Progress in the mechanism of radiation-induced lung injury

Hang-Jie Ying^{1,2,3}, Min Fang^{1,2,3}, Ming Chen^{1,2,3}

¹Department of Radiation Oncology, Cancer Hospital of the University of Chinese Academy of Sciences (Zhejiang Cancer Hospital), Hangzhou, Zhejiang 310022, China;

³Zhejiang Key Laboratory of Radiation Oncology, Zhejiang Cancer Hospital, Hangzhou, Zhejiang 310022, China.

Radiation therapy is widely used to treat various thoracic tumors. However, X-rays inevitably cause damage to normal lung tissues while killing tumor cells, leading to the occurrence of radiation-induced lung injury (RILI). Recent data showed that lung cancer has the highest incidence of RILI (5–25%), followed by mediastinal lymphoma (5–10%), and breast cancer (1-5%).^[1] With the progress in research, our understanding of the mechanism of RILI has changed from the traditional hypothesis of "target cell death" to "a continuous process involving multiple cells," which is dynamic and evolving [Figure 1]. In the present review, we summarize the current knowledge of RILI, and discuss the potential limitations of combined radio-immunotherapy.

Single-nucleotide polymorphisms (SNPs) are the most common human heritable variations, accounting for more than 90% of all known polymorphisms.^[2] Studies have found that SNPs are associated with susceptibility to RILI. It was reported that lung cancer patients with the CT+TT genotype of interleukin (IL)-4 SNP rs2243250 had a lower risk of developing radiation pneumonia \geq grade 3 after radiotherapy.^[3] However, patients with autophagy related 16 like 2 (ATG16L2) rs10898880 CC variant genotype had a higher risk.^[4]

Inherited illness caused by gene mutations will also lead to an increase in sensitivity to radiotherapy, such as ataxia telangiectasia and Nijmegen breakage syndrome, which are caused by the mutation of ataxia telangiectasia mutated (ATM) and Nijmegen breakage syndrome 1, respectively. These patients will have severe reactions even if they are exposed to a small dose of radiation.^[5,6]

Large amounts of reactive oxygen species (ROS) are produced rapidly in lung tissue after irradiation, which is mainly caused by ionization of water molecules. ROS

Access this article online	
Quick Response Code:	Website: www.cmj.org
	DOI: 10.1097/CM9.000000000001032

produced by X-ray irradiation disappear in a very short time. However, Hao *et al*^[7] found that a significant decrease of blood oxygen partial pressure could be detected 180 days after lung irradiation with 15 Gy in dogs. Similarly, Yin *et al*^[8] found that ROS showed dynamic changes after a single dose of 18 Gy irradiation on the right lung of dogs, which reached a peak after 4 weeks of irradiation. Those results support the theory that radiation-induced chronic oxidative stress occurs in the lungs. This process involves a variety of cytokines, cell types, and gene products. These secondary productions of large amounts of ROS deplete the body's antioxidant capacity rapidly, further aggravating tissue damage and allowing lung lesions to persist over time.

The alveolar capillary barrier, which comprises vascular endothelial cells (VECs) and alveolar epithelial cells (AECs), is very sensitive to ionizing radiation. Ionizing radiation can cause the increase of vascular permeability and inflammatory infiltration of VECs in of the early stage. With extended irradiation time and increased irradiation dose, VECs will rupture and fall off, accompanied by platelet attachment, resulting in capillary embolism. Type I AECs lack proliferative ability, and undergo necrosis or apoptosis directly after irradiation. Type II AECs (AEC IIs) activate fibroblasts through epithelial-mesenchymal transition, and then differentiate into myofibroblasts, which then form characteristic fibroblast foci and secrete the extracellular matrix, ultimately leading to fibrosis. The abnormal proliferation of AEC IIs also reduces the secretion of alveolar surfactant, which decreases the alveolar surface tension, resulting in pulmonary edema and atelectasis.^[9]

The essence of RILI is lymphocytic alveolitis, which is closely related to the immune response mediated by lymphocytes. According to the different cytokines secreted, $CD4^+$ T cells can be divided into T_h1 and T_h2 cells. T_h1 cells mainly mediate cellular immune response by secreting T_h1 cytokines, such as interferon gamma (IFN- γ), tumor

Correspondence to: Dr. Ming Chen, No. 1 East Banshan Road, Gongshu District, Hangzhou, Zhejiang 310022, China

E-Mail: chenming@zjcc.org.cn

Copyright © 2020 The Chinese Medical Association, produced by Wolters Kluwer, Inc. under the CC-BY-NC-ND license. This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Chinese Medical Journal 2021;134(2) Received: 12-03-2020 Edited by: Pei-Fang Wei

²Institute of Cancer and Basic Medicine (IBMC), Chinese Academy of Sciences, Hangzhou, Zhejiang 310022, China;



necrosis factor alpha, IL-2, and IL-12. T_h2 cells mainly mediate the humoral immune response by secreting T_h2 cytokines, such as IL-4, IL-5, IL-13, and monocyte chemoattractant protein-1. The imbalance of immune function of T_h1/T_h2 cells is one of the important pathogeneses of RILI. For example, compared with C57BL/6J mice, the fibrotic response of IFN- γ -/- mice, deficient in T_h1 cells, was significantly enhanced.^[10] IL-4 levels increased in the lungs of mice within 3 weeks after irradiation, and inhibiting the expression of IL-4 could effectively inhibit the progress of radiation-induced lung fibrosis (RILF).^[11]

Macrophages play a key role in the development of RILI because they are the first line of defense against external invasion. Macrophages can be divided into two types: T_h 1-derived cytokine IFN- γ can promote the expression of inducible nitric oxide synthase, which is a marker for classically activated macrophages (CAMs or M1). T_h 2-derived cytokines IL-4 and IL-13 can active the activity of arginase-1 (Arg-1), which is a marker for alternatively activated macrophages (AAMs or M2). M1 macrophages have been shown to prevent the development

of pulmonary fibrosis, and M2 macrophages are the most prominent type of macrophages in pulmonary fibrosis. Therefore, the balance transition from M1 macrophages that promote inflammation to M2 macrophages that promote fibrosis and wound healing is one of the important reasons for the formation of pulmonary fibrosis after radiation.

Macrophages can also be divided into alveolar macrophages (AMs) and interstitial macrophages (IMs), according to their different anatomical sites. Chen *et al*^[12] found that AMs are the primary inflammatory cells that infiltrate irradiated lung tissues, and a single high dose of chest irradiation led to the depletion of AMs, but not IMs. Moreover, AMs possibly reflect a more pro-inflammatory phenotype. IMs are mainly distributed in the alveolar septum, bronchus, and blood vessels, and play an important role in the regulation of Arg-1 was found in IMs isolated from lung tissue of RILF mice. IMs, but not AMs, were able to induce myofibroblast activation *in vitro*.^[13]

Radiotherapy can induce immunogenic death of tumor cells, expose tumor antigens, produce damage-associated molecular patterns, and promote the maturation of dendritic cells (DCs). The phenotype of mature DCs changed and migrated to local draining lymph nodes. Tumor antigens are delivered to CD8⁺ T cells through major histocompatibility complex I (MHC I), and CD8⁺ T cells are activated to differentiate into cytotoxic T lymphocytes (CTLs). These tumor-specific CTLs enter the blood circulation and home to the tumor, specifically identifying all cells expressing similar antigens, mediating target cell death through perforin, granzyme, and other pathways, then removing tumors inside and outside the radiotherapy field.^[14] Radiotherapy combined with immunotherapy can regulate and magnify this process. In the model of spontaneous lung metastasis and double leg transplanted tumor in mice, Vanpouille-Box et al^[15] found that radiotherapy combined with transforming growth factor- β (TGF- β) inhibitor could produce abscopal effect, which may be related to the promotion of DCs activation and enhancement of T-cell immune response to tumorspecific antigen after the inhibition of TGF-β. However, uncontrolled pro-inflammatory signals induced by immunotherapy alone can cause toxic effects in normal tissues, such as immune-related adverse effects, which often occur in liver, heart, lung, and so on. Radiotherapy combined with immunotherapy increases the risk of radiation-induced side effects, including RILI.^[16] Therefore, radiotherapy combined with immunotherapy must be regarded as a "double-edged sword" that needs to be handled carefully. How to choose the treatment time, treatment sequence and dose to achieve the best effect of combined therapy needs further research.

Clinically, corticosteroids are mainly used to treat radiation pneumonitis, and early intervention is expected to lead to recovery. Antioxidant drugs have also proven to prevent and treat radiation pneumonitis. For example, amifostine, which scavenges tissue-free radicals and has antioxidation effects, shows a good protective effect on radiation damage in the human body, and was the first broad-spectrum cell protector recognized by international authorities.^[17] Although the risk of RILF can be predicted by detecting many blood biochemical indexes, such as TGF-β, IL-6, krebs von den lungen-6 (KL-6), surfactant protein, and interleukin-1 receptor antagonist (IL-1ra),^[18] most drugs for RILF are still undergoing preclinical research. Thus, it could be said that there are no effective treatments for RILF. Pirfenidone and nintedanib have been approved by FDA for the treatment of idiopathic pulmonary fibrosis. The pathological features of idiopathic pulmonary fibrosis and RILF are very similar, and both go through the process of early inflammation, lung parenchyma injury, damaged alveolar repair, and fibrosis.^[19] At present, clinical trials on the effects of pirfenidone (NCT03902509) and nintedanib (NCT02496585) on RILI are ongoing. We expect that these drugs will have promising clinical prospects in the management of RILI.

Conflicts of interest

None.

References

- Hanania AN, Mainwaring W, Ghebre YT, Hanania NA, Ludwig M. Radiation-induced lung injury: assessment and management. Chest 2019;156:150–162. doi: 10.1016/j.chest.2019.03.033.
- Chaudhary R, Singh B, Kumar M, Gakhar SK, Saini AK, Parmar VS, et al. Role of single nucleotide polymorphisms in pharmacogenomics and their association with human diseases. Drug Metab Rev 2015;47:281–290. doi: 10.3109/03602532.2015.1047027.
- Tang Y, Yang L, Qin W, Yi M, Liu B, Yuan X. Validation study of the association between genetic variant of IL4 and severe radiation pneumonitis in lung cancer patients treated with radiation therapy. Radiother Oncol 2019;141:86–94. doi: 10.1016/j.radonc.2019.09.002.
- Wen J, Liu H, Wang L, Wang X, Gu N, Liu Z, *et al.* Potentially functional variants of ATG16L2 predict radiation pneumonitis and outcomes in patients with non-small cell lung cancer after definitive radiotherapy. J Thorac Oncol 2018;13:660–675. doi: 10.1016/j. jtho.2018.01.028.
- Amirifar P, Ranjouri MR, Yazdani R, Abolhassani H, Aghamohammadi A. Ataxia-telangiectasia: a review of clinical features and molecular pathology. Pediatr Allergy Immunol 2019;30:277–288. doi: 10.1111/pai.13020.
- Chrzanowska KH, Gregorek H, Dembowska-Bagińska B, Kalina MA, Digweed M. Nijmegen breakage syndrome (NBS). Orphanet J Rare Dis 2012;7:13. doi: 10.1186/1750-1172-7-13.
- Hao Y, Ran Y, Lu B, Li J, Zhang J, Feng C, *et al.* Therapeutic effects of human umbilical cord-derived mesenchymal stem cells on canine radiation-induced lung injury. Int J Radiat Oncol Biol Phys 2018;102:407–416. doi: 10.1016/j.ijrobp.2018.05.068.
- Yin Z, Yang G, Deng S, Wang Q. Oxidative stress levels and dynamic changes in mitochondrial gene expression in a radiation-induced lung injury model. J Radiat Res 2019;60:204–214. doi: 10.1093/jrr/rry105.
- 9. Huang Y, Zhang W, Yu F, Gao F. The cellular and molecular mechanism of radiation-induced lung injury. Med Sci Monit 2017;23:3446–3450. doi: 10.12659/msm.902353.
- Paun A, Bergeron ME, Haston CK. The Th1/Th17 balance dictates the fibrosis response in murine radiation-induced lung disease. Sci Rep 2017;7:11586. doi: 10.1038/s41598-017-11656-5.
- Zhang C, Zhao H, Li BL, Gao F, Liu H, Cai JM, et al. CpGoligodeoxynucleotides may be effective for preventing ionizing radiation induced pulmonary fibrosis. Toxicol Lett 2018;292:181– 189. doi: 10.1016/j.toxlet.2018.04.009.
- Chen C, Yang S, Zhang M, Zhang Z, Zhang SB, Wu B, et al. Triptolide mitigates radiation-induced pneumonitis via inhibition of alveolar macrophages and related inflammatory molecules. Oncotarget 2017;8:45133–45142. doi: 10.18632/oncotarget.16456.
- Meziani L, Mondini M, Petit B, Boissonnas A, Thomas de Montpreville V, Mercier O, *et al.* CSF1R inhibition prevents radiation pulmonary fibrosis by depletion of interstitial macrophages. Eur Respir J 2018;51. doi: 10.1183/13993003.02120-2017.
- 14. Ng J, Dai T. Radiation therapy and the abscopal effect: a concept comes of age. Ann Transl Med 2016;4:118. doi: 10.21037/atm.2016.01.32.
- 15. Vanpouille-Box C, Diamond JM, Pilones KA, Zavadil J, Babb JS, Formenti SC, *et al.* TGFbeta is a master regulator of radiation therapy-induced antitumor immunity. Cancer Res 2015;75:2232–2242. doi: 10.1158/0008-5472.CAN-14-3511.
- Antonia SJ, Villegas A, Daniel D, Vicente D, Murakami S, Hui R, et al. Durvalumab after chemoradiotherapy in stage III non-small-cell lung cancer. N Engl J Med 2017;377:1919–1929. doi: 10.1056/ NEJMoa1709937.
- Antonadou D, Coliarakis N, Synodinou M, Athanassiou H, Kouveli A, Verigos C, *et al.* Randomized phase III trial of radiation treatment +/- amifostine in patients with advanced-stage lung cancer. Int J Radiat Oncol Biol Phys 2001;51:915–922. doi: 10.1016/s0360-3016 (01)01713-8.
- Kong FM, Ao X, Wang L, Lawrence TS. The use of blood biomarkers to predict radiation lung toxicity: a potential strategy to individualize thoracic radiation therapy. Cancer Control 2008;15:140–150. doi: 10.1177/107327480801500206.
- Heukels P, Moor CC, von der Thüsen JH, Wijsenbeek MS, Kool M. Inflammation and immunity in IPF pathogenesis and treatment. Respir Med 2019;147:79–91. doi: 10.1016/j.rmed.2018.12.015.

How to cite this article: Ying HJ, Fang M, Chen M. Progress in the mechanism of radiation-induced lung injury. Chin Med J 2021;134:161–163. doi: 10.1097/CM9.00000000001032