

RESEARCH ARTICLE

Prevalence and novel genetic characteristics of *Cryptosporidium* spp. in wild rodents in the northern foothills of the Dabie Mountains, southeast Henan Province, China

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Abstract

Background

Cryptosporidium spp. are prevalent zoonotic pathogens that affect both humans and animals. The pathogens are spread through feces and represent a major cause of diarrhea. As they are both abundant and widely distributed, wild rodents play a significant role in the transmission of Cryptosporidium spp. The Dabie Mountains in southeast Henan Province are rich in wildlife resources as well as various species of livestock. However, the epidemiological characteristics of Cryptosporidium spp. among local wild rodents remain poorly understood. Therefore, the infection rate and genetic characteristics of Cryptosporidium spp. in wild rodents within this region should be determined.

Methods

Between March 2023 and April 2024, a total of 267 wild rodents were captured in the northern foothills of the Dabie Mountains, and fecal samples were collected from their intestines for DNA extraction. Species identification of wild rodents was conducted using PCR amplification of the universal vertebrate cytochrome b (cytb) gene. Nested PCR was subsequently used to amplify the small subunit (SSU) rRNA, actin, heat shock protein 70 (HSP70), and 60 kDa glycoprotein (gp60) genes for the analysis of *Cryptosporidium* species, genotypes, and subtypes in the fecal samples



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Results

The infection rate of *Cryptosporidium* spp. in wild rodents from the northern foothills of the Dabie Mountains was 21.3% (57/267). Seven species of wild rodents were identified, and the infection rates for *Cryptosporidium* spp. varied among host species. In particular, the infection rate was 21.4% (25/117) in *Niviventer lotipes*, 22.4% (22/98) in *Apodemus agrarius*, 17.2% (5/29) in *Rattus nitidus*, 22.2% (4/18) in *Apodemus draco*, and 33.3% (1/3) in *Rattus tanezumi*. The identification results indicated the presence of five *Cryptosporidium* species: *Cryptosporidium apodemi* (n = 12), *C. ubiquitum* (n = 11), *C. viatorum* (n = 7), *C. ratti* (n = 2), and *C. occultus* (n = 2). Moreover, two novel genotypes were identified: *Cryptosporidium sp.* rat genotype VI (n = 8) and *Cryptosporidium sp.* rat genotype VI (n = 8) and *Cryptosporidium sp.* rat genotype VI (n = 15). Notably, a new subtype of *C. viatorum* designated as XVgA4 was discovered.

Conclusions

This study revealed the prevalence of *Cryptosporidium* spp. in wild rodents in the northern foothills of the Dabie Mountains and identified two novel *Cryptosporidium* genotypes, along with a new subtype, *C. viatorum*-XVgA4. The findings highlight the genetic diversity of *Cryptosporidium* spp., underscoring the increased risk of *Cryptosporidium* spp. transmission posed by local wild rodents population. It suggests that host-specific factors should be considered in epidemiological surveillance and control strategies of *Cryptosporidium* spp., which is of great significance for the prevention and control of Cryptosporidiosis.

Author summary

Cryptosporidium spp. are zoonotic pathogens that have a significant impact on both human and animal health, commonly causing diarrhea, abdominal pain, and even dehydration leading to death. To investigate the infection rate and genetic characteristics of Cryptosporidium spp. in wild rodents of a specific region, we gathered samples from the northern foothills of the Dabie Mountains in southeast Henan Province. A total of 267 rodents were sampled, revealing an infection rate of 21.3%. Notably, we mark the first identification of Cryptosporidium viatorum in Niviventer lotipes and Apodemus agrarius, as well as the detection of Cryptosporidium occultus in Rattus nitidus. Furthermore, we uncovered two novel Cryptosporidium genotypes and introduced a new subtype family within C. viatorum. We observed that wild rodents exhibited a higher infection rate of Cryptosporidium spp. during autumn and in field habitats, suggesting a close correlation between geographical environment and climate with the transmission dynamics of Cryptosporidium spp.



Introduction

Cryptosporidium spp. are important zoonotic pathogens affecting humans as well as domestic and wild animals. The parasites rank as the second leading cause of diarrhea and mortality among children, following rotavirus infection [1,2]. Cryptosporidiosis is primarily transmitted through the fecal—oral route, spreading via contaminated water and food, as well as through person-to-person and human-to-animal contact [3]. As the most abundant and diverse group of mammals, rodents are widely distributed [4] and play an important role in the transmission of various pathogens, including Cryptosporidium spp. [5, 6].

The global prevalence of *Cryptosporidium* infection shows considerable variation [7]. At present, the overall global infection rate of *Cryptosporidium* spp. in rodents is approximately 17.0% [8], with prevalence rates ranging from 6.8% to 50.7% [9–16]. To date, at least 31 *Cryptosporidium* species have been identified in rodents [8, 17–19], and 18 *Cryptosporidium* species are known to infect humans. These include *Cryptosporidium hominis*, *C. parvum*, *C. meleagridis*, *C. felis*, *C. canis*, *C. ubiquitum*, *C. viatorum*, *C. mortiferum*, *C. muris*, *C. andersoni*, *C. suis*, *C. scrofarum*, *C. equi*, *C. erinacei*, *C. tyzzeri*, *C. occultus*, *C. ditrichi*, and *C. wrairi* [3]. Notably, *C. viatorum*, *C. muris*, *C. tyzzeri*, *C. ditrichi*, and *C. mortiferum* were initially discovered in rodents [20]. Other common *Cryptosporidium* species identified in rodents include *C. apodemi*, *C. ratti*, and *Cryptosporidium* sp. rat genotypes II-IV [9, 10, 21, 22].

The diverse terrain of the Dabie Mountains in China creates an ideal environment for the survival and reproduction of various flora and fauna, resulting in a region rich in wildlife resources. The Dabie Mountains host a rich diversity of wild rodent species, with variations in species composition across different habitats. In forested areas, the dominant species include *Apodemus draco*, *Apodemus agrarius*, and *Niviventer confucianus*. In agricultural fields and farmlands, the rodent community is primarily composed of species from the genus *Apodemus*, with *A. agrarius* and *A. draco* being the most prevalent. The region also supports a substantial animal husbandry sector.

This study aimed to examine the prevalence of *Cryptosporidium* species among wild rodents in the northern foothills of the Dabie Mountain and to identify the specific species present. Furthermore, we explored the potential zoonotic risks associated with *Cryptosporidium* species carried by wild rodents, thereby providing information for local health authorities for the prevention and control of *Cryptosporidium*. infection in the region.

Results

Identification of wild rodent species

Molecular biological identification and analysis identified seven species among the 267 captured wild rodents: *N. lotipes* (n=117), *A. agrarius* (n=98), *R. nitidus* (n=29), *A. draco* (n=18), *R. tanezumi* (n=3), *Micromys minutus* (n=1), and *Arvicola terrestris* (n=1) (Table 1).

Identification of Cryptosporidium spp.

Among 267 rodent feces samples, 57 positive samples for *Cryptosporidium* spp. were identified using nested PCR targeting the SSU rRNA, actin, and HSP70 genes. This led to an infection rate of 21.3% (57/267) (Table 1). The infection rate of *Cryptosporidium* spp. among wild rodents varied across seasons, reaching a peak in autumn (35.6%, 37/104), followed by summer (14.9%, 15/101). The infection rates were the lowest in winter (8.3%, 1/12) and spring (8.0%, 4/50) (P<0.01) (Table 2). The wild rodents captured in fields exhibited a higher infection rate of *Cryptosporidium* spp. (26.9%, 39/145) than those captured in forested areas (14.8%, 18/122) (P<0.05) (Table 2). Among the wild rodent species, *R. tanezumi* had the highest infection rate of *Cryptosporidium* spp., at 33.3% (1/3), followed by *A. chinensis* at 22.2% (4/18), *N. lotipes* at 21.4% (25/117), *A. agrarius* at 20.4% (20/98), and *R. nitidus* at 17.2% (5/29) (P>0.05) (Table 1).



Table 1. Infection and distribution of *Cryptosporidium* spp. in wild rodents in the northern foothills of the Dabie Mountains, southeast Henan Province.

Samples	No. positive/no. examined (%)	Species/Genotypes (n)	Subtypes (n)	OR (95% CI)	P
By rodent species		0.982			
Niviventer lotipes	25/117 (21.4)	Cryptosporidium sp. rat genotype VII (14), Cryptosporidium sp. rat genotype VI (5), C. viatorum (5), C. ubiquitum (1)	C. viatorum- XVgA4 (5)	Reference	
Apodemus agrarius	20/98 (20.4)	C. apodemi (12), C. ubiquitum (7), Cryptosporidium sp. rat genotype VI (2), C. viatorum (1)	C. viatorum- XVgA4 (1)	1.060 (0.547-2.052)	
Rattus nitidus	5/29 (17.2)	C. ratti (2), C. occultus (2), Cryptosporidium sp. rat genotype VI (1)	1	1.304 (0.452-3.765)	
Apodemus draco	4/18 (22.2)	C. ubiquitum (3), C. viatorum (1)	C. viatorum- XVgA4 (1)	0.951 (0.288-3.145)	
Rattus tanezumi	1/3 (33.3)	Cryptosporidium sp. rat genotype VII (1)	1	0.543 (0.047-6.240)	
Micromys minutus	0/1 (0)	1	1	0.786 (0.715-0.864)	
Arvicola terrestris	0/1 (0)	1	1	0.786 (0.715-0.864)	
Total	57/267 (21.3)	Cryptosporidium sp. rat genotype VII (15), C. apodemi (12), C. ubiquitum (11), Cryptosporidium sp. rat genotype VI (8), C. viatorum (7), C. occultus (2), C. ratti (2)	C. viatorum- XVgA4 (7)		

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Table 2. Infection and distribution of *Cryptosporidium* spp. in wild rodents in different factors in the northern foothills of the Dabie Mountains, southeast Henan Province.

Samples	No. positive/no. examined (%)	Species/Genotypes (n)	Subtypes (n)	OR (95% CI)	P
Season					<0.01
Spring	4/50 (8.0)	C. viatorum (2), Cryptosporidium sp. rat genotype VI (1), C. apodemi (1)	C. viatorum-XVgA4 (2)	6.351 (2.119-19.036)	
Summer	15/101 (14.9)	Cryptosporidium sp. rat genotype VII (5), C. apodemi (5), Cryptosporidium sp. rat genotype VI (2), C. viatorum (1), C. occultus (1), C. ubiquitum (1)	C. viatorum-XVgA4 (1)	3.166 (1.605-6.247)	
Autumn	37/104 (35.6)	Cryptosporidium sp. rat genotype VII (10), C. ubiquitum (10), C. apodemi (6), Cryptosporidium sp. rat genotype VI (4), C. viatorum (4), C. ratti (2), C. occultus (1)	C. viatorum-XVgA4 (4)	Reference	
Winter	1/12 (8.3)	Cryptosporidium sp. rat genotype VI (1)	1	6.075 (0.754-48.922)	
Habitat					0.016
Field	39/145 (26.9)	Cryptosporidium sp. rat genotype VI (8), C. ubiquitum (8), Cryptosporidium sp. rat genotype VII (7), C. apodemi (7), C. viatorum (6), C. ratti (2), C. occultus (1)	C. viatorum-XVgA4 (6)	Reference	
Forest	18/122 (14.8)	Cryptosporidium sp. rat genotype VII (8), C. apodemi (5), C. ubiquitum (3), C. viatorum (1), C. occultus (1)	C. viatorum-XVgA4 (1)	2.126 (1.143-3.954)	
Host gender					0.837
Female	31/142 (21.8)	Cryptosporidium sp. rat genotype VII (10), C. viatorum (5), Cryptosporidium sp. rat genotype VI (4), C. ubiquitum (4), C. apodemi (4), C. ratti (2), C. occultus (2)	C. viatorum-XVgA4 (5)	Reference	
Male	26/125 (20.8)	C. apodemi (8), C. ubiquitum (7), Cryptosporidium sp. rat genotype VII (5), Cryptosporidium sp. rat genotype VI (4), C. viatorum (2)	C. viatorum-XVgA4 (2)	1.063 (0.591-1.913)	

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Cryptosporidium species and genotypes

The sequencing and phylogenetic analysis using the SSU rRNA, actin, and HSP70 genes identified five known *Cryptosporidium* species with zoonotic potential, along with two novel genotypes. The study identified *C. apodemi* (n = 12), *C. ubiquitum* (n = 11), *C. viatorum* (n = 7), *C. ratti* (n = 2), and *C. occultus* (n = 2), along with two novel genotypes named as *Cryptosporidium sp.* rat genotype VI (n = 8) and *Cryptosporidium sp.* rat genotype VII (n = 15).

Among these *Cryptosporidium* species and genotypes, the nucleotide sequences generated from *C. ubiquitum* and *C. apodemi* were identical to the GenBank sequences MT507486 and MG266041, respectively. In addition, the nucleotide sequences from *C. ubiquitum*, *C. apodemi*, *C. viatorum*, *C. occultus*, *C. occultus*, and *C. ratti* had 1–5 single nucleotide polymorphisms (SNPs) compared to the GenBank reference sequences [MT507486, MG266041, JX978269, MT507491, MT507489]. *Cryptosporidium sp.* rat genotype VI and *Cryptosporidium sp.* rat genotype VII were located on independent branches and were identified as two new genotypes. *Cryptosporidium sp.* rat genotype VI had closer genetic relationships with *C. suis* and *C. occultus*, while *Cryptosporidium sp.* rat genotype VII was closely related to *C. ratti* (Fig 1).

Subtypes of zoonotic Cryptosporidium spp

The zoonotic *Cryptosporidium* species *C. viatorum* was further characterized by the gp60 gene. Seven *C. viatorum* samples were successfully amplified. According to the established nomenclature [23], a new subtype family was identified and named XVg. In the phylogenetic analysis, XVg clustered with the known subtype families XVe and XVf and exhibited genetic relatedness to the subtype family XVa (Fig 2). Based on the number of contiguous TCA trinucleotides in the serine repeat region, a new subtype, XVgA4, was identified within the new subtype family.

Discussion

Wild rodents are widely distributed and have strong reproductive capacity, and they are often in close contact with both humans and domestic animals. The infection rate of *Cryptosporidium* spp. in rodents varies significantly among world regions. In a global meta-analysis, the overall prevalence of *Cryptosporidium* spp. in rodents was 19.8% (4589/23142) [19]. Currently, *Cryptosporidium* infection in rodents have been recorded in 19 countries/regions, with prevalence rates of 0.7%–100%. At 28.0%, Europe has the highest prevalence rate, whereas South American countries exhibit the lowest at 5.0% [8, 19]. Conversely, the infection rate of wild rodents in China varies widely, from 4.0% to 73.9% [24]. The present study is the first to investigate and research the *Cryptosporidium* infection status in wild rodents in the northern foothills of the Dabie Mountains, and the overall infection rate was 21.3% (57/267), which is generally consistent with previous reports.

Several studies have shown that the infection rate of *Cryptosporidium* spp. varies among rodent species. According to previous reports, prevalence rates of *Cryptosporidium* spp. in *Apodemus* spp. range from 13.7% to 31.8% [12, 24], from 21.3% to 22.6% in voles [12, 25], and 14.3% in shrews [12, 22]. In our study, different species of wild rodents exhibited little variation in infection rates. The highest infection rate was observed in *R. tanezumi*(33.3%, 1/3), which was followed by *A. chinensis* (22.2%, 4/18), *N. lotipes* (21.4%, 25/117), *A. agrarius* (20.4%, 20/98), and *R. nitidus* (17.2%, 5/29).

Differences in the distributions of *Cryptosporidium* spp. have been observed among rodent species. *C. parvum*, *C. ubiquitum* and *C. muris* are the predominant species infecting rodents [19]. In the present study, *Cryptosporidium sp.* rat genotype VI, *C. apodemi*, *C. ubiquitum*, and *Cryptosporidium sp.* rat genotype VI showed the highest infection rates. *Cryptosporidium sp.* rat genotype VII was primarily found in rat species such as *N. lotipes* and *R. tanezumi*, while *C. apodemi* exhibited strong host specificity, infecting only *A. agrarius*. *Cryptosporidium ubiquitum* was predominant in *Apodemus* spp., including *A. agrarius* and *A. chinensis*. Furthermore, *Cryptosporidium sp.* rat genotype VI was found in *N. lotipes*, *A. agrarius*, and *R. nitidus*. Nevertheless, we did not find any local wild rodents infected with *C. parvum* and *C. muris*, and the specific reasons warrant further exploration. Notably, this study analyzed the infection rate of *Cryptosporidium* spp. in wild rodents across different seasons. The highest *Cryptosporidium* infection rate was observed in autumn (35.6%,



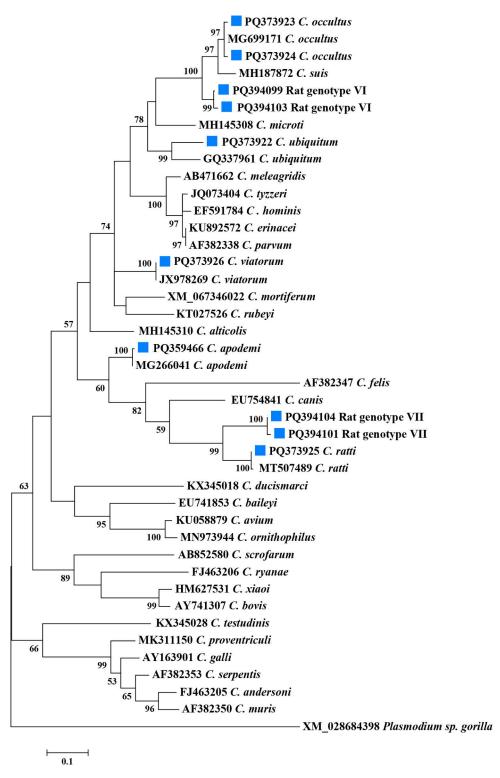


Fig 1. Phylogenetic relationships of *Cryptosporidium* **spp.** Phylogenetic trees based on the maximum likelihood analyses of the actin gene. Bootstrap values > 50% from 1000 replicates are displayed. The blue squares represent the newly submitted sequences of *Cryptosporidium* species and genotypes identified in this study. The scale bar indicates 0.1 nucleotide substitutions per site.

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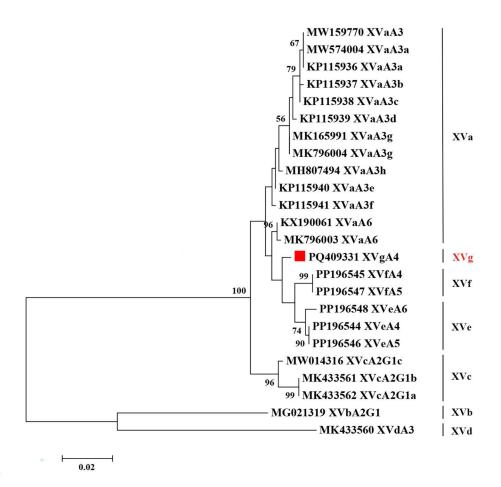


Fig 2. Phylogenetic relationships between various subtypes of *C. viatorum.* Phylogenetic trees based on the gp60 gene sequences of *C. viatorum* were constructed using the maximum likelihood method with 1000 bootstrap replicates. Novel subtypes are indicated by red squares. The scale bar indicates 0.02 nucleotide substitutions per site.

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37/104), followed by summer (14.9%, 15/101). Winter (8.3%, 1/12) and spring (8.0%, 4/50) recorded the lowest infection rate. Previous studies have stated that the infection rates of *Cryptosporidium* spp. in wild rodents generally show little variation between summer and autumn [22, 26, 27]. However, in this study, the infection rate in autumn was significantly higher than in summer. This discrepancy could be due to several factors. The mild temperatures and humid climate in the Dabie Mountains during autumn create favorable conditions for the survival and dispersal of *Cryptosporidium* oocysts. Moreover, wild rodents tend to store large amounts of food and water in autumn to prepare for winter, and this leads to increased activity during this period. These factors may collectively contribute to a significant increase in the *Cryptosporidium* infection rate among wild rodents in autumn.

Furthermore, topography has an impact on the infection rate of *Cryptosporidium* spp. in wild rodents. The sampling sites in this study were primarily divided into field and forest areas. In accordance with previously reported findings, the infection rate of *Cryptosporidium* spp. in wild rodents captured in field was significantly higher at 26.9% (39/145) compared to 14.8% (18/122) in forest areas [22]. The infection rate of *Cryptosporidium* spp. did not differ significantly between male and female rodents.



A total of five Cryptosporidium species and two novel genotypes were successfully identified: C. apodemi, C. ubiquitum, C. viatorum, C. ratti, C. occultus, as well as Cryptosporidium sp. rat genotype VI and Cryptosporidium sp. rat genotype VII. Among these species. C. ubiquitum. C. viatorum, and C. occultus are known to be zoonotic. Cryptosporidium ubiquitum, as an important zoonotic pathogen, can infect a wide range of domestic and wild animals [28]. It has been reported in various wild rodents, such as A. agrarius, A. draco, Mus musculus, and Rattus norvegicus [1, 29,30]. This study is the first to report the presence of C. ubiquitum in N. lotipes, which contributes to a more comprehensive understanding of the ecology and host adaptability of C. ubiquitum and provides valuable insights for assessing its zoonotic risks. In China, Cryptosporidium viatorum was initially only found in humans and urban wastewater or combined sewage overflow [23, 31–36]. Nevertheless, in 2018, C. viatorum was reported in non-human hosts such as Rattus lutreolus in Australia, R. norvegicus, Leopoldamys edwardsi, Berylmys bowersi, Niviventer coninga, and A. draco in China, and Rattus rattus in France [1, 37–40]. This study is the first to report that N. lotipes and A. agrarius can be infected by C. viatorum, thereby expanding its host range. This finding also supports the hypothesis that wild rodents may serve as hosts for the zoonotic species C. viatorum. Recent studies suggest that the C. viatorum-XVa subtype family isolated from wild rodents is genetically similar to subtypes found in humans [40, 41]. The newly discovered XVg subtype family is genetically closely related to the XVa subtype family, although further research is needed. The present study also reports for the first time that C. occultus can infect R. nitidus. First discovered in rats in 2018, Cryptosporidium occultus was later reported in 2020 to infect humans [37, 42]. In addition to the previously mentioned zoonotic Cryptosporidium species, C. apodemi and C. ratti identified in this study have to date only been found in wild rodents, and both of these Cryptosporidium species exhibit strong host specificity [21, 24, 43]. The newly discovered genotypes Cryptosporidium sp. rat genotype VI and Cryptosporidium sp. rat genotype VII were particularly significant in this study, accounting for 23 out of 57 Cryptosporidium-positive samples, indicating that these two genotypes may be prevalent among local wild rodents. In the phylogenetic analysis, Cryptosporidium sp. rat genotype VI was closely related to C. suis and C. occultus, while Cryptosporidium sp. rat genotype VII was more closely related and belonged to the same clade as C. ratti. The Dabie Mountains, which are located in the southeastern region of Henan Province, serve as a natural "gene bank" for species, featuring a diverse array of wild animals and livestock. Cryptosporidiosis has harmed both local public health and animal husbandry. The identification of these two new Cryptosporidium genotypes has improved our understanding of the epidemiological characteristics of Cryptosporidium spp. in wild rodents in the northern foothills of the Dabie Mountains, providing valuable insights into the development of effective prevention and control strategies in the region.

The infection rate of *Cryptosporidium* spp. in wild rodents is influenced by multiple factors, such as host specificity, geographical distribution, climatic conditions, sampling topography, and host gender [39]. This study revealed that the infection rate of *Cryptosporidium* spp. differed significantly among different seasons and land use forms; however, there were relatively small differences among species of rodents and rodent genders. Therefore, to better understand the epidemiology of *Cryptosporidium* spp. in wild rodents in the northern foothills of the Dabie Mountains, additional sampling of wild rodents within the region is needed.

Conclusions

This study shows that wild rodents in the northern foothills of the Dabie Mountains, southeast Henan Province, harbor a diverse range of *Cryptosporidium* spp., including five species and two novel genotypes. Notably, this is the first report of *C. ubiquitum* infections *N. lotipes*, *C. viatorum* infections in *N. lotipes* and *A. agrarius*, as well as the identification of *C. occultus* in *R. nitidus*. The discovery of a novel subtype family, XVg, within *C. viatorum* highlights the genetic diversity of these pathogens. These findings emphasize the heightened risk of *Cryptosporidium* spp. transmission posed by wild rodents in the Dabie Mountains. Owing to the large numbers and wide distribution of rodents, the pathogens they carry are a significant source of infection for both humans and domestic animals. Hence, more extensive sampling surveys should be conducted in diverse regions to determine the transmission risk of Cryptosporidiosis from wild rodents.



Materials and methods

Ethics statement

All research procedures used in this work were approved by the Institutional Review Board of Henan Agricultural University (approval no. IRB-HENAU-20190820–02).

Sample collection and processing

Between March 2023 and March 2024, a total of 267 wild rodents were collected from the northern foothills of the Dabie Mountains in southeastern Henan Province (Latitude range: approximately 31.42°N to 31.90°N. Longitude range: approximately 114.74°E to 115.44°E) (Fig 3 and S1 Table). Featuring a diverse range of landforms, the Dabie Mountains are rich in both wildlife and domestic animal resources. The rodents were captured in rural, field, and forested areas, including farming areas where they frequently interact with domestic animals. After capturing the wild rodents, euthanasia was performed using an overdose of inhaled anesthetic. Subsequently, approximately 500 mg of feces was collected for sampling from the rectal contents of each rodent and stored for subsequent analysis.

DNA extraction

The whole-genome DNA from wild rodent feces was extracted using a fecal DNA extraction kit (Tiangen Biotech (Beijing) Co., Ltd.) Following the manufacturer's protocol. DNA was extracted from a total of 267 fecal samples. The extracted DNA was then stored at −20°C for subsequent analyses.

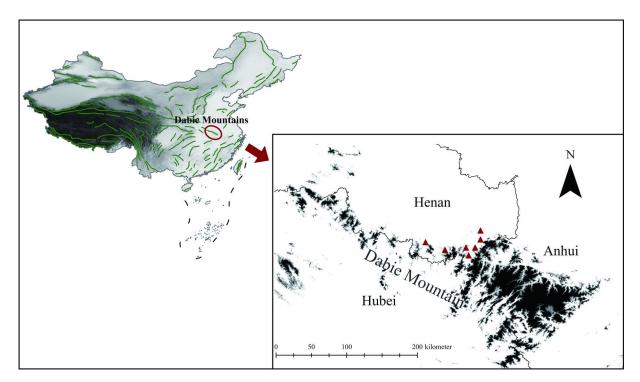


Fig 3. Geographical locations of wild rodents examined in the present study in the northern foothills of the Dabie Mountains, southeast Henan Province, China (red triangle). The map sources are from the National Platform for Common Geospatial Information Services (https://www.tianditu.gov.cn/) and the Geospatial Data Cloud (https://www.gscloud.cn/).

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PCR amplification

Wild rodent species were examined by PCR amplification of the cytochrome b (cytb) gene [44]. *Cryptosporidium* spp. were analyzed through nested PCR amplification of the small subunit (SSU) rRNA, actin, and heat Shock Protein 70 (HSP70) genes [45–47]. *C. viatorum* subtypes were determined using PCR and sequence analysis of the 60 kDa glycoprotein (gp60) gene [23]. Genomic DNA from *Cryptosporidium parvum* (from cattle) and *C. viatorum* (from *Rattus* species) served as positive controls in the PCR analyses.

Sequencing and phylogenetic analysis

All positive secondary PCR amplicons were sequenced bi-directionally on an ABI 3730 platform by Sangon Biotech (Henan, China). The successfully bidirectionally sequenced DNA fragments were aligned and assembled into complete DNA sequences using Mega 10.0 (http://www.megasoftware.net/). The assembled complete DNA sequences were then subjected to BLAST comparisons, with sequences in the National Center for Biotechnology Information (NCBI) (https://pubmed.ncbi.nlm.nih.gov/) database to identify the species of the parasite. Phylogenetic analysis was conducted using the maximum composite likelihood model, and bootstrap values were calculated by analyzing 1000 replicates. The other parameters were the default values of MEGA 10.0.

Statistical analysis

Chi-squared tests and 95% confidence intervals (CIs) were calculated using Crosstab in SPSS version 24.0 (SPSS Inc., Chicago, IL, USA). The chi-squared test was used to determine differences in the prevalence of *Cryptosporidium* spp. between rodent species, habitats, seasons, and host genders. The results were considered statistically significant at P<0.05.

Supporting information

S1 Table. The latitude and longitude information of the wild rodent sampling sites in the northern foothills of the Dabie Mountains, southeast Henan Province.

(DOCX)

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Funding acquisition: Longxian Zhang.

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Methodology: Yin Fu, Longxian Zhang. **Project administration:** Longxian Zhang.



Resources: Longxian Zhang.
Software: Pitambar Dhakal.
Supervision: Longxian Zhang.

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References

- Feng K, Yang S, Xu Y, Wen L, Chen J, Zhang W, et al. Molecular characterization of Cryptosporidium spp., Giardia spp. and Enterocytozoon bieneusi in eleven wild rodent species in China: Common distribution, extensive genetic diversity and high zoonotic potential. One Health. 2024;18:100750. https://doi.org/10.1016/j.onehlt.2024.100750 PMID: 38798737
- 2. Innes EA, Chalmers RM, Wells B, Pawlowic MC. A One Health Approach to Tackle Cryptosporidiosis. Trends Parasitol. 2020;36(3):290–303. https://doi.org/10.1016/j.pt.2019.12.016 PMID: 31983609
- 3. Zhou S, Jiang Y, Cao J. Cryptosporidium infection in rodents in China. Chin J Parasitol Parasit Dis. 2024;42:512–20.
- 4. Islam MM, Farag E, Hassan MM, Jaffrey SS, Atta M, Al-Marri AM, et al. Rodent-borne zoonoses in Qatar: A possible One-Health framework for the intervention of future epidemic. One Health. 2023;16:100517. https://doi.org/10.1016/j.onehlt.2023.100517 PMID: 37363248
- García-Livia K, Fernández-Álvarez Á, Feliu C, Miquel J, Quilichini Y, Foronda P. Cryptosporidium spp. in wild murids (Rodentia) from Corsica, France. Parasitol Res. 2022;121(1):345–54. https://doi.org/10.1007/s00436-021-07369-4 PMID: 34816301
- Meerburg BG, Singleton GR, Kijlstra A. Rodent-borne diseases and their risks for public health. Crit Rev Microbiol. 2009;35(3):221–70. https://doi.org/10.1080/10408410902989837 PMID: 19548807
- 7. Feng Y. Cryptosporidium in wild placental mammals. Exp Parasitol. 2010;124(1):128–37. https://doi.org/10.1016/j.exppara.2008.11.005 PMID: 19084007
- 8. Taghipour A, Olfatifar M, Foroutan M, Bahadory S, Malih N, Norouzi M. Global prevalence of Cryptosporidium infection in rodents: A systematic review and meta-analysis. Prev Vet Med. 2020;182:105119. https://doi.org/10.1016/j.prevetmed.2020.105119 PMID: 32823254
- 9. Lv C, Zhang L, Wang R, Jian F, Zhang S, Ning C, et al. Cryptosporidium spp. in wild, laboratory, and pet rodents in china: prevalence and molecular characterization. Appl Environ Microbiol. 2009;75(24):7692–9. https://doi.org/10.1128/AEM.01386-09 PMID: 19820152
- 10. Tan TK, Low VL, Ng WH, Ibrahim J, Wang D, Tan CH, et al. Occurrence of zoonotic Cryptosporidium and Giardia duodenalis species/genotypes in urban rodents. Parasitol Int. 2019;69:110–3. https://doi.org/10.1016/j.parint.2018.12.007 PMID: 30590124
- 11. Backhans A, Jacobson M, Hansson I, Lebbad M, Lambertz S, Gammelgård E. Occurrence of pathogens in wild rodents caught on Swedish pig and chicken farms. Epidemiol Infect. 2013;141:1885–91.
- Danišová O, Valenčáková A, Stanko M, Luptáková L, Hatalová E, Čanády A. Rodents as a reservoir of infection caused by multiple zoonotic species/genotypes of C. parvum, C. hominis, C. suis, C. scrofarum, and the first evidence of C. muskrat genotypes I and II of rodents in Europe. Acta Trop. 2017;172:29–35. https://doi.org/10.1016/j.actatropica.2017.04.013 PMID: 28433573
- 13. Feliu C, López M, Gómez MS, Torres J, Sánchez S, Miquel J, et al. Parasite fauna of rodents (Murinae) from El Hierro (Canary Islands, Spain): a multidisciplinary approach. Acta Parasitol. 2012;57(2):171–8. https://doi.org/10.2478/s11686-012-0016-7 PMID: 22807054
- 14. Ng-Hublin JSY, Singleton GR, Ryan U. Molecular characterization of Cryptosporidium spp. from wild rats and mice from rural communities in the Philippines. Infect Genet Evol. 2013;16:5–12. https://doi.org/10.1016/j.meegid.2013.01.011 PMID: 23376410
- 15. Song J, Kim C-Y, Chang S-N, Abdelkader TS, Han J, Kim T-H, et al. Detection and Molecular Characterization of Cryptosporidium spp. from Wild Rodents and Insectivores in South Korea. Korean J Parasitol. 2015;53(6):737–43. https://doi.org/10.3347/kjp.2015.53.6.737 PMID: 26797442
- 16. Stenger BLS, Horčičková M, Clark ME, Kváč M, Čondlová Š, Khan E, et al. Cryptosporidium infecting wild cricetid rodents from the subfamilies Arvicolinae and Neotominae. Parasitology. 2018;145(3):326–34. https://doi.org/10.1017/S0031182017001524 PMID: 28870264
- 17. Huang J, Chen M, He Y, Chen H, Huang M, Li N. Cryptosporidium equi n. sp. (Apicomplexa: Cryptosporidiidae): biological and genetic characterisations. Int J Parasitol. 2023;53:545–55.
- 18. Tůmová L, Ježková J, Prediger J, Holubová N, Sak B, Konečný R, et al. Cryptosporidium mortiferum n. sp. (Apicomplexa: Cryptosporididae), the species causing lethal cryptosporidiosis in Eurasian red squirrels (Sciurus vulgaris). Parasit Vectors. 2023;16(1):235. https://doi.org/10.1186/s13071-023-05844-8 PMID: 37454101
- 19. Zhang K, Fu Y, Li J, Zhang L. Public health and ecological significance of rodents in Cryptosporidium infections. One Health. 2021;14:100364. https://doi.org/10.1016/j.onehlt.2021.100364 PMID: 34984218
- 20. Ryan U, Zahedi A, Feng Y, Xiao L. An Update on Zoonotic Cryptosporidium Species and Genotypes in Humans. Animals (Basel). 2021;11(11):3307. https://doi.org/10.3390/ani11113307 PMID: 34828043



- 21. Čondlová Š, Horčičková M, Sak B, Květoňová D, Hlásková L, Konečný R. Cryptosporidium apodemi sp. n. and Cryptosporidium ditrichi sp. n. (Apicomplexa: Cryptosporidiidae) in Apodemus spp. Eur J Protistol. 2018;63:1–1.
- 22. Kivistö R, Kämäräinen S, Huitu O, Niemimaa J, Henttonen H. Zoonotic cryptosporidium spp. in wild rodents and shrews. Microorganisms. 2021;9.
- 23. Stensvold CR, Elwin K, Winiecka-Krusnell J, Chalmers RM, Xiao L, Lebbad M. Development and application of a gp60-based typing assay for Cryptosporidium viatorum. J Clin Microbiol. 2015;53:1891–7.
- 24. Čondlová Š, Horčičková M, Havrdová N, Sak B, Hlásková L, Perec-Matysiak A, et al. Diversity of Cryptosporidium spp. in Apodemus spp. in Europe. Eur J Protistol. 2019;69:1–13. https://doi.org/10.1016/j.ejop.2019.02.005 PMID: 30826667
- 25. Horčičková M, Čondlová Š, Holubová N, Sak B, Květoňová D, Hlásková L, et al. Diversity of Cryptosporidium in common voles and description of Cryptosporidium alticolis sp. n. and Cryptosporidium microti sp. n. (Apicomplexa: Cryptosporidiidae). Parasitology. 2019;146(2):220–33. https://doi.org/10.1017/S0031182018001142 PMID: 30012231
- 26. Ni H-B, Sun Y-Z, Qin S-Y, Wang Y-C, Zhao Q, Sun Z-Y, et al. Molecular Detection of Cryptosporidium spp. and Enterocytozoon bieneusi Infection in Wild Rodents From Six Provinces in China. Front Cell Infect Microbiol. 2021;11:783508. https://doi.org/10.3389/fcimb.2021.783508 PMID: 34900760
- 27. Jiang Y, Jiang A, Ren G, Wang L, Xin X, Yuan Z, et al. Cross-species transmission of Cryptosporidium in wild rodents from the southern region of Zhejiang Province of China and its possible impact on public health. Parasite. 2024;31:34. https://doi.org/10.1051/parasite/2024033 PMID: 38949636
- 28. Li N, Xiao L, Alderisio K, Elwin K, Cebelinski E, Chalmers R, et al. Subtyping Cryptosporidium ubiquitum,a zoonotic pathogen emerging in humans. Emerg Infect Dis. 2014;20(2):217–24. https://doi.org/10.3201/eid2002.121797 PMID: 24447504
- 29. Liu L, Xu Q, Jiang A, Zeng F, Zhao W, Tan F. Molecular characterization of *Cryptosporidium* in wild rodents from the Inner Mongolian Autonomous Region and Liaoning Province, China: assessing host specificity and the potential for zoonotic transmission. Front Vet Sci. 2024;11:1406564.
- Trogu T, Formenti N, Marangi M, Viganò R, Bionda R, Giangaspero A, et al. Detection of Zoonotic Cryptosporidium ubiquitum in Alpine Wild Ruminants. Pathogens. 2021;10(6):655. https://doi.org/10.3390/pathogens10060655 PMID: 34070669
- 31. Elwin K, Hadfield S, Robinson G, Crouch N, Chalmers R. Cryptosporidium viatorum n. sp. (Apicomplexa: Cryptosporidiidae) among travellers returning to Great Britain from the Indian subcontinent, 2007-2011. Int J Parasitol. 2012;42:675–82.
- 32. Insulander M, Silverlås C, Lebbad M, Karlsson L, Mattsson JG, Svenungsson B. Molecular epidemiology and clinical manifestations of human cryptosporidiosis in Sweden. Epidemiol Infect. 2013;141(5):1009–20. https://doi.org/10.1017/S0950268812001665 PMID: 22877562
- 33. Adamu H, Petros B, Zhang G, Kassa H, Amer S, Ye J, et al. Distribution and clinical manifestations of Cryptosporidium species and subtypes in HIV/AIDS patients in Ethiopia. PLoS Negl Trop Dis. 2014;8(4):e2831. https://doi.org/10.1371/journal.pntd.0002831 PMID: 24743521
- **34.** Ukwah BN, Ezeonu IM, Ezeonu CT, Roellig D, Xiao L. Cryptosporidium species and subtypes in diarrheal children and HIV-infected persons in Ebonyi and Nsukka, Nigeria. J Infect Dev Ctries. 2017;11:173–9.
- 35. Sánchez A, Munoz M, Gómez N, Tabares J, Segura L, Salazar Á, et al. Molecular Epidemiology of Giardia, Blastocystis and Cryptosporidium among Indigenous Children from the Colombian Amazon Basin. Front Microbiol. 2017;8:248. https://doi.org/10.3389/fmicb.2017.00248 PMID: 28270802
- **36.** Huang C, Hu Y, Wang L, Wang Y, Li N, Guo Y. Environmental transport of emerging human-pathogenic cryptosporidium species and subtypes through combined sewer overflow and wastewater. Appl Environ Microbiol. 2017;83:e00682-17.
- 37. Xu N, Liu H, Jiang Y, Yin J, Yuan Z, Shen Y, et al. First report of Cryptosporidium viatorum and Cryptosporidium occultus in humans in China, and of the unique novel C. viatorum subtype XVaA3h. BMC Infect Dis. 2020;20(1):16. https://doi.org/10.1186/s12879-019-4693-9 PMID: 31910816
- 38. Koehler AV, Wang T, Haydon SR, Gasser RB. Cryptosporidium viatorum from the native Australian swamp rat Rattus lutreolus An emerging zoonotic pathogen?. Int J Parasitol Parasites Wildl. 2018;7(1):18–26. https://doi.org/10.1016/j.ijppaw.2018.01.004 PMID: 29556470
- 39. Zhao W, Zhou H, Huang Y, Xu L, Rao L, Wang S, et al. Cryptosporidium spp. in wild rats (Rattus spp.) from the Hainan Province, China: Molecular detection, species/genotype identification and implications for public health. Int J Parasitol Parasites Wildl. 2019;9:317–21. https://doi.org/10.1016/j.ijppaw.2019.03.017 PMID: 31338292
- **40.** Chen Y-W, Zheng W-B, Zhang N-Z, Gui B-Z, Lv Q-Y, Yan J-Q, et al. Identification of Cryptosporidium viatorum XVa subtype family in two wild rat species in China. Parasit Vectors. 2019;12(1):502. https://doi.org/10.1186/s13071-019-3763-6 PMID: 31661007
- 41. Stensvold CR, Elwin K, Winiecka-Krusnell J, Chalmers RM, Xiao L, Lebbad M. Development and Application of a gp60-Based Typing Assay for Cryptosporidium viatorum. J Clin Microbiol. 2015;53(6):1891–7. https://doi.org/10.1128/JCM.00313-15 PMID: 25832304
- 42. Kváč M, Vlnatá G, Ježková J, Horčičková M, Konečný R, Hlásková L, et al. Cryptosporidium occultus sp. n. (Apicomplexa: Cryptosporidiidae) in rats. Eur J Protistol. 2018;63:96–104. https://doi.org/10.1016/j.ejop.2018.02.001 PMID: 29506004
- **43.** Ježková J, Prediger J, Holubová N, Sak B, Konečný R, Feng Y, et al. Cryptosporidium ratti n. sp. (Apicomplexa: Cryptosporidiidae) and genetic diversity of Cryptosporidium spp. in brown rats (Rattus norvegicus) in the Czech Republic. Parasitology. 2021;148(1):84–97. https://doi.org/10.1017/S0031182020001833 PMID: 32981543
- **44.** Robins JH, Hingston M, Matisoo-smith E, Ross HA. Identifying Rattus species using mitochondrial DNA. Molecular Ecology Notes. 2007;7(5):717–29. https://doi.org/10.1111/j.1471-8286.2007.01752.x



- **45.** Xiao L, Escalante L, Yang C, Sulaiman I, Escalante A, Montali R. Phylogenetic analysis of cryptosporidium parasites based on the small-subunit rRNA gene locus. Appl Environ Microbiol. 1999;65:1578–83.
- **46.** Sulaiman IM, Lal AA, Xiao L. Molecular phylogeny and evolutionary relationships of cryptosporidium parasites at the actin locus. J Parasitol. 2002;88:388–94.
- 47. Sulaiman IM, Morgan UM, Thompson RC, Lal AA, Xiao L. Phylogenetic relationships of Cryptosporidium parasites based on the 70-kilodalton heat shock protein (HSP70) gene. Appl Environ Microbiol. 2000;66(6):2385–91. https://doi.org/10.1128/AEM.66.6.2385-2391.2000 PMID: 10831415