **Rastogi A, Novak E, Platts AE, Mann DL.** Epidemiology, pathophysiology and clinical outcomes for heart failure patients with a mid-range ejection fraction. Eur J Heart Fail. 2017 Dec;19(12):1597-1605. doi: [10.1002/ejhf.879](https://doi.org/10.1002/ejhf.879)
- 92. Alexanian M, Przytycki PF, Micheletti R, et al. A transcriptional switch governs fibroblast activation in heart disease. Nature. 2021 Jul;595(7867):438-443. doi: [10.1038/](https://doi.org/10.1038/s41586-021-03674-1) [s41586-021-03674-1](https://doi.org/10.1038/s41586-021-03674-1)
- 93. **Garcia S, Kandar F, Boyle A,** et al. Effects of pulsatile- and continuous-flow left ventricular assist devices on left ventricular unloading. J Heart Lung Transplant. 2008 Mar;27(3):261-7. doi: [10.1016/j.healun.2007.12.001](https://doi.org/10.1016/j.healun.2007.12.001)
- 94. **Kato TS, Chokshi A, Singh P,** et al. Effects of continuousflow versus pulsatile-flow left ventricular assist devices on myocardial unloading and remodeling. Circ Heart Fail. 2011 Sep;4(5):546-53. doi: [10.1161/](https://doi.org/10.1161/CIRCHEARTFAILURE.111.962142) [CIRCHEARTFAILURE.111.962142](https://doi.org/10.1161/CIRCHEARTFAILURE.111.962142)
- 95. **Klotz S, Deng MC, Stypmann J,** et al. Left ventricular pressure and volume unloading during pulsatile versus nonpulsatile left ventricular assist device support. Ann Thorac Surg. 2004 Jan;77(1):143-9; discussion 149-50. doi: [10.1016/s0003-4975\(03\)01336-5](https://doi.org/10.1016/S0003-4975(03)01336-5)
- 96. **Linde C, Leclercq C, Rex S,** et al. Long-term benefits of biventricular pacing in congestive heart failure: results from the MUltisite STimulation in cardiomyopathy (MUSTIC) study. J Am Coll Cardiol. 2002 Jul 3;40(1):111-8. doi: [10.1016/](https://doi.org/10.1016/S0735-1097(02)01932-0) [s0735-1097\(02\)01932-0](https://doi.org/10.1016/S0735-1097(02)01932-0)
- 97. **St John Sutton MG, Plappert T, Abraham WT,** et al. Effect of cardiac resynchronization therapy on left ventricular size and function in chronic heart failure. Circulation. 2003 Apr 22;107(15):1985-90. doi: [10.1161/01.](https://doi.org/10.1161/01.CIR.0000065226.24159.E9) [CIR.0000065226.24159.E9](https://doi.org/10.1161/01.CIR.0000065226.24159.E9)
- 98. Merlo M, Stolfo D, Anzini M, et al. Persistent recovery of normal left ventricular function and dimension in idiopathic dilated cardiomyopathy during long‐term follow‐up: does real healing exist? J Am Heart Assoc. 2015 Jan 13;4(1):e001504. doi: [10.1161/JAHA.114.000570](https://doi.org/10.1161/JAHA.114.000570)
- 99. **Halliday BP, Wassall R, Lota AS,** et al. Withdrawal of pharmacological treatment for heart failure in patients with recovered dilated cardiomyopathy (TRED-HF): an open-label, pilot, randomised trial. Lancet. 2019 Jan 5;393(10166):61-73. doi: [10.1016/S0140-6736\(18\)32484-X](https://doi.org/10.1016/S0140-6736(18)32484-X)

TO CITE THIS ARTICLE:

Park AC, Mann DL. The Pathobiology of Myocardial Recovery and Remission: From Animal Models to Clinical Observations in Heart Failure Patients. Methodist DeBakey Cardiovasc J. 2024;20(4):16-30. doi: [10.14797/mdcvj.1389](https://doi.org/10.14797/mdcvj.1389)

Submitted: 08 April 2024 **Accepted:** 21 June 2024 **Published:** 20 August 2024

COPYRIGHT:

© 2024 The Author(s). This is an open-access article distributed under the terms of the Attribution-NonCommercial 4.0 International (CC BY-NC 4.0), which permits unrestricted use, distribution, and reproduction in any noncommercial medium, provided the original author and source are credited. See https://creativecommons.org/licenses/by-nc/4.0/.

Methodist DeBakey Cardiovascular Journal is a peer-reviewed open access journal published by Houston Methodist DeBakey Heart & Vascular Center.

