Supporting information for

Snowball: a novel gene family required for developmental patterning in fruiting bodies of mushroom-forming fungi (Agaricomycetes)

Csenge Földi¹², Zsolt Merenyi¹, Bálint Balázs¹, Árpád Csernetics¹, Nikolett Miklovics¹, Hongli Wu¹, Botond Hegedüs¹, Máté Virágh¹, Zhihao Hou¹, Xiao-Bin Liu¹, László Galgóczy¹³, László G. Nagy¹*

- Synthetic and Systems Biology Unit, Institute of Biochemistry, HUN-REN Biological Research Centre,, Temesvári krt. 62, Szeged H-6726, Hungary
- ² Doctoral School of Biology, Faculty of Science and Informatics, University of Szeged, Szeged H-6726, Hungary
- Department of Biotechnology, Faculty of Science and Informatics, University of Szeged, Szeged H-6726, Hungary
- * Corresponding author (Inagy@fungenomelab.com)

Contents:

- Supplementary figures 1-7.
- Legends for supplementary tables

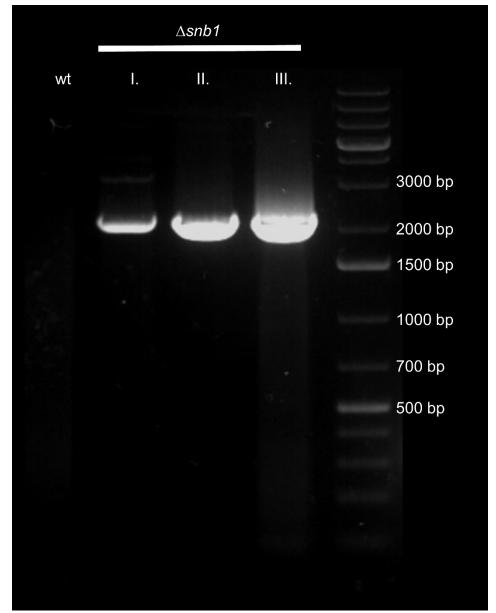


Fig. S1. - PCR analysis of snb1 knockout strains

For the primer pair snb1_out_check_fwd/pab1_inner_250bp_rev, the expected amplicon size if the repair template integrated into the Cas9 cleavage site is 2090bp. I., II. and III. are corresponding to the three obtained $\Delta snb1$ strains.

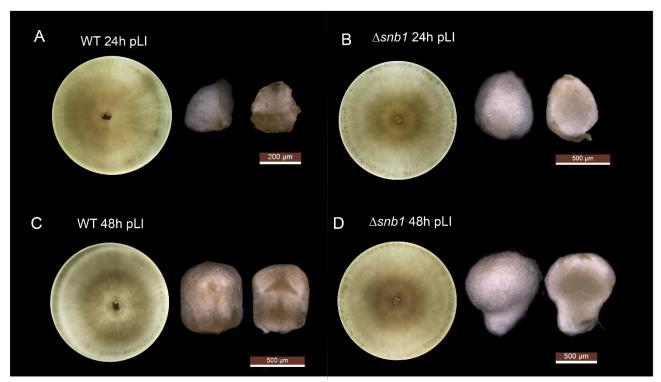


Fig. S2. - Time points sampled for RNA-Seq experiments showing the morphology of wt and \triangle snb1 at early developmental stages. Hyphal knots (**A**, **B**) and stage 1 primordia (**C**, **D**) after 24 hours and 48 hours of light induction, respectively. Scale bars are shown under the corresponding stages.

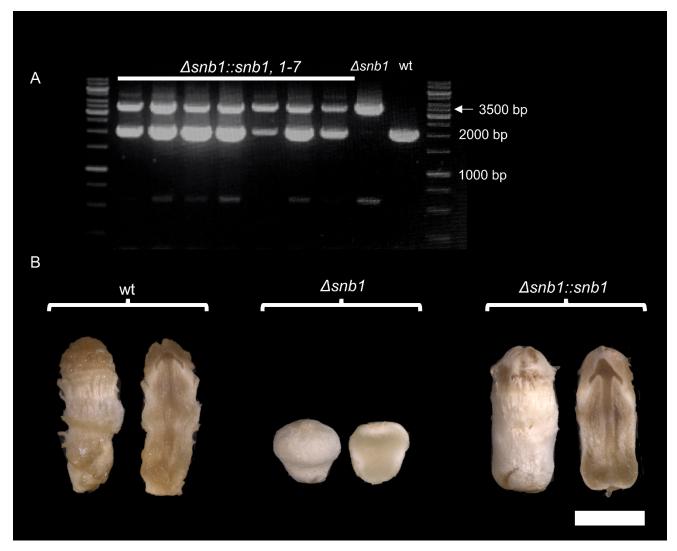


Fig. S3. - Complementation of Δsnb1 strain by the wild type gene. **A)** PCR validation of gene complementation using an inner primer pair of snb1 (snb1_inner_fwd/snb1_inner_rev). The expected size of Δsnb1 is larger due to the insertion of pab1 gene (expected amplicon size in the presence of the intact snb1 gene is 2122bp, expected amplicon size in the presence of disrupted snb1 gene is 3602bp). **B)** Cross sections of developing fruiting bodies of wt, Δsnb1, and complemented Δsnb1 C. cinerea strains three days post-light induction (pLI). Strains were grown on YMG medium with halved glucose content at 28°C. Scale bar = 2mm.

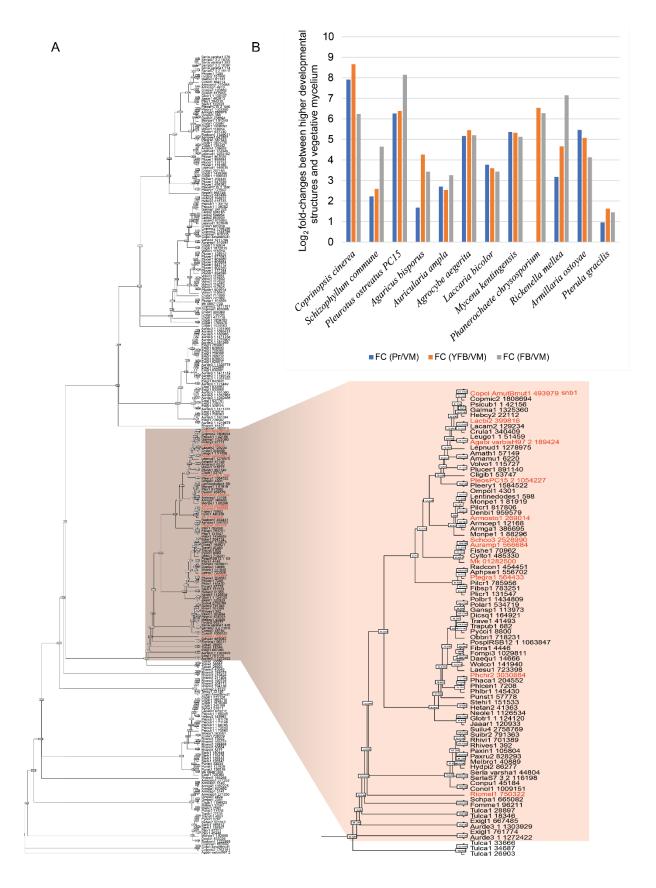


Fig. S4. - A) *Phylogenetic tree of SNB1 family.* Orthogroup with orthologs of SNB1 are highlighted with brownish background. Orthologs with red text are represented in Fig. S4/b. B) Log₂ fold-change differences in expression of *C. cinerea* SNB1 and its orthologues in further agaricomycetes species between vegetative

mycelia (VM) and different developmental stages of fruiting body (Pr: primordium, YFB: young fruiting body, FB: matured fruiting body)

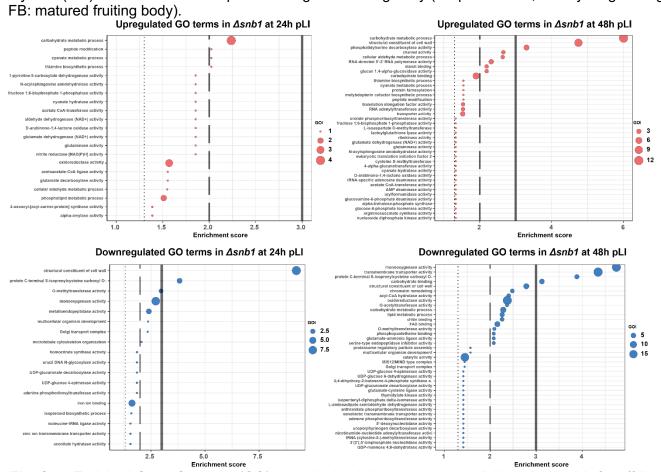


Fig. S5. - Enriched Gene Ontology (GO) terms in Δsnb1 compared to wt 24h and 48h pLI. Cutoff lines are drawn at enrichment scores corresponding to p=0.05, p=0.01, and p=0.001 (from left to right). GO terms are ordered by Kolmogorov–Smirnov p-values. See Table S3. for details on GO enrichment.

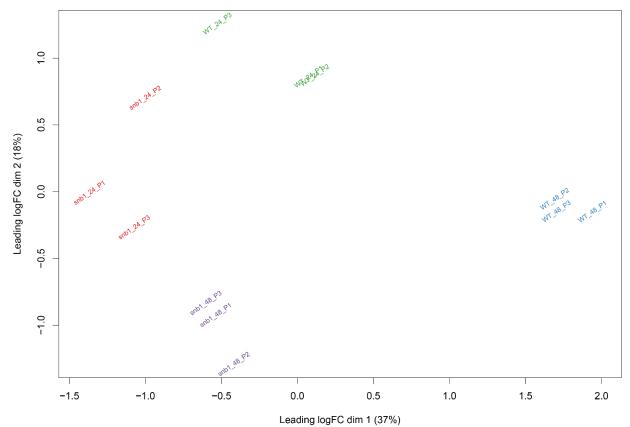


Fig. S6. - MDS-plot of the RNA-sequencing results.

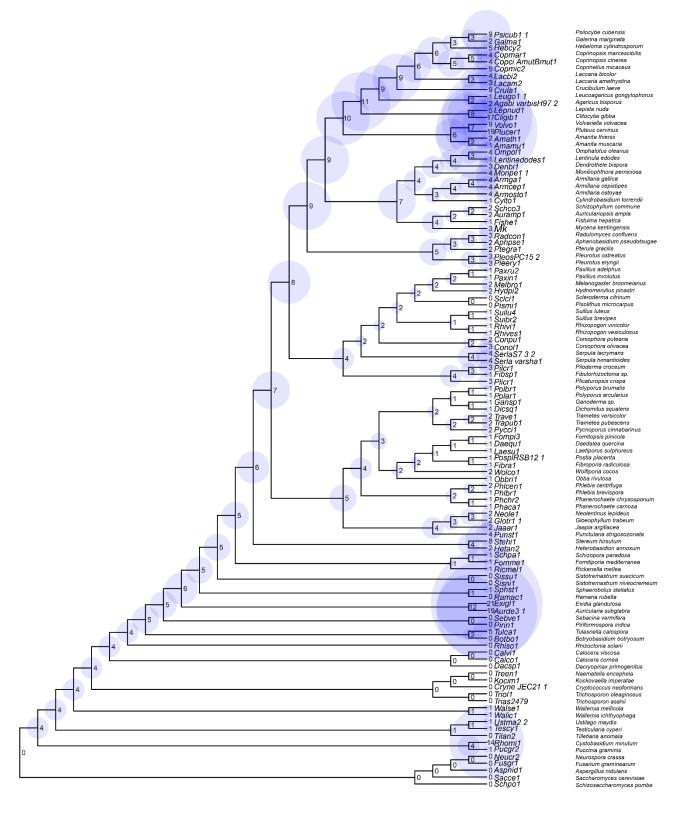


Fig. **S7.** - Inferred ancestral copy numbers of the SNB1 homologs in the proteome of 109 examined species. The size of the circles is proportional to the number of copy numbers.

Table contents:

- Table S1. List of primers used in this study.
- **Table S2.** All differentially expressed genes (DEGs) from the comparison of wild-type and $\Delta snb1$ mutant strains after 24h and 48h post-light induction (pLI).
- Table S3. Detailed list of enriched Gene Ontology (GO) terms in ∆snb1 compared to wt 24h and 48h pLI.
- **Table S4.** Mapping statistics for $\triangle snb1$ and wild type.