

Article

Isolation and Cytotoxicity Evaluation of the Chemical Constituents from *Cephalantheropsis gracilis*

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Abstract: *Cephalantheropsis gracilis* afforded five new compounds: cephalanthrin-A (**1**), cephalanthrin-B (**2**), cephathrene-A (**3**), cephathrene-B (**4**), methyl 2-(aminocarbonyl) phenylcarbamate (**5**), and 52 known compounds. The structures of the new compounds were determined by spectroscopic analysis. Among the compounds isolated, tryptanthrin (**6**), phaitanthrin A (**7**), cephalinone D (**19**), and flavanthrin (**30**) showed significant cytotoxicity against MCF-7, NCI-H460, and SF-268 cell lines.

Keywords: *Cephalantheropsis gracilis*; Orchidaceae; quinazoline; tryptanthrin; indolotryptanthrin; dihydrophenanthrene; cytotoxicity

1. Introduction

The genus *Cephalantheropsis* (also known as *Cephalanceropsis*) belongs to the Orchidaceae family and is comprised of eight species distributed in Southeast Asia. The plant, *Cephalantheropsis gracilis* (Lindl.) Shiu-Ying Hu var. *gracilis*, is an orchid native to Taiwan and grows in forests at altitudes of 500–1500 m throughout the island [1]. The crude methanol extract of *C. gracilis* showed significant cytotoxicity against human breast carcinoma (MCF-7), lung carcinoma (NCI-H460), and central nervous system carcinoma (SF-268) cell lines in our preliminary screening. In earlier papers, the isolation of indole alkaloids was reported from *C. gracilis*, but they are unlikely to be responsible for such anticancer activity [2,3]. In the course of continuing the search for bioactive molecules from *C. gracilis*, two new quinazolines, cephalanthrin-A (**1**) and cephalanthrin-B (**2**), two new dihydrophenanthrenes, cephathrene-A (**3**) and cephathrene-B (**4**), and a methyl 2-(aminocarbonyl)phenylcarbamate (**5**) [4] (Figure 1) as well as 52 known compounds were obtained and identified from a methanol extract (in addition to common long-chain fatty acids, chlorophylls, and steroids). Herein, we describe the structural elucidation of these new compounds and the cytotoxic properties of all compounds identified toward several human cancer cell lines.

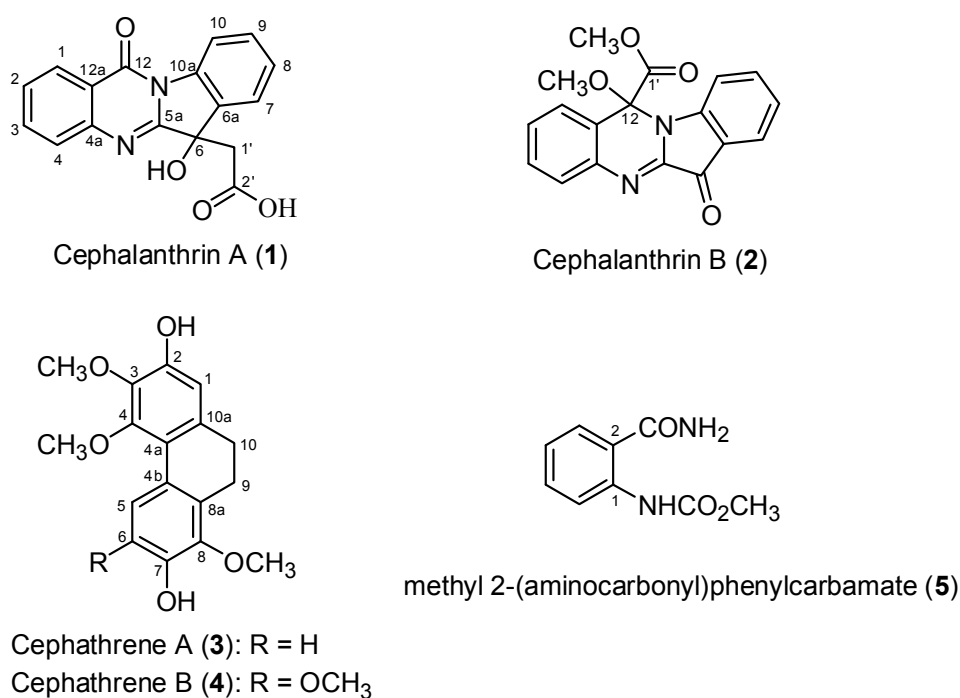


Figure 1. Structures of five new compounds 1–5.

2. Results and Discussion

Cephalanthrin-A (**1**) was isolated as an optically active white amorphous powder. The molecular formula was determined to be C₁₇H₁₂N₂O₄ from a molecular ion of *m/z* 308.0794 by HREIMS. In the IR spectrum, a very broad band at 3000 cm⁻¹ and an absorption band at 1652 cm⁻¹ both indicated the presence of a carboxylic acid. The 1D ¹H and ¹³C NMR data (Table 1, Figure S1), together with 2D COSY, HMQC, and HMBC spectra, revealed two sets of *o*-disubstituted benzene rings, one at δ_H 7.59 (t, *J* = 7.5 Hz, H-2), 7.77 (d, *J* = 7.5 Hz, H-4), 7.83 (t, *J* = 7.5 Hz, H-3), and 8.34 (1H, d, *J* = 7.5 Hz, H-1);

δ_C 123.0 (C-12a), 127.4 (C-1), 128.0 (C-2), 128.5 (C-4), 135.2 (C-3), and 148.4 (C-4a). The other was at δ_H 7.38 (t, $J = 7.7$ Hz, H-8), 7.51 (t, $J = 7.7$ Hz, H-9), 7.73 (1H, d, $J = 7.7$ Hz, H-7), and 8.49 (1H, d, $J = 7.7$ Hz, H-10); δ_C 117.2 (C-10), 124.6 (C-7), 127.3 (C-8), 130.8 (C-9), 134.2 (C-6a), and 140.8 (C-10a). The ^{13}C NMR data also indicated the presence of an imino C-5a (δ 161.9) and an amidic C-12 (δ 160.0). These signals appear to be very closely related to indolo[2,1-*b*]quinazoline-6,12-dione (tryptanthrin, **6**), except that the carbonyl C-6 is replaced by a saturated quaternary carbon (δ 75.9). NMR spectroscopic data also allowed us to determine the remaining two substituents on C-6 being an –OH group (δ 3.81) and a –CH₂COOH group (δ_H 3.45, 3.55 (each 1H, d, $J = 16.5$ Hz, H-1'); δ_C 43.6 (C-1') and 170.9 (C-2')). The methylene protons adjacent to a quaternary carbon (C-6) could be split due to either chirality or steric hindrance. HMBC correlations from H-1 to C-12, H-7 to C-6 and H-1' to C-5a, C-6 and C-6a, as well as the NOE correlations between H-1' and H-7, supported a structure of 6-hydroxy-6-(carboxymethyl)-tryptanthrin for cephalanthrin-A (**1**).

Table 1. 1H and ^{13}C NMR Spectroscopic Data and HMBC Correlations for Cephalanthrins **1** and **2**.

Position	1 in Acetone-<i>d</i>₆ (300 MHz/75 MHz)			2 in CDCl₃ (300 MHz/75 MHz)		
	δ_H (<i>J</i> in Hz)	δ_C	HMBC	δ_H (<i>J</i> in Hz)	δ_C	HMBC
1	8.34 d (7.5)	127.4	C-3, -4a, -12	7.60 d (7.6)	126.1	C-3, -4a, -12
2	7.59 t (7.5)	128.0	C-4, -12a	7.41 t (7.6)	129.3	C-4, -12a
3	7.83 t (7.5)	135.2	C-1, -4a	7.52 t (7.6)	131.4	C-1, -4a
4	7.77 d (7.5)	128.5	C-2, -12a	7.72 d (7.6)	129.7	C-2, -12a
4a		148.4			141.9	
5a		161.9			144.7	
6		75.9			184.0	
6a		134.2			120.5	
7	7.73 d (7.7)	124.6	C-6, -9, -10a	7.83 d (7.6)	125.5	C-6, -9, -10a
8	7.38 t (7.7)	127.3	C-6a, -10	7.20 t (7.6)	124.0	C-6a, -10
9	7.51 t (7.7)	130.8	C-7, -10a	7.59 t (7.6)	137.9	C-7, -10a
10	8.49 d (7.7)	117.2	C-6a, -8	7.23 d (7.6)	112.3	C-6a, -8
10a		140.8			148.9	
12		160.0			87.4	
12a		123.0			120.5	
1'	3.45 d (16.5) 3.55 d (16.5)	43.6	C-5a, -6, -6a, -2'		167.8	
2'		170.9				
6-OH	3.81 s					
1'-OCH ₃				3.71 s	53.8	C-1'
12-OCH ₃				3.07 s	50.6	C-12

Cephalanthrin-B (**2**), also isolated as an optically active yellow amorphous powder, was determined to have a molecular formula of C₁₈H₁₄N₂O₄. By comparison of the 1H and ^{13}C NMR spectra of **2** (Figure S2) with those of tryptanthrin (**6**) and cephalanthrin-A (**1**), an oxygenated quaternary carbon (δ 87.4) was shown to replace the carbonyl C-12 to form a 12,12-disubstituted tryptanthrin (Table 1). Methoxyl (δ_H 3.07; δ_C 50.6) and methoxycarbonyl (δ_H 3.71; δ_C 53.8 and 167.8) substituents were also identified, and their presence was confirmed by HMBC correlations from both H-1 and 12-OCH₃ to

C-12 and the NOE correlations between H-1, H-10 and 1'-OCH₃, 12-OCH₃. Hence, a structure of 12-methoxy-12-(methoxycarbonyl)-tryptanthrin was deduced for cephalanthrin-B (**2**). Although compound **2** has been synthesized by Cornforth *et al.* [5], this represents the first isolation of a pure compound from a natural source.

Due to the small specific rotations of compounds **1** ($[\alpha]_D +8.0^\circ$) and **2** ($[\alpha]_D +3.0^\circ$), we suspected they might be not optically pure compounds. The configuration of compounds **1** and **2** has not been determined due to the isolation of insufficient amounts of these materials. However, we adopted the similar structure of phaitanthrin A (**7**) as a model. First, our attempts to synthesize a pair of diastereomeric esters by acylating **7** with (+)- α -methoxy- α -trifluoromethylphenylacetyl chloride [(+)-MTPACl] [6], even with acetyl chloride, were unsuccessful. This most likely is due to steric inhibition at the tertiary alcohol, which is present in the tryptanthrin skeleton. We then tried to analyze the C-6 chemical shift behaviors using the chiral shift reagents, tris[3-trifluoroacetyl-D- and L-camphorato]europium(III) [(*R*)- and (*S*)-Eu(tfc)₃] [7], again to no avail.

Cephatrene-A (**3**) was isolated as a white amorphous powder with a molecular formula of C₁₇H₁₈O₅ as determined by the molecular ion peak at m/z 302.1153 in HREIMS data. UV absorptions at 267 and 304 nm indicated the presence of a benzene system, and the IR spectrum revealed an OH absorption band at 3404 cm⁻¹. From the ¹H NMR spectrum, two mutually-coupled aromatic protons at δ 6.86 (1H, d, $J = 8.7$ Hz, H-6) and 7.93 (1H, d, $J = 8.7$ Hz, H-5) and a proton at δ 6.63 (1H, s, H-1) suggested the presence of 1,2,3,4-tetrasubstituted and pentasubstituted benzene rings, respectively (Table 2, Figure S3). We also found two mutually coupled aliphatic signals at δ 2.66 (2H, m, H-10) and 2.79 (2H, m, H-9), which were assigned to an ethylene group. HMBC correlations of H-9 with C-4b, C-8, C-8a, C-10, and C-10a, as well as H-10 with C-1, C-4a, C-8a, C-9, and C-10a indicated that the two benzene rings are linked together by the ethylene group. Furthermore, the HMBC correlation between H-5 and C-4a established the existence of a bond between C-4a and C-4b. Thus, compound **3** possessed a 9,10-dihydrophenanthrene skeleton. The HMBC correlations of H-1, H-5, H-6, H-9, and H-10 allowed for the identification of the quaternary aromatic carbons, C-2, C-3, C-4a, C-4b, C-7, C-8, C-8a, and C-10a. We determined the identity of five other substituents, two hydroxyls and three methoxyls. The hydroxyl signal at δ 5.65 showed HMBC correlations with C-6, C-8, and C-7, and the other hydroxyl signal at δ 5.70 showed HMBC correlations with C-1, C-3, and C-2, indicating that the two hydroxyl groups are attached to C-7 and C-2, respectively. Whereas the three methoxyl signals at δ 3.75, 3.79, and 3.96 were shown to be located at C-4, C-8, and C-3, respectively, owing to the HMBC correlations of 3-OCH₃ with C-3, 4-OCH₃ with C-4, and 8-OCH₃ with C-8. Additional evidence for the positions of these substituents came from the NOE correlations between 2-OH and H-1, 3-OCH₃; H-5 and 4-OCH₃; H-1 and H-10; 8-OCH₃ and 7-OH, H-9. Therefore, cephatrene-A (**3**) was assigned the structure 2,7-dihydroxy-3,4,8-trimethoxy-9,10-dihydrophenanthrene.

Cephatrene-B (**4**) was isolated as a white amorphous powder. The HREIMS showed a molecular ion consistent with the molecular formula C₁₈H₂₀O₆. The spectral data showed a resemblance to compound **3** (Table 2). The ¹H NMR spectrum (Figure S4) disclosed the presence of two singlet aromatic protons at 6.63 (H-1) and 7.75 (H-5), indicating a hexasubstituted 9,10-dihydrophenanthrene. The regiochemistries of the substituents, two hydroxyls and four methoxyls, were determined by HMQC, HMBC, and NOESY experiments. As in the case of **3**, the two hydroxyls at δ 5.59 and 5.70 are located at C-7 and C-2, respectively, whereas three of the four methoxyls at δ 3.75, 3.84, and 3.97

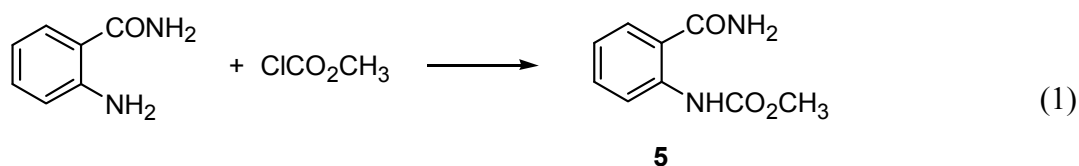
are at C-4, C-8, and C-3, respectively. The remaining methoxyl at δ 3.93 was thought to be at C-6 due to NOE correlations between H-5 and 4- and 6-OCH₃. Thus, the structure of 2,7-dihydroxy-3,4,6,8-tetramethoxy-9,10-dihydrophenanthrene was established for cephatrene-B (**4**).

Table 2. ¹H and ¹³C NMR Spectroscopic Data and HMBC Correlations for Cephatrenes **3** and **4**.

Position	3 in CDCl ₃ (400 MHz/100 MHz)			4 in CDCl ₃ (300 MHz/75 MHz)		
	δ_H (J in Hz)	δ_C	HMBC	δ_H (J in Hz)	δ_C	HMBC
1	6.63 s	109.9	C-2, -3, -4a, -10	6.63 s	110.1	C-2, -3, -4a, -10
2		147.6			147.7	
3		139.0			139.0	
4		150.5			150.4	
4a		120.3			120.3	
4b		125.9			124.2 *	
5	7.93 d (8.7)	124.2	C-4a, -7, -8a	7.75 s	106.6	C-4a, -4b, -6, -7, -8a
6	6.86 d (8.7)	112.8	C-4b, -8		145.5	
7		147.1			137.1	
8		143.6			143.7	
8a		130.8			124.3 *	
9	2.79 m	22.4	C-4b, -8, -8a, -10, -10a	2.73 m	21.5	C-4b, -8, -10, -10a
10	2.66 m	29.5	C-1, -4a, -8a, -9, -10a	2.64 m	29.8	C-1, -4a, -8a, -9
10a		134.5			135.2	
2-OH	5.70 s		C-1, -2, -3	5.70 s		
3-OCH ₃	3.96 s	61.1	C-3	3.97 s	61.2	C-3
4-OCH ₃	3.75 s	60.1	C-4	3.75 s	60.1	C-4
6-OCH ₃				3.93 s	56.3	C-6
7-OH	5.65 s		C-6, -7, -8	5.59 s		
8-OCH ₃	3.79 s	61.3	C-8	3.84 s	60.8	C-8

* Assignments may be interchangeable.

Compound **5**, with molecular formula C₉H₁₀N₂O₃, was isolated as a white amorphous powder. It had UV absorptions at 257, 289, and 311 nm, indicative of an aromatic system. The ¹H NMR spectrum (Figure S5) showed resonances at δ 7.05 (1H, t, J = 7.8 Hz, H-4), 7.49 (1H, t, J = 7.8 Hz, H-5), 7.83 (1H, d, J = 7.8 Hz, H-3), and 8.38 (1H, d, J = 7.8 Hz, H-6) for an *o*-disubstituted benzene. The HMBC correlations from H-3 to an amide carbon (δ 172.0) and from OCH₃ (δ 3.71) to a carbamate carbon (δ 154.6) and the NOE correlation between H-6 and an amine proton (δ 11.29) suggested the structure of methyl 2-(aminocarbonyl)phenylcarbamate for **5**. The ¹H and ¹³C NMR spectra of **5** and the acylation product of *o*-aminobenzamide with methyl chloroformate (ClCO₂CH₃) were identical and further confirmed the structure of **5** (Equation (1)).



Other known compounds were also isolated from *C. grilis* including 7 quinazolines, tryptanthrin (**6**) [8], phaitanthrin A (**7**) [8], phaitanthrin B (**8**) [8], methylisatoid (**9**) [8], candidine (**10**) [8],

1*H*-quinazoline-2,4-dione (**11**) [9], and 3-(2-hydroxy-phenyl)-3*H*-quinazolin-4-one (**12**) [10]; 15 indole alkaloids, cephalandole A (**13**) [2], cephalandole B (**14**) [2], cephalandole C (**15**) [2], cephalinone A (**16**) [2], cephalinone B (**17**) [2], cephalinone C (**18**) [2], cephalinone D (**19**) [2], (*S*)-3-(2-oxopropyl)-3-hydroxyindolin-2-one (**20**) [2], methyl dioxindole-3-acetate (**21**) [2], isatan (**22**) [2], indigo (**23**) [2], indirubin (**24**) [2], isatin (**25**) [2], indole-3-carbaldehyde (**26**) [2], and indole-3-carboxylic acid (**27**) [2]; 2 indolotryptanthrins cephatrindole A (**28**) [3] and cephatrindole B (**29**) [3]; 5 dihydrophenanthrenes, flavanthrin (**30**) [11], coelonin (**31**) [11], 6-*O*-methylcoelonin (**32**) [12], 2,7-dihydroxy-3,4-dimethoxy-9,10-dihydrophenanthrene (**33**) [13], and 2,7-dihydroxy-3,4,6-trimethoxy-9,10-dihydrophenanthrene (**34**) [11]; 1 lignin, secoisolariciresinol (**35**) [14]; 1 flavonoid, kaempferol 3-rutinoside (**36**) [15]; 1 ionol, blumenol A (**37**) [16]; and 20 benzenoids, 2-aminobenzoic acid (**38**), methyl 2-aminobenzoate (**39**), *N*-cinnamoyltyramine (**40**), *N*-*p*-coumaroyltyramine (**41**), *N*-*trans*-feruloyltyramine (**42**), dihydroconiferyl dihydro-*p*-coumarate (**43**), 4-hydroxybenzaldehyde (**44**), 1-(4-hydroxy-phenyl)ethanone (**45**), 4-hydroxy-phenethyl alcohol (**46**), 3-(4-hydroxy-phenyl)-propionic acid (**47**), vanillin (**48**), vanillic acid (**49**), 4-hydroxy-3-methoxybenzyl alcohol (**50**), syringaldehyde (**51**), 3,5-dimethyl-4-hydroxypropiophenone (**52**), *trans*-*p*-coumaric acid (**53**), *trans*-ferulic acid (**54**), *cis*-ferulic acid (**55**), methyl *trans*-4-hydroxy-3-methoxycinnamate (**56**), and methylsinapat (**57**).

All the isolated compounds were subjected to cytotoxic evaluation against MCF-7, NCI-H460, and SF-268 cell lines. Tryptanthrin (**6**), phaitanthrin A (**7**), cephalinone D (**19**), and flavanthrin (**30**) showed significant cytotoxicity against MCF-7, NCI-H460, and SF-268 cell lines with IC₅₀ values of 7.6–42.9 μM (Table 3).

Table 3. Cytotoxicity of active compounds toward three cancer lines.

Compound	IC ₅₀ (μM)		
	MCF-7	NCI-H460	SF-268
Tryptanthrin (6)	9.4 ± 0.3	8.5 ± 0.8	22.6 ± 1.1
Phaitanthrin A (7)	17.8 ± 0.8	17.3 ± 1.2	42.9 ± 1.0
Cephalinone D (19)	7.6 ± 0.7	7.8 ± 1.0	12.2 ± 1.3
Flavanthrin (30)	21.9 ± 1.5	22.8 ± 2.3	23.0 ± 2.0

Values were mean ± SD (*n* = 3–8); MCF-7 = human breast tumor cell line; NCI-H460 = human lung tumor cell line; SF-268 = human entral nervous system tumor cell line.

3. Experimental Section

3.1. General

Optical rotations were measured on a Jasco DIP-370 digital polarimeter (JASCO, Tokyo, Japan). UV spectra were recorded on an Agilent 8453 spectrophotometer (Agilent Technologies, Palo Alto, CA, USA). IR spectra were recorded on a Nicolet Magna FT-IR spectrophotometer (Nicolet Instrument, Inc., Madison, WI, USA). NMR spectra were recorded on a Bruker Avance 300 (Bruker, Karlsruhe, Germany) and AMX 400 spectrometers (Bruker, Karlsruhe, Germany), and all chemical shifts are given in ppm using tetramethylsilane (δ 0.00) as an internal standard. Mass spectra were obtained on a VG 70-250S spectrometer by a direct inlet system (Micromass Corp., Manchester, UK).

3.2. Plant Material

Whole *Cephalantheropsis gracilis* plants were collected from Pingtung Hsien, Taiwan, in December 2004, as authenticated by Chang-Sheng Kuoh, Department of Biology, National Cheng Kung University, Tainan, Taiwan. A voucher specimen (No: PLW-0401) was deposited in the Herbarium of National Cheng Kung University, Tainan, Taiwan.

3.3. Extraction and Isolation

The dried aerial parts of *C. gracilis* (2.4 kg) were extracted with MeOH (8 L) under reflux 8 times. The combined extracts were concentrated under reduced pressure to produce a dark brown syrup. The syrup was then suspended in H₂O and then partitioned with hexane, CHCl₃, and EtOAc, successively. The concentrated hexane layer (47 g) was chromatographed on a silica gel column by eluting with a gradient of hexane-Me₂CO (10:1 to pure Me₂CO) to give six fractions. Fraction 3 was chromatographed on silica gel by elution with hexane-*i*-Pr₂O (1:3 to pure *i*-Pr₂O) to give **48** (9.6 mg). Fraction 4 was chromatographed on silica gel using the same solvent mixture to yield **3** (4.3 mg), **34** (7.0 mg), **52** (0.7 mg), and **57** (1.4 mg). Fraction 5 was chromatographed on silica gel eluting with *i*-Pr₂O (pure *i*-Pr₂O to 30:1 of *i*-Pr₂O-Me₂CO to pure Me₂CO) to give **4** (6.6 mg), **32** (3.5 mg), and **29** (5 mg).

The CHCl₃ extract (30 g) was chromatographed on a silica gel column by eluting with a gradient of hexane-Me₂CO (1:2 to pure Me₂CO) to yield twelve fractions. Fraction 1 was subjected to chromatography on a silica gel column eluting with a gradient of hexane-Me₂CO (10:1 to pure Me₂CO) to give **10** (61.8 mg) and **39** (2.2 mg). Fraction 2 was chromatographed on a silica gel column eluting with a gradient of hexane-Me₂CO (6:1 to pure Me₂CO) to give **7** (39.4 mg), **2** (3.6 mg), **6** (543.1 mg), **13** (8.6 mg), and **17** (5.8 mg). Similarly, fraction 3 was chromatographed with a gradient of hexane-Me₂CO (4:1 to pure Me₂CO) to give **51** (2.2 mg) and **56** (22.7 mg). Fraction 5 was subjected to chromatography over silica gel eluting with a gradient of hexane-*i*-Pr₂O (1:4 to pure *i*-Pr₂O) to give **14** (20.0 mg) and **24** (48.0 mg). Fraction 6 was further purified on a silica gel column eluting with a gradient of *i*-Pr₂O-MeOH (50:1 to pure MeOH) to give **9** (34.4 mg), **19** (30.0 mg), and **28** (12 mg). Fraction 7 was chromatographed on a silica gel column eluting with a gradient of hexane-CHCl₃ (6:1 to pure CHCl₃) to give **25** (62.1 mg), **33** (0.7 mg), **5** (2.9 mg), **43** (11.4 mg), **44** (8 mg), and **45** (1.6 mg). Fraction 8 was subjected to chromatography over silica gel eluting with a gradient of CHCl₃-Me₂CO (30:1 to pure Me₂CO) to give **12** (50.4 mg), **26** (2.7 mg), **31** (39.5 mg), **40** (3.4 mg), and **50** (8.3 mg). Fraction 9 was further chromatographed on a silica gel column eluting with a gradient of hexane-EtOAc (15:1 to pure EtOAc) to give **11** (54.1 mg) and **18** (6.2 mg). Fraction 10 was chromatographed on a silica gel column eluting with a gradient of *i*-Pr₂O-MeOH (9:1 to pure MeOH) to give **27** (2.2 mg), **35** (2.3 mg), **42** (103 mg), **46** (1.2 mg), **49** (33.3 mg), and **37** (5.1 mg). Finally, fraction 11 was chromatographed on a silica gel column eluting with a gradient of CHCl₃-MeOH (10:1 to pure MeOH) to give **53** (2.1 mg), **54** (37.4 mg), **55** (5.4 mg), and Fraction 12 yielded **23** (328.2 mg) as a pure crystalline material.

The EtOAc extract (20 g) was subjected to column chromatography using Cosmosil 75 C18 and eluted with a gradient of H₂O-MeOH (from pure H₂O to pure MeOH) to give nine fractions. Fraction 2

was subjected to further chromatography on a Cosmosil 75 C18 column eluting with a gradient of H₂O–MeOH (from pure H₂O to pure MeOH) to give **21** (3.6 mg) and **47** (12.5 mg). Fraction 3 was chromatographed on a silica gel column eluting with a gradient of *i*-Pr₂O–MeOH (15:1 to pure MeOH) to give **22** (2.2 mg). Fraction 4 was subjected to repeated chromatography on a silica gel column eluting with a gradient of *i*-Pr₂O–MeOH (10:1 to pure MeOH) to give **1** (1.4 mg), **16** (3.5 mg), **36** (12.6 mg), and **41** (44.5 mg). Fraction 5 was chromatographed on a silica gel column eluting with a gradient of *i*-Pr₂O–MeOH (20:1 to pure MeOH) to give **8** (9.8 mg), **15** (22.2 mg), and **38** (2.9 mg). Fraction 6 was further purified over silica gel eluting with a gradient of CHCl₃–MeOH (15:1 to pure MeOH) to give **20** (20.9 mg). Fraction 7 was further chromatographed on a silica gel column eluting with a gradient of *i*-Pr₂O–MeOH (8:1 to pure MeOH) to give **30** (4.0 mg).

Cephalanthrin-A (1). White amorphous powder, mp 212–214 °C; [α]_D +8.0° (c 0.07, CH₃OH); UV λ_{\max} (log ϵ) CH₃OH 206 (4.0), 261 (3.3), 302 (3.0), 314 (3.1), 329 (3.0) nm; IR ν_{\max} (KBr) 3000 (br), 1652, 1464 cm⁻¹; EIMS m/z (rel. int.) 308 (M⁺, 13), 250 (100), 219 (8), 119 (19); HREIMS m/z 308.0794 [M]⁺ (Calcd for C₁₇H₁₂N₂O₄, 308.0797).

Cephalanthrin-B (2). Yellow amorphous powder, mp 215–217 °C; [α]_D +3.0° (c 0.11, CHCl₃); UV λ_{\max} (log ϵ) CHCl₃ 259 (3.2), 276 (3.0), 316 (2.9), 442 (2.9) nm; IR ν_{\max} (KBr) 1754, 1722, 1643, 1607, 1592 cm⁻¹; EIMS m/z (rel. int.) 322 (M⁺, 2), 291 (5), 263 (100); HREIMS m/z 322.0955 [M]⁺ (Calcd For C₁₈H₁₄N₂O₄, 322.0953).

Cephathrene-A (3). White amorphous powder, mp 96–98 °C; UV λ_{\max} (log ϵ) CHCl₃ 267 (3.3), 304 (3.2) nm; IR ν_{\max} (KBr) 3404, 1582, 1483, 1467 cm⁻¹; EIMS m/z (rel. int.) 302 (M⁺, 100), 255 (35), 184 (10); HREIMS m/z 302.1153 [M]⁺ (Calcd for C₁₇H₁₈O₅, 302.1154).

Cephathrene-B (4). White amorphous powder; UV λ_{\max} (log ϵ) CHCl₃: 265 (3.2), 316 (3.0) nm; IR ν_{\max} (KBr) 3400, 1606, 1501, 1464 cm⁻¹; EIMS m/z (rel. int.) 332 (M⁺, 20), 331 (100), 302 (24), 285 (17); HREIMS m/z 332.1262 [M]⁺ (Calcd For C₁₈H₂₀O₆, 332.1260).

Methyl 2-(Aminocarbonyl)phenylcarbamate (5). White amorphous powder, mp 200–202 °C; UV λ_{\max} (log ϵ) CH₃OH 212 (2.9), 228 (3.0), 257 (2.9), 289 (2.7), 311 (2.7) nm; IR ν_{\max} (KBr) 3417, 3211, 1726, 1686, 1626, 1598, 1531 cm⁻¹; ¹H NMR (acetone-*d*₆) δ 3.71 (3H, s, OCH₃), 6.99 and 7.74 (each 1H, br s, NH₂), 7.05 (1H, t, *J* = 7.8 Hz, H-4), 7.49 (1H, t, *J* = 7.8 Hz, H-5), 7.83 (1H, d, *J* = 7.8 Hz, H-3), 8.38 (1H, d, *J* = 7.8 Hz, H-6), 11.29 (1H, br s, 1-NH); ¹³C NMR (acetone-*d*₆) δ 52.2 (OCH₃), 119.2 (C-2), 119.6 (C-6), 122.2 (C-4), 129.1 (C-3), 133.4 (C-5), 141.7 (C-1), 154.6 (NC=O), 172.0 (2-C=O); EIMS m/z (rel. int.) 194 (M⁺, 31), 162 (32), 146 (100), 118 (9); HREIMS m/z 194.0693 [M]⁺ (calcd for C₉H₁₀N₂O₃, 194.0692).

3.4. Cytotoxicity Assay

The cytotoxicity assay was carried out according to the procedure described in the literature [17].

4. Conclusions

Five new compounds, cephalanthrin-A (**1**), cephalanthrin-B (**2**), cepathrene-A (**3**), cepathrene-B (**4**), methyl 2-(aminocarbonyl)phenylcarbamate (**5**), and 52 known compounds were isolated from

Cephalantheropsis gracilis. Cephalinone D (**19**) showed the strongest cytotoxicity against the tested tumor cell lines, with IC₅₀ values ranging from 7.6 to 42.9 μM. The modifications using Cephalinone D as template are being studied in our laboratories, aiming to discover the derivatives with strong anticancer activity.

Supplementary Materials

Supplementary materials can be found at <http://www.mdpi.com/1422-0067/16/02/3980/s1>.

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Author Contributions

Chi-Fen Chang performed experiments, analyzed data and wrote the paper; Yu-Lin Hsu, Chao-Ying Lee and Chia-Hua Wu performed experiments and analyzed data; Yang-Chang Wu revised the paper; and Ta-Hsien Chuang analyzed data and wrote the paper.

Conflicts of Interest

The authors declare no conflict of interest.

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