



## Draft Genome Sequence of *Falsirhodobacter* sp. Strain alg1, an Alginate-Degrading Bacterium Isolated from Fermented Brown Algae

Tetsushi Mori,<sup>a,d</sup> Mami Takahashi,<sup>a,d</sup> Reiji Tanaka,<sup>b,d</sup> Toshiyuki Shibata,<sup>b,d</sup> Kouichi Kuroda,<sup>c,d</sup> Mitsuyoshi Ueda,<sup>c,d</sup> Haruko Takeyama<sup>a</sup>

Faculty of Science and Engineering, Waseda University Center for Advanced Biomedical Sciences, Shinju-ku, Tokyo, Japan<sup>a</sup>; Department of Life Sciences, Graduate School of Bioresources, Mie University, Tsu, Mie, Japan<sup>b</sup>; Division of Applied Life Sciences, Graduate School of Agriculture, Kyoto University, Kitashirakawa Oiwake–cho, Sakyo–ku, Kyoto, Japan<sup>c</sup>; JST–CREST, Chiyoda–ku, Tokyo, Japan<sup>d</sup>

*Falsirhodobacter* sp. alg1 is an alginate-degrading bacterium, the second species from the nonphototrophic bacterial genus *Falsirhodobacter*. We report the first draft genome of a bacterium from this genus and point out possible important features related to alginate assimilation and its evolutionary aspects.

Received 23 July 2014 Accepted 30 July 2014 Published 21 August 2014

Citation Mori T, Takahashi M, Tanaka R, Shibata T, Kuroda K, Ueda M, Takeyama H. 2014. Draft genome sequence of *Falsirhodobacter* sp. strain alg1, an alginate-degrading bacterium isolated from fermented brown algae. Genome Announc. 2(4):e00826-14. doi:10.1128/genomeA.00826-14.

Copyright © 2014 Mori et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Tetsushi Mori, moritets@aoni.waseda.jp

A lginate-degrading bacteria are considered important targets since they harbor significant genes for the assimilation of alginate, a major component for the production of bioethanol from brown algae (1, 2). These bacteria are also important for evolutionary studies to understand the as yet unclear mechanisms behind alginate assimilation (3). Here, we introduce the draft genome sequence of a novel alginate-degrading bacterium, *Falsirhodobacter* sp. alg1 (AB916500), isolated from the supernatant of fermented brown macroalgae, *Eisenia bicyclis*, collected off the shore of Arasaki, Kanagawa Prefecture, Japan. We classified this bacterium in the *Falsirhodobacter* genus (4), because although it showed 97% similarity to bacteria from the phototrophic *Rhodobacter* genus (5), this bacterium also showed nonphototrophic characteristics and had a similarity of 96.6% to the recently reported *Falsirhodobacter halotolerans* (4).

The genome of this bacterium was sequenced by shotgun sequencing using the GS Junior benchtop system (Roche Diagnostics). Two sequence runs were conducted on the bacterial genome, extracted on separate occasions, and the sequence reads attained from both runs were 205,121 bp (average read length, 483 bp) and 170,198 bp (average read length, 413 bp). All sequencing reads from both runs were pooled and de novo assembled using the Geneious R7 v. 7.1.2 (Biomatters) (6) to reveal a total of 2,952,029 bp with an average GC content of 60.2%, consisting of 205 contigs ( $N_{50}$ , 413,630 bp). Two of the contigs represented circular plasmids sized at 177,174 bp (FRA\_CON006) and 10,356 bp (FRA\_CON010). Automatic gene annotation was performed by Rapid Annotation using Subsytem Technology (RAST) (7), and an overview of the annotated genome was viewed using the SEED viewer (8). The genome sequence contains 3,096 coding sequences (CDS), in which 1,599 CDS (52%) were classified into 385 subsystems, while 1,497 (48%) were uncategorized. In addition, 49 tRNA genes for 18 amino acids and 11 rRNA genes (small subunit [SSU], 3; large subunit [LSU], 5; and 5S, 3) were identified.

Analyzing the pathways within the draft genome, we deter-

mined the genes related to alginate degradation. Two CDS were shown to encode an alginate lyase precursor (algFR1) (located on FRA\_CON002) and an oligonucleotide alginate lyase (algFR2) (located on FRA\_CON003), each classified as members of the CAZy (9) alginate lyase families PL-7 and PL-15, respectively. Functional protein analysis by thin-layer chromatography and liquid chromatography-mass spectrometry (LC-MS) showed that both of these alginate lyases have exolytic properties, suggesting that this bacterium has the ability to efficiently degrade alginate to its monomeric unsaturated sugar, 4-deoxy-L-erythro-5hexoseulose uronate. Subsequently, the search for alginolytic clusters was conducted using the FGENESB online prediction tool (Softberry) (10). Interestingly, we found that algFR2 formed a small operon with several ABC transporter-related genes and an acetoin reductase. In addition, several candidate CDS distributed throughout the genome were also identified that showed similarity to genes found within the alginolytic clusters of other alginatedegrading bacterial strains (3, 11). We believe that Falsirhodobacter sp. alg1 may serve as an important target for future research in alginate degradation and evolutionary studies.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number BBJC00000000. The version described in this paper is the first version, BBJC01000000.

## ACKNOWLEDGMENTS

We thank Naoko Midorikawa and Yumiko Yamada for their assistance in sequencing the genome.

This work was financially supported by the Japan Science and Technology Agency, CREST.

## REFERENCES

 Wargacki AJ, Leonard E, Win MN, Regitsky DD, Santos CN, Kim PB, Cooper SR, Raisner RM, Herman A, Sivitz AB, Lakshmanaswamy A, Kashiyama Y, Baker D, Yoshikuni Y. 2012. An engineered microbial platform for direct biofuel production from brown macroalgae. Science 335:308–313. http://dx.doi.org/10.1126/science.1214547.

- Kawai S, Ohashi K, Yoshida S, Fujii M, Mikami S, Sato N, Murata K. 2014. Bacterial pyruvate production from alginate, a promising carbon source from marine brown macroalgae. J. Biosci. Bioeng. 117:269–274. http://dx.doi.org/10.1016/j.jbiosc.2013.08.016.
- Thomas F, Barbeyron T, Tonon T, Génicot S, Czjzek M, Michel G. 2012. Characterization of the first alginolytic operons in a marine bacterium: from their emergence in marine *Flavobacteriia* to their independent transfers to marine *Proteobacteria* and human gut *Bacteroides*. Environ. Microbiol. 14:2379–2394. http://dx.doi.org/10.1111/j.1462 -2920.2012.02751.x.
- Subhash Y, Tushar L, Sasikala Ch, Ramana ChV. 2013. Falsirhodobacter halotolerans gen. nov., sp. nov., isolated from dry soils of a solar saltern. Int. J. Syst. Evol. Microbiol. 63:2132–2137. http://dx.doi.org/10.1099/ ijs.0.044107-0.
- Hoff WD, van der Horst MA, Nudel CB, Hellingwerf KJ. 2009. Prokaryotic phototaxis. Methods Mol. Biol. 571:25–49. http://dx.doi.org/ 10.1007/978-1-60761-198-1\_2.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics 28:1647–1649. http://dx.doi.org/10.1093/ bioinformatics/bts199.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V,

Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.

- Overbeek R, Begley T, Butler RM, Choudhuri JV, Chuang HY, Cohoon M, de Crécy-Lagard V, Diaz N, Disz T, Edwards R, Fonstein M, Frank ED, Gerdes S, Glass EM, Goesmann A, Hanson A, Iwata-Reuyl D, Jensen R, Jamshidi N, Krause L, Kubal M, Larsen N, Linke B, McHardy AC, Meyer F, Neuweger H, Olsen G, Olson R, Osterman A, Portnoy V, Pusch GD, Rodionov DA, Rückert C, Steiner J, Stevens R, Thiele I, Vassieva O, Ye Y, Zagnitko O, Vonstein V. 2005. The subsystems approach to genome annotation and its use in the project to annotate 1000 genomes. Nucleic Acids Res. 33:5691–5702. http://dx.doi.org/10.1093/ nar/gki866.
- Lombard V, Golaconda Ramulu H, Drula E, Coutinho PM, Henrissat B. 2014. The carbohydrate-active enzymes database (CAZy) in 2013. Nucleic Acids Res. 42:D490–D495. http://dx.doi.org/10.1093/nar/gkt1178.
- Solovyev V, Salamov A. 2011. Automatic annotation of microbial genomes and Metagenomic sequences. *In* Metagenomics and its applications in agriculture, biomedicine and environmental studies. Nova Science Publishers, Hauppauge, NY.
- 11. Mann AJ, Hahnke RL, Huang S, Werner J, Xing P, Barbeyron T, Huettel B, Stüber K, Reinhardt R, Harder J, Glöckner FO, Amann RI, Teeling H. 2013. The genome of the alga-associated marine flavobacterium *formosa agariphila* KMM 3901<sup>T</sup> reveals a broad potential for degradation of algal polysaccharides. Appl. Environ. Microbiol. 79:6813–6822. http://dx.doi.org/10.1128/AEM.01937-13.