

Research article

Open Access

Galph_s-coupled receptor signaling actively down-regulates α₄β₁-integrin affinity: A possible mechanism for cell de-adhesion

Alexandre Chigaev*^{1,2}, Anna Waller², Or Amit² and Larry A Sklar*^{1,2}

Address: ¹Department of Pathology, University of New Mexico Health Sciences Center, Albuquerque, NM 87131, USA and ²Cancer Research and Treatment Center, University of New Mexico Health Sciences Center, Albuquerque, NM 87131, USA

Email: Alexandre Chigaev* - achigaev@salud.unm.edu; Anna Waller - awaller@salud.unm.edu; Or Amit - oamit@salud.unm.edu; Larry A Sklar* - lsklar@salud.unm.edu

* Corresponding authors

Published: 5 June 2008

Received: 17 December 2007

BMC Immunology 2008, 9:26 doi:10.1186/1471-2172-9-26

Accepted: 5 June 2008

This article is available from: <http://www.biomedcentral.com/1471-2172/9/26>

© 2008 Chigaev et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Activation of integrins in response to inside-out signaling serves as a basis for leukocyte arrest on endothelium, and migration of immune cells. Integrin-dependent adhesion is controlled by the conformational state of the molecule (i.e. change in the affinity for the ligand and molecular unbending (extension)), which is regulated by seven-transmembrane Guanine nucleotide binding Protein-Coupled Receptors (GPCRs). α₄β₁-integrin (CD49d/CD29, Very Late Antigen-4, VLA-4) is expressed on leukocytes, hematopoietic stem cells, hematopoietic cancer cells, and others. Affinity and extension of VLA-4 are both rapidly up-regulated by inside-out signaling through several Gα_i-coupled GPCRs. The goal of the current report was to study the effect of Gα_s-coupled GPCRs upon integrin activation.

Results: Using real-time fluorescent ligand binding to assess affinity and a FRET based assay to probe α₄β₁-integrin unbending, we show that two Gα_s-coupled GPCRs (H₂-histamine receptor and β₂-adrenergic receptor) as well as several cAMP agonists can rapidly down modulate the affinity of VLA-4 activated through two Gα_i-coupled receptors (CXCR4 and FPR) in U937 cells and primary human peripheral blood monocytes. This down-modulation can be blocked by receptor-specific antagonists. The Gα_s-induced responses were not associated with changes in the expression level of the Gα_i-coupled receptors. In contrast, the molecular unbending of VLA-4 was not significantly affected by Gα_s-coupled GPCR signaling. In a VLA-4/VCAM-1-specific myeloid cell adhesion system, inhibition of the VLA-4 affinity change by Gα_s-coupled GPCR had a statistically significant effect upon cell aggregation.

Conclusion: We conclude that Gα_s-coupled GPCRs can rapidly down modulate the affinity state of VLA-4 binding pocket through a cAMP dependent pathway. This plays an essential role in the regulation of cell adhesion. We discuss several possible implications of this described phenomenon.

Background

Integrins, one of the largest families of cell adhesion molecules play an important role in the regulation of immune responses and leukocyte traffic, development, hemostasis,

and cancer [1]. A unique feature of integrins is in their ability to rapidly and reversibly regulate cell adhesion, and thereby modulate cell recruitment and homing, without a significant change in the expression of the molecule.

Understanding the molecular mechanisms that regulate rapid changes in cell adhesion avidity is essential, since integrins are known to play roles in many human diseases. They represent an attractive target for several existing and emerging drugs for treatment of inflammatory diseases, anti-angiogenic cancer therapy, anti-thrombotic therapy, and others [2-5].

$\alpha_4\beta_1$ -integrin (CD49d/CD29, Very Late Antigen-4, VLA-4) is expressed on peripheral blood leukocytes, hematopoietic progenitors/stem cells, hematopoietic cancer cells, and others [6-8]. On circulating lymphocytes, VLA-4 has the potential to exist in multiple affinity states that can mediate tethering, rolling, and arrest on a ligand (CD106, Vascular Cell Adhesion Molecule-1, VCAM-1) that is up-regulated on inflamed endothelia [9,10]. The ability of leukocytes to arrest, extravasate, and navigate is directly related to the activation of VLA-4 integrin through $G\alpha_i$ -coupled GPCRs. Classical chemoattractants and chemokines that include formyl peptide and SDF-1 (CXCL12) are examples of $G\alpha_i$ -coupled GPCR ligands [11]. $G\alpha_i$ -coupled GPCRs activate integrin by triggering an inside-out signaling pathway. The change in the affinity for the ligand as well as the unbending (extension) of the integrin molecule are believed to be a part of this activation process [12-14].

We have developed a VLA-4 specific fluorescent probe that allows us to monitor VLA-4 affinity changes in real-time on live cells. Our model cell line is undifferentiated U937 cells, which do not attach to plastic surfaces nor exhibit homotypic adhesion [15], and constitutively express VLA-4 integrins in a low affinity state [6]. In these undifferentiated U937 cells VLA-4 affinity was rapidly and transiently triggered by several different $G\alpha_i$ -coupled receptors, and the kinetics of the affinity change reflected the desensitization kinetics of a particular GPCR [6]. Moreover, in a VLA-4/VCAM-1 specific cell adhesion system, the time course of avidity changes in response to receptor activation coincided with the time course of the affinity changes [16]. The increased cell aggregation could be accounted for by VLA-4 affinity changes rather than integrin clustering or increasing number of VLA-4/VCAM-1 bonds [17]. Using the same VLA-4 specific fluorescent probe, we have also developed a FRET based assay for the detection of VLA-4 unbending (extension) upon inside-out activation [18], which was validated in a series of publications [19-21]. We found that rather than being intrinsically related to the affinity change of the integrin binding pocket, the conformational unbending of the VLA-4 molecule is regulated by a different signaling pathway, in a different temporal fashion [20].

Recently, it has been proposed that in addition to well-known pro-adhesive signaling events (modulated by $G\alpha_i$ -

coupled GPCRs), anti-adhesive signaling events can participate in regulation of cell adhesion, diapedesis and chemotaxis [11]. We noted previously that anti-inflammatory signals could be derived from $G\alpha_s$ -coupled signaling pathway [22], and recently that U-73122, a suspected histamine analog [23], could inhibit the VLA-4 affinity increase [20]. The goal of the current work was to explore whether the $G\alpha_s$ -coupled GPCR related pathway can affect the affinity or conformational state of the integrin. We found that two $G\alpha_s$ -coupled receptors constitutively expressed on U937 cells can actively down regulate VLA-4 affinity, after activation through two $G\alpha_i$ -coupled GPCRs. This affinity down regulation was not modulated by the change in the intracellular Ca^{2+} concentration. Furthermore, the unbending of VLA-4 integrin, as detected using the FRET based assay, was not significantly affected by $G\alpha_s/cAMP$ signaling pathway. Using a VLA-4/VCAM-1-specific myeloid cell adhesion model system, we found that blocking the integrin affinity change by the activation of the $G\alpha_s/cAMP$ signaling pathway had a statistically significant effect upon cell adhesion. However, we also detected an activation of cell adhesion by $G\alpha_i$ -coupled GPCR under these conditions (when the affinity change was blocked). In the absence of the affinity change a rapid increase in cell adhesion was attributed to the unbending (extension) of the integrin molecule.

Results

Signaling through $G\alpha_s$ -coupled receptors blocks $G\alpha_i$ -coupled receptor-induced VLA-4 affinity up-regulation

The LDV-FITC probe, which mimics binding of a natural ligand to VLA-4, was used to study the effect of $G\alpha_s$ -coupled receptor signaling on the VLA-4 affinity change. Because the concentration of the LDV-FITC probe in these experiments (4 nM) has been chosen to be lower than K_d for the resting state of VLA-4 and higher than the K_d for the activated state, the transition from the low to the high affinity state was accompanied by additional binding of the probe. As shown previously, activation of $G\alpha_i$ -coupled GPCRs leads to up-regulation of VLA-4 affinity, and the LDV-FITC binding kinetics reflects the activation kinetics for a particular GPCR. LDV-FITC binding is rapid and transient for wild type receptors (because of the receptor desensitization), and rapid and sustained for a non-desensitizing mutant of FPR [6].

To study the effect of $G\alpha_s$ -coupled receptors we took advantage of two $G\alpha_s$ -coupled receptors that constitutively expressed on U937 cells: histamine H2 receptor and β_2 adrenergic receptors [24,25]. U937 cells stably transfected with CXCR4 receptor, preincubated with 4 nM LDV-FITC, were treated with receptor-specific agonists at saturating concentration (amthamine or isoproterenol) and DMSO (control). Next, cells were activated with CXCL12 (SDF-1). While control cells showed rapid and

transient activation of VLA-4, cells preactivated through $G\alpha_s$ -coupled receptor showed a significantly lower response (Fig. 1A). Although, a slight decrease in the LDV-FITC binding to the resting cells after $G\alpha_s$ -coupled receptor stimulation was observed, amthamine or isoproterenol alone did not affect the resting VLA-4 affinity as measured by dissociation rate analysis (data not shown). The majority of integrin molecules on resting U937 cells exhibit low affinity (see Fig. 5 in [6]), however a small population of high affinity sites that cannot be detected by a dissociation rate analysis could exist. $G\alpha_s$ -coupled receptor signalling can eliminate this small fraction of active integrins.

The use of a non-desensitizing mutant of FPR allowed us to study the effect of $G\alpha_s$ -coupled activation in real-time. In the absence of $G\alpha_i$ -coupled GPCR desensitization, VLA-4 maintained its high affinity state for a longer period of time [6,19]. Addition of specific $G\alpha_s$ -coupled receptor ligands in this case led to a dose dependent down regulation of integrin affinity that was reflected in the rapid dissociation of the LDV-FITC probe (Fig. 1B). Thus, according to the analysis of LDV-FITC ligand binding, activation of $G\alpha_s$ -coupled GPCRs prevents VLA-4 affinity up-regulation, if done prior to $G\alpha_i$ -coupled receptor signaling, or it rapidly down regulates VLA-4 affinity, if performed during continuous $G\alpha_i$ -coupled receptor signaling.

The effect of $G\alpha_s$ -coupled receptor agonists is reversed by specific receptor antagonists

Next, to verify whether the observed effects are receptor-specific, we used two specific $G\alpha_s$ -coupled receptor antagonists: tiotidine (histamine H2 receptor antagonist), and ICI-118,551 (β_2 adrenergic receptor antagonist) [26,27]. Addition of receptor antagonists after U937 cells were treated with $G\alpha_s$ -coupled receptor agonist completely reversed a suppressive effect of $G\alpha_s$ -coupled receptor signaling (blue lines, Fig. 2A,B). In this case, at the end of the experiment the binding of the LDV-FITC probe returned back to the level of the untreated control (red lines, labeled fMLFF only, Fig. 2A,B). It is worth noting that $G\alpha_s$ -coupled receptor antagonists by themselves did not have any effect on fMLFF activated samples (see Fig. 2B, black line labeled fMLFF, ICI-118,551), nor did they activate VLA-4 when added alone (data not shown). Thus, $G\alpha_s$ -coupled receptors provide a signal that results in VLA-4 affinity down regulation, and this effect is receptor specific since specific receptor antagonists reversed the effects of $G\alpha_s$ -coupled receptor agonists.

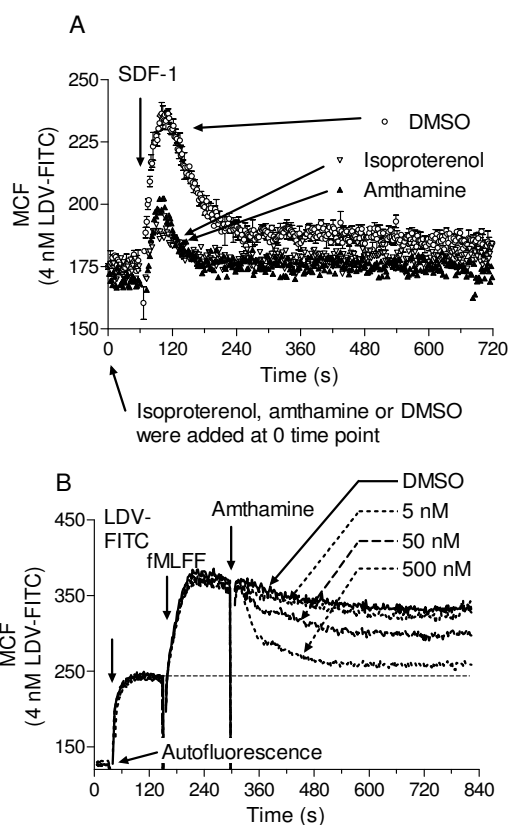


Figure 1
Binding and dissociation of the LDV-FITC probe in response to $G\alpha_s$ -coupled receptor agonists. Experiments were conducted as described under "Methods". A, LDV-FITC probe binding and dissociation on U937 cells stably transfected with CXCR4 receptor plotted as mean channel fluorescence (MCF) versus time. The experiment involved sequential additions of $G\alpha_s$ -coupled receptor ligands (isoproterenol (100 nM) or amthamine (500 nM)), then, fluorescent LDV-FITC probe (4 nM, below saturation, added 3 min prior to addition of $G\alpha_i$ -coupled receptor ligands). Control cells were treated with vehicle (DMSO). Next, cells were activated with SDF-1 (25 nM, arrows). Rapid and reversible binding of the probe reflects VLA-4 affinity change [6]. Curves are means out of two independent runs calculated on a point-by-point basis. B, LDV-FITC probe binding and dissociation on U937 cells stably transfected with the non-desensitizing mutant of FPR (Δ ST) [33] plotted as mean channel fluorescence (MCF) versus time. The experiment involved sequential additions of fluorescent LDV-FITC probe (4 nM), fMLFF (100 nM), and different concentrations of amthamine, or DMSO (control) (arrows). The MCF value corresponding to cell autofluorescence is indicated by the horizontal arrow. One representative experiment out of three experiments is shown. Experiments shown in the different panels were performed using different instruments, and therefore MCF values are not identical.

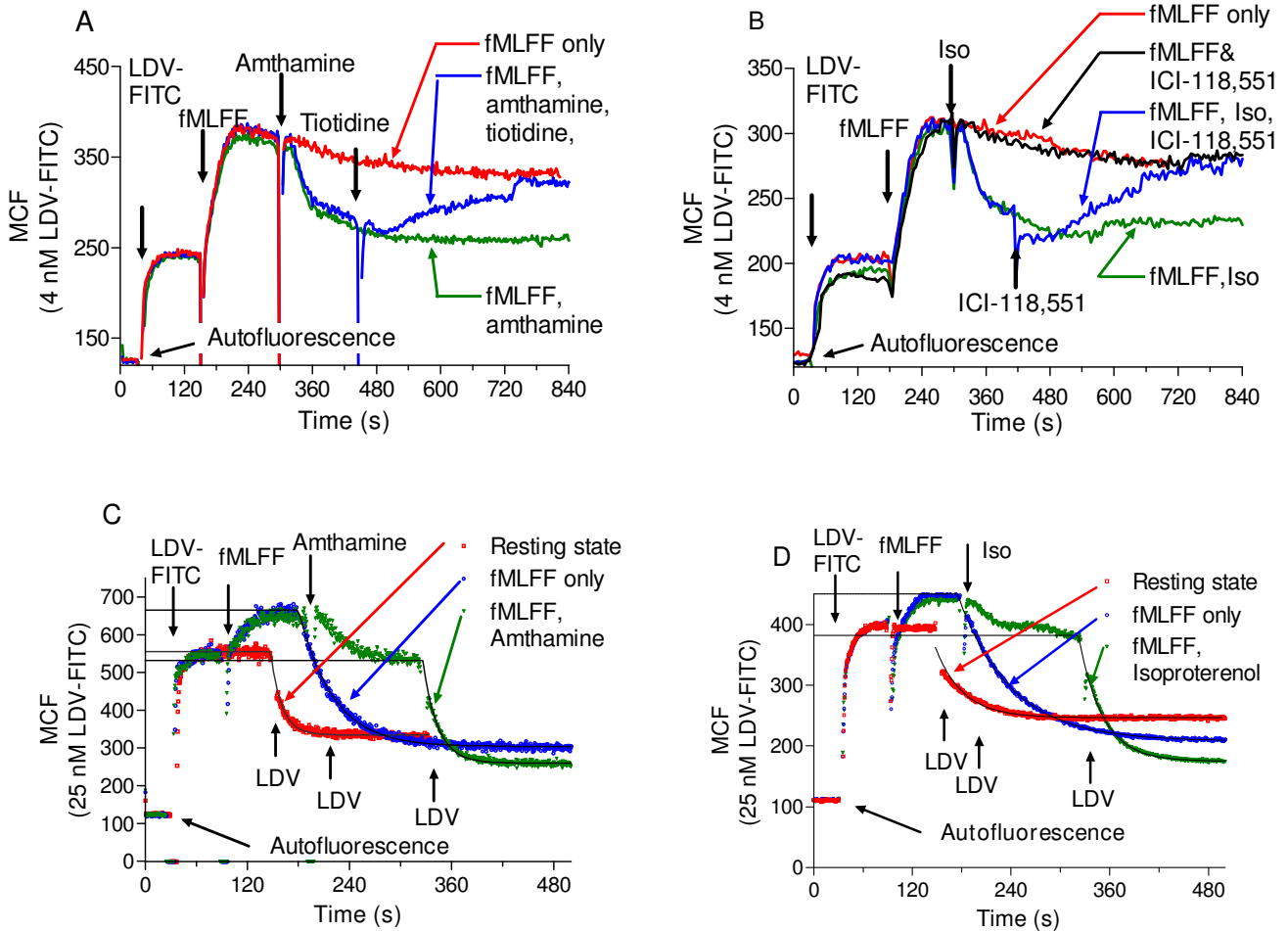


Figure 2
Binding and dissociation of the LDV-FITC probe in response to $G\alpha_s$ -coupled receptor agonists and antagonists.
 A, B, The effect of $G\alpha_s$ -coupled receptor antagonists. U937 cells stably transfected with the non-desensitizing mutant of FPR were sequentially treated with the LDV-FITC probe (4 nM, below saturation), fMLFF (100 nM), and A, amthamine (500 nM), tiotidine (10 μ M), or B, isoproterenol (1 nM), ICI-118,551 (10 μ M). B, Additional control sample was treated with the same concentration of LDV-FITC, fMLFF, and ICI-118,551 without addition of isoproterenol to show the effect of $G\alpha_s$ -coupled receptor antagonist without addition of the agonist. The MCF value corresponding to cell autofluorescence is indicated by the horizontal arrow. Data are plotted as MCF versus time. One representative experiment out of three experiments is shown.
 C, D, Kinetic analysis of binding and dissociation of LDV-FITC probe on U937 cells stably transfected with the non-desensitizing mutant of FPR. Cells were sequentially treated with the LDV-FITC probe (25 nM, near saturation), $G\alpha_s$ -coupled receptor ligand (fMLFF, 100 nM), $G\alpha_s$ -coupled receptor ligands, C, amthamine (500 nM), or D, isoproterenol (100 nM). At time points indicated by arrows, cells were treated with excess unlabeled LDV containing small molecule (2 μ M), and the dissociation of the fluorescent molecule was followed. Dissociation rate constants (k_{off}) were obtained by fitting dissociation curves to a single exponential decay equation (as described in the text). Experiments shown in the different panels were performed using different instruments, and therefore MCF values are not identical.

Dissociation rate analysis confirmed that changes in the LDV-FITC probe binding could be attributed to the change in the affinity of the binding pocket

For different affinity states of VLA-4, the equilibrium dissociation constant (K_d) of the LDV-FITC probe varied inversely with the dissociation rate constant (k_{off}), suggesting that the association rate constant was essentially

independent of receptor conformation [6,10]. Therefore, we routinely use dissociation rate analysis to quantitatively probe the affinity state of the VLA-4 binding pocket. Cells were preincubated with a higher concentration of LDV-FITC (25 nM) to saturate the majority of low affinity sites. Next, a large excess of the unlabeled LDV competitor is added to initiate a dissociation of labeled probe. After

activation through $G\alpha_i$ -coupled GPCR, little additional binding of the probe was seen (since the K_d for the low affinity state is ~ 12 nM (Table I in [6]), at 25 nM $\sim 70\%$ of sites are occupied before activation) (Fig. 2C,D). At rest, the majority of VLA-4 molecules present on U937 cells exhibit the low affinity state ($k_{off} \sim 0.1-0.05$ s $^{-1}$). After $G\alpha_i$ -coupled receptor activation by fMLFF the dissociation rate was significantly slower ($k_{off} \sim 0.01-0.025$ s $^{-1}$, slower k_{off} indicates higher affinity). After additional stimulation through two different $G\alpha_s$ -coupled receptors, dissociation rates were comparable to the rate at the resting state. These k_{off} values for the resting and activated VLA-4 receptor were quantitatively similar to previously published data [6]. Thus, the analysis of dissociation rates at rest, after $G\alpha_i$ -coupled receptor activation, and after $G\alpha_s$ -coupled receptor stimulation indicated that the $G\alpha_s$ -coupled receptor related signaling pathway is decreasing VLA-4 affinity by increasing the dissociation rate of the ligand.

G α_s -coupled receptor activation has no significant effect upon expression of G α_i -coupled receptors and VLA-4 integrin

Next, in order to exclude the possibility that stimulation of $G\alpha_s$ -coupled receptors dramatically changes surface expression of $G\alpha_i$ -coupled receptors and VLA-4, U937 cells were treated with $G\alpha_s$ -coupled receptor agonists, and fMLFF in a manner that was identical to experiments described above (Fig. 1, 2). Then, cells were stained with anti-CXCR4, anti-FPR, and anti-CD49d antibodies (Fig. 3). No statistically significant changes in a surface expression were detected. Thus, the effect of down-regulation of integrin affinity is not related to the expression of $G\alpha_i$ -coupled GPCRs or VLA-4 integrin itself.

Earlier data suggest that FPR signalling typically requires occupancy of a fraction of the receptor sites [28]. In our experiments with saturating ligand concentration termination of signaling is expected to require a significant down-modulation of receptor expression within a few minutes. Furthermore, the ability of $G\alpha_s$ -coupled receptor antagonists to completely reverse the effect of $G\alpha_s$ -coupled receptor agonists (Fig. 2A, B, blue lines) indicates that integrin affinity down-regulation is triggered by the $G\alpha_s$ -coupled GPCRs activation rather than by $G\alpha_i$ -coupled deactivation. In addition, the use of the non-desensitizing FPR mutant emphasizes the role of $G\alpha_s$ -coupled signalling in the down-regulation of integrin affinity.

Forskolin and dbcAMP mimic the effect of G α_s -coupled receptor signaling

$G\alpha_s$ -coupled receptors are well known to activate adenylyl cyclase, which increases the concentration of intracellular cyclic adenosine monophosphate (cAMP). Also, treatment of U937 cells with β -adrenoceptor agonist induced a several fold elevation of intracellular cAMP concentra-

tion [29]. Therefore, we have tested whether activation of adenylyl cyclase by forskolin or dibutyryl cAMP would mimic the effects of $G\alpha_s$ -coupled receptors activation on VLA-4 affinity regulation. We found that preincubation of U937 cells with either compound abolished VLA-4 affinity up-regulation induced through CXCR4 receptor (Fig. 4A). We also detected affinity down regulation in real-time after activation of U937 cells through a non-desensitizing mutant of FPR (Fig. 4B,C). It is worth noting that the overall kinetics of the LDV-FITC ligand binding and dissociation in these cases was comparable to the case of $G\alpha_s$ -coupled receptor activation (compare Fig. 1 and Fig. 4). Thus, our data suggest that the down regulation of VLA-4 affinity after $G\alpha_s$ -coupled receptor activation is modulated by the increased concentration of cAMP.

Signaling through G α_s -coupled receptors does not block VLA-4 unbending (extension)

A second aspect of integrin activation through "inside-out" signaling is a large conformational change, which results in the molecular unbending (extension) of the molecule, and has been associated with cell rolling [30]. The unbent integrin conformation results in faster cell aggregation rates, potentially because of the accessibility of the VLA-4 binding site [20]. Moreover, according to our recent findings, VLA-4 molecular extension is regulated by a mechanism, independent of a regulation of the affinity for the ligand [20]. Therefore, we studied the effect of $G\alpha_s$ -coupled receptor signaling on integrin extension using the FRET based real-time unbending assay (Fig. 5) [18-20].

Activation of U937 cell through a non-desensitizing mutant of FPR results in the rapid and sustained unquenching of the fluorescent signal, which is interpreted as a change in the distance of closest approach between the surface of the plasma membrane and the headgroup of the integrin molecule (molecular unbending) (see Methods and Fig. 1 in [18], or Fig. 5 in [19]). Addition of $G\alpha_s$ -coupled receptor ligands resulted in a rapid decrease of the fluorescent signal to about 40-50% of the control within ~ 60 seconds. Thereafter, the fluorescent signal slowly recovered back to $\sim 70-90\%$ of the control at the end of the experiment for the case of histamine H2 receptor stimulation (Fig. 5A,B). The recovery phase was less definitive for the case of β_2 adrenergic receptor stimulation (data not shown). Thus, the overall kinetics of the affinity change as detected by LDV-FITC probe binding and dissociation was entirely different from the kinetics of the molecular unbending as detected using FRET (compare Figs. 1, 2, 4 and Fig. 5A,B). In the case of the affinity change it takes up to 10 min to reach "full deactivation". The signal returns to the baseline value at 200-800 s after the beginning of the experiment, and no significant recovery was observed at this time scale (see Fig. 1B, Fig. 2A,B, Fig. 4B,C). In contrast, for the FRET signal, the

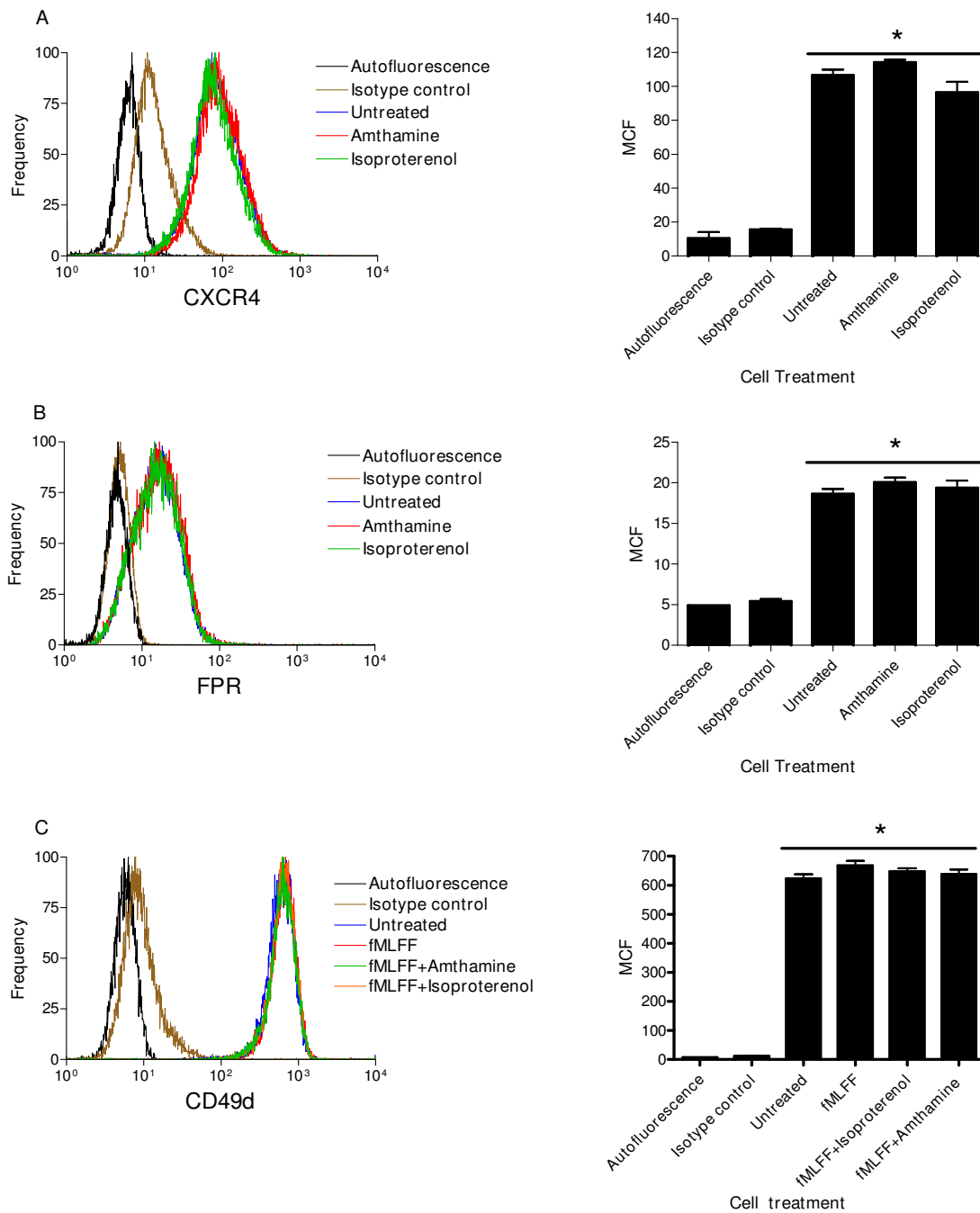


Figure 3
Effect of $G\alpha_s$ -coupled receptor activation upon surface expression of CXCR4 receptor, formyl peptide receptor, and VLA-4. U937 cells were treated with vehicle (untreated control), amthamine, isoproterenol, or sequentially with fMLFF, and vehicle (untreated control), amthamine or isoproterenol as described for experiments shown in Fig. 1 and 2. Next, cells were placed on ice and stained with anti-CXCR4 antibodies, A, anti-FPR antibodies, B, or CD49d antibodies, C. Histograms and bar graphs of mean channel fluorescence (MCF) \pm SEM (n = 3) for unstained cell (autofluorescence), nonspecific binding (isotype control), treated with vehicle (untreated), and treated with different ligands are shown. One representative experiment out of two experiments is shown. * indicates means are not significantly different (P > 0.05) as calculated by one-way ANOVA analysis using GraphPad Prism software.

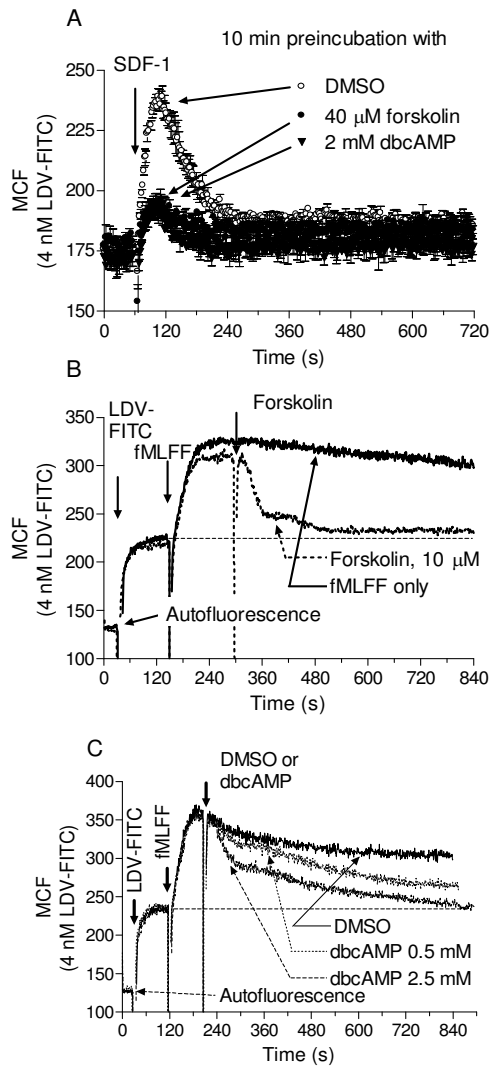


Figure 4
Binding and dissociation of the LDV-FITC probe in response to cAMP agonists. A, LDV-FITC probe binding and dissociation on U937 cells stably transfected with CXCR4 receptor plotted as mean channel fluorescence (MCF) versus time. Prior to SDF-I addition, cells were preincubated with forskolin (40 μ M), dibutyryl-cAMP (2 mM), or DMSO (control) for 10 min at 37°C. Next, cells were preincubated with 4 nM LDV-FITC, and treated with SDF-I (25 nM). Mean and standard error of mean are shown (n = 3). B, C, LDV-FITC probe binding and dissociation on U937 cells stably transfected with the non-desensitizing mutant of FPR plotted as mean channel fluorescence (MCF) versus time. After sequential additions of fluorescent LDV-FITC probe (4 nM), and fMLFF (100 nM), cells were treated with B, forskolin (10 μ M), C, different concentration of dibutyryl-cAMP, or DMSO (control). The MCF value corresponding to cell autofluorescence is indicated by the horizontal arrow. One representative experiment out of three experiments is shown.

response was maximal at \sim 1 min after $G\alpha_s$ -coupled receptor activation, and the signal did not return back to the baseline as in the case of the affinity change. This partial quenching result does not distinguish whether all VLA-4 molecules were still partially unbent (intermediate extended state) or whether a fraction of the VLA-4 molecules was still fully extended, while the rest of the integrins returned back to the bent conformation. However, the different kinetics of the FRET change and LDV-FITC binding support the idea that unbending and affinity are regulated through different signaling mechanisms [20].

Next, to study the effect of a sustained elevation of cAMP concentration upon integrin unbending we preincubated U937 with forskolin (10 μ M for 5 min). This treatment was sufficient to completely inhibit (Fig. 4A) or reverse (Fig. 4B) the effect of $G\alpha_i$ -coupled receptors activation upon VLA-4 affinity up-regulation. However, we could not find any statistically significant difference in the FRET based real-time unbending assay between forskolin treated and DMSO (vehicle) treated cells (Fig. 5C,D). Thus, a sustained elevation of cAMP by forskolin had no effect upon VLA-4 unbending after $G\alpha_i$ -coupled receptor activation.

The effect of $G\alpha_s$ -coupled receptor on VLA-4 activation is independent of Ca^{2+} -signaling

Conformational unbending of VLA-4 is dependent on the elevation of cytosolic Ca^{2+} concentration, while affinity up-regulation can be observed in the absence of Ca^{2+} increase [20]. Therefore, we tested whether in our model system $G\alpha_s$ -coupled receptor activation would significantly change intracellular Ca^{2+} -concentration (Fig. 6). Activation through a non-desensitizing mutant of FPR resulted in sustained elevation of intracellular Ca^{2+} . None of the $G\alpha_s$ -coupled receptor agonists significantly lowered the intracellular Ca^{2+} concentration. In fact, we have observed a small increase of intracellular Ca^{2+} after amthamine treatment (Fig. 6B). Thus, activation of $G\alpha_s$ -coupled receptors decreased VLA-4 affinity without significantly changing (decreasing) intracellular Ca^{2+} -concentration.

These data additionally support the finding that the affinity state of the binding pocket is largely independent of cytoplasmic Ca^{2+} level (Fig. 2 in [20]). Furthermore, partial quenching of the FRET signal (Fig. 5B) together with elevated intracellular Ca^{2+} (Fig. 6) is analogous to the previously described condition, which arises several minutes after wild type $G\alpha_i$ -coupled receptor activation. There, the affinity state was already low because of rapid receptor desensitization, while VLA-4 integrins remained partially extended, as detected in the FRET based assay (see Fig. 7 in [20]).

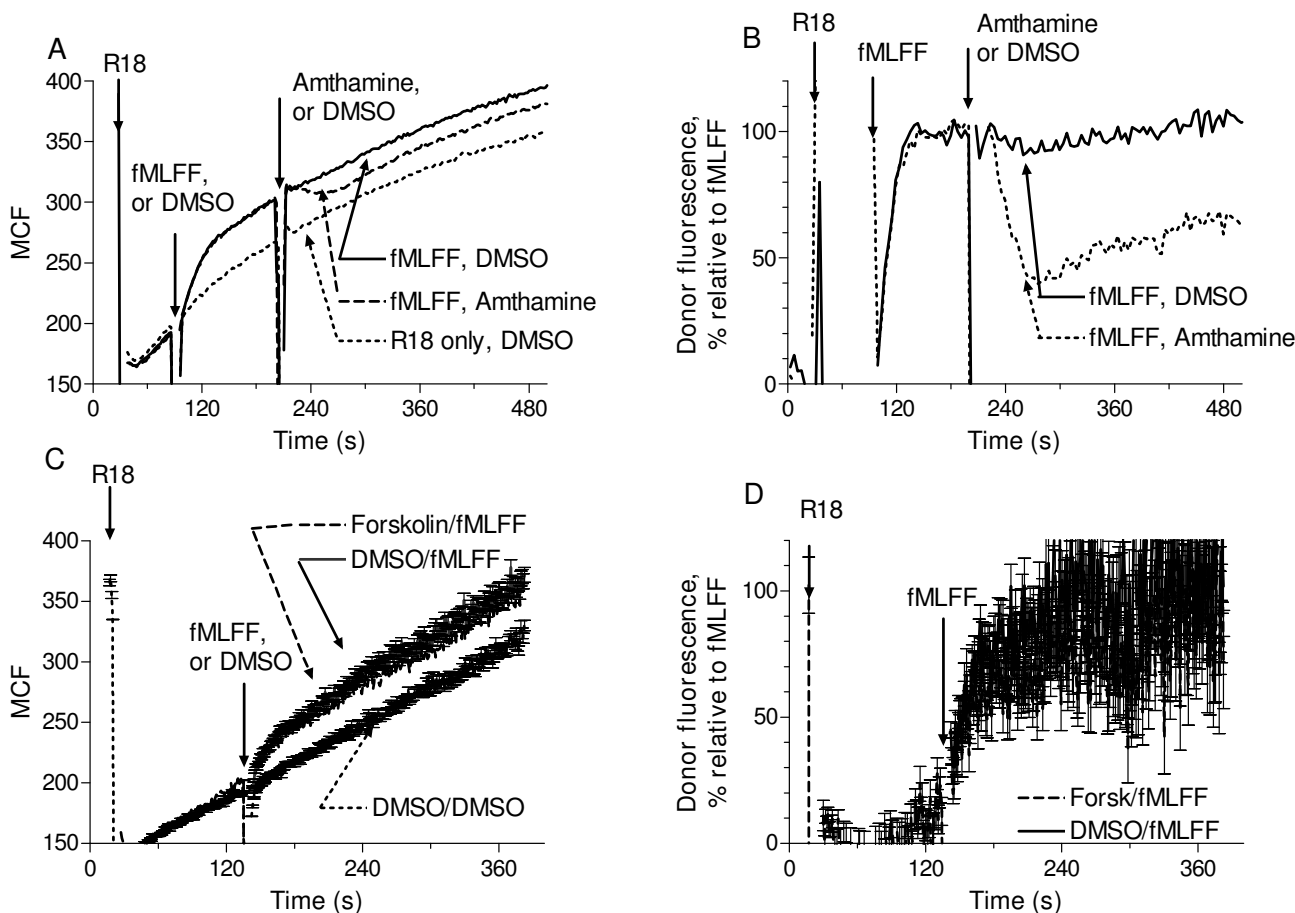


Figure 5
Energy transfer, on U937 cells stably transfected with the non-desensitizing mutant of FPR, between the LDV-FITC donor probe and octadecylrhodamine (R18) acceptor probe. Experiments were conducted as described under "Methods", and in [18,19]. A, Cells were preincubated at 37°C with 100 nM LDV-FITC probe to saturate high and low affinity sites. Next, LDV-FITC fluorescence was quenched after addition of octadecyl rhodamine (R18, 10 μM). Then, cells were activated by addition of fMLFF (100 nM). Data are plotted as MCF versus time for three conditions: quenched and then activated by $G\alpha_i$ -coupled receptor agonist (fMLFF (solid line)), quenched, then activated by $G\alpha_i$ -coupled receptor agonist (fMLFF) and then activated by $G\alpha_s$ -coupled receptor agonist (amthamine, 500 nM (dashed line)), and quenched only (R18 only, DMSO vehicle, dotted line). B, Data from panel A were re-plotted by subtracting the baseline data (R18 only) from activated cell data. The data are normalized assuming that average MCF value for FPR activated cells is equal to 100%; therefore, the Y-axis is labeled as "Donor fluorescence, % relative to fMLFF". As shown previously, activation of the cells transfected with the non-desensitizing mutant of FPR [18] led to the rapid unquenching of the FITC signal, which was interpreted as a change in the distance of closest approach between the VLA-4 ligand binding site occupied by LDV-FITC and the membrane surface (so called unbending or extension of the integrin molecule, see Fig. 1 in [18], or Fig. 5C in [19]). Curves are means out of five independent runs calculated on a point-by-point basis. C, Cells were preincubated for 5 min at 37°C with 10 μM of forskolin or DMSO (control). Next, cells were stained with 100 nM LDV-FITC probe and fluorescence was quenched after addition of octadecyl rhodamine (R18, 10 μM). Cells were activated by addition of 100 nM fMLFF, or DMSO (control). Notice the absence of the difference between forskolin treated (forskolin/fMLFF) and DMSO treated (DMSO/fMLFF) cells after addition of fMLFF. Mean and standard error of mean are shown (n = 3). D, Data from panel C were re-plotted as described for panel B.

The effect of $G\alpha_s$ /cAMP signaling pathway activation upon cell adhesion

Next, to study the implications of the $G\alpha_s$ -coupled signaling pathway on VLA-4 dependent cell adhesion we uti-

lized a VLA-4/VCAM-1-specific real-time cell adhesion assay. The specificity of cell aggregation was tested using anti- α_4 integrin mAb (HP2/1) [16] as well as the unlabeled LDV small molecule that completely blocked cell

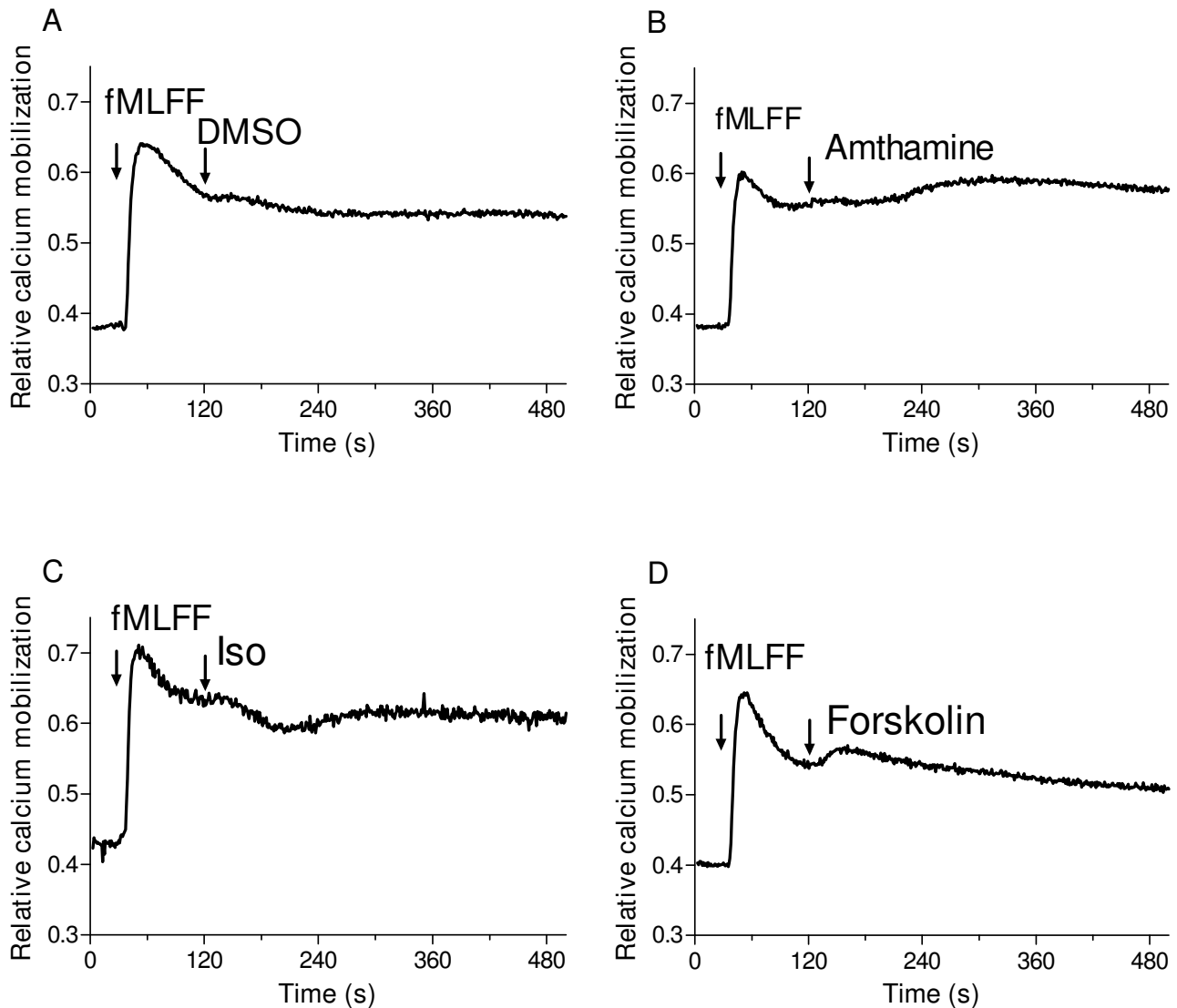


Figure 6

Kinetics of intracellular Ca^{2+} response, detected using Indo-1, AM, in U937 cells stably transfected with the non-desensitizing mutant of FPR in response to activation by $\text{G}\alpha_i/\text{G}\alpha_s$ -coupled receptor agonists. Experiments were conducted as described under "Methods". A, Control cells loaded with Indo-1 were treated with fMLFF (100 nM) and DMSO. B, Cells were sequentially treated with fMLFF (100 nM) and amthamine (500 nM); C, cells were treated with fMLFF (100 nM) and isoproterenol (100 nM); D, cells were treated with fMLFF and Forskolin (40 μM). One representative experiment out of three experiments is shown.

aggregation ([20] and Fig. 7A, solid squares). Prior to mixing, pre-stained (red, PKH26GL) U937 cells, stably transfected with non-desensitizing mutant of FPR, were treated with forskolin (20 μM , 10 min, 37°C). This treatment was sufficient to fully block/reverse the effect of $\text{G}\alpha_i$ -coupled receptors on the VLA-4 affinity change (Fig. 4A,B). However, it was insufficient to completely prevent rapid cellular aggregation induced by fMLFF (Fig. 7A, compare open and solid triangles after 5 min). A decrease in the number of free singlets in the media (singlets depletion) results

from an increase in the number of aggregates, i.e. aggregation. It is worthwhile noting that we were also able to detect a statistically significant difference between forskolin treated and vehicle (DMSO) treated cells after fMLFF activation (Fig. 7A, compare open triangles and solid circles). Similar data were obtained using $\text{G}\alpha_s$ -coupled receptor activation (data not shown). This suggests that the cAMP pathway was able to reduce the potential of cells to form aggregates in response to $\text{G}\alpha_i$ -coupled receptor activation.

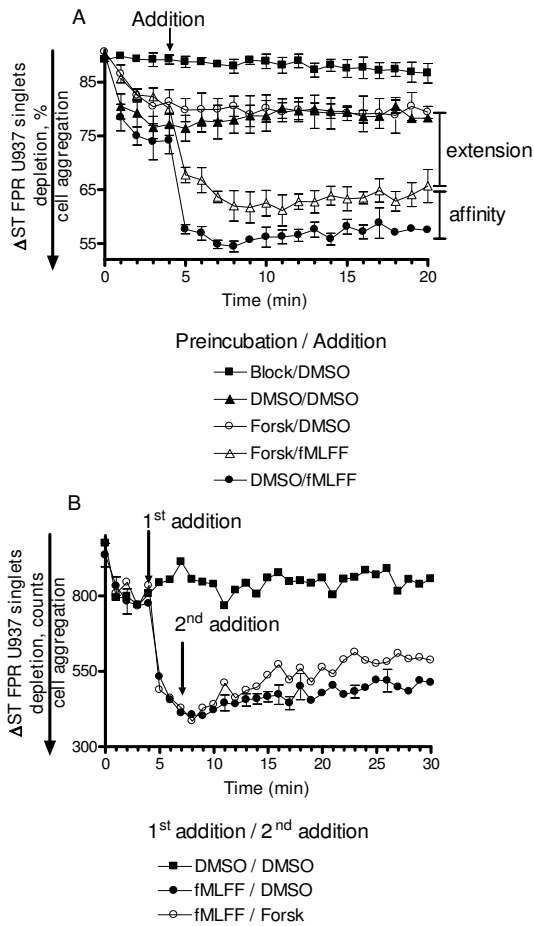


Figure 7
Changes in cell adhesion between U937 FPR (Δ ST) and VCAM-1-transfected B78H1 cells at resting state and in response to receptor stimulation. A, U937 FPR (Δ ST) cells were preincubated for 10 min at 37°C with forskolin (20 μ M), or DMSO (vehicle). Cells were stimulated (arrow) with fMLFF (100 nM), or DMSO (vehicle). Data are plotted as % of U937 FPR (Δ ST) singlets versus time. Five different experimental conditions are shown: Block/DMSO (preincubation with blocking 2 μ M LDV small molecule (addition of DMSO, 1 μ l), DMSO/DMSO, resting state (preincubation with DMSO/addition of DMSO), Forsk/DMSO (preincubation with forskolin/addition of DMSO), Forsk/fMLFF (preincubation with 20 μ M forskolin/stimulation with 100 nM fMLFF), DMSO/fMLFF (preincubation with DMSO, stimulation with 100 nM fMLFF). The non-desensitizing FPR mutant is used to maintain VLA-4 in a state of constant high affinity. Mean and standard error of mean of three independent experiments are shown (n = 3). Bars on the right side of the panel indicate a relative contribution of affinity and extension in the overall cell adhesion. B, Effect of forskolin addition on adhesion in real-time. Cells were stimulated with fMLFF, or DMSO (1st addition). Next, forskolin (20 μ M) was added. Representative experiment out of two experiments is shown.

Since G_{α_s} /cAMP signaling pathway blocks only one aspect of integrin activation, the affinity state of VLA-4 binding pocket, the difference between forskolin treated/fMLFF activated (open triangles) and untreated/fMLFF activated samples (solid circles) appears to be attributable to the impact of the affinity change on cell aggregation. The difference between unstimulated cells (solid triangles) and forskolin pretreated cells after fMLFF stimulation (open triangles) appears to be attributable to molecular unbending (extension).

The effect of G_{α_s} /cAMP signaling pathway activation upon cell aggregation can be also observed in real-time (Fig. 7B). Addition of forskolin to cellular aggregates formed after fMLFF activation resulted in an increase in the number of free U937 singlets (Fig. 7B, compare solid and open circles after second addition). An increase in the number of free singlets in the media is interpreted as a decrease in a number of aggregates, i.e. disaggregation. Thus, rapid down regulation of VLA-4 affinity through G_{α_s} /cAMP signaling pathway plays an important role in the regulation of VLA-4/VCAM-1 specific cell adhesion.

VLA-4 affinity down-modulation can be detected on peripheral blood monocytes

Finally, to confirm that VLA-4 affinity down regulation can be detected in primary cells, we have used purified human peripheral blood monocytes. Monocytes express CXCR4 and FPR receptors. Rapid and transient binding of LDV-FITC can be observed in response to cell activation through these GPCRs [31]. Because, monocytes also express the β_2 -adrenergic receptor, we have used isoproterenol as G_{α_s} -coupled receptor ligand. Experiments were performed in a manner identical to Fig. 1A. Human peripheral blood monocytes pretreated for 2.5 min with saturating isoproterenol had significantly reduced amplitude of LDV-FITC binding response after addition of SDF-1 (Fig. 8A), and fMLFF (Fig. 8B). In addition, pretreatment with isoproterenol affected the resting monocytes, resulting in lower LDV-FITC binding. These results were very similar to U937 cells (compare Fig. 1A and Fig. 8A). Thus, the effect of G_{α_s} -dependent affinity down regulation of VLA-4 can be observed on human peripheral blood monocytes.

As shown previously, the number of α_4 -integrin molecules detected on peripheral blood cells is significantly lower than on the cell lines, which represents a less-differentiated phenotype. On peripheral blood human monocytes we have detected only ~6,000 sites. For U937 cells, we estimated about 40,000–100,000 sites [6]. Therefore, the overall signal from LDV-FITC on monocytes is ~10 fold lower than for U937 cells.

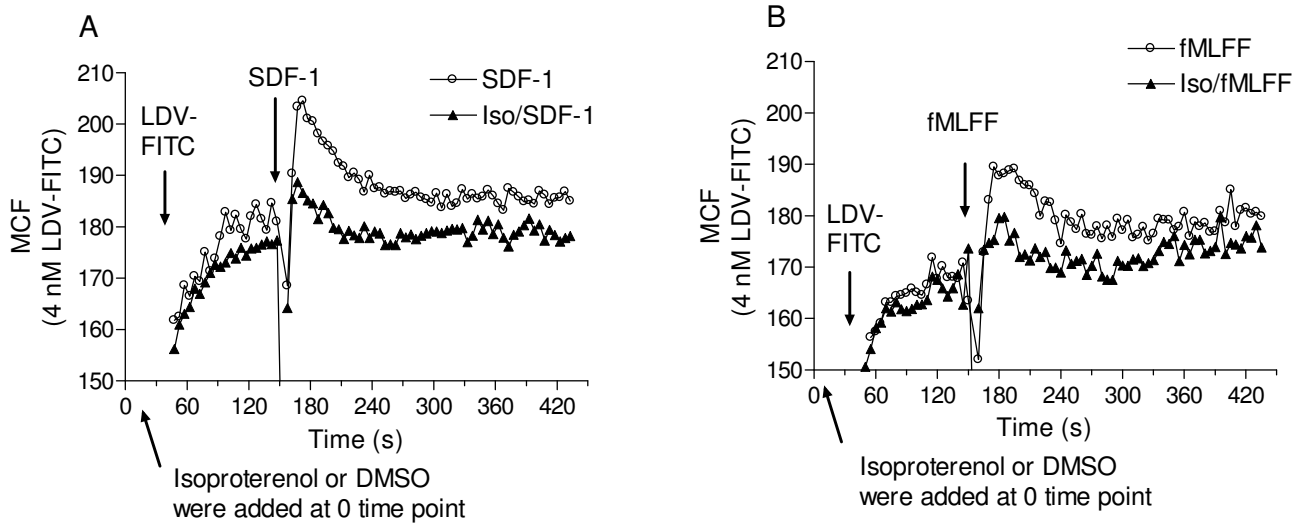


Figure 8

Binding and dissociation of the LDV-FITC probe in response to $G\alpha_s$ -coupled receptor agonist. Experiments were conducted as described under "Methods". A, LDV-FITC probe binding and dissociation on peripheral blood monocytes plotted as mean channel fluorescence (MCF) versus time. The experiment analogous to the one shown in Fig. 1A involved sequential additions of $G\alpha_s$ -coupled receptor ligand (isoproterenol (100 nM)) or DMSO (control), next, fluorescent LDV-FITC probe (4 nM, below saturation, added 2 min prior to addition of $G\alpha_i$ -coupled receptor ligand). Cells were activated with A, SDF-1 (25 nM, arrow), or B, fMLFF (100 nM, arrow). Rapid and reversible binding of the probe reflects a transient VLA-4 affinity change. Curves are means out of four independent runs calculated on a point-by-point basis. One representative experiment out of two independent experiments for each ligand is shown. Experiments shown in the different panels were performed using different instruments, and therefore MCF values are not identical.

Discussion

LDV-FITC probe

The VLA-4 specific fluorescent probe was based on a highly specific $\alpha_4\beta_1$ -integrin inhibitor BIO1211, which contains the Leu-Asp-Val (LDV) ligand binding motif from the alternatively spliced connecting segment-1 (CS-1) peptide of cellular fibronectin [10,32]. Because carboxy-terminal amino acids did not affect ligand binding affinity (see Table 1 in [32]), they were replaced with Pro-Ala-Ala-Lys-FITC. This resulted in a fluorescent probe that we used to study VLA-4 integrin affinity/conformational changes in a conventional flow cytometer. One of the major advantages of this probe is that because of its low molecular weight, diffusion-limited rates of binding allow binding of the probe on the time scale relevant to dynamic cell responses. Therefore, this probe can be used to detect rapid and transient VLA-4-integrin affinity changes in real-time on live cells [6,16,19,20]. We have also used this probe to develop a FRET based assay for the detection of VLA-4 unbending (extension) in real-time after inside-out activation through GPCRs [18-20].

VLA-4 affinity, extension, and cell adhesion

Previously, we established that affinities of the probe and native VLA-4 ligand (VCAM-1) vary in parallel when the

integrin affinity state (natural lifetime) is modulated [16]. This affinity modulation results in a rapid change in cell adhesion efficiency. Moreover, molecular dissociation rates and cellular disaggregation rates in the absence of shear are comparable, suggesting that only few bonds are needed to form cellular aggregates. Therefore, we proposed that the increased aggregation is driven by integrin affinity changes, and not by clustering of the molecules or by increased bond numbers [17]. Moreover, using GPCR-transfected cells, we showed that the time course of avidity changes in response to the $G\alpha_i$ -coupled receptor activation coincides with the time course of the affinity changes. Affinity up-regulation led to rapid cell aggregation [6,16], and down regulation of affinity and extension resulted in the cells disaggregation [20].

Here we show that in addition to affinity up-regulation, the affinity of the VLA-4 binding pocket can be actively down-regulated, and this down regulation is in response to the $G\alpha_s$ -coupled receptors/cAMP signaling pathway. Moreover, affinity down regulation can be seen under continuous signaling conditions, after cell activation through a non-desensitizing mutant of $G\alpha_i$ -coupled GPCR. This mutant is known to produce normal $G\alpha_i$ -coupled signaling, but it cannot be phosphorylated, and thus

it does not desensitize and internalize [33]. As the affinity of the VLA-4 binding pocket regulates the life-time of the VLA-4/VCAM bond [17], down regulation of integrin affinity would decrease the lifetime of the aggregate, and possibly reduce the transition to firm adhesion.

Another aspect of integrin activation is unbending (or extension) of the molecule [12,34]. Here we show that VLA-4 unbending was not inhibited by the $G\alpha_s$ /cAMP signaling pathway. In fact, even in the presence of a high concentration of forskolin, which blocked the VLA-4 affinity change completely, cells were able to respond to fMLFF activation by rapidly aggregating. This increase in cell adhesion is interpreted to arise solely from the unbending of the integrin molecule. Thus, for the first time in this model system we have established conditions under which cell adhesion was up-regulated exclusively due to the extension of the integrin molecule.

The discovery, that affinity of the integrin binding pocket and the molecular unbending are regulated by two different signalling pathways and modulate lifetime of adhesive interaction and cell capture efficiency, has already been documented [20]. We proposed that the low affinity unbent conformation represents an ideal rolling conformation (see Fig. 10B in [20]). We postulated that "several different proteins could participate in the "inside-out" activation of the integrins; some of them would regulate affinity, others – the conformational unbending" [20]. It is possible that affinity regulation by $G\alpha_s$ /cAMP signalling is directly related to Epac/Rap1 pathway, which is reported to affect integrin dependent cell adhesion [35]. For integrin unbending the molecular mechanism is still unclear. However, as shown previously, VLA-4 molecular unbending requires intracellular Ca^{2+} increase [20]. Current data showed that activation via $G\alpha_s$ -coupled GPCRs did not significantly decrease intracellular Ca^{2+} nor affect molecular unbending. This supports our current model of integrin conformation regulation in response to "inside-out" signalling.

Active down regulation of VLA-4 affinity as a possible mechanism of cell trafficking

Cancer cell metastasis, role of NK cells

Tumor cell migration is essential in metastasis. As with hematopoietic progenitor cells, release and homing of certain cancer cells require adhesion molecules and $G\alpha_i$ -coupled GPCRs (such as VLA-4, CXCL12/CXCR4) [7]. Since human tissue microarrays showed that 70–90% of breast, colon, and prostate carcinoma tissues express the β 2-adrenergic receptor [36], these cells may possess the extracellular signaling machinery needed for $G\alpha_i$ / $G\alpha_s$ -coupled to regulate cell adhesion. It has already been shown that β 2-adrenergic receptor agonist activates, and β 2-receptor antagonist blocks migration of prostate, colon and breast

carcinoma cells in vitro [37-39]. In vivo, a β 2-adrenergic receptor agonist significantly increased lymph node metastases, while an antagonist inhibited this effect, tumor growth was not affected by either treatment [36]. Thus, it is possible that down regulation of integrin affinity would significantly change the adhesive properties of tumor cells, facilitate cell migration/de-adhesion and lymphatic drainage, and thereby alter their overall metastatic potential.

NK cells play an important role in tumor cell clearance and resistance of organs to metastases. To effectively kill tumor cells, NK cells must localize at tumor sites. The VLA-4/VCAM-1 interaction is critical for recruitment of NK cells into lung, liver, and tumor sites [40]. As noted above, VLA-4 on NK cells exhibits constitutively high affinity for VCAM-1 as opposed to the low affinity state of VLA-4 on other circulating leukocytes [41,42]. Under stress, or treatment with β -adrenergic receptor agonist, a rapid rise in blood NK cells has been attributed to a reduction of their adherence to endothelia [43-45]. Since up-regulation of integrin affinity is essential for firm adhesion on vascular endothelia [9], affinity down-modulation would dramatically affect proper cell localization or recruitment. Affinity down-regulation could account for the phenomenon of β -adrenoceptor- or catecholamine-dependent suppression of NK cell activity, and a decrease in the overall resistance of an organism to metastasis [46].

Selective recruitment of leukocyte subsets

The multistep paradigm of leukocyte recruitment, where the interplay between selectins, chemokines, GPCRs, and integrins determines the specificity of leukocyte recruitment and trafficking, is broadly accepted [11,47-49]. GPCRs, through $G\alpha_i$ coupled inside-out signaling pathways, rapidly activate integrins, and trigger integrin dependent adhesion. Recently, it has been shown that in addition to pro-adhesive $G\alpha_i$ -coupled GPCRs, anti-adhesive $G\alpha_s$ -coupled receptors can modulate leukocyte transendothelial migration in vitro and recruitment and homing in vivo ([11] and references therein). That rapid affinity down-regulation can account for this modulation enriches our current understanding of selective recruitment of leukocyte subsets. It is worth noting that the same ligand can induce a pro-adhesive signal in one cell type ($G\alpha_i$ -coupled), and at the same time provide an anti-adhesive signal in another cell type ($G\alpha_s$ -coupled). For example, the H4 histamine receptor, selectively expressed on mast cells and eosinophils, is $G\alpha_i$ -coupled, and could serve as a chemotactic receptor for these cell types [50]. The $G\alpha_s$ -coupled H2 histamine receptor expressed on lymphocytes, macrophages and monocytes [51-53] could provide an anti-adhesive signal. Thus, active down-regulation of integrin affinity through $G\alpha_s$ -coupled GPCRs

might be an essential mechanism regulating trafficking and selective recruitment of different leukocyte subsets.

Methods

Materials

The VLA-4 specific ligand [6,16,18] 4-((N'-2-methylphenyl)ureido)-phenylacetyl-L-leucyl-L-aspartyl-L-valyl-L-prolyl-L-alanyl-L-alanyl-L-lysine (LDV containing small molecule), and its FITC-conjugated analog (LDV-FITC) were synthesized at Commonwealth Biotechnologies. Octadecyl rhodamine B chloride (R18), and Indo-1, AM were from Invitrogen. Restriction enzymes were purchased from New England BioLabs. Human recombinant CXCL12/SDF-1 α was from R&D Systems. Mouse anti-human CD14/R-PE antibodies, clone UCHM1, and isotype control mouse IgG2a/R-PE, clone RPC 5 were from Ancell Corp. PE mouse anti-human CXCR4 (CD184 PE) antibodies, clone 12G5, isotype control (mouse IgG2a κ PE) clone G155-178, PE mouse anti-human formyl peptide receptor, clone 5F1, isotype control (mouse IgG1 κ PE) clone MOPC-21, PE mouse anti-human CD49d (α_4 -integrin subunit PE) clone 9F10 were purchased from BD Biosciences and used according to manufacturer instructions. All other reagents were from Sigma-Aldrich. Stock solutions were prepared in DMSO, at concentrations \sim 1000 fold higher than the final concentration. Usually, 1 μ l of stock solution was added to 1 ml of cell suspension yielding a final DMSO concentration of 0.1%. Control samples were treated with equal amount of pure DMSO (vehicle).

Cell Lines and Transfectant Construct

The human histiocytic lymphoma cell line U937 was purchased from ATCC. Wild type CXCR4 (CD184) receptor stably transfected U937 cells, and site-directed mutants of the FPR (non-desensitizing mutant of FPR Δ ST) in U937 cells were prepared as described [54] and were a gift of Dr. Eric Prossnitz (University of New Mexico). High receptor expressing cells were selected using the MoFlo Flow Cytometer (DakoCytomation). Cells were grown at 37°C in a humidified atmosphere of 5% CO₂ and 95% air in RPMI 1640 (supplemented with 2 mM L-glutamine, 100 units/mL penicillin, 100 μ g/mL streptomycin, 10 mM HEPES, pH 7.4, and 10% heat inactivated fetal bovine serum). Cells were then harvested and resuspended in 1 ml of HEPES buffer (110 mM NaCl, 10 mM KCl, 10 mM glucose, 1 mM MgCl₂, 1.5 mM CaCl₂ and 30 mM HEPES, pH 7.4) containing 0.1% HSA and stored on ice. The buffer was depleted of lipopolysaccharide by affinity chromatography over polymyxin B sepharose (Detoxigel; Pierce Scientific). Cells were counted using the Coulter Multisizer/Z2 analyzer (Beckman Coulter). For experiments, cells were suspended in the same HEPES buffer at 1 \times 10⁶ cells/ml and warmed to 37°C. Alternatively, cell

were resuspended in serum/dye-free warm RPMI (37°C) and used immediately.

Purification of human peripheral blood monocytes

Human peripheral blood mononuclear leukocytes were isolated by density gradient centrifugation. Venous blood anticoagulated with heparin or EDTA was obtained from healthy volunteers, and the buffy coat was layered over Ficoll Hypaque (Amersham Biosciences), and centrifuged at 1162 g to obtain mononuclear leukocytes. Leukocytes were washed in 2 mM EDTA in PBS without calcium/magnesium chloride (Gibco), resuspended in RPMI 1640 supplemented with 10% heat-inactivated fetal bovine serum and used immediately. Monocytes were purified from mononuclear leukocytes using AutoMACS automated magnetic cell sorter (Miltenyi Biotec) by negative depletion using magnetic beads with anti-human CD3, CD16, CD19, CD56 specific antibodies (Miltenyi Biotec) according to manufacturer instructions. Purified monocytes were resuspended in HHB buffer supplemented with 0.1% HSA at 1 \times 10⁶ cells/ml and used within 1–3 hours. Monocyte purity was tested using anti-human CD14 PE antibodies (clone UCHM1, Ancell Corp.). For LDV-FITC binding experiments the population of cells was gated on forward vs. side scatter intensity based on CD14⁺ cells.

Kinetic Analysis of Binding and Dissociation of VLA-4 Specific Ligand

Kinetic analysis of the binding and dissociation of the LDV-FITC probe was described previously [6]. Briefly, cells (1 \times 10⁶ cells/ml) were preincubated in HEPES buffer containing 0.1% HSA at different conditions for 10–20 min at 37°C. Flow cytometric data were acquired for up to 1024 s at 37°C while the samples were stirred continuously at 300 rpm with a 5 \times 2 mm magnetic stir bar (Bell-Art Products). Samples were analyzed for 30–120 s to establish a baseline. The fluorescent ligand was added and acquisition was re-established, creating a 5–10 s gap in the time course. For real-time affinity activation experiments, 4 nM LDV-FITC was added after establishing a baseline for unstained cells marked on figures as "autofluorescence". Then, data were acquired for 2–3 minutes, and cells were treated with different GPCR ligands at saturating concentration (10 times or higher than K_d). In several experiments cells were treated sequentially with two or more different compounds. Acquisition was re-established, and data were acquired continuously for up to 1024 s. The concentration of the LDV-FITC probe used in the experiments (4 nM) was below the dissociation constant (K_d) for its binding to resting VLA-4 (low affinity state, K_d \sim 12 nM), and above the K_d for physiologically activated VLA-4 (high affinity state, K_d \sim 1–2 nM) [6]. Therefore, the transition from the low affinity to the high affinity receptor state led to increased binding of the probe (from \sim 25% to \sim 70–80% of receptor occupancy, as calculated based on the

one site binding equation), which was detected as an increase in the mean channel fluorescence (MCF). For kinetic dissociation measurements, cell samples were pre-incubated with the fluorescent probe (25 nM), treated with excess unlabeled LDV containing small molecule (2 μ M) and the dissociation of the fluorescent molecule was followed. The resulting data were converted to MCF *versus* time using FCSQuery software developed by Dr. Bruce Edwards (University of New Mexico).

GPCR ligand concentrations were chosen based upon known binding affinities [55-60]. Ligands were used at the concentrations of two or more fold higher than respective dissociation constants (K_d). This results in more than 66% coverage of the receptor sites. For G α_s -coupled receptor binding site competition experiments (Fig. 2A,B) lower agonist concentrations were chosen to limit potential rebinding after addition of a large excess of antagonist (20 fold or more). The concentrations of antagonists were chosen based on their respective affinities, to cover more than 99% of receptor binding sites (200 fold greater than K_d). By themselves, these high concentrations of G α_s -coupled receptor antagonists did not affect cells activation after stimulation by G α_i -coupled receptor agonists (see Fig. 2B, black line).

FRET Detection of VLA-4 Unbending (Extension)

The fluorescence resonance energy transfer (FRET) assay used the LDV-FITC probe as a donor, which specifically binds to the α_4 -integrin headgroup, and octadecyl rhodamine B (R18) as an acceptor incorporated into the plasma membrane was previously described in [18] and validated in several publications [19,20]. Briefly, U937 cells stably transfected with CXCR4 or the non-desensitizing mutant (Δ ST) of the formyl peptide receptor [33,61], were preincubated with 100 nM LDV-FITC probe in HEPES buffer containing 1.5 mM CaCl₂, 1 mM MgCl₂, and 0.1% HSA at 37°C. Alternatively, 100 nM LDV-FITC was added after establishing a baseline for unstained cells (autofluorescence). Next, samples were analyzed for 1–3 minutes to establish a baseline for 100 nM LDV-FITC, and then a saturating amount of octadecyl rhodamine B (R18, 10 μ M final) was added to yield maximal quenching of donor fluorescence. 1–3 min after R18 was added; cells were treated with different GPCR ligands at saturating concentration (10 times or higher than K_d). In several experiments cells were treated sequentially with two or more different compounds as in the LDV-FITC probe binding experiments. Donor intensities (FL1 in MCF units) were measured using a Becton-Dickinson FACScan flow cytometer at 37°C. Changes in the donor fluorescence intensity were interpreted as changes in the distance of closest approach between LDV-FITC ligand binding site on VLA-4 and the surface of the plasma membrane [18,19].

Calcium Mobilization

Calcium mobilization in response to ligand stimulation was measured as described previously [61]. Briefly cells were harvested by centrifugation, washed, and resuspended at 5 \times 10⁶ cells/ml in HEPES buffer containing 1.5 mM CaCl₂, 1 mM MgCl₂. Next, cells were incubated with 5 μ M Indo-1, AM for 30 min at 37°C, washed, and resuspended at 1 \times 10⁶ cells/ml in HEPES buffer, and stored on ice. Cells were allowed to equilibrate at 37°C for 2 min, stimulated with different GPCR ligands, and continuous fluorescence was monitored using a spectrofluorometer (Photon Technologies International). Excitation at 350 nm, detection at 405 nm and 490 nm. Fluorescence intensity ratio (405 nm/490 nm) was computed in real-time as a measure of intracellular Ca²⁺.

Cell Adhesion Assay

The cell suspension adhesion assay has been described previously [16]. Briefly, U937/ Δ ST FPR stably transfected cells were labeled with red fluorescent PKH26GL dye, and B78H1/VCAM-1 transfectants were stained with green fluorescent PKH67GL dye (Sigma-Aldrich). Labeled cells were washed, resuspended in HEPES buffer supplemented with 0.1% HSA and stored on ice until used in assays. Control U937 cells were preincubated with the LDV-containing small molecule for blocking. Prior to data acquisition, cells were warmed to 37°C for 10 min separately and then mixed. During data acquisition, the samples were stirred with a 5 \times 2-mm magnetic stir bar (Bel-Art Products, Pequannock, NJ) at 300 rpm and kept at 37°C. For stimulation, cells were treated with different GPCR ligands at saturating concentration (10 times or higher than K_d) as described in "Kinetic Analysis of Binding and Dissociation of VLA-4 Specific Ligand". In several experiments cells were treated sequentially with two different compounds. The number of cell aggregates containing U937 adherent to B78H1/VCAM-1 (red and green co-fluorescent particles) as well as a number of singlets (red or green fluorescent particles, FL2 and FL1 in FACScan flow cytometer) were followed in real time. As shown previously, cell aggregation as detected using cell aggregates (red and green co-fluorescent particles) did not discriminate between doublets and higher order aggregates [17]. Therefore, the analysis of cell aggregation was done by using singlet cell depletion methodology [62]. The percentage of singlets was calculated as follows: singlets depletion, % = (number of singlets/(number of aggregates + number of singlets)) \times 100. Experiments were done using a FACScan flow cytometer and Cell Quest software (Becton Dickinson, San Jose, CA). The data were converted to number of singlets/aggregates *versus* time using FCSQuery software developed by Dr. Bruce Edwards (University of New Mexico).

Statistical analysis

Curve fits and statistics were performed using GraphPad Prism (GraphPad). Each experiment was repeated at least three times. The experimental curves represent the mean of two or more independent runs. SEM was calculated using GraphPad Prism.

Abbreviations

cAMP (adenosine 3',5'-cyclophosphate), dbcAMP (N-6,2'-O-dibutyryladenine 3',5'-cyclic monophosphate), fMLFF (N-formyl-L-methionyl-L-leucyl-L-phenylalanyl-L-phenylalanine, formyl peptide), FPR (formyl peptide receptor 1), FRET (fluorescence resonance energy transfer), GPCR (guanine nucleotide binding protein coupled receptor), HSA (human serum albumin), HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), LDV containing small molecule (4-((N'-2-methylphenyl)ureido)-phenylacetyl-L-leucyl-L-aspartyl-L-valyl-L-prolyl-L-alanyl-L-alanyl-L-lysine), LDV-FITC containing small molecule (4-((N'-2-methylphenyl)ureido)-phenylacetyl-L-leucyl-L-aspartyl-L-valyl-L-prolyl-L-alanyl-L-alanyl-L-lysine-FITC), mAb (monoclonal antibody) LFA-1 (lymphocyte function-associated antigen-1, CD11a/CD18, $\alpha_1\beta_2$ integrin), MCF (mean channel fluorescence, equivalent of mean fluorescence intensity), NK cell (natural killer cells), SDF-1 (stromal cell-derived factor-1, CXCL12), VCAM-1 (vascular cell adhesion molecule 1, CD106), VLA-4 (very late antigen 4, CD49d/CD29, $\alpha_4\beta_1$ integrin).

Authors' contributions

AC designed the study, carried out ligand binding experiments, cell adhesion studies, and wrote the manuscript. AW carried out β_2 -adrenergic receptor experiments, participated in study design. OA performed antibody staining experiments and helped with monocytes experiments. LAS contributed to the study concept, and participated in study design and coordination. All authors read and approved the final manuscript.

Acknowledgements

We thank Eric R. Prossnitz for providing U937 cells and plasmids, and Bruce S. Edwards for providing FCSQuery software. This work was supported by National Institutes of Health Grants HL56384, MH074425, CA118100, and HL081062 (to L.A.S.), Leukemia and Lymphoma Society Grant 7388-06 (to L.A.S.), and by Dedicated Health Research Funds of the University of New Mexico School of Medicine grant C-2297-RAC (to A.C.). We are grateful for the peripheral blood monocytes preparation by Cheryl A. Miller and Elizabeth N. McKown from Dr. C. Rick Lyons' laboratory at the University of New Mexico Cancer Center.

References

- Hynes RO: **Integrins: bidirectional, allosteric signaling machines.** *Cell* 2002, **110**:673-687.
- Cai W, Chen X: **Anti-angiogenic cancer therapy based on integrin $\alpha_5\beta_3$ antagonism.** *Anticancer Agents Med Chem* 2006, **6**:407-428.

- De Meyer SF, Vanhoorelbeke K, Ulrichts H, Staelens S, Feys HB, Salles I, Fontayne A, Deckmyn H: **Development of monoclonal antibodies that inhibit platelet adhesion or aggregation as potential anti-thrombotic drugs.** *Cardiovasc Hematol Disord Drug Targets* 2006, **6**:191-207.
- Hilden TJ, Nurmi SM, Fagerholm SC, Gahmberg CG: **Interfering with leukocyte integrin activation--a novel concept in the development of anti-inflammatory drugs.** *Ann Med* 2006, **38**:503-511.
- O'Connor P: **Natalizumab and the role of alpha 4-integrin antagonism in the treatment of multiple sclerosis.** *Expert Opin Biol Ther* 2007, **7**:123-136.
- Chigaev A, Blenc AM, Braaten JV, Kumaraswamy N, Kepley CL, Andrews RP, Oliver JM, Edwards BS, Prossnitz ER, Larson RS, Sklar LA: **Real time analysis of the affinity regulation of alpha 4-integrin. The physiologically activated receptor is intermediate in affinity between resting and Mn(2+) or antibody activation.** *J Biol Chem* 2001, **276**:48670-48678.
- Gazit Y: **Homing and mobilization of hematopoietic stem cells and hematopoietic cancer cells are mirror image processes, utilizing similar signaling pathways and occurring concurrently: circulating cancer cells constitute an ideal target for concurrent treatment with chemotherapy and antineoplastic-specific antibodies.** *Leukemia* 2004, **18**:1-10.
- Lapidot T, Petit I: **Current understanding of stem cell mobilization: the roles of chemokines, proteolytic enzymes, adhesion molecules, cytokines, and stromal cells.** *Exp Hematol* 2002, **30**:973-981.
- Feigelson SW, Grabovsky V, Winter E, Chen LL, Pepinsky RB, Yednock T, Yablonski D, Lobb R, Alon R: **The Src kinase p56(lck) up-regulates VLA-4 integrin affinity. Implications for rapid spontaneous and chemokine-triggered T cell adhesion to VCAM-1 and fibronectin.** *J Biol Chem* 2001, **276**:13891-13901.
- Chen LL, Whitty A, Lobb RR, Adams SP, Pepinsky RB: **Multiple activation states of integrin $\alpha_4\beta_1$ detected through their different affinities for a small molecule ligand.** *J Biol Chem* 1999, **274**:13167-13175.
- Laudanna C, Kim JY, Constantin G, Butcher E: **Rapid leukocyte integrin activation by chemokines.** *Immunol Rev* 2002, **186**:37-46.
- Beglova N, Blacklow SC, Takagi J, Springer TA: **Cysteine-rich module structure reveals a fulcrum for integrin rearrangement upon activation.** *Nat Struct Biol* 2002, **9**:282-287.
- Laudanna C, Alon R: **Right on the spot. Chemokine triggering of integrin-mediated arrest of rolling leukocytes.** *Thromb Haemost* 2006, **95**:5-11.
- Takagi J, Petre BM, Walz T, Springer TA: **Global conformational rearrangements in integrin extracellular domains in outside-in and inside-out signaling.** *Cell* 2002, **110**:599-11.
- Garcia A, Serrano A, Abril E, Jimenez P, Real LM, Canton J, Garrido F, Ruiz-Cabello F: **Differential effect on U937 cell differentiation by targeting transcriptional factors implicated in tissue- or stage-specific induced integrin expression.** *Exp Hematol* 1999, **27**:353-364.
- Chigaev A, Zwart G, Graves SW, Dwyer DC, Tsuji H, Foutz TD, Edwards BS, Prossnitz ER, Larson RS, Sklar LA: **Alpha4beta1 integrin affinity changes govern cell adhesion.** *J Biol Chem* 2003, **278**:38174-38182.
- Zwart G, Chigaev A, Foutz T, Larson RS, Posner R, Sklar LA: **Relationship between Molecular and Cellular Dissociation Rates for VLA-4/VCAM-1 Interaction in the Absence of Shear Stress.** *Biophys J* 2004, **86**:1243-1252.
- Chigaev A, Buranda T, Dwyer DC, Prossnitz ER, Sklar LA: **FRET detection of cellular alpha4-integrin conformational activation.** *Biophys J* 2003, **85**:3951-3962.
- Chigaev A, Zwart G, Buranda T, Edwards BS, Prossnitz ER, Sklar LA: **Conformational regulation of alpha 4 beta 1-integrin affinity by reducing agents. "Inside-out" signaling is independent of and additive to reduction-regulated integrin activation.** *J Biol Chem* 2004, **279**:32435-32443.
- Chigaev A, Waller A, Zwart G, Buranda T, Sklar LA: **Regulation of cell adhesion by affinity and conformational unbending of alpha4beta1 integrin.** *J Immunol* 2007, **178**:6828-6839.
- Larson RS, Davis T, Bologna C, Semenuk G, Vijayan S, Li Y, Oprea T, Chigaev A, Buranda T, Wagner CR, Sklar LA: **Dissociation of I domain and global conformational changes in LFA-1: refine-**

- ment of small molecule-I domain structure-activity relationships. *Biochemistry* 2005, **44**:4322-4331.
22. Tecoma ES, Motulsky HJ, Traynor AE, Omann GM, Muller H, Sklar LA: **Transient catecholamine modulation of neutrophil activation: kinetic and intracellular aspects of isoproterenol action.** *J Leukoc Biol* 1986, **40**:629-644.
 23. Hughes SA, Gibson WJ, Young JM: **The interaction of U-73122 with the histamine H1 receptor: implications for the use of U-73122 in defining H1 receptor-coupled signalling pathways.** *Naunyn Schmiedeberg Arch Pharmacol* 2000, **362**:555-558.
 24. Liggett SB: **Beta-adrenoceptor-effector system of the human macrophage U937 cell line.** *Eur J Pharmacol* 1989, **163**:171-174.
 25. Shayo C, Davio C, Brodsky A, Mladovan AG, Legnazzi BL, Rivera E, Baldi A: **Histamine modulates the expression of c-fos through cyclic AMP production via the H2 receptor in the human promonocytic cell line U937.** *Mol Pharmacol* 1997, **51**:983-990.
 26. van der GH, Timmerman H: **Selective ligands as tools to study histamine receptors.** *Eur J Med Chem* 2000, **35**:5-20.
 27. Zhang L, Jensen RT, Maton PN: **Characterization of beta-adrenoceptors on smooth muscle cells from guinea pig stomach.** *Am J Physiol* 1990, **259**:G436-G442.
 28. Sklar LA, Omann GM, Painter RG: **Relationship of actin polymerization and depolymerization to light scattering in human neutrophils: dependence on receptor occupancy and intracellular Ca⁺⁺.** *J Cell Biol* 1985, **101**:1161-1166.
 29. Torphy TJ, Zhou HL, Cieslinski LB: **Stimulation of beta adrenoceptors in a human monocyte cell line (U937) up-regulates cyclic AMP-specific phosphodiesterase activity.** *J Pharmacol Exp Ther* 1992, **263**:1195-1205.
 30. Salas A, Shimaoka M, Kogan AN, Harwood C, von Andrian UH, Springer TA: **Rolling adhesion through an extended conformation of integrin alphaLbeta2 and relation to alpha I and beta I-like domain interaction.** *Immunity* 2004, **20**:393-406.
 31. Chan JR, Hyduk SJ, Cybulsky MI: **Detecting rapid and transient upregulation of leukocyte integrin affinity induced by chemokines and chemoattractants.** *J Immunol Methods* 2003, **273**:43-52.
 32. Lin K, Ateeq HS, Hsiung SH, Chong LT, Zimmerman CN, Castro A, Lee WC, Hammond CE, Kalkunte S, Chen LL, Pepinsky RB, Leone DR, Sprague AG, Abraham WM, Gill A, Lobb RR, Adams SP: **Selective, tight-binding inhibitors of integrin alpha4beta1 that inhibit allergic airway responses.** *J Med Chem* 1999, **42**:920-934.
 33. Prossnitz ER: **Desensitization of N-formylpeptide receptor-mediated activation is dependent upon receptor phosphorylation.** *J Biol Chem* 1997, **272**:15213-15219.
 34. Takagi J, Springer TA: **Integrin activation and structural rearrangement.** *Immunol Rev* 2002, **186**:141-163.
 35. Bos JL, de Bruyn K, Enserink J, Kuiperij B, Rangarajan S, Rehmann H, Riedl J, de Rooij J, van Mansfeld F, Zwartkruis F: **The role of Rap1 in integrin-mediated cell adhesion.** *Biochem Soc Trans* 2003, **31**:83-86.
 36. Palm D, Lang K, Niggemann B, Drell TL, Masur K, Zaenker KS, Entschladen F: **The norepinephrine-driven metastasis development of PC-3 human prostate cancer cells in BALB/c nude mice is inhibited by beta-blockers.** *Int J Cancer* 2006, **118**:2744-2749.
 37. Drell TL, Joseph J, Lang K, Niggemann B, Zaenker KS, Entschladen F: **Effects of neurotransmitters on the chemokinesis and chemotaxis of MDA-MB-468 human breast carcinoma cells.** *Breast Cancer Res Treat* 2003, **80**:63-70.
 38. Lang K, Drell TL, Lindecke A, Niggemann B, Kaltschmidt C, Zaenker KS, Entschladen F: **Induction of a metastatogenic tumor cell type by neurotransmitters and its pharmacological inhibition by established drugs.** *Int J Cancer* 2004, **112**:231-238.
 39. Masur K, Niggemann B, Zanker KS, Entschladen F: **Norepinephrine-induced migration of SW 480 colon carcinoma cells is inhibited by beta-blockers.** *Cancer Res* 2001, **61**:2866-2869.
 40. Fogler WE, Volker K, McCormick KL, Watanabe M, Ortaldo JR, Wiltrot RH: **NK cell infiltration into lung, liver, and subcutaneous B16 melanoma is mediated by VCAM-1/VLA-4 interaction.** *J Immunol* 1996, **156**:4707-4714.
 41. Rose DM, Cardarelli PM, Cobb RR, Ginsberg MH: **Soluble VCAM-1 binding to alpha4 integrins is cell-type specific and activation dependent and is disrupted during apoptosis in T cells.** *Blood* 2000, **95**:602-609.
 42. Hyduk SJ, Oh J, Xiao H, Chen M, Cybulsky MI: **Paxillin selectively associates with constitutive and chemoattractant-induced high-affinity alpha4beta1 integrins: implications for integrin signaling.** *Blood* 2004, **104**:2818-2824.
 43. Shakhbar G, Ben Elyahu S: **In vivo beta-adrenergic stimulation suppresses natural killer activity and compromises resistance to tumor metastasis in rats.** *J Immunol* 1998, **160**:3251-3258.
 44. Benschop RJ, Nijkamp FP, Ballieux RE, Heijnen CJ: **The effects of beta-adrenoceptor stimulation on adhesion of human natural killer cells to cultured endothelium.** *Br J Pharmacol* 1994, **113**:1311-1316.
 45. Schedlowski M, Hosch W, Oberbeck R, Benschop RJ, Jacobs R, Raab HR, Schmidt RE: **Catecholamines modulate human NK cell circulation and function via spleen-independent beta 2-adrenergic mechanisms.** *J Immunol* 1996, **156**:93-99.
 46. Ben Elyahu S, Shakhbar G, Page GG, Stefanski V, Shakhbar K: **Suppression of NK cell activity and of resistance to metastasis by stress: a role for adrenal catecholamines and beta-adrenoceptors.** *Neuroimmunomodulation* 2000, **8**:154-164.
 47. Kunkel EJ, Butcher EC: **Chemokines and the tissue-specific migration of lymphocytes.** *Immunity* 2002, **16**:1-4.
 48. Springer TA: **Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm.** *Cell* 1994, **76**:301-314.
 49. Weber C: **Novel mechanistic concepts for the control of leukocyte transmigration: specialization of integrins, chemokines, and junctional molecules.** *J Mol Med* 2003, **81**:4-19.
 50. Hofstra CL, Desai PJ, Thurmond RL, Fung-Leung WP: **Histamine H4 receptor mediates chemotaxis and calcium mobilization of mast cells.** *J Pharmacol Exp Ther* 2003, **305**:1212-1221.
 51. Mirossay L, Chastre E, Callebort J, Launay JM, Housset B, Zimmer A, Abita JP, Gespach C: **Histamine H2 receptors and histidine decarboxylase in normal and leukemic human monocytes and macrophages.** *Am J Physiol* 1994, **267**:R602-R611.
 52. Sachs B, Hertl M, Merk HF: **Histamine receptors on lymphocytes: distribution and functional significance.** *Skin Pharmacol Appl Skin Physiol* 2000, **13**:313-323.
 53. Smit MJ, Leurs R, Shukrula SR, Bast A, Timmerman H: **Rapid desensitization of the histamine H2 receptor on the human monocyte cell line U937.** *Eur J Pharmacol* 1994, **288**:17-25.
 54. Kew RR, Peng T, DiMartino SJ, Madhavan D, Weinman SJ, Cheng D, Prossnitz ER: **Undifferentiated U937 cells transfected with chemoattractant receptors: a model system to investigate chemotactic mechanisms and receptor structure/function relationships.** *J Leukoc Biol* 1997, **61**:329-337.
 55. Booze RM, Crisostomo EA, Davis JN: **Species differences in the localization and number of CNS beta adrenergic receptors: rat versus guinea pig.** *J Pharmacol Exp Ther* 1989, **249**:911-920.
 56. Ezeamuzie CI, Phillips E: **Histamine H(2) receptors mediate the inhibitory effect of histamine on human eosinophil degranulation.** *Br J Pharmacol* 2000, **131**:482-488.
 57. Foreman JC, Norris DB, Rising TJ, Webber SE: **A study of the H2-receptor for histamine stimulating adenylate cyclase in homogenates of guinea-pig lung parenchyma.** *Br J Pharmacol* 1986, **87**:37-44.
 58. Freer RJ, Day AR, Muthukumaraswamy N, Pinon D, Wu A, Showell HJ, Becker EL: **Formyl peptide chemoattractants: a model of the receptor on rabbit neutrophils.** *Biochemistry* 1982, **21**:257-263.
 59. Green SA, Cole G, Jacinto M, Innis M, Liggett SB: **A polymorphism of the human beta 2-adrenergic receptor within the fourth transmembrane domain alters ligand binding and functional properties of the receptor.** *J Biol Chem* 1993, **268**:23116-23121.
 60. Ueda H, Siani MA, Gong W, Thompson DA, Brown GG, Wang JM: **Chemically synthesized SDF-1alpha analogue, N33A, is a potent chemotactic agent for CXCR4/Fusin/LESTR-expressing human leukocytes.** *J Biol Chem* 1997, **272**:24966-24970.
 61. Hsu MH, Chiang SC, Ye RD, Prossnitz ER: **Phosphorylation of the N-formyl peptide receptor is required for receptor internalization but not chemotaxis.** *J Biol Chem* 1997, **272**:29426-29429.
 62. Edwards BS, Kuckuck FW, Prossnitz ER, Okun A, Ransom JT, Sklar LA: **Plug flow cytometry extends analytical capabilities in cell adhesion and receptor pharmacology.** *Cytometry* 2001, **43**:211-216.