


## COMMENT

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# A role for the rare endogenous retrovirus $\beta 4$ in development of Japanese fancy mice

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Two coat-color mutations, *nonagouti*, which changes coat color from wild-type agouti to black, and *piebald*, which induces irregular white spotting, are the characteristics of Japanese fancy mouse strain JF1/Ms. In our *Communications Biology* article, we reported that insertion of a rare type of endogenous retrovirus  $\beta 4$  has caused both coat color mutations. Although there are some reports on the roles of  $\beta 4$  in the mouse genome, further studies on  $\beta 4$  will uncover new features of endogenous retrovirus sequences.

### Genetic mutations in coat color genes

Coat color mutation in animals remains one of the most attractive phenomena for both research scientists and the general public. Rudimental genetics of coat color mutations in mice has been reported in historic literature from the Edo period of Japan<sup>1</sup>. Since phenotype is determined by the appearance and the mode of inheritance is relatively simple, various coat color mutations have been identified and documented to date<sup>2</sup>. In the early days of understanding mice genetics, researchers had investigated the coat color mutations as a model of heredity. Many researchers obtained coat color mutations from a stock of fancy mice which was kept by fanciers at that time. This series of mutations is now known as classical mutations.

Laboratory mice have some classical coat color mutations such as *nonagouti* (*a*), *brown* (*Tyrp1<sup>b</sup>*), *albino* (*Tyr<sup>c</sup>*), *dilution* (*Myo5a<sup>d</sup>*), and *piebald* (*Ednrb<sup>s</sup>*). Most genetic causes of these mutations have been identified. The *Tyrp1<sup>b</sup>* and *Tyr<sup>c</sup>* mutations caused by point mutations in the protein-coding sequences, would disrupt the protein function and reduce the pigmentation<sup>3,4</sup>. On the other hand, insertional mutations in *a*, *Myo5a<sup>d</sup>*, and *Ednrb<sup>s</sup>* alleles have been found to prevent gene transcription and result in hypomorphic phenotype<sup>5,6</sup>. These insertional mutations were caused by insertions of some types of endogenous retrovirus sequence<sup>7</sup>.

In the classical *nonagouti* (*a*) mutation with black coat color, the causal genetic mutation was first identified by Bultman and co-workers in 1994. They reported an insertion of sequence consisted of a 5.4 kb VL30 retroviral sequence containing 5.5 kb unknown sequence in the first

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intron in *nonagouti* (*a*) allele<sup>8</sup>. Several spontaneous reverse-mutations from nonagouti to agouti coat colors were found to be completely linked with an exclusion of the inserted sequence caused by homologous recombination between the long terminal repeat (LTR) sequences of the VL30 element. Based on these evidences, the cause of *nonagouti* mutation was believed to be the insertion of VL30.

#### **$\beta$ 4 retroviral insertion in *nonagouti* allele**

A quarter century later, we noticed that Japanese wild-derived mouse strain, MSM/Ms (MSM), has an insertion of VL30 in the *agouti* gene even though they exhibit agouti coat color. In order to resolve the inconsistency between the nonagouti phenotype and the insertion of VL30, we re-examined the genetic analyses of the *nonagouti* allele. In the work published in *Communications Biology*<sup>9</sup>, we reported that an insertion of a poorly characterized 9.3 kb endogenous betaretrovirus group 4 ( $\beta$ 4, also named as ERVB4, ERV- $\beta$ 4, or MmERV- $\beta$ 4) element, which was previously presented as 5.5 kb unknown sequence, into the VL30 element in the *nonagouti* allele<sup>8</sup>. Similar to other retroviral mutations, the  $\beta$ 4 element caused abnormal expression of the *agouti* gene by preventing transcription and interrupting mRNA splicing. Interestingly, a solo-LTR in the *nonagouti* allele developed via homologous recombination between the two  $\beta$ 4-LTR sequences has resulted in partial reverse mutation of the coat color to black-and-tan which has yellow coat only on the ventral side. From these results and the precise deletion experiment of  $\beta$ 4 by CRISPR/Cas9,  $\beta$ 4 was revealed to be the true cause of the *nonagouti* mutation.

As one of the endogenous retroviruses,  $\beta$ 4 elements in the mouse genome were first reported by Baillie and co-workers in 2004<sup>10</sup>. They identified a repetitive sequence within the genome of murid rodents as one of the groups of Betaretrovirus including type B and type D retroviruses, such as the mouse mammary tumor virus and *Mus musculus* type D retrovirus, respectively. Apart from that, LTR sequences of the  $\beta$ 4 in the *nonagouti* allele had been described as a repetitive sequence in mouse genome<sup>4,6</sup>. There are at least 2000 copies of  $\beta$ 4-LTR sequences, a few disrupted or truncated  $\beta$ 4 sequences and one possibly intact  $\beta$ 4 full-length sequence distributed in the reference mouse genome. However, there were limited reports on the phenotypic mutation caused by the insertion of  $\beta$ 4 element. To our understanding, the *piebald* (*s*) mutation was the only case, which was caused by  $\beta$ 4 insertion into the first intron of *endothelin receptor type B* gene (*Ednrb*) in Japanese fancy mice<sup>6,11</sup>, until our report on the *nonagouti* mutation.

#### **Two $\beta$ 4 elements inserted in coat color genes of Japanese fancy mice**

Japanese fancy mouse strain, JF1/Ms (JF1), is known to have two classical coat color mutations, *nonagouti* and *piebald* alleles, leading to characteristic black spots on the white coat, with black eyes<sup>11</sup>. The insertional mutation in the *piebald* allele was identified by Yamada and co-workers in 2006<sup>6</sup>, in which a retroposon-like element was found in the first intron of *Ednrb* gene and had disrupted the normal expressions of the gene. This inserted element was annotated as a truncated type of  $\beta$ 4 element in our study<sup>9</sup>. Similar to the case of nonagouti phenotype, spontaneous reverse mutation of the piebald phenotype caused by the homologous recombination between the two  $\beta$ 4 LTR sequences was found in JF1 strain, and resulted in nonagouti coat color in a revertant strain JF1-s<sup>+</sup> (Fig. 1). Moreover, we also found that the insertion of  $\beta$ 4 within the VL30 in the *nonagouti* allele has originally occurred in a lineage of Japanese fancy mice<sup>9</sup>. All these discoveries indicate that both coat color mutations are insertional



**Fig. 1 Coat color phenotypes caused by insertion of  $\beta$ 4 elements in Japanese fancy mice.** Japanese fancy mouse strain, JF1, shows the characteristic combination of nonagouti and piebald phenotypes, in which the  $\beta$ 4 elements were inserted in *a* and *Ednrb*<sup>s</sup> alleles (right), respectively. A spontaneous revertant mouse of JF1, JF1-s<sup>+</sup>, which lacks large part of the inserted  $\beta$ 4 sequence in the *Ednrb*<sup>s</sup> allele and with restored piebald phenotype (left).

mutations caused by retrotransposition of the  $\beta$ 4 elements in a specific line of Japanese fancy mice.

This intriguing coincidence implies the possibility of a series of certain mutation processes via retroviral insertion, such as the retroviral expansion in host mouse genome. Although the transposition activity of  $\beta$ 4 is still unclear, we and other groups have observed especially high expression level of  $\beta$ 4 transcripts in testes<sup>9,12,13</sup>, suggesting a high frequency of germ-line transposition of the  $\beta$ 4 elements. The intact viral genes of  $\beta$ 4 found in the *nonagouti* allele also indicates sustained transposition activity of the  $\beta$ 4 elements<sup>9,10</sup>. Based on these findings, we speculate that the  $\beta$ 4 elements were expanded during the domestication process of Japanese fancy mice. In fact, the number of  $\beta$ 4 insertions in JF1 mouse genome is about three to seven times higher than in common inbred strains. This high number of  $\beta$ 4 insertions is not observed in the closely related wild-derived mouse strain, MSM. These data clearly demonstrate that the insertion of  $\beta$ 4 has contributed to the characteristic coat color of Japanese fancy mice.

From an evolutionary perspective, the current known endogenous betaretroviruses including the  $\beta$ 4 in the mouse genome are believed to have emerged in the genome of murid rodents at least 20 million years ago<sup>10</sup>. Therefore, some  $\beta$ 4 elements that are shared in the homologous loci among related species seems to be orthologs inherited from common ancestors. However, the origin of the  $\beta$ 4 element recently inserted in the *nonagouti* allele is unclear except that the element has emerged in the lineage of Japanese fancy mice<sup>9</sup>. Although it is highly possible that it comes from a copy that is retrotransposed from another locus, no other identical sequence has been found in the reference mouse genome. Given the intact viral genes, it is also possible that the element has derived from exogenous infection of  $\beta$ 4 retrovirus into founder mouse of the Japanese fancy mice.

### The role of $\beta 4$ elements in mouse genome

Although the role of  $\beta 4$  elements in mouse genome is still unclear, there are several examples indicating a contribution of the  $\beta 4$  to functional genes. For example, the murine-specific C-repeat region of XIST sequence is known to be originally derived from the ERVB4 sequence<sup>14</sup>. This C-repeat sequence interacts with YY1 protein to target XIST RNA to a specific genomic locus<sup>15</sup>. A second example is the *Serpina3a* gene, the first exon of which consists of one part of  $\beta 4$ -LTR sequence. Similar to other  $\beta 4$  elements, *Serpina3a* gene is also expressed strongly in testes<sup>16</sup>. The original first exon of *Serpina3a* gene is located on the downstream of the  $\beta 4$ -LTR, but its promoter activity is weak and the *Serpina3a* mRNA is transcribed mainly from the  $\beta 4$ -LTR sequence. This  $\beta 4$ -LTR sequence is shared among the related species and seemed to be inserted before the divergence of *M. musculus*, implying a certain role of this alternative promoter. Therefore, we suggest that *cis*-regulatory elements were introduced in the reproductive system through insertion of  $\beta 4$  in the murine lineage.

Beside these significant contributions, high methylation in H3K9me3 and H4K20me3 marks was also found in the  $\beta 4$  sequences<sup>17</sup>, implying a repressive epigenetic state on the  $\beta 4$  similar to the well-known intra-cisternal A-type particle elements. We also observed that a part of the  $\beta 4$  sequences is enriched for CCCTC-binding factor, which regulates three-dimensional chromatin architecture. These characteristics may be related to a role of the  $\beta 4$  in regulating gene expression similar to other LTR retrotransposons<sup>18</sup>, but more detailed analyses are crucial to further elucidate actual functions of  $\beta 4$ .

### Concluding remarks

Coat color is an excellent index for studying how mutation alters gene functions that are linked with phenotype. As such, we investigated the insertion of  $\beta 4$  in nonagouti mice and solo-LTR of  $\beta 4$  in black-and-tan mice. There is limited evidence that  $\beta 4$  elements in mouse genome controls gene function. Our findings showed that the retrovirus insertion caused not only a null phenotype but also partial and reversible alternation of the phenotype. Whether  $\beta 4$  had contributed in the domestication and evolution of mouse remains an intriguing issue, and further studies are crucial to understand the roles of  $\beta 4$ . Ensuing the studies of coat color mutations, functional studies focused on certain genes containing  $\beta 4$  sequence together with genomic studies using high-throughput sequencing technologies will allow us to uncover how  $\beta 4$  controls gene functions and reveal other roles of  $\beta 4$ .

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### References

1. Jo-en-shi. *Chingan-sodategusa* (Zeniya Chobei press, Kyoto, 1787).
2. Silvers, W. K. *The Coat Colors of Mice. A Model for Mammalian Gene Action and Interaction* (Springer-Verlag, New York, 1979).
3. Zdarsky, E., Favor, J. & Jackson, I. J. The molecular basis of *brown*, an old mouse mutation, and of an induced revertant to wild type. *Genetics* **126**, 443–449 (1990).
4. Beermann, F. et al. Rescue of the albino phenotype by introduction of a functional tyrosinase gene into mice. *EMBO J.* **9**, 2819–2826 (1990).

5. Jenkins, N. A., Copeland, N. G., Taylor, B. A. & Lee, B. K. Dilute (*d*) coat colour mutation of DBA/2J mice is associated with the site of integration of an ecotropic MuLV genome. *Nature* **293**, 370–374 (1981).
6. Yamada, T. et al. Reduced expression of the endothelin receptor type B gene in piebald mice caused by insertion of a retroposon-like element in intron 1. *J. Biol. Chem.* **281**, 10799–10807 (2006).
7. Gagnier, L., Belancio, V. P. & Mager, D. L. Mouse germ line mutations due to retrotransposon insertions. *Mob. DNA* **10**, 15 (2019).
8. Bultman, S. J. et al. Molecular analysis of reverse mutations from nonagouti (*a*) to black-and-tan (*a'*) and white-bellied agouti (*A<sup>w</sup>*) reveals alternative forms of agouti transcripts. *Genes Dev.* **8**, 481–490 (1994).
9. Tanave, A., Imai, Y. & Koide, T. Nested retrotransposition in the East Asian mouse genome causes the classical *nonagouti* mutation. *Commun. Biol.* **2**, 283 (2019).
10. Baillie, G. J., van de Lagemaat, L. N., Baust, C. & Mager, D. L. Multiple groups of endogenous betaretroviruses in mice, rats, and other mammals. *J. Virol.* **78**, 5784–5798 (2004).
11. Koide, T. et al. A new inbred strain JF1 established from Japanese fancy mouse carrying the classic piebald allele. *Mamm. Genome* **9**, 15–19 (1998).
12. Gnanakkan, V. P. et al. TE-array—a high throughput tool to study transposon transcription. *BMC Genomics* **14**, 869 (2013).
13. Jin, Y., Tam, O. H., Paniagua, E. & Hammell, M. TETranscripts: a package for including transposable elements in differential expression analysis of RNA-seq datasets. *Bioinformatics* **31**, 3593–3599 (2015).
14. Elisaphenko, E. A. et al. A dual origin of the Xist gene from a protein-coding gene and a set of transposable elements. *PLoS ONE* **3**, e2521 (2008).
15. Jeon, Y. & Lee, J. T. YY1 tethers Xist RNA to the inactive X nucleation center. *Cell* **146**, 119–133 (2011).
16. Horvath, A. J., Forsyth, S. L. & Coughlin, P. B. Expression patterns of murine antichymotrypsin-like genes reflect evolutionary divergence at the *Serpina3* locus. *J. Mol. Evol.* **59**, 488–497 (2004).
17. Day, D. S., Luquette, L. J., Park, P. J. & Kharchenko, P. V. Estimating enrichment of repetitive elements from high-throughput sequence data. *Genome Biol.* **11**, R69 (2010).
18. Thompson, P. J., Macfarlan, T. S. & Lorincz, M. C. Long terminal repeats: from parasitic elements to building blocks of the transcriptional regulatory repertoire. *Mol. Cell.* **62**, 766–776 (2016).

### Author contributions

A.T. conceived and A.T. and T.K. wrote the commentary.

### Competing interests

The authors declare no competing interests.

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