



## **Complete Genome Sequence of Sin4, a Siphophage Infecting Carbapenemase-Producing Klebsiella pneumoniae**

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**ABSTRACT** Klebsiella pneumoniae is a commonly antibiotic-resistant human pathogen. This report describes the complete genome sequence and important features of Sin4, a siphophage infecting carbapenemase-producing K. pneumoniae. By its genome size, predicted packaging mechanism, protein similarity, and classification given to its closest relatives, Sin4 was determined to be a T1-like phage.

Carbapenemase-producing K. pneumoniae is a pathogenic Gram-negative bacterium<br>within the Enterobacteriaceae family of microorganisms [\(1\)](#page-1-0). Sequence type 258 (ST258), a multidrug-resistant clade spreading in hospital settings around the world, was used here to isolate new bacteriophages from the environment [\(2\)](#page-1-1). We report here the complete genome sequence of the K. pneumoniae siphophage Sin4.

Bacteriophage Sin4 was isolated from a filtered (0.2- $\mu$ m pore size) sample collected at a wastewater treatment plant in College Station, TX, based on its ability to grow on a pKpQIL plasmid-cured derivative of K. pneumoniae strain 1776c [\(2\)](#page-1-1). The K. pneumoniae host was grown aerobically in tryptic soy broth or agar (Difco) at 37°C, and phage propagation was done using the soft agar overlay method [\(3\)](#page-1-2). Morphology was determined using transmission electron microscopy performed at the Texas A&M University Microscopy and Imaging Center on 2% (wt/vol) uranyl acetate-stained samples [\(4\)](#page-1-3). Phage genomic DNA was prepared by the shotgun library preparation protocol modification of the Promega Wizard DNA clean-up system [\(5\)](#page-1-4). Library preparation was done using a TruSeq Nano low-throughput kit, and sequencing occurred on an Illumina MiSeq platform with v2 500-cycle chemistry. There were 707,310 total paired-end 250-bp reads in the phage-containing index. The reads were quality controlled using FastQC [\(http://www.bioinformatics.babraham.ac.uk/projects/fastqc/\)](http://www.bioinformatics.babraham.ac.uk/projects/fastqc/). Af-ter trimming with the FASTX-Toolkit 0.0.14 [\(http://hannonlab.cshl.edu/fastx\\_toolkit/\)](http://hannonlab.cshl.edu/fastx_toolkit/), a single contig was assembled at 983-fold coverage using SPAdes v3.5.0 [\(6\)](#page-1-5). The genome was confirmed to be complete and accurate by Sanger sequencing of a PCR product off the contig ends (forward primer, 5'-CCGAAAGGCCTGGTATAGTT-3', and reverse primer, 5'-CAGTCTGCTTGTCGTTGATTTG-3'). The program PhageTerm predicts that Sin4 uses a headful packaging mechanism [\(7\)](#page-1-6). To identify Rho-independent terminators, the program TransTermHP v2.09 was used [\(8\)](#page-1-7). Protein-coding genes were predicted using Glimmer v3.0 and MetaGeneAnnotator v1.0 and corrected using tools available on the Center for Phage Technology Galaxy instance with Web Apollo [\(https://cpt.tamu.edu/](https://cpt.tamu.edu/galaxy-pub) [galaxy-pub\)](https://cpt.tamu.edu/galaxy-pub) [\(9](#page-1-8)[–](#page-1-9)[12\)](#page-1-10). According to an ARAGORN v2.36 scan, Sin4 does not contain tRNA genes [\(13\)](#page-1-11). The prediction of protein function was performed using primarily Inter-ProScan v5.22-61 and BLAST v2.2.31 with the NCBI nonredundant and UniProtKB Swiss-Prot/TrEMBL databases [\(14](#page-1-12)[–](#page-1-13)[16\)](#page-1-14). Additionally, TMHMM v2.0 for transmembrane domains and HHpred (multiple sequence alignment [MSA] generation with the HHblits ummiclus30\_2018\_08 database and modeling with PDB\_mmCIF70, HHSuite v3.0) predictions provided further support for the annotation [\(17,](#page-1-15) [18\)](#page-2-0). Unless otherwise stated, default parameters were used for all tools listed.

**Citation** Castillo M, Tran R, Newkirk H, Liu M, Gill JJ, Ramsey J. 2019. Complete genome sequence of Sin4, a siphophage infecting carbapenemase-producing Klebsiella pneumoniae. Microbiol Resour Announc 8:e01048-19. [https://doi.org/10.1128/MRA](https://doi.org/10.1128/MRA.01048-19) [.01048-19.](https://doi.org/10.1128/MRA.01048-19)

**Editor** Catherine Putonti, Loyola University Chicago

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**Received** 26 August 2019 **Accepted** 7 September 2019 **Published** 26 September 2019

Sin4 is a siphophage with a 49,916-bp genome, a coding density of 91.6%, and a G-C content of 50.3%. Of the 78 predicted genes in Sin4, 42 are not assigned a function. Sin4 is most closely related to Klebsiella phage 1513 (GenBank accession number [KP658157\)](https://www.ncbi.nlm.nih.gov/nuccore/KP658157), with which it shares 85.4% nucleotide sequence identity across the entire genome according to progressiveMauve v2.4.0 [\(19\)](#page-2-1). Phages 1513 and Sin4 also share 69 proteins.

By genome size, predicted packaging mechanism, and the classification given to its closest relatives, Sin4 is a T1-like phage with 42 proteins similar to phage T1 (GenBank accession number [NC\\_005833\)](https://www.ncbi.nlm.nih.gov/nuccore/NC_005833) [\(20\)](#page-2-2). Notable Sin4 genes include those for a DNA cytosine methyltransferase (NCBI accession number [QEG07100\)](https://www.ncbi.nlm.nih.gov/protein/QEG07100) and a DNA adenine methyltransferase (NCBI accession number [QEG07085\)](https://www.ncbi.nlm.nih.gov/protein/QEG07085).

**Data availability.** The genome sequence and associated data for phage Sin4 were deposited under GenBank accession number [MK931442,](https://www.ncbi.nlm.nih.gov/nuccore/MK931442) BioProject accession number [PRJNA222858,](https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA222858) SRA accession number [SRR8869237,](https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR8869237) and BioSample accession number [SAMN11360409.](https://www.ncbi.nlm.nih.gov/biosample/SAMN11360409)

## **ACKNOWLEDGMENTS**

This work was supported by funding from the National Science Foundation (awards EF-0949351 and DBI-1565146) and by the National Institutes of Health (NIAID award number AI121689). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Department of Biochemistry and Biophysics of Texas A&M University.

We thank Thomas Walsh of the Weill Cornell Medical School and Karen Frank of the National Institutes of Health for the provision of bacterial isolates. We are grateful for the advice and support of the CPT staff.

This announcement was prepared in partial fulfillment of the requirements for BICH464 Bacteriophage Genomics, an undergraduate course at Texas A&M University.

## <span id="page-1-0"></span>**REFERENCES**

- 1. Logan LK, Weinstein RA. 2017. The epidemiology of carbapenemresistant Enterobacteriaceae: the impact and evolution of a global menace. J Infect Dis 215:S28 –S36. [https://doi.org/10.1093/infdis/jiw282.](https://doi.org/10.1093/infdis/jiw282)
- <span id="page-1-1"></span>2. Satlin MJ, Chen L, Patel G, Gomez-Simmonds A, Weston G, Kim AC, Seo SK, Rosenthal ME, Sperber SJ, Jenkins SG, Hamula CL, Uhlemann A-C, Levi MH, Fries BC, Tang Y-W, Juretschko S, Rojtman AD, Hong T, Mathema B, Jacobs MR, Walsh TJ, Bonomo RA, Kreiswirth BN. 2017. Multicenter clinical and molecular epidemiological analysis of bacteremia due to carbapenem-resistant Enterobacteriaceae (CRE) in the CRE epicenter of the United States. Antimicrob Agents Chemother 61:e02349-16. [https://doi.org/10.1128/AAC.02349-16.](https://doi.org/10.1128/AAC.02349-16)
- <span id="page-1-3"></span><span id="page-1-2"></span>3. Adams MH. 1956. Bacteriophages. Interscience Publishers, Inc., New York, NY.
- <span id="page-1-4"></span>4. Valentine RC, Shapiro BM, Stadtman ER. 1968. Regulation of glutamine synthetase. XII. Electron microscopy of the enzyme from Escherichia coli. Biochemistry 7:2143–2152. [https://doi.org/10.1021/bi00846a017.](https://doi.org/10.1021/bi00846a017)
- <span id="page-1-5"></span>5. Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. Methods Mol Biol 502:27– 46. [https://doi.org/10.1007/978-1-60327-565-1\\_4.](https://doi.org/10.1007/978-1-60327-565-1_4)
- 6. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455– 477. [https://doi.org/10.1089/cmb.2012.0021.](https://doi.org/10.1089/cmb.2012.0021)
- <span id="page-1-6"></span>7. Garneau JR, Depardieu F, Fortier L-C, Bikard D, Monot M. 2017. PhageTerm: a tool for fast and accurate determination of phage termini and packaging mechanism using next-generation sequencing data. Sci Rep 7:8292. [https://doi.org/10.1038/s41598-017-07910-5.](https://doi.org/10.1038/s41598-017-07910-5)
- <span id="page-1-7"></span>8. Kingsford CL, Ayanbule K, Salzberg SL. 2007. Rapid, accurate, computational discovery of Rho-independent transcription terminators illuminates their relationship to DNA uptake. Genome Biol 8:R22. [https://doi](https://doi.org/10.1186/gb-2007-8-2-r22) [.org/10.1186/gb-2007-8-2-r22.](https://doi.org/10.1186/gb-2007-8-2-r22)
- <span id="page-1-8"></span>9. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved

microbial gene identification with GLIMMER. Nucleic Acids Res 27: 4636 – 4641. [https://doi.org/10.1093/nar/27.23.4636.](https://doi.org/10.1093/nar/27.23.4636)

- 10. Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. DNA Res 15:387–396. [https://doi.org/10.1093/dnares/dsn027.](https://doi.org/10.1093/dnares/dsn027)
- <span id="page-1-9"></span>11. Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Grüning BA, Guerler A, Hillman-Jackson J, Hiltemann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. Nucleic Acids Res 46:W537–W544. [https://doi.org/10.1093/nar/gky379.](https://doi.org/10.1093/nar/gky379)
- <span id="page-1-10"></span>12. Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elsik CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. Genome Biol 14:R93. [https://doi](https://doi.org/10.1186/gb-2013-14-8-r93) [.org/10.1186/gb-2013-14-8-r93.](https://doi.org/10.1186/gb-2013-14-8-r93)
- <span id="page-1-12"></span><span id="page-1-11"></span>13. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. [https://doi.org/10.1093/nar/gkh152.](https://doi.org/10.1093/nar/gkh152)
- 14. Jones P, Binns D, Chang H-Y, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A, Scheremetjew M, Yong S-Y, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. Bioinformatics 30: 1236 –1240. [https://doi.org/10.1093/bioinformatics/btu031.](https://doi.org/10.1093/bioinformatics/btu031)
- <span id="page-1-14"></span><span id="page-1-13"></span>15. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. [https://doi.org/10.1186/1471-2105-10-421.](https://doi.org/10.1186/1471-2105-10-421)
- <span id="page-1-15"></span>16. The UniProt Consortium. 2018. UniProt: the universal protein knowledgebase. Nucleic Acids Res 46:2699. [https://doi.org/10.1093/nar/](https://doi.org/10.1093/nar/gky092) [gky092.](https://doi.org/10.1093/nar/gky092)
- 17. Krogh A, Larsson B, Heijne von G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: appli-

cation to complete genomes. J Mol Biol 305:567–580. [https://doi.org/10](https://doi.org/10.1006/jmbi.2000.4315) [.1006/jmbi.2000.4315.](https://doi.org/10.1006/jmbi.2000.4315)

- <span id="page-2-0"></span>18. Zimmermann L, Stephens A, Nam S-Z, Rau D, Kübler J, Lozajic M, Gabler F, Söding J, Lupas AN, Alva V. 2018. A completely reimplemented MPI Bioinformatics Toolkit with a new HHpred server at its core. J Mol Biol 430:2237–2243. [https://doi.org/10.1016/j.jmb.2017.12.007.](https://doi.org/10.1016/j.jmb.2017.12.007)
- <span id="page-2-1"></span>19. Darling AE, Mau B, Perna NT. 2010. ProgressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5:e11147. [https://doi.org/10.1371/journal.pone.0011147.](https://doi.org/10.1371/journal.pone.0011147)
- <span id="page-2-2"></span>20. Roberts MD, Martin NL, Kropinski AM. 2004. The genome and proteome of coliphage T1. Virology 318:245–266. [https://doi.org/10.1016/j.virol](https://doi.org/10.1016/j.virol.2003.09.020) [.2003.09.020.](https://doi.org/10.1016/j.virol.2003.09.020)