

Polar Profile of Antiviral Peptides from AVPpred Database

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Abstract Diseases of viral origin in humans are among the most serious threats to health and the global economy. As recent history has shown the virus has a high pandemic potential, among other reasons, due to its ability to spread by air, hence the identification, investigation, containment, and treatment of viral diseases should be considered of paramount importance. In this sense, the bioinformatics research has focused on finding fast and efficient algorithms that can identify highly toxic antiviral peptides and to serve as a first filter, so that trials in the laboratory are substantially reduced. The work presented here contributes to this effort through the use of an algorithm already published by this team, called polarity index method, which identifies with high efficiency antiviral peptides from the exhaustive analysis of the polar profile, using the linear sequence of the peptide. The test carried out included

all peptides in APD2 Database and 60 antiviral peptides identified by Kumar and co-workers (*Nucleic Acids Res* 40:W199–204, 2012), to build its AVPpred algorithm. The validity of the method was focused on its discriminating capacity so we included the 15 sub-classifications of both Databases.

Keywords Polarity index method · Antiviral peptides · AVPpred algorithm

Abbreviations

SCAAP	Selective cationic amphipathic antibacterial peptides
APD2	Antimicrobial peptide database http://aps.unmc.edu/AP/ [1] accessed December 19, 2012. AVPpred, http://crdd.osdd.net/servers/avppred [2] accessed March 10, 2013
QSAR	Quantitative structure activity relationships
SVM	Support vector machine
SARS	Severe acute respiratory syndrome
HIV	Human immunodeficiency virus
AAindex	Amino acid indices, substitution matrices and pair-wise contact potentials database http://www.genome.jp/aaindex/ [3] accessed March 10, 2013

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Introduction

Within the antimicrobial peptides there is a group of particular importance, the antiviral peptides. These peptides are potential drugs to face the increased incidence of chronic viral infections caused by HIV and hepatitis [4, 5]; for that reason there is a need to accelerate the development of synthetic antiviral peptides. Inclusive, the testing of vaccines against HIV and hepatitis [6, 7] that are under

Table 1 Q[i,j] Polarity matrix

	P+	P-	N	NP
P+	0.0354861617	0.0184528027	0.0354861617	0.0681334287
P-	0.0163236335	0.0177430809	0.0298083741	0.0709723234
N	0.0404542238	0.0326472670	0.0872959569	0.0908445716
NP	0.0667139813	0.0617459193	0.0915542915	0.1937544346

Incidences of 60 AVPpred antiviral peptides extracted from Kumar [2], and one antiviral peptide from APD2 [1]

development offers no guarantee of success regardless of the millions of infected people. In counterpart, antiviral medications are not oriented to the most acute infections that cause serious diseases, such as hemorrhagic fever and cancer [8–10]; they have limited effectiveness and serious side effects. Perhaps more importantly, the antiviral chemotherapy is producing a rapid development of drug-resistant strains, as a result of the high rate of virus replication, due to the low resistance to replication [11].

In such a scenario, the work in proteomics and bioinformatics should focus on the generation of fast and robust algorithms [12, 13] identifying the antiviral action, from the linear or three-dimensional structure of the peptide. However, the simulation of the three-dimensional structure of the peptide is very complex, without taking into account other factors involved such as: The dynamics of the membrane and toxicity. In this work, we use the Polarity index method [14], already published by our team to identify SCAAP [15–17] and which only uses the linear peptide sequence to identify the same group of antiviral peptides that were identified by AVPpred algorithm [2].

This method [14] generates an exhaustive analysis of the peptide polarity through its polarity matrix. In this sense, the method apparently does not consider other factors, but indirectly it does, because it requires being calibrated by “a set of peptides” that are characteristic of the profile.

Materials and Methods

The identification of antiviral peptides performed by the polarity index method [14] requires the modifications below (Supplementary Material).

Polarity Index method. Updates

Modifications

- Replacing the Q[i,j] matrix in the source program [14] by the Table 1, which represents the incidences of antiviral sequences with a unique pathogenic action. Table 1 considers 60 AVPpred antiviral peptides

extracted from Kumar and co workers [2], and 1 antiviral peptide extracter from APD2 database [1].

- Replacing the rule in the source program [14] by $P[i,j] + Q[i,j]$ vector complying with the next rule: polar interactions 15 or 16 are present in the 1st position, polar interaction 14 is not present from 3th to 8th positions, polar interaction 2 is not present in the first seven positions, polar interaction 5 is not present in the first five positions, polar interactions 4 or 8 are not present from 12th to 16th positions, polar interactions 14, 15 or 16 are not present from 11th to 16th positions, polar interaction 9 is not present in the 1st, 4th, 5th or 9th positions, polar interaction 1 is not present in the 10th or 11th positions, polar interaction 11 is not present in the first position, polar interaction 1 is not present in 10th or 11st positions, polar interaction 14 is not present in the 5th position, and polar interaction 4 is not present in the 4th position (Table 2).

APD2 Database Trial Data Preparation

We use 3,636 peptides in the APD2 Database [1] classified by their *multiple* action against: 149 Gram– ONLY, 1711 Gram +/Gram– ONLY, 315 Gram+ ONLY, 141 cancer cells, 9 sperms, 88 HIV, 744 fungi, 21 insects, 244 mammalian cells, 47 parasites, 3 protists, 39 chemotaxis, 0 SCAAP, 125 virus; and also 1,059 by their *unique* action against: 111 Gram– ONLY, 213 Gram+ ONLY, 518 Gram+/Gram– ONLY, 20 cancer cells, 0 HIV, 88 fungi, 35 H1N1, 2 insects, 11 mammalian cells, 9 parasites, 1 protists, 0 chemotaxis, 30 SCAAP, 0 sperms and 21 virus.

AVPpred Algorithm Trial Data Preparation

From Kumar and co workers [2] work about antiviral peptides, we took 60 validated and experimental peptides from 1,245 antiviral peptides. His work evaluated these 60 peptides with 25 physicochemical properties (Table 3) out of 144 properties from the same database and called it AAindex database [3]. These peptides were used to build the SVM AVPpred algorithm [2], and we are using them here to validate the Polarity index method

Table 2 Polarity index method rules

Position P[i,j] + Q[i,j] vector of study.	1 (1,1)	2 (1,2)	3 (1,3)	4 (1,4)	5 (2,1)	6 (2,2)	7 (2,3)	8 (2,4)	9 (3,1)	10 (3,2)	11 (3,3)	12 (3,4)	13 (4,1)	14 (4,2)	15 (4,3)	16 (4,4)
<i>Rule # 1</i>	✓															
Polar interactions 15 or 16 are present in the 1st position																
<i>Rule # 2</i>					×	×	×	×	×	×	×					
Polar interaction 14 is not present from 3th to 8th positions																
<i>Rule # 3</i>		×	×	×	×	×	×	×	×	×	×					
Polar interaction 2 is not present in the first seven positions																
<i>Rule # 4</i>		×	×	×	×	×										
Polar interaction 5 is not present in the first five positions																
<i>Rule # 5</i>													×	×	×	×
Polar interactions 4 or 8 are not present from 12th to 16th positions																
<i>Rule # 6</i>													×	×	×	×
Polar interactions 14, 15 or 16 are not present from 11th to 16th positions																
<i>Rule # 7</i>		×			×	×							×			
Polar interaction 9 is not present in the 1st, 4th, 5th or 9th positions																
<i>Rule # 8</i>													×	×		
Polar interaction 1 is not present in the 10th or 11th positions																
<i>Rule # 9</i>		×														
Polar interaction 11 is not present in the first position																
<i>Rule # 10</i>													×	×		
Polar interaction 1 is not present in 10th or 11st positions																
<i>Rule # 11</i>					×											
Polar interaction 14 is not present in the 5th position																
<i>Rule # 12</i>						×										
Polar interaction 4 is not present in the 4th position																

Identification Rules in Polarity index method for antiviral peptides. (✓) The polar interaction is present in the position. (✗) The polar interaction is not present in the position

Results

Polarity index method is an algorithm that determines the probable antiviral pathogen action of peptides by

using the peptide polarity sequence. It was applied to the APD2 database and the experimental peptides from AVPpred algorithm [2] with the following results.

Table 3 Physicochemical properties from AVPpred

#	AAindex ID	Description	Polarity	Reference
1	BEGF750103	Conformational parameter of beta-turn	✓	[18]
2	BULH740102	Apparent partial specific volume		[19]
3	CHAM810101	Steric parameter	✓	[20]
4	CHOP780204	Normalized frequency of N-terminal helix	✓	[21]
5	CHOP780206	Normalized frequency of N-terminal non helical region	✓	[21]
6	CHOP780215	Frequency of the 4th residue in turn	✓	[21]
7	CIDH920104	Normalized hydrophobicity scales for alpha/beta-proteins	✓	[21]
8	COHE430101	Partial specific volume		[22]
9	FASG760105	PK-C	✓	[23]
10	FAUJ880104	STERIMOL length of the side chain		[24]
11	FINA770101	Helix-coil equilibrium constant	✓	[25]
12	FINA910101	Helix initiation parameter at position $i - 1$		[26]
13	GEIM800101	Alpha-helix indices	✓	[27]
14	GEIM800102	Alpha-helix indices for alpha-proteins	✓	[27]
15	GEIM800104	Alpha-helix indices for alpha/beta-proteins	✓	[27]
16	KARP850101	Flexibility parameter for no rigid neighbors	✓	[28]
17	KARP850102	Flexibility parameter for one rigid neighbor	✓	[28]
18	AURR980101	Normalized positional residue frequency at helix termini N4'	✓	[29]
19	AURR980118	Normalized positional residue frequency at helix termini C"	✓	[29]
20	AURR980120	Normalized positional residue frequency at helix termini C4'	✓	[29]
21	AVBF000107	Slopes tripeptide FDPB PARSE neutral		[30]
22	GEOR030102	Linker propensity from 1-linker dataset		[31]
23	KIDA850101	Hydrophobicity-related index	✓	[32]
24	GUYH850102	Apparent partition energies calculated from Wertz-Scheraga index	✓	[33]
25	CASG920101	Hydrophobicity scale from native protein structures	✓	[34]

Selected physicochemical properties to build the AVPpred algorithm by Kumar [2]. AAindex amino acid index database [3]. Polarity (✓) physicochemical properties are directly or indirectly related to the polarity

Table 4 Polarity index matches by linear sequence in virus

No	Code	ID PUBMED	Sequence	#1	#2	References
1	AVP_0618	6096849	GPPISLERLDVGTNLGNIAKLEAKEAKELLESSDQI			[35]
2	AVP_0629	3788062	KVLHLEGEVNKIALLSTNKAVVSLNSNGVSVLTS	N		[36]
3	AVP_0607	2893293	DFLEENITALLEEAQIQQEKNMYELQKLNSWDVFG			[37]
4	AVP_0168	8382405	EGPTLGNWAREIWATLFGKA	N		[38]
5	AVP_0179	8382405	NWAREIWATLFKA	N		[38]
6	AVP_0467	10390360	FAIKWEYVLLLFL			[39]
7	AVP_0512	1848704	SWLRDIWDWKCEVLSDFK			[40]
8	AVP_0514	1848704	SWLRDIWDWLCEVLSDFK			[40]
9	AVP_0373	1848704	SWLRDIWDWICEVLSDFK			[40]
10	AVP_0372	52472831	SWLRDIWDWICEVLSD			[41]
11	AVP_0387	9223423	TWLRAIWDWVCTALTDFK			[42]
12	AVP_0323	8822631	PPVYTKDVKDISSQISSMNQLQQSKDYIKEAQKILDtvNPSL			[43]
13	AVP_0328	3012869	VANDPIDISIELNKAKSDEESKEWIRRSNQKLDSD	N		[44]
14	AVP_0210	11118300	ANTAFVSSAHNTQKIPAGAPFNRLNRLAMLADLRQNAAFAG			[45]
15	AVP_0024	1695254	CGGNLLRAIEAQHQHLLQLTVWGIKQLQARILAVERYLKQ			[46]
16	AVP_0312	16667080	EQCREEEDDR	N		[47]
17	AVP_0358	15893660	GGTIFDCGETCFLGTCYTPGCSCGNYGFCYGTN	N		[48]

Table 4 continued

No	Code	ID PUBMED	Sequence	#1	#2	References
18	AVP_0019	7841460	GICRCICGKGICRCICGR			[49]
19	AVP_0284	21685289	GICRCICGRGICRCICGR			[50]
20	AVP_0397	7529412	GIKEWKRVQRIKDFRLNLV	N		[51]
21	AVP_0361	16872274	GLPVCGETCVGGTCNTPGCTCSWPVCTR	N		[52]
22	AVP_0286	10521339	GVCRCLCRRGVCRICRR			[53]
23	AVP_0304	15949629	KTCENLADTY			[54]
24	AVP_0684	7031661	LEAIPCSIPPCFLFGKPFVF			[55]
25	AVP_0692	7031661	LEAIPISIPPELAFAKPFVF			[55]
26	AVP_0703	7031661	LEAIPMSIPPEVFFGKPFVF			[55]
27	AVP_0155	9777331	LSYRCPCR	N		[56]
28	AVP_0409	1433527	WMEWDREIEELAKKAELAKKAELAKKAWASLWNWF			[57]
29	AVP_0222	3031048	YALLIRMIYKNI			[58]
30	AVP_0224	19468303	YQLLARMYKNI			[59]
31	AVP_0225	9504927	YQLLIAMIYKNI			[60]
32	AVP_0584	2578615	YTSLIHSLIEESQNQQEKNEQELLEFDKWASLWNWF	N		[61]
33	AVP_0591	3040055	YTSLIHSLIEESQNQQEKNEQELLELNKWASLWNWF	N		[62]
34	AVP_0310	9516047	GLFGVLGSIAKHVLPHVVVIAEKL	N		[63]
35	AVP_0183	7826699	DWHLGQGVSIWRKK			[64]
36	AVP_0444	9784389	FLFPLITSFLSKVL			[65]
37	AVP_0452	10951191	GLFDIICKKIAESW			[66]
38	AVP_0459	12089438	GWLKKIESIIDAF			[67]
39	AVP_0460	11053427	HVDKKVADKVLLKQLRIMRLTRL	N		[68]
40	AVP_0348	7826699	TTWEAWDRAIAEYAARIEALIRAAQEQQEKNEAILREL			[64]
41	AVP_0354	7826699	TTWEAWDRAIAEYAARIEALIRASQEQQEKNEAILREL			[64]
42	AVP_0612	3012869	ELNKAKSDLEESKEWIRRSNQKLDSIGNWHQSSTT	N		[44]
43	AVP_0623	3012869	IENLKAKSDLEESKEWIRRSNQKLDSIGNWHQSST	N		[44]
44	AVP_0390	3012465	AAHLIDALYAEFLGGRVLT			[69]
45	AVP_0068	10077657	HRWRKRWRKHRWRKRWK			[70]
46	AVP_0061	10077657	KRWRKRWRKWRWRKRWK			[70]
47	AVP_0055	15095345	RTRKGRKRTRKGRK			[71]
48	AVP_0191	14648299	RGGKIAGKIAKIAGKIAKIAGKIA			[72]
49	AVP_0134	3012869	ISIELNKAKSDEESKEWIRRSNQKLDSIGNWHQS	N		[44]
50	AVP_0482	2959959	RRKKAVALLPAVLLA			[73]
51	AVP_0483	2959959	RRKKAVALLPAVLLAL			[73]
52	AVP_0302	2661722	KDLLFK	N		[74]
53	AVP_0113	3788062	GPPISLERLDVGTNLGNIAIKLEDAKELLESSDQI			[35]
54	AVP_0114	3788062	HRIDLGPPISLERLDVGTNLGNIAIKLEDAKELLE			[35]
55	AVP_0116	3788062	ISLERLDVGTNLGNIAIKLEDAKELLESSDQILRS			[35]
56	AVP_0288	12810954	CGECGGGHIVGRFCMVRFLRLVFI			[75]
57	AVP_0289	9634230	CRCCELKSLCPTLMRVVRLGLVLL	N		[76]
58	AVP_0138	6096849	LVFPSDEFDASISQVNEKINQSLAFIRKSDELLHN			[77]
59	AVP_0278	2833514	KWKLFFKIGIGKFLHVAKKF			[78]
60	AVP_0335	12886019	CDVIALLCHLNT			[79]
61	APD2_01209	7744961	RICRCICGRRICRCICGR			[80]

Subject sequences identified by polarity index method in APD2 database [1] and by AVPpred [2], where peptides have action only on virus. #1: (N) rejected peptide by SVM AVPpred algorithm. #2: (N) rejected peptide by polarity index method. Source: National Center for Biotechnology Information, US National Library of Medicine <http://blast.ncbi.nlm.nih.gov/Blast.cgi?PAGE=Proteins> in database: Swiss-Prot (swissprot), accessed March 20, 2013

Table 5 Polarity index matches by pathogenic action

Number of hits	Gram+ only	Gram− only	Gram+/−	Virus %	HIV	Fungi	Protists	Parasites	Insects	Sperms	Cancer cells	Mammalian cells	SCAAP	AVPpred	%	
Unique action	38	19	134	1	100	0	6	0	1	0	0	2	2	0	13	42
	213	111	518	1	0	88	1	9	2	0	20	11	0	51	60	
Multiple action	66	27	426	19	16	163	0	12	3	1	28	81	8	0	0	

Polarity index method matches for both APD2 [1] and AVPpred antiviral peptide groups [2]. Unique action peptides with pathogenic action against only one group. Multiple action peptides with pathogenic action against two or more groups. (%): Percentage of hits/total peptides

From the 25 physicochemical properties used to design the SVM AVPpred algorithm [2], 21 are directly or indirectly ($18/25 = 84\%$) related to the polarity (Table 3, column Polarity with entries with figure ✓).

Polarity index method had over $43/60 = 71\%$ efficiency detecting the 60 validated and experimental anti-viral peptides from Kumar and coworkers [2] (Table 4, entries 1–60 and Table 5 column AVPpred), and $1/1 = 100\%$ detecting the antiviral peptides from APD2 database (Table 4, entry 61 and Table 5, column Virus). There are no coincidences between both excluding sets (Table 4, columns #1 and #2).

Discussion

When reviewing different databases of antimicrobial peptides, we detected a peptide with predominantly toxic action toward a pathogenic group; this allows us to assume that nature considers only small changes in the primary structure of the peptide to induce its possible pathogenic action. In that sense, the peptide linear structure plays an important role in the identification of its pathogenic action, when we use algorithms that use “training sets” with the desired profile. Although the physicochemical property called polarity is involved in most of the algorithms that predict anti-virus peptides, this method has an innovative aspect as it expresses the metric through a polarity matrix that includes 16 interactions. We see this matrix as a picture of the polar dynamics of the peptide. The polarity matrix clearly shows a pattern that led us to achieve an identification efficiency of 70 % on the AVPpred database. The same pattern also rejected other groups of peptides in APD2 database, with the exception of the anti-virus set. We assume that if we built the matrix with one digit, perhaps we did not have enough information to focus the method correctly.

We believe the effectiveness of the polarity index method in terms of the computing resources required makes it suitable candidate for a more detailed analysis related to the sub-domains of peptides. In this regards, we have initiated a comprehensive classification of the APD2 database gathering, from published manuscripts, the toxicity values of antimicrobial peptides and thus explore peptide sub-domains with specific and very toxic pathogenic action. Our team is working on this as we consider it of vital importance to strongly support basic scientific research. We have also published work where the same method is used to understand the profile of the “first proteins” from 4 billion years ago. For this task, we are using clusters of GPU coprocessors, which will allow the analysis of 15 amino acids in length peptides.

Finally, within the antiviral peptide group, there are two sub-groups not approached in this work as they constitute by themselves an independent topic for the importance they

have and the degree of subject matter expertise required: the influenza A type H1N1 and HIV peptides. Given their potential to provoke a world pandemic, these two groups of peptides concentrate the efforts of several research groups, as they can undoubtedly become a problem of enormous proportions. Our team is now directing the method toward the identification of these two sub-groups.

Conclusions

In summary, we report an implementation of a polarity index method in the exhaustive prediction of antiviral peptides from AVPPred and APD2 databases, with high level of discriminative efficiency ($44/61 = 72$), from the reading of its linear sequence.

Availability

The test files and source code are given as “Supplementary Material”

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Conflict of interest We declare that we do not have any financial and personal interest with other people or organizations that could inappropriately influence (bias) our work.

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