

Research Article

Using pCO₂ Gap in the Differential Diagnosis of Hyperlactatemia Outside the Context of Sepsis: A Physiological Review and Case Series

Petr Waldauf,¹ Katerina Jiroutkova,^{1,2} and Frantisek Duska ^{1,2}

¹Department of Anaesthesia and Intensive Care Medicine, The Third Faculty of Medicine, Charles University and FNKV University Hospital, Prague, Czech Republic

²Oxylab: Lab of Mitochondrial Physiology, The Third Faculty of Medicine, Charles University, Prague, Czech Republic

Correspondence should be addressed to Frantisek Duska; frantisek.duska@lf3.cuni.cz

Received 5 May 2019; Accepted 17 October 2019; Published 4 December 2019

Academic Editor: Samuel A. Tisherman

Copyright © 2019 Petr Waldauf et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction. There is an inverse relationship between cardiac output and the central venous-arterial difference of partial pressures of carbon dioxide (pCO₂ gap), and pCO₂ gap has been used to guide early resuscitation of septic shock. It can be hypothesized that pCO₂ gap can be used outside the context of sepsis to distinguish type A and type B lactic acidosis and thereby avoid unnecessary fluid resuscitation in patients with high lactate, but without organ hypoperfusion. **Methods.** We performed a structured review of the literature enlightening the physiological background. Next, we retrospectively selected a series of case reports of nonseptic critically ill patients with elevated lactate, in whom both arterial and central venous blood gases were simultaneously measured and the diagnosis of either type A or type B hyperlactatemia was conclusively known. In these cases, we calculated venous-arterial CO₂ and O₂ content differences and pCO₂ gap. **Results.** Based on available physiological data, pCO₂ can be considered as an acceptable surrogate of venous-arterial CO₂ content difference, and it should better reflect cardiac output than central venous saturation or indices based on venous-arterial O₂ content difference. In our case report of nonseptic patients, we observed that if global hypoperfusion was present (i.e., in type A lactic acidosis), pCO₂ gap was elevated (>1 kPa), whilst in the absence of it (i.e., in type B lactic acidosis), pCO₂ gap was low (<0.5 kPa). **Conclusion.** Physiological rationale and a small case series are consistent with the hypothesis that low pCO₂ gap in nonseptic critically ill is suggestive of the absence of tissue hypoperfusion, mandating the search for the cause of type B lactic acidosis rather than administration of fluids or other drugs aimed at increasing cardiac output.

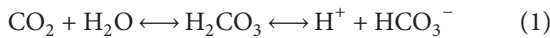
1. Introduction

Differential diagnosis of elevated blood lactate is the daily bread and butter of all clinicians looking after critically ill patients. In principle, hyperlactatemia can either be caused by increased production in tissues (type A) or impaired lactate uptake (type B) or by these two mechanisms combined [1, 2]. Correctly determining the cause(s) of hyperlactatemia is of utmost importance, indeed, because it determines the treatment, which can be life saving for a patient with one underlying cause, but harmful for another. A classical example of this concept is fluid resuscitation that can be helpful in correcting tissue hypoperfusion, but

harmful for patients with other causes of elevated lactate. Nice as it sounds, recognizing patients that would benefit from fluid administration (or other ways of increasing cardiac output) is often very difficult at the bedside and often results in fluid abuse, particularly in patients with elevated lactate and on vasopressors [3]. It was recognized in sepsis that pCO₂ gap (or its mathematical derivatives) outperformed other markers in detecting tissue hypoperfusion [4–7]. We hypothesize that the difference in carbon dioxide partial pressure between central venous and arterial blood (pCO₂ gap) can be a useful aid in the differential diagnosis of elevated lactate also outside the context of sepsis. In this paper, we present the theoretical

rationale for this hypothesis and review published data and confront them with observations of $p\text{CO}_2$ gaps in a case series of nonseptic patients with known cause of lactate elevation.

1.1. Physiological Background. During the process of cellular respiration, carbon dioxide is produced mostly by decarboxylation reactions in citric acid cycle and diffuses into the bloodstream through the extracellular fluid. Whilst approximately 10% of the carbon dioxide (CO_2) in the blood remains dissolved in the plasma, the remaining CO_2 diffuses rapidly into the red blood cells, where it is either bound to terminal NH_2 groups (30%) forming carbaminohaemoglobin or reacts with water to form carbonic acid (60%) that dissociates to bicarbonate and a proton (H^+) [8]:



The production of HCO_3^- occurs rapidly because of catalysis by carbonic anhydrase. The HCO_3^- then leaves the red blood cells in exchange of Cl^- (the process known as chloride or Hamburger shift), thereby promoting the entry of more CO_2 into the red blood cell. In the bloodstream, CO_2 in all 3 forms is conveyed back to the respiratory surfaces at a rate, which is directly proportional to cardiac output.

Cardiac output (CO) calculated using the Fick principle applied to CO_2 agrees well with cardiac output calculated from O_2 -derived parameters in normal subjects at rest and during exercise [9]. The Fick equation (for indirect CO calculation) applied to CO_2 is

$$\text{CO} = \frac{\text{VCO}_2}{\text{VctCO}_2(\text{B}) - \text{ActCO}_2(\text{B})}, \quad (2)$$

where VCO_2 is the CO_2 production, CO is the cardiac output, and $\text{VctCO}_2 - \text{ActCO}_2$ is the venous-to-arterial CO_2 content difference. After substituting ctCO_2 gap into the abovementioned equation, we obtain

$$\text{ctCO}_2(\text{B})\text{gap} = \frac{\text{VCO}_2}{\text{CO}}, \quad (3)$$

where the $\text{ctCO}_2(\text{B})$ gap is inversely related to CO and proportional to VCO_2 . The value for ctCO_2 is not directly measured; instead, it is calculated from measured pH and $p\text{CO}_2$, which is a mathematically complex and error-prone process, whereas $p\text{CO}_2$ is a directly measured parameter that is readily available to clinicians. In this paper, we will determine the $p\text{CO}_2$ gap by subtracting the peripheral arterial $p\text{CO}_2$ from the central venous $p\text{CO}_2$. This is because central venous $p\text{CO}_2$ has been shown to be a reliable substitute for mixed-venous $p\text{CO}_2$ [10], and pulmonary artery is rarely catheterized in contemporary ICU practice because of the invasiveness of the procedure [11].

Over the physiologic range of $p\text{CO}_2$, the relationship between $p\text{CO}_2$ and the total blood CO_2 content is close to linear, so $p\text{CO}_2$ may be considered a reliable substitute for CO_2 content [12, 13]. Factors that disturb the linearity between $p\text{CO}_2$ and CTCO_2 tend to offset each other for a

given CtCO_2 , and $p\text{CO}_2$ is higher in metabolic acidosis and lower for lower saturation of haemoglobin with oxygen (see Figure 1). During low flow states, organic acids (mainly lactate) are released from hypoperfused tissues, causing base excess of venous blood to be more negative as compared with the arterial blood. On the other hand, venous blood leaving the hypoperfused tissue tend to be more deoxygenated, causing $p\text{CO}_2$ to be lower for a given CtCO_2 (Haldane effect [14]). In turn, it can be hypothesized that A-V $p\text{CO}_2$ gap is representative of CTCO_2 gap under a wide range of clinical situations.

In turn, when $p\text{CO}_2$ replaces CtCO_2 in equation (3), we get

$$(\text{PvCO}_2 - \text{PaCO}_2) = p\text{CO}_2(\text{B})\text{gap} = \frac{\text{VCO}_2}{\text{CO}} * k, \quad (4)$$

where $(\text{PvCO}_2 - \text{PaCO}_2)$ is the venous-to-arterial PCO_2 difference and k is the PCO_2 to CTCO_2 correlation (assumed to be constant). In line, both CTCO_2 and $p\text{CO}_2$ gaps were found to increase with the decrease in CO, and the relationship follows a hyperbolic pattern (see Figure 2) [15, 16].

Moreover, elevated CO_2 gap can be considered as the marker of the cardiac output in relation of peripheral metabolic requirements. Hypoxic hypoxia in the presence of adequate cardiac output cannot cause $p\text{CO}_2$ gap elevation as demonstrated by elegant experiments of Vallet et al. [17]. On the other hand, if there is an inhomogeneity of distribution of perfusion as it is the case in sepsis, the $p\text{CO}_2$ gap is more sensitive than ScvO_2 drop in detecting patients who would benefit from measures aimed at increasing cardiac output [4–6, 18–20]. The cut-off value of $p\text{CO}_2$ gap in a septic patient was found to be 0.8 kPa (6 mmHg) [20, 21]. The superiority of $p\text{CO}_2$ gap over ScvO_2 desaturation in detecting hypoperfusion likely reflects the fact that in the presence of microvascular shunting, central venous blood is a mixture of arterialized blood from shunts and desaturated blood from the hypoperfused regions. Because of 20 times higher diffusibility of CO_2 as compared with O_2 [22, 23] central venous blood can have normal or high ScvO_2 (due to arterialized blood from shunts), whilst CO_2 content is elevated proportionally to the degree of peripheral tissue hypoperfusion as shunting capillaries are still capable to drain CO_2 from the hypoperfused tissues. It has been proposed (L. Gattinoni–personal communication) that local tissue acidosis caused by hypoperfusion releases free CO_2 from bicarbonate, thereby increasing venous $p\text{CO}_2$ even further, a phenomenon called “Coca Cola effect” in analogy to releasing bubbles by adding piece of lemon into a carbonated drink. In addition, some CO_2 can be produced anaerobically [18].

Tissue metabolism also influences the respiratory exchange ratio (RER), i.e., the amount of CO_2 produced per each mole of O_2 consumed. Oxidation of lipids releases less CO_2 (0.7 moles) than oxidation of amino acids (0.84 moles) or carbohydrates (1.0 mole) for 1 mole of consumed O_2 . After reaching anaerobic threshold, RER increases well above 1.0. In analogy, anaerobic metabolism in peripheral tissues can be detected by calculating a surrogate of tissue RER:

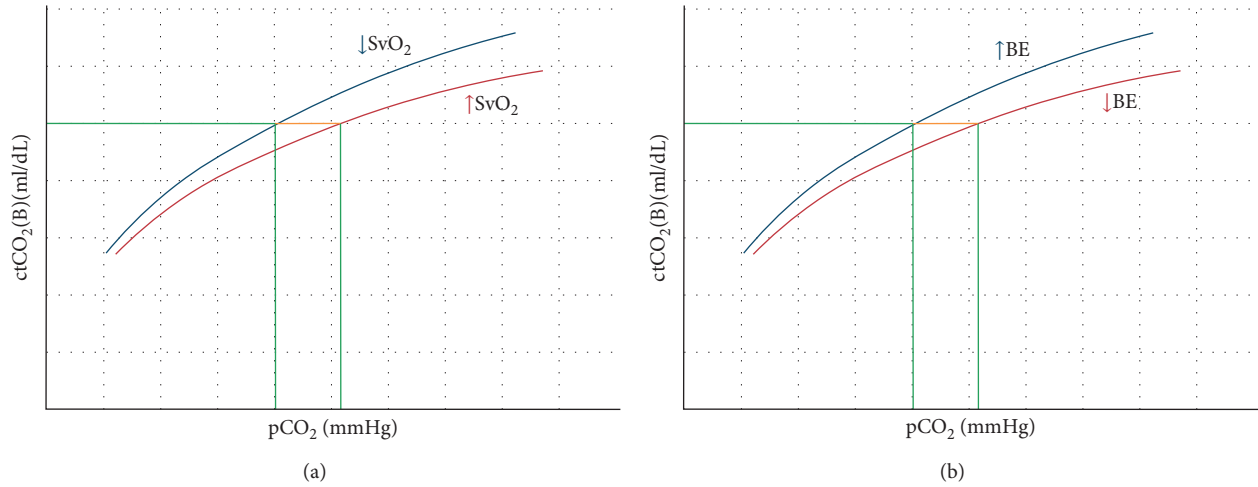


FIGURE 1: The relationship between partial pressure of CO₂ (pCO₂) and whole-blood CO₂ content (ctCO₂) and the influence of saturation of venous blood with oxygen (SvO₂, left) (a) and base excess (BE, right) (b), respectively.

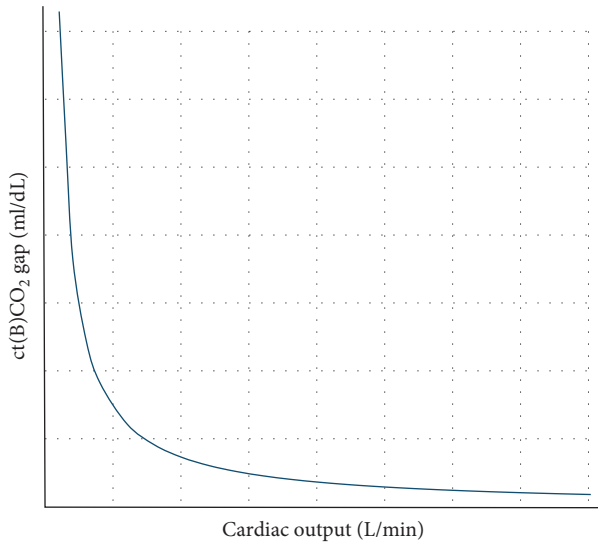


FIGURE 2: Relationship between cardiac output and central venous-to-arterial difference of CO₂ content in blood (ct(B)CO₂ gap).

$$RER_{sur} = \frac{pCO_2 \text{ gap}}{Act(B)O_2 - Vct(B)O_2} \quad (5)$$

The value of $RER_{sur} > 1.4$ (when pCO₂ gap is in (mmHg) and ctO₂ in (ml/dL)) was found to be associated with the presence of tissue hypoxia [24]. This is the analogy of the increase of whole-body respiratory quotient when a person exercising on a treadmill overcomes the aerobic threshold. Indeed, RER can be elevated even with normal pCO₂ gap, if venoarterial difference of oxygen content is very low, as could be the case when significant left-to-right shunting (e.g., at microcirculation level) is accompanying tissue ischaemia.

To summarize, unlike oxygen in the opposite direction, CO₂ is able to reach the bloodstream regardless of the status of microcirculation. According to the Fick principle, mixed

venous-to-arterial CtCO₂ gap is inversely related to cardiac output and central venous-to-arterial pCO₂ gap seems to be its acceptable surrogate. In the light of this, it can be hypothesized that even outside the context of sepsis, pCO₂ gap can be a useful aid in the differential diagnosis of lactic acidosis. In particular, elevated pCO₂ gap can identify patients who would benefit from fluids and/or other measures aimed at increasing cardiac output. In order to support this hypothesis, we present a series of patients in whom pCO₂ gap was measured, and the cause of lactic acidosis was conclusively known.

2. Methods

In a clinical information system (MetaVision ver. 6, IMD Soft, Israel) that contains data of 5251 patients admitted to 22 bed ICU of the Department of Anaesthesia and Intensive Care of FNKV University Hospital since 2012, we retrospectively searched for patients who had central venous and arterial blood gases measured simultaneously (within 2 min) and also had elevated lactate within 24 hours of paired blood gas measurement. From the list of patients fulfilling these criteria, we selected those where 2 clinicians independently agreed on lactic acidosis being either type A or B, and the diagnosis was either beyond all doubts (e.g., a young fit trauma victim with active bleeding and hemorrhagic shock) or supported by additional evidence (e.g., findings at autopsy). All the rest of the cases were labeled as undetermined.

In those cases, we calculated arterial and venous blood CO₂ content as

$$ctCO_2(B) = 9.286 * 10^{-3} * pCO_2 * ctHb * \left[1 + 10^{(1+10^{(pH_{ERY} - pK_{ERY}))})} \right] + ctCO_2(P) * \left(\frac{ctHb}{21} \right), \quad (6)$$

where

$$\begin{aligned}
 \text{ctCO}_2(P) &= 0.23 * \text{pCO}_2 + \text{cHCO}_3^-(P) \\
 \text{cHCO}_3^-(P) &= 0.23 * \text{pCO}_2 * 10^{(\text{pH}-\text{pK}_p)}, \\
 \text{pK}_p &= 6.125 - \log(1 + 10^{(\text{pH}-8.7)}), \\
 \text{pH}_{\text{ERY}} &= 7.19 + 0.77 * (\text{pH} - 7.4) + 0.035 * (1 - \text{sO}_2), \\
 \text{pK}_{\text{ERY}} &= 6.125 - \log\left(1 + 10^{(\text{pH}_{\text{ERY}}-7.84-0.06*\text{sO}_2)}\right).
 \end{aligned}
 \tag{7}$$

These equations were from the manual of ABL-800 blood gas machine (Radiometer, Denmark), and indices “ERY” indicate erythrocyte (red blood cell)/.

Oxygen content in arterial blood was calculated as

$$\begin{aligned}
 \text{Act(B)O}_2\left(\frac{\text{mL}}{\text{dL}}\right) &= \left[1.34 * \text{Hb}\left(\frac{\text{g}}{\text{dL}}\right) * \text{SaO}_2 * 0.01\right] \\
 &+ [0.0225 * \text{PaO}_2 \text{ (kPa)}]
 \end{aligned}
 \tag{8}$$

Oxygen content in venous blood was correspondingly calculated as

$$\begin{aligned}
 \text{Vct(B)O}_2\left(\frac{\text{mL}}{\text{dL}}\right) &= \left[1.34 * \text{Hb}\left(\frac{\text{g}}{\text{dL}}\right) * \text{SvO}_2 * 0.01\right] \\
 &+ [0.0225 * \text{PvO}_2 \text{ (kPa)}]
 \end{aligned}
 \tag{9}$$

The study was performed in accordance with the Declaration of Helsinki. Because of retrospective and epidemiological nature of the study, informed consent was not required.

3. Results

Out of all 5,251 patients in the database, we have found 23 cases with nonseptic patients with elevated lactate and both arterial and venous gases measured. Out of these, there were 6 cases where the diagnosis of either type A or B was beyond any reasonable doubt as independently agreed by 2 clinicians.

3.1. Case Series

3.1.1. Example Case 1: Type A Lactacidaemia due to Global Hypoperfusion. A 40-year-old male attempted suicide by jumping out of 3rd floor window. He was intubated at scene, brought in, and diagnosed complex pelvic fracture and compressive fracture of L3. Parenchymatous organs were without signs of injury. After volume resuscitation and blood transfusions, he was haemodynamically stable and remained sedated with plan to operate fractures the next day. In very early hours of the next morning, he suddenly developed signs of haemorrhagic shock with tachycardia 150/min, haemoglobin drop from 129 to 92 g/L, and hypotension with an increase of noradrenalin dose from 0.4 to 1.1 $\mu\text{g}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$. Lactate at this stage was 1.4 mM (only 90 min later increasing to 3.4 mM), ScvO₂ 68%, pCO₂ gap was 1.02 kPa, and RER 1.86. CT scan was repeated and showed a haemoperitoneum and R-sided haemothorax due

to right-sided diaphragmatic injury that included rupture and bleeding from the teres hepatis ligament. He was classified as having type A lactic acidosis due to haemorrhagic shock.

3.1.2. Example Case 2: Type A Lactacidaemia due to Local Ischaemia. A 53-year-old female, previously fit and well, underwent Whipple’s pancreatoduodenectomy due to pancreatic tumour. The operation and the immediate postoperative course were uneventful. She was extubated and haemodynamically stable, passing urine and not requiring vasopressors. Three hours after surgery, she developed severe abdominal pain despite functional epidural analgesia. Her lactate increased to 5.3 mM, ScvO₂ was 71%, pCO₂ gap 1.06 kPa, and RER 1.58. She was diagnosed small bowel ischaemia due to occlusion of superior mesenteric artery on angio-CT and underwent relaparotomy and an aortomesenteric bypass operation. She was classified as type A lactic acidosis based on perioperative finding of ischaemic bowel.

3.1.3. Example Case 3: Type A Lactacidaemia due to Combination of Global and Local Hypoperfusion. A 25-year-old man, previously fit and well, completely transected his brachial artery by a broken glass. He suffered massive blood loss and was found in profound shock by emergency services. He was brought in after having received 2 L of crystalloids and 1 g of tranexamic acid. This patient with evident tissue hypoperfusion due to acute blood loss, with lactate 7.2 mM and ScvO₂ 58%, had pCO₂ gap 1.38 kPa (10 mmHg) and RER of 2.5. In addition, CT scan performed at admission revealed a nonocclusive ischaemia and necrosis of the small bowel that required laparotomy and bowel resection. He was classified as type A lactic acidosis based on perioperative finding of ischaemic bowel.

3.1.4. Example Case 4: Type B Lactacidaemia due to Metformin Overdose. A 71-year-old female with type 2 diabetes was admitted after suicidal attempt committed due to metformin overdose. On admission, she was unconscious, intubated and ventilated, haemodynamically unstable (noradrenaline dose 0.53 $\mu\text{g}\cdot\text{kg}^{-1}\cdot\text{min}^{-1}$), and in profound acidosis (pH 6.58), with lactate 17 mmol/L, SvO₂ was 78.7%, pCO₂ gap 0.37 kPa (2.8 mmHg), and RER 1.14. She was classified as type B lactic acidosis due to metformin overdose [25] by consensus of clinicians.

3.1.5. Example Case 5: Type B Lactacidaemia due to High Dose Steroids and Betamimetics. A 66-year-old obese female with a history of asthma was hit by a car whilst crossing the road. At scene, she was confused and complaining of shortness of breath. She was intubated and air-lifted to the trauma centre. She was stable and found to have no injuries on the CT scan, and the decision was made to wake her up and extubate. Her pre-extubation lactate was 1.7 mM. Immediately after extubation, she developed severe bronchospasm, which did not respond to inhalatory betamimetics

TABLE 1: Case series of patients with different causes of elevated lactate.

Case	Lac (a) (mmol/L)	SaO ₂ (%)	ScvO ₂ (%)	pCO ₂ gap (kPa)	RER	Type of lactic acidosis and timing of diagnosis	Evidence for classifying hyperlactataemia as either A or B
#1	3.4	99	68	1.02	1.86	A—a posteriori	Peroperative finding
#2	5.3	99	71	1.06	1.58	A—a posteriori	Peroperative finding
#3	7.2	100	58	1.38	2.5	A—a priori	Consensus of clinicians
#4	17.0	99	79	0.37	1.14	B—a priori	Consensus of clinicians
#5	7.4	94	76	0.30	0.78	B—a posteriori	Consensus of clinicians
#6	12.8	99	72	0.30	0.71	B—a posteriori	Consensus of clinicians

Note: Lac a = lactate (arterial); RER = respiratory exchange ratio calculated as per equation (5).

(salbutamol 5 + 5 mg), IV steroids (methylprednisolone 125 mg), and mandated reintubation. Oxygenation was maintained above 93% throughout. Lactate 30 min after intubation, whilst she was sedated and still paralysed, was 7.4 mM, ScvO₂ 76%, pCO₂ gap 0.3 kPa, and RER 0.78. Steroids were discontinued for the next few hours, lactate decreased again, and the lady was successfully extubated. She was classified as posteriori by consensus of clinicians as having type B lactic acidosis due to a combined effect of betamimetics [26] and steroids [27].

3.1.6. Example Case 6: Type B Lactacidaemia in Patient with Advanced Multiple Myeloma. A 75-year-old male with known advanced multiple myeloma was referred for sudden-onset paraplegia. He was found to have T8 fracture and underwent an urgent decompressive spinal surgery. Perioperatively, he also received spinal dose steroids. Blood loss was estimated to 1.6L and he required perioperative transfusion of blood products, but remained haemodynamically stable. Perioperative blood gas showed lactate 12.8 mM, ScvO₂ 72%, pCO₂ gap 0.3 kPa, and RER 0.71. He was referred to ICU after operation. Despite persistent elevation of lactate, he remained stable and was extubated. He was discharged to ward after 5 days, in much improved condition but with lactate still in range of 5–10 mM, a phenomenon that has been described in patients with multiple myeloma [28–30]. He was diagnosed by a posteriori consensus of clinicians as having type B lactic acidosis due to multiple myeloma.

Data from cases of type A and B lactic acidosis are described in Table 1.

3.2. Differences between Patients with Type A and B Lactacidaemia. As summarised in Table 1, all patients with typical type A lactic acidosis had pCO₂ gap well above 0.8 kPa (6 mmHg) including patients who had normal ScvO₂. On the contrary, all patients with type B lactic acidosis had pCO₂ gap around 0.4 kPa (3 mmHg). Similar or even better discrimination is obtained when RER is used (Table 1). We found a close correlation between ctCO₂ gap and pCO₂ gap ($R^2 = 0.71$ data not shown), but not so close correlation between ctCO₂ gap and ctO₂ gap ($R^2 = 0.54$, data not shown). Venoarterial lactate difference [31, 32] was in range of -0.3 to +0.2 mM and we have not found any difference between patients with type A and B lactacidaemia.

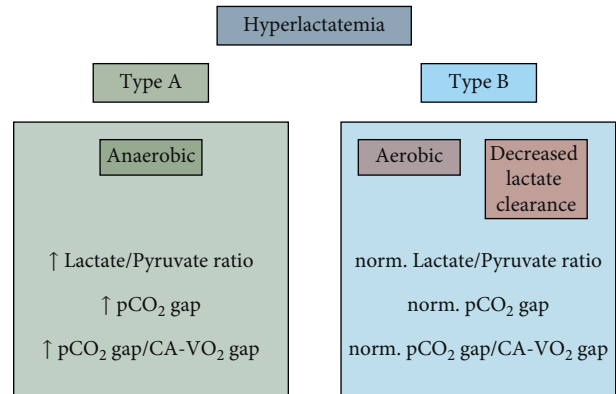


FIGURE 3: Main physiological features of type A and B hyperlactataemias.

4. Discussion

The presented review of physiology and the small case series generate the hypothesis that the use of venous-arterial pCO₂ gap can be a useful aid in the differential diagnosis of hyperlactataemia. Type A lactic acidosis caused by global or local tissue hypoperfusion with a switch to anaerobic metabolism, whilst type B lactic acidosis is mostly characterized by the absence of tissue hypoxia, and the cause of elevated lactate is either aerobic production or decreased lactate clearance (Figure 3).

Type B lactic acidosis is seen in the heterogeneous group of diseases that are of common occurrence in a typical general ICU [33]. Aerobic lactate production can occur as a paraneoplastic phenomenon (by Warburg effect [34, 35]) or more often as the result of β -adrenergic receptor stimulation either during a sympathetic surge (e.g., acute severe asthma, subarachnoidal haemorrhage, and pheochromocytoma) or after administration of β -receptor agonists (typically β_2 -mimetics or adrenaline). Decreased lactate clearance is seen in metformin or ethanol poisoning or in liver failure. Steroids cause hyperlactaemia [36] by a combination of increased aerobic lactate production and decreased lactate clearance [27]. Indeed, many patients would have a combination of type A and B causes of lactic acidosis (e.g., those with septic shock [37, 38], but also many others [39]) (Table 2).

Because the principal difference of type A and B of hyperlactataemia is the presence or absence of tissue hypoxia, the ratio of lactate/pyruvate has been proposed to aid

TABLE 2: Overview of causes of elevated lactate.

Group	Mechanism	Condition/disease	Expected finding	
A (increased lactate production)	Severe hypoxia	Any cause (pO ₂ < 4 kPa)	High pCO ₂ gap fluids and ↑ cardiac output are likely to help	
	Low O ₂ transport capacity	CO poisoning Severe anaemia Low preload (hypovolaemia)		
	Low global oxygen delivery leading to excessive anaerobic glycolysis	Low cardiac output = hypodynamic shock		
	Low cardiac output = hypodynamic shock	Low contractility (cardiogenic) High afterload (obstructive)		
	Local ischaemia leading to excessive anaerobic glycolysis	Normal or high cardiac output, but demand even higher	Strenuous exercise Shivering or seizures	Fluids and ↑ cardiac output may or may not help
		Inflow occlusion	Limb ischaemia Mesenteric ischaemia	
		Decreased perfusion pressure	Compartment syndromes	
		Increased glycolysis in the presence of enough oxygen	Local ischaemia (Wartburg effect)	Cancer
			Stimulation of muscle and liver glycogenolysis	Beta-2-mimetics Adrenalin (exogenous or excessive stress) Electrical muscle stimulation [1] Cocaine Theophylline Metformin
				Blocked oxidative phosphorylation (cytopathic hypoxia)
B (decreased lactate uptake)	Decreased lactate uptake	Production of L- and D-lactate by colon bacteria	Short bowel +	
		Liver failure	Acute liver failure Liver ischaemia	
		Failed conversion of pyruvate to AcCoA	Thiamine deficiency	
		Failed conversion of lactate to pyruvate	Alcohol intoxication	
Mixed	Sepsis	Element of hypoxia, aerobic glycolysis, and splanchnic ischaemia	Complex condition	
	Propylen glycol poisoning	Mix of D- and L-lactate overproduction and element of oxidative phosphorylation block		

the differential diagnosis of elevated lactate. Due to the ubiquitous presence of cytosolic lactate dehydrogenase lactate/pyruvate ratio reflects the redox situation of cells, i.e., [NADH]/[NAD⁺] ratio, tissue hypoxia would lead to a block of reoxidation of NADH back to NAD⁺. Accumulated NADH would cause leftward shift of equilibrium shown in the following equation:



Hence, in the presence of any cessation of electron transfer chain (of which by far the most common cause is hypoxia), the lactate/pyruvate ratio increases above normal value of 10:1, which is considered a hallmark of type A lactic acidosis [33, 39] In line, type B lactic acidosis would have

both lactate and pyruvate increased, with the ration 1:10 remaining constant. Nonetheless, the lactate/pyruvate ratio remains of very limited clinical use because currently it is impossible to measure pyruvate concentration by point-of-care techniques, and its laboratory assay is complex and prone to preanalytical errors [39]. Despite technical complexity, the study of lactate/pyruvate ration in critically ill patients with elevated lactate demonstrated that a large proportion of them do not have the biochemical signs of tissue hypoxia and have in general a better prognosis. Indeed, those patients are unlikely to benefit from measures aimed to increase systemic oxygen delivery and more fluids or inotropes may actually cause harm. Unnecessary fluid loading is performed very often in the critically ill (as

evidenced in FENICE study [3]), and we can only speculate how often this is triggered by elevated lactate being misinterpreted as a marker of tissue hypoperfusion. Anecdotally, ordering of fluid boluses as automatic response to elevated lactate has been termed “fluid reflex”. We suggest that the role of pCO₂ gap in detecting tissue hypoperfusion is further studied as a possible marker of tissue hypoperfusion in situations of elevated lactate outside the context of sepsis. We demonstrate in our series of case reports that, in pCO₂ gap reflected ctCO₂ gap and was elevated (>1 kPa) in patients with global or local hypoxia, but remained low (<0.5 kPa) in patients with type B lactic acidosis, perhaps due in part to the hyperkinetic circulation that often accompanies diseases associated with type B hyperlactataemia. The pCO₂ gap cut-off of 0.8 kPa, used in studies on early-goal-directed therapy in septic shock [18, 19] seems to be applicable to our nonseptic cases, too, but indeed, the cut-off with best sensitivity and specificity remains to be determined. Unlike ScvO₂, the use of pCO₂ gap is less distorted by the presence of microcirculatory shunts, and hence it should have higher sensitivity to detect tissue hypoperfusion.

Nonetheless, our small observational case series is by no means the proof of the hypothesis outlined above. The next step would be a prospective trial looking at lactate dynamics after a fluid bolus in relation to pCO₂ gap at baseline in patients with elevated lactate and with cardiac output continuously monitored. Also, much larger series of patients is needed to determine what is the proportion of hyperlactatemic patients in which pCO₂ gap might be helpful as compared with those with mixed type A+B hyperlactatemias. Lastly, it remains unclear how specific and sensitive pCO₂ gap is in detecting organ hypoperfusion in patients with mixed etiologies of lactic acidosis.

In conclusion, in this paper, we present a hypothesis that venous-arterial pCO₂ gap may be a useful aid in the differential diagnosis of elevated lactate in critically ill patients and that it has potential to avoid administration of unnecessary fluids and ionotropics in patients, who have lactate elevated in the absence of tissue hypoperfusion. We demonstrate, for the first time in the literature, that pCO₂ gap is elevated in nonseptic patients with type A lactic acidosis and normal in type B lactic acidosis.

Abbreviations

Act(B)O ₂ :	Content of oxygen in whole arterial blood
CaCO ₂ :	Total carbon dioxide content in whole arterial blood
Central venous blood:	Sampled from a central venous catheter (inserted into vena cava superior)
CO:	Cardiac output
ct(B)CO ₂ gap:	Arterial-to-venous carbon dioxide content difference
ct(B)O ₂ gap:	Arterial-to-venous oxygen content difference
CvCO ₂ :	Total carbon dioxide content in whole venous blood
DO ₂ :	Oxygen delivery
EGDT:	Early goal-directed therapy

Hb:	Haemoglobin
MALA:	Metformin-associated lactic acidosis
Mixed venous blood:	Sampled from a pulmonary artery catheter
pCO ₂ gap:	Central venous-to-arterial carbon dioxide partial pressure difference (in the literature ΔPCO ₂ is also used)
RQ:	Respiratory quotient
S _v O ₂ :	Central venous oxygen saturation
Vct(B)O ₂ :	Content of oxygen in whole venous blood.

Data Availability

The deidentified patient’s data used to support the findings of this study are *in extenso* provided in the manuscript.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors’ Contributions

PW and KJ performed the literature review and PW and FD collected and analysed the patients’ data. All authors contributed to writing the paper and read and approved its final version.

Acknowledgments

The work was supported by Charles University grant Q37 and by Institutional support of FNKV University Hospital.

References

- [1] R. D. Cohen and H. F. Woods, *Clinical and Biochemical Aspects of Lactic Acidosis*, Blackwell Scientific Publications, Hoboken, NJ, USA, 1976.
- [2] J. A. Kraut and N. E. Madias, “Lactic acidosis,” *New England Journal of Medicine*, vol. 371, no. 24, pp. 2309–2319, 2014.
- [3] M. Cecconi, C. Hofer, J.-L. Teboul et al., “Fluid challenges in intensive care: the FENICE study,” *Intensive Care Medicine*, vol. 41, no. 9, pp. 1529–1537, 2015.
- [4] B. Vallet, M. R. Pinsky, and M. Cecconi, “Resuscitation of patients with septic shock: please “mind the gap”!,” *Intensive Care Medicine*, vol. 39, no. 9, pp. 1653–1655, 2013.
- [5] G. A. Ospina-Tascón, M. Umaña, W. F. Bermúdez et al., “Can venous-to-arterial carbon dioxide differences reflect microcirculatory alterations in patients with septic shock?,” *Intensive Care Medicine*, vol. 42, no. 2, pp. 211–221, 2016.
- [6] P. A. V. Beest, M. C. Lont, and N. D. Holman, “Central venous-arterial pCO₂ difference as a tool in resuscitation of septic patients,” *Intensive Care Medicine*, vol. 39, no. 6, pp. 1034–1039, 2013.
- [7] J. Mallat, M. Lemyze, M. Meddour et al., “Ratios of central venous-to-arterial carbon dioxide content or tension to arteriovenous oxygen content are better markers of global anaerobic metabolism than lactate in septic shock patients,” *Annals of Intensive Care*, vol. 6, no. 1, pp. 1–9, 2016.
- [8] J. B. West, “Gas transport to the periphery: how gases are moved to the peripheral tissues?,” in *Respiratory Physiology: The Essentials*, J. B. West, Ed., pp. 69–85, Amazon, Seattle, WA, USA, 4th edition, 1990.

- [9] X.-G. Sun, J. E. Hansen, H. Ting et al., "Comparison of exercise cardiac output by the fick principle using oxygen and carbon dioxide," *Chest*, vol. 118, no. 3, pp. 631-640, 2000.
- [10] J. Cuschieri, E. P. Rivers, M. W. Donnino et al., "Central venous-arterial carbon dioxide difference as an indicator of cardiac index," *Intensive Care Medicine*, vol. 31, no. 6, pp. 818-822, 2005.
- [11] T. A. Bowdle, "Complications of invasive monitoring," *Anesthesiology Clinics of North America*, vol. 20, no. 3, pp. 571-588, 2002.
- [12] B. Lamia, X. Monnet, and J. L. Teboul, "Meaning of arteriovenous PCO₂ difference in circulatory shock," *Minerva Anestesiol*, vol. 72, no. 6, pp. 597-604, 2006.
- [13] I. Giovannini, C. Chiarla, G. Boldrini, and M. Castagneto, "Calculation of venoarterial CO₂ concentration difference," *Journal of Applied Physiology*, vol. 74, no. 2, pp. 959-964, 1993.
- [14] J.-L. Teboul and T. Scheeren, "Understanding the haldane effect," *Intensive Care Medicine*, vol. 43, no. 1, pp. 91-93, 2017.
- [15] H. Zhang and J.-L. Vincent, "Arteriovenous differences in PCO₂ and pH are good indicators of critical hypoperfusion," *American Review of Respiratory Disease*, vol. 148, no. 4_pt_1, pp. 867-871, 1993.
- [16] P. Van der Linden, I. Rausin, A. Deltell et al., "Detection of tissue hypoxia by arteriovenous gradient for PCO₂ and pH in anesthetized dogs during progressive hemorrhage," *Anesthesia & Analgesia*, vol. 80, no. 2, pp. 269-275, 1995.
- [17] B. Vallet, J.-L. Teboul, S. Cain, and S. Curtis, "Venous-arterial CO₂ difference during regional ischemic or hypoxic hypoxia," *Journal of Applied Physiology*, vol. 89, no. 4, pp. 1317-1321, 2000.
- [18] F. Vallée, B. Vallet, O. Mathe et al., "Central venous-to-arterial carbon dioxide difference: an additional target for goal-directed therapy in septic shock?," *Intensive Care Medicine*, vol. 34, no. 12, pp. 2218-2225, 2008.
- [19] J. Bakker, J.-L. Vincent, P. Gris, M. Leon, M. Coffernils, and R. J. Kahn, "Veno-arterial carbon dioxide gradient in human septic shock," *Chest*, vol. 101, no. 2, pp. 509-515, 1992.
- [20] J. Mallat and B. Vallet, "Difference in venous-arterial carbon dioxide in septic shock," *Minerva Anestesiol*, vol. 81, no. 4, pp. 419-425, 2015.
- [21] J. Mallat, M. Lemyze, L. Tronchon et al., "Use of venous-to-arterial carbon dioxide tension difference to guide resuscitation therapy in septic shock," *World Journal of Critical Care Medicine*, vol. 5, no. 1, pp. 47-56, 2016.
- [22] H. Bartels and R. Wrbitzky, "Bestimmung des CO₂-Absorptionskoeffizienten zwischen 15 und 38°C in wasser und plasma," *Pflüger's Archiv für die gesamte Physiologie des Menschen und der Tiere*, vol. 271, no. 2, pp. 162-168, 1960.
- [23] C. Christoforides, L. H. Laasberg, and J. Hedley-Whyte, "Effect of temperature on solubility of O₂ in human plasma," *Journal of Applied Physiology*, vol. 26, no. 1, pp. 56-60, 1969.
- [24] A. Mekontso-Dessap, V. Castelain, N. Anguel et al., "Combination of venoarterial PCO₂ difference with arteriovenous O₂ content difference to detect anaerobic metabolism in patients," *Intensive Care Medicine*, vol. 28, no. 3, pp. 272-277, 2002.
- [25] A. Moioli, B. Maresca, A. Manzione et al., "Metformin associated lactic acidosis (MALA): clinical profiling and management," *Journal of Nephrology*, vol. 29, no. 6, pp. 783-789, 2016.
- [26] D. B. Cotton, H. T. Strassner, L. G. Lipson, and D. A. Goldstein, "The effects of terbutaline on acid base, serum electrolytes, and glucose homeostasis during the management of pre term labor," *American Journal of Obstetrics and Gynecology*, vol. 141, no. 5, pp. 617-624, 1981.
- [27] T. H. Ottens, M. Nijsten, J. Hofland et al., "Effect of high-dose dexamethasone on perioperative lactate levels and glucose control: a randomized controlled trial," *Critical Care*, vol. 19, no. 1, p. 41, 2015.
- [28] P. Sia, T. J. Plumb, and J. A. Fillaus, "Type B lactic acidosis associated with multiple myeloma," *American Journal of Kidney Diseases*, vol. 62, no. 3, pp. 633-637, 2013.
- [29] C. Ustun, P. Fall, H. M. Szerlip et al., "Multiple myeloma associated with lactic acidosis," *Leukemia & Lymphoma*, vol. 43, no. 12, pp. 2395-2397, 2002.
- [30] S. Abdullah, M. Ali, and M. Sabha, "Type-B lactic acidosis associated with progressive multiple myeloma," *Saudi Medical Journal*, vol. 36, no. 2, pp. 239-242, 2015.
- [31] J. G. Younger, J. L. Falk, and S. G. Rothrock, "Relationship between arterial and peripheral venous lactate levels," *Academic Emergency Medicine*, vol. 3, no. 7, pp. 730-733, 1996.
- [32] M. H. Weil, S. Michaels, and E. C. Rackow, "Comparison of blood lactate concentrations in central venous, pulmonary artery, and arterial blood," *Critical Care Medicine*, vol. 15, no. 5, pp. 489-490, 1987.
- [33] M. Suistomaa, E. Ruokonen, A. Kari, and J. Takala, "Time-pattern of lactate and lactate to pyruvate ratio in the first 24 hours of intensive care emergency admissions," *Shock*, vol. 14, no. 1, pp. 8-12, 2000.
- [34] M. G. Vander Heiden, L. C. Cantley, and C. B. Thompson, "Understanding the Warburg effect: the metabolic requirements of cell proliferation," *Science*, vol. 324, no. 5930, pp. 1029-1033, 2009.
- [35] C.-H. Lee, G. Gundem, W. Lee et al., "Persistent severe hyperlactatemia and metabolic derangement in lethal SDHB-mutated metastatic kidney cancer: clinical challenges and examples of extreme Warburg effect," *JCO Precision Oncology*, vol. 1, pp. 1-14, 2017.
- [36] D. H. Henneman and J. P. Bunker, "The pattern of intermediary carbohydrate metabolism in Cushing's syndrome," *The American Journal of Medicine*, vol. 23, no. 1, pp. 34-45, 1957.
- [37] N. E. Madias, "Lactic acidosis," *Kidney International*, vol. 29, no. 3, pp. 752-774, 1986.
- [38] B. Levy, O. Desebbe, C. Montemont, and S. Gibot, "Increased aerobic glycolysis through β_2 stimulation is a common mechanism involved in lactate formation during shock states," *Shock*, vol. 30, no. 4, pp. 417-421, 2008.
- [39] R. Rimachi, F. B. D. Carvalho, C. Orellano-Jimenez, F. Cotton, J. L. Vincent, and D. D. Backer, "Lactate/pyruvate ratio as a marker of tissue hypoxia in circulatory and septic shock," *Anaesth Intensive Care*, vol. 40, no. 3, pp. 427-432, 2012.