

Genome Sequence of the 2,4,5-Trichlorophenoxyacetate-Degrading Bacterium *Burkholderia phenoliruptrix* Strain AC1100

Ping Xu,^{a,b} Hao Yu,^b Ananda M. Chakrabarty,^c Luying Xun^a

School of Molecular Biosciences, Washington State University, Pullman, Washington, USA^a; State Key Laboratory of Microbial Metabolism, Shanghai Jiao Tong University, Shanghai, China^b; Department of Physiology and Biophysics, University of Illinois College of Medicine, Chicago, Illinois, USA^c

Burkholderia phenoliruptrix strain AC1100 (ATCC 53867) degrades a variety of recalcitrant xenobiotics, including 2,4,5trichlorophenoxyacetate. The molecular mechanism of 2,4,5-trichlorophenoxyacetate degradation has been extensively studied. Here we present a 7.8-Mb assembly of the genome sequence of this 2,4,5-trichlorophenoxyacetate-degrading strain, which may provide useful information related to the degradation of chlorinated aromatic compounds.

Received 4 July 2013 Accepted 11 July 2013 Published 8 August 2013

Citation Xu P, Yu H, Chakrabarty AM, Xun L. 2013. Genome sequence of 2,4,5-trichlorophenoxyacetate-degrading bacterium *Burkholderia phenoliruptrix* strain AC1100. Genome Announc. 1(4):e00600-13. doi:10.1128/genomeA.00600-13.

Copyright © 2013 Xu et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Ping Xu, pingxu@sjtu.edu.cn, or Luying Xun, luying_xun@wsu.edu.

trains of the genus Burkholderia occupy a wide range of ecological niches and have versatile properties of bioremediation, biocontrol, and plant growth promotion (1). The herbicide 2,4,5trichlorophenoxyacetic acid (2,4,5-T), a suspected carcinogen, is a component of Agent Orange that was used in the Vietnam War and created long-term health problems (2, 3). Burkholderia phenoliruptrix AC1100 (ATCC 53867) (formerly Pseudomonas cepacia AC1100 and Burkholderia cepacia AC1100) was the first pure culture that used 2,4,5-trichlorophenoxyacetate as a sole source of carbon and energy (4-6). Three gene clusters, tftAB, tftCD, and tftEFGH, are involved in converting 2,4,5-trichlorophenoxyacetate to 3-oxoadipate in B. phenoliruptrix AC1100 (6-10). An IS element adjacent to the *tftAB* gene cluster provides the promoter for *tftAB*. The 2,4,5-trichlorophenoxyacetate-degrading ability is unstable due to the loss of the promoter created by the IS element (6, 11, 12). Thus, 2,4,5-trichlorophenoxyacetate is usually used as the sole carbon and energy source to cultivate B. phenoliruptrix AC1100 to preserve its ability for 2,4,5-trichlorophenoxyacetate degradation. Here, we present the genome sequence of B. phenoliruptrix AC1100, providing genomic contents for the biodegradation of 2,4,5-trichlorophenoxyacetate and other chlorinated aromatic compounds.

The genome sequence of *B. phenoliruptrix* AC1100 was obtained using the Illumina HiSeq 2000 system (100-bp paired-end sequencing). The reads were *de novo* assembled with Velvet software to 286 contigs (>500 bp), providing 44-fold coverage (13). The contig N_{50} was 81,158 bp, and the largest contig was 305,883 bp. Gene prediction and genome annotation were performed using of the RAST autoannotation server and NCBI PAPPC pipeline (14, 15). The tRNA genes were predicted using tRNAscan software (16). The gene function and classification were performed using the KEGG and Clusters of orthologous Groups (COG) databases (17).

The draft genome sequence of strain AC1100 comprises 7,811,030 bp, with a G+C content of 63.1%. There are 7,443 predicted protein coding sequences (CDS) (877 bp average length, 83.5% coding density). The genome of strain AC1100 has 1 rRNA operon and 52 tRNA loci. There are 493 subsystems represented in the genome sequence (2,844 CDS in total), and the metabolic network of AC1100 (determined by the RAST server) was reconstructed (14). We have predicted a rich set of genes (189 CDS) responsible for the degradation of aromatic compounds and 176 CDS for stress responses. Operons of tftAB, tftCD, and tftEFGH were found located on three contigs, and characteristics of the sequence are different from the core genome, indicating that they might belong to mobile regions of the genome (4-6). The genes for the degradation of catechol (catechol 1,2-dioxygenase), benzoate (benzoate 1,2-dioxygenase and benzoate-4-monooxygenase), toluene (toluene 4-monooxygenase), and the 3-oxoadipate pathway, homogentisate pathway, and central metacleavage pathway were predicted. The bacterium may have powerful degradation potentials for aromatic compounds. The genomic information of strain AC1100 will provide new insights into the genetic versatility of Burkholderia species and the metabolism of complex aromatic compounds.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ASXI00000000. The version described in this paper is the first version, ASXI01000000.

ACKNOWLEDGMENTS

The first author acknowledges financial support from the China Scholarship Council (2010831174) for visiting U.S. universities and a grant from the National Natural Science Foundation of China (31121064).

REFERENCES

- 1. Coenye T, Vandamme P. 2003. Diversity and significance of *Burkholderia* species occupying diverse ecological niches. Environ. Microbiol. 5:719–729.
- Kellogg ST, Chatterjee DK, Chakrabarty AM. 1981. Plasmid-assisted molecular breeding: new technique for enhanced biodegradation of persistent toxic chemicals. Science 214:1133–1135.
- Kilbane J, Chatterjee D, Karns J, Kellogg S, Chakrabarty A. 1982. Biodegradation of 2, 4, 5-trichlorophenoxyacetic acid by a pure culture of *Pseudomonas cepacia*. Appl. Environ. Microbiol. 44:72–78.

- Sangodkar UM, Chapman PJ, Chakrabarty AM. 1988. Cloning, physical mapping and expression of chromosomal genes specifying degradation of the herbicide 2,4,5-T by *Pseudomonas cepacia* AC1100. Gene 71:267–277.
- Haugland RA, Sangodkar UM, Sferra PR, Chakrabarty AM. 1991. Cloning and characterization of a chromosomal DNA region required for growth on 2,4,5-T by *Pseudomonas cepacia* AC1100. Gene 100:65–73.
- Hubner A, Danganan CE, Xun L, Chakrabarty AM, Hendrickson W. 1998. Genes for 2,4,5-trichlorophenoxyacetic acid metabolism in *Burkholderia cepacia* AC1100: characterization of the tftC and tftD genes and locations of the tft operons on multiple replicons. Appl. Environ. Microbiol. 64:2086–2093.
- Danganan CE, Ye RW, Daubaras DL, Xun L, Chakrabarty AM. 1994. Nucleotide sequence and functional analysis of the genes encoding 2,4,5trichlorophenoxyacetic acid oxygenase in *Pseudomonas cepacia* AC1100. Appl. Environ. Microbiol. 60:4100–4106.
- Daubaras DL, Hershberger CD, Kitano K, Chakrabarty AM. 1995. Sequence analysis of a gene cluster involved in metabolism of 2,4,5trichlorophenoxyacetic acid by *Burkholderia cepacia* AC1100. Appl. Environ. Microbiol. 61:1279–1289.
- Zaborina O, Daubaras DL, Zago A, Xun L, Saido K, Klem T, Nikolic D, Chakrabarty AM. 1998. Novel pathway for conversion of chlorohydroxyquinol to maleylacetate in *Burkholderia cepacia* AC1100. J. Bacteriol. 180: 4667–4675.
- 10. Gisi MR, Xun L. 2003. Characterization of chlorophenol 4-monooxygenase (TftD) and NADH: flavin adenine dinucleotide oxi-

doreductase (TftC) of *Burkholderia cepacia* AC1100. J. Bacteriol. 185: 2786-2792.

- Haugland RA, Sangodkar UM, Chakrabarty AM. 1990. Repeated sequences including RS1100 from *Pseudomonas cepacia* AC1100 function as IS elements. Mol. Gen. Genet. 220:222–228.
- 12. Hubner A, Hendrickson W. 1997. A fusion promoter created by a new insertion sequence, IS1490, activates transcription of 2,4,5-trichlorophenoxyacetic acid catabolic genes in *Burkholderia cepacia* AC1100. J. Bacteriol. **179**:2717–2723.
- 13. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829.
- 14. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. doi:10.1186/1471-2164-9-7 5.
- Pruitt KD, Tatusova T, Klimke W, Maglott DR. 2009. NCBI reference sequences: current status, policy and new initiatives. Nucleic Acids Res. 37:D32–D36. doi:10.1093/nar/gkn721.
- Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. Nucleic Acids Res. 33:W686–W689. doi:10.1093/nar/gki366.
- 17. Kanehisa M, Goto S. 2000. KEGG: Kyoto encyclopedia of genes and genomes. Nucleic Acids Res. 28:27–30.