

Review

# MicroRNAs as Biomarkers in Colorectal Cancer

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**Abstract:** MicroRNAs (miRs) are small RNAs that repress mRNA translation, resulting in the degradation of mRNAs and regulation of the expression levels of various genes. Recent studies have shown that aberrant miR expression has a functional role in the initiation and progression of various malignancies, including colorectal cancer (CRC), which is one of the leading causes of cancer-related death worldwide. miRs have also been shown to have applications as diagnostic, prognostic, and predictive biomarkers because of their high tissue specificity, stability, and altered expression in tumor development. In this report, we examined the role of miRs as biomarkers in CRC through a review of meta-analyses and large-scale analyses having strong statistical confidence in the study outcomes. We also discuss current issues in the clinical application of these miRs.

**Keywords:** microRNA; biomarker; colorectal cancer

## 1. Introduction

The incidence of colorectal cancer (CRC) is increasing worldwide. In Japan, CRC is the leading cause of death in women. Many researchers are actively pursuing molecular biological analyses of the mechanisms involved in the onset and progression of CRC. We now know that genetic and epigenetic abnormalities lead to characteristic gene expression profiles that are strongly linked to clinical outcomes [1,2].

Recent reports have demonstrated the presence of noncoding (nontranslational) RNAs (ncRNAs), which do not carry any genetic information for protein synthesis, in the genome. Many subtypes of ncRNAs have been discovered [3]. MicroRNAs (miRs) are small ncRNAs comprised of 18–25 bases. They bind to the 3' noncoding region of the target mRNA and inhibit the expression of multiple target genes, thereby regulating various cell functions in the body (e.g., differentiation, cell proliferation, and apoptosis) [4,5]. Currently, over 2500 types of miRs have been identified in humans. Associations with various diseases, infectious diseases, central nervous system disorders, and cardiovascular disease have been reported. The initial evidence implicating miRs in CRC pathogenesis was provided in a study by Michael et al. [6], which indicated that *miR-143* and *miR-145* are downregulated in CRC. Since then, many additional studies have demonstrated the tumor suppressive or oncogenic functions of miRs and have described the applications of miRs in the clinical setting as biomarkers or therapeutic targets for CRC.

In the current report, we examined the role of miRs as biomarkers in CRC through a review of meta-analyses and large-scale analyses having strong statistical confidence in the study outcomes. We also discuss current issues in the clinical application of these miRs.

## 2. Methods

For the literature search using PubMed, we used the following search terms on 9 August 2017: “microRNA or miR” AND “colorectal cancer or colon cancer” AND “biomarker.” We then extracted papers related to miRs as a diagnostic, prognostic, or predictive biomarker with meta-analyses or large-scale analyses (total number of samples >100). We found four papers with meta-analyses of prognostic or diagnostic markers. These papers are described in the Tables. We did not list the papers which are described in these papers with meta-analyses in the Tables.

## 3. Biomarkers for the Early Detection and Prediction of Prognosis in CRC

In order to achieve favorable outcomes, early detection of CRC is critical. To this end, clinicians often use fecal occult blood tests (FOBTs) and colonoscopy together or separately [7–9]. FOBTs are performed annually and are noninvasive screening tests that are used to detect blood in stool; these tests have yielded an estimated 24–39% reduction in CRC-related mortality, but are not highly sensitive. Indeed, positive FOBT results lead to frequent colonoscopy screening [9], which is the most common method for detection of CRC. Colonoscopy reduces the risk of CRC by 30–75%; however, an estimated 25% of polyps are not detected during colonoscopy, and the technique is expensive and invasive [9–11]. Given these caveats, cheaper, less-invasive, and more quantitative tests would provide an attractive alternative to the current standard screening methods.

Recently, miRs have been highlighted as diagnostic, prognostic, and predictive biomarkers because of the high tissue specificity, stability, and altered expression in tumor development [12,13]. Thus, miR analysis could offer a less-invasive and more cost-effective alternative to supplement existing screening approaches.

### 3.1. miRs in Tumor Tissues

Tumor tissue is the most important source for the identification of CRC-related miRs. miRs are expressed differently in cancer and normal cells and therefore may constitute a reliable diagnostic biomarker. The roles of miRs in CRC were initially identified by Michael et al. [6], who discovered that *miR-143/miR-145* were significantly reduced in CRC [14].

Various miRs, including *miR-101* [15], the *let7* family, *miR-133b*, *miR-126* [16], and *miR-142-3p* [17], have been found to act as tumor suppressors (tumor-suppressive miRs). Some miRs are highly expressed in CRC cells, playing a major role in creating a microenvironment that allows the cancer cells to thrive. These miRs are known as oncomirs (oncogenic miRs). One of the earliest miRs to be identified as an oncomir was *miR-21*, which plays an important role in the initiation, progression, and metastasis of CRC [18]. Recent studies have identified many miRs as prognostic biomarkers in CRC tissues (Table 1).

**Table 1.** Tissue and blood microRNAs as prognostic biomarkers for CRC.

| microRNA | Tissues                          |                              |
|----------|----------------------------------|------------------------------|
|          | Population with Poor Prognosis * | References                   |
| miR-7    | up                               | [19]                         |
| miR-20a  | up                               | [20,21]                      |
|          | -                                | [22]                         |
| miR-21   | up                               | [23], Meta-analysis: [24,25] |
| miR-22   | down                             | [26]                         |
| miR-31   | -                                | [15]                         |
| miR-92a  | up                               | [27]                         |
|          | -                                | [15]                         |
| miR-93   | down                             | [19]                         |
| miR-101  | -                                | [15]                         |
| miR-155  | up                               | [28]                         |
| let-7c   | down                             | [29]                         |
| miR-126  | down                             | [30]                         |
| miR-130b | up                               | [31]                         |

Table 1. Cont.

| Tissues            |                                  |            |
|--------------------|----------------------------------|------------|
| microRNA           | Population with Poor Prognosis * | References |
| miR-132            | down                             | [32]       |
| miR-139-3p         | down                             | [33]       |
| miR-15             | down                             | [34]       |
| miR-16             | down                             | [34]       |
| miR-181b           | up                               | [20]       |
|                    | -                                | [22]       |
| miR-183            | up                               | [35]       |
| miR-195            | down                             | [19,36]    |
| miR-196a           | up                               | [37]       |
| miR-196b           | up                               | [37]       |
| miR-203            | up                               | [20]       |
|                    | -                                | [22,38]    |
| miR-215            | up                               | [39]       |
| miR-224            | up                               | [40]       |
| miR-340            | down                             | [41]       |
| miR-34-5p          | down                             | [42]       |
| miR-320e           | up                               | [43]       |
| miR-17-5p          | up                               | [44]       |
| miR-106a           | down                             | [44]       |
|                    | up                               | [20]       |
|                    | -                                | [15]       |
| miR-150            | down                             | [45]       |
| miR-29a            | up                               | [46]       |
| miR-372            | up                               | [47]       |
| miR-141            | up                               | [19]       |
| miR-144            | down                             | [48]       |
| miR-145            | -                                | [15]       |
| miR-181a           | up                               | [49]       |
| miR-494            | up                               | [19]       |
| Blood/Serum/Plasma |                                  |            |
| microRNA           | Population with Poor Prognosis * | References |
| miR-21             | up                               | [50]       |
| miR-96             | up                               | [51]       |
| miR-124-5p         | down                             | [52]       |
| miR-141            | up                               | [53]       |
| miR-155            | up                               | [54]       |
| miR-200b           | up                               | [51]       |
| miR-200c           | up                               | [55]       |
| miR-218            | down                             | [56]       |
| miR-221            | up                               | [57]       |
| miR-29a            | up                               | [46]       |
| miR-148a           | down                             | [58]       |
| miR-183            | up                               | [59]       |
| miR-345            | up                               | [60]       |
| miR-19a            | up                               | [61]       |
| miR-203            | up                               | [62,63]    |

\* up, upregulated expression in patients with poor prognosis compared with that in patients with good prognosis. down, downregulated expression in patients with poor prognosis compared with that in patients with good prognosis. -, no significant difference in prognosis between patients with upregulated and downregulated miRs.

### 3.2. miRs in Stool

CRC-specific miRs can also be detected in stools. Analysis of stool miRs has attracted much interest in recent years as a potential noninvasive diagnostic tool for early CRC screening. Tumor cells that are shed from CRC tissues can provide valuable genetic and epigenetic information to facilitate tumor detection [64].

Many studies have reported differential expression of miRs in the stools of patients with CRC [65–73]. The feasibility of isolating stool RNAs and quantifying mature miRs by reverse transcription polymerase chain reaction (RT-PCR) was initially demonstrated in a study by Ahmed et al., in which seven miRs, including *miR-320*, *miR-126*, *miR-484-5p*, *miR-143*, *miR-145*, *miR-16*,

and *miR-125b*, were downregulated in stool collected from patients with CRC when compared with those of healthy controls [65].

A study by Link et al. demonstrated the presence of higher levels of *miR-21* and *miR-106a* in the stool of patients with CRC and colorectal adenoma compared with that in healthy controls by RT-PCR and microarray analysis [68]. Yau et al. found that the expression of *miR-221* and *miR-18a* increased steadily with tumor initiation and progression using an miR expression array [73]. Koga et al. reported that the overall sensitivity and specificity of stool analysis based on upregulation of the *miR-17-92* cluster, *miR-21*, and *miR-135* were 74% and 79%, respectively, suggesting that miR expression analysis in stool samples could be a useful method for CRC screening [72].

Notably, stool *miR-29* was found to be significantly more abundant in patients with rectal cancer than in those with colon cancer, suggesting the feasibility of using differential miR expression patterns as cancer fingerprints [71]. Recent studies have identified many miRs in the stool as diagnostic biomarkers for CRC (Table 2). However, despite the promise of stool miRs for detection of CRC, there are challenges associated with obtaining and handling the samples.

**Table 2.** Blood and stool microRNAs as diagnostic biomarkers for CRC.

| Blood/Serum/Plasma |                       |                           |
|--------------------|-----------------------|---------------------------|
| microRNA           | Population with CRC * | References                |
| miR-18a            | up                    | [74,75]                   |
| miR-19b            | up                    | [74]                      |
| miR-15a            | up                    | [74]                      |
| miR-21             | up                    | Meta-analysis: [24,76–78] |
| miR-24             | down                  | [79]                      |
| miR-29a            | up                    | [74,75,80]                |
| miR-34a            | down                  | [81]                      |
| miR-375            | down                  | [82]                      |
| miR-409-3p         | up                    | [83]                      |
| miR-7              | down                  | [83]                      |
| miR-93             | down                  | [83]                      |
| miR-17-3p          | up                    | [84]                      |
| let-7g             | up                    | [85]                      |
| miR-31             | down                  | [85]                      |
| miR-181b           | down                  | [85]                      |
| miR-203            | down                  | [85]                      |
| miR-378            | up                    | [86]                      |
| miR-20a            | up                    | [75]                      |
| miR-143            | up                    | [75]                      |
| miR-145            | up                    | [75]                      |
| miR-133a           | up                    | [75]                      |
| miR-106b           | up                    | [75]                      |
| miR-335            | up                    | [74]                      |
| miR-601            | down                  | [87]                      |
| miR-223            | up                    | [88,89]                   |
| miR-92             | up                    | [75,80,84,88,89]          |
|                    | down                  | [85]                      |
| miR-760            | down                  | [87]                      |
| miR-423-5p         | down                  | [79]                      |
| miR-320a           | down                  | [79]                      |
| miR-19a            | up                    | [74,78,88]                |
| miR-425-5p         | up                    | [78]                      |
| miR-422a           | down                  | [88]                      |

Table 2. Cont.

| Stool             |                       |                     |
|-------------------|-----------------------|---------------------|
| microRNA          | Population with CRC * | References          |
| miR-17-92 cluster | up                    | [72]                |
| miR-20a           | up                    | [90]                |
| miR-21            | up                    | Meta-analysis: [76] |
| miR-135           | up                    | [70,72]             |
| miR-144 *         | up                    | [91]                |
| miR-29a           | down                  | [71]                |
| miR-223           | up                    | [89]                |
| miR-221           | up                    | [73]                |
| miR-92a           | up                    | [69,89]             |
| miR-224           | down                  | [71]                |

\* up, upregulated expression in population with CRC compared with healthy volunteers. down, downregulated expression in population with CRC compared with healthy volunteers.

### 3.3. miRs in Serum/Plasma

Cancer cells release miRs into the peripheral blood [92]. Thus, circulating miRs could be detected from serum and plasma; these molecules are currently being exploited aggressively as potential biomarkers for the diagnosis and monitoring of cancer progression or treatment in patients with CRC.

The first report of miRs detected in the serum of patients with CRC was published by Chen et al. in 2008 [93]. They identified 69 miRs in the serum of patients with CRC but not in the serum of healthy controls.

Circulating miRs are present in various forms and have even been found within exosomes. As extracellular vesicles secreted from cells by exocytosis, exosomes are found in most circulating body fluids and contain proteins, mRNAs, and miRs [94]. Notably, exosomal miRs are more stable than other miR forms because they are not degraded by endogenous RNase. Accordingly, exosomal miRs may have potential applications as cancer-specific biomarkers.

Exosomes are key contributors to intercellular communication. Exosomes can carry a number of molecules, such as DNAs, mRNAs, proteins, and miRs, to recipient cells [95]. Hence, exosomes and their cargo transfer specific messages to recipient cells and change the behavior of these cells. Many studies have shown that exosomes released from cancer cells are important players in tumor progression in several diseases, including CRC [95,96]. We previously reported that abundant expression of exosomal *miR-19a* in serum was a prognostic biomarker for recurrence in patients with CRC [61]. Recent studies have identified many miRs in blood as prognostic or diagnostic biomarkers in CRC (Tables 1 and 2).

## 4. Biomarkers for Prediction of Drug Efficacy

Drug resistance is a major obstacle to effective cancer therapy. Selecting patients who would benefit from drug treatment will help to promote therapeutic efficacy and avoid resource waste. miRs are expected to have applications as predictive biomarkers for therapeutic responses because some miRs have been shown to induce chemoresistance and to be associated with poor prognosis in various malignancies, including CRC [97–100].

Cohort research in the United States of America and China has shown that increases in *miR-21* expression are associated with resistance to 5-fluorouracil (5-FU) chemotherapy [22]. In this in vitro study, resistance was induced by inhibiting the DNA repair protein MutS homolog2 (MSH2). Additionally, *miR-140*, *miR-215*, *miR-224*, and *miR-20a* have also been reported to contribute to drug resistance. Currently, studies are underway to determine if the expression of these miRs can be used to predict chemotherapy efficacy or as treatment targets.

The list of miRs as predictive biomarkers of response to vascular endothelial growth factor- or epidermal growth factor receptor (EGFR)-targeted therapy and chemotherapy, which are used in the standard treatment regimen for CRC, is shown in Table 3.

**Table 3.** Tissue and blood microRNA biomarkers for therapeutic response in CRC.

| Tissues                   |            |                |               |            |
|---------------------------|------------|----------------|---------------|------------|
| Therapy                   | microRNA   | Non-responders | Pre-therapy * | References |
| Anti-angiogenetic therapy | miR-126    | up             |               | [101]      |
| Anti-EGFR therapy         | miR-7      | down           |               | [102]      |
| Chemotherapy              | miR-31-5p  | up             |               | [103]      |
|                           | Let-7c     | down           |               | [104]      |
|                           | miR-99a    | down           |               | [104]      |
|                           | miR-125b   | down           |               | [104]      |
|                           | miR-17-5p  | up             |               | [105]      |
|                           | miR-21     | up             |               | [106]      |
|                           | miR-143    | down           |               | [107]      |
|                           | miR-148a   | down           |               | [108]      |
|                           | miR-148b   | down           |               | [109]      |
|                           | miR-150    | down           |               | [45]       |
|                           | miR-200c   | down           |               | [110]      |
|                           | miR-320    | down           |               | [111]      |
|                           | miR-625-3p | up             |               | [112]      |
|                           | miR-181b   | up             |               | [112]      |
|                           | miR-27b    | up             |               | [112]      |
|                           | miR-664-3p | down           |               | [113]      |
|                           | miR-455-5p | up             |               | [113]      |
| miR-196b-5p               | down       |                | [113]         |            |
| miR-592                   | down       |                | [113]         |            |
| Blood/Serum/Plasma        |            |                |               |            |
| Therapy                   | microRNA   | Non-responders | Pre-therapy * | References |
| Anti-angiogenetic therapy | miR-126    | up             |               | [114]      |
| Anti-EGFR therapy         | miR-345    | up             |               | [60]       |
| Chemotherapy              | miR-126    | up             |               | [114]      |
|                           | miR-143    | down           |               | [107]      |
|                           | miR-345    | up             |               | [60]       |
|                           | miR-1914 * | down           |               | [115]      |
|                           | miR-1915   | down           |               | [115]      |

\* up, upregulated expression at pre-therapy in non-responders compared with responders. down, downregulated expression at pre-therapy in non-responders compared with responders.

## 5. Limitations

### 5.1. Conflicting Functions of miRs

Recently, studies have focused on the contrasting roles of miRs in oncogenesis and tumor suppression [38,116]. Some miRs have been reported to be upregulated in one report and downregulated in another report, indicating the contradictory functions of these miRs in CRC. For example, *miR-27a* was found to be downregulated and showed tumor-suppressive functions in CRC, targeting *Stat3* and *Smad2* [117]; in other studies [118,119], this same miR was found to be upregulated and showed oncogenic functions in CRC [120]. Additionally, *miR-155* has also been shown experimentally to have conflicting functions in mouse breast cancer [121].

Tumor-derived miRs may have site-dependent functions that promote tumor development. Thus, miR localization should be considered for clinical cancer screening or anticancer therapy for targeting miRs [63].

### 5.2. Sensitivity/Specificity

Despite the potential of individual miRs as biomarkers or therapeutic targets, combinations of miRs are expected to enhance the sensitivity and specificity for cancer diagnosis and increase the intensity of treatment [122]. Indeed, combinations of miRs have been shown to provide high diagnostic accuracy, with a high area under the curve, in lung [123], pancreatic [124], liver [125], and breast cancers [126,127].

Evaluation of miR panels using large, independent patient cohorts must be performed before miR biomarkers can be implemented in the clinical setting in CRC. Furthermore, combinations of some miR panels and cancer antigens or molecular biomarkers should be assessed as reported in breast cancer [127,128] and pancreatic cancer [124].

### 5.3. Internal Control

Endogenous control is crucial for the normalization, reliability, and reproducibility of diagnostic results because it helps to normalize differences among sample qualities and variations during the detection process. In pooled studies, internal controls have included *miR-16*, *U6*, and *miR-451*, of which *U6* and *miR-16* have been most popular [122]. However, it is still unclear whether these miRs are good internal references [129]. The suitability of the miR as a reference may depend on the type of organ or tissue. Caution is required when selecting an internal control gene for evaluating the expression profiles of miRs in patients with CRC.

## 6. Development of Novel Diagnostic Methods Using miRs

In Japan, the New Energy and Industrial Technology Development Organization (NEDO; <http://www.nedo.go.jp/english/index.html>) has carried out a collaborative industry/government/academia project since 2014 to develop a cutting-edge, next-generation cancer diagnostic system (Project Number: 14009). Since 2015, this project has been now supported by the Japan Agency for Medical Research and Development (AMED). The objective is to construct a database of miR expression in body fluids, conduct a comprehensive analysis, discover early expression markers for 13 cancers (including breast cancer and CRC) or other diseases (such as dementia), and create biological tools that could be practically applied for the detection of these markers.

## 7. Conclusions

As highlighted in this review, miRs have considerable potential as biomarkers and therapeutic targets because miRs can drive and modulate tumorigenesis and tumor progression in CRC. However, the clinical significance of miRs as biomarkers is still not conclusive, and independent validation studies are needed for clinical application.

The use of miRs as biomarkers for CRC would provide a new, less-invasive technique to screen for CRC and determine prognosis. A screening panel consisting of multiple miRs may provide the most precise and effective screening tool for CRC.

Interestingly, recent studies have demonstrated that miRs may be critical regulators of immune responses, and aberrant expression or dysfunction of miRs in the immune system is associated with cancers [130–132]. Moreover, emerging evidence has demonstrated that immune-associated miRs are dysregulated in both tumor cells and immune cells, suggesting that miRs could be involved in communication between tumor cells and immune cells [133]. Much effort has been made to discover the precise role of miRs in the regulation of antitumor immune responses.

In this review, we found that most papers reporting the clinical significance of miRs in CRC were small-scale studies, and a meta-analysis was reported only for *miR-21* in CRC. More meta-analyses or large-scale analyses are required to provide reliable data for clarification of the clinical significance of miRs for clinical applications. A large prospective clinical study may be the best approach.

Finally, we hope that elucidation of the molecular mechanisms of miRs in CRC will lead to the discovery of early diagnostic methods or allow the development of next-generation oligonucleotide drugs and other novel therapies.

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## References

1. The Cancer Genome Atlas Network. Comprehensive molecular characterization of human colon and rectal cancer. *Nature* **2012**, *487*, 330–337.
2. Uchi, R.; Takahashi, Y.; Niida, A.; Shimamura, T.; Hirata, H.; Sugimachi, K.; Sawada, G.; Iwaya, T.; Kurashige, J.; Shinden, Y.; et al. Integrated multiregional analysis proposing a new model of colorectal cancer evolution. *PLoS Genet.* **2016**, *12*, e1005778. [[CrossRef](#)] [[PubMed](#)]
3. Chu, C.Y.; Rana, T.M. Small RNAs: Regulators and guardians of the genome. *J. Cell. Physiol.* **2007**, *213*, 412–419. [[CrossRef](#)] [[PubMed](#)]
4. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* **2004**, *116*, 281–297. [[CrossRef](#)]
5. Lin, S.; Gregory, R.I. MicroRNA biogenesis pathways in cancer. *Nat. Rev. Cancer* **2015**, *15*, 321–333. [[CrossRef](#)] [[PubMed](#)]
6. Michael, M.Z.; SM, O.C.; van Holst Pellekaan, N.G.; Young, G.P.; James, R.J. Reduced accumulation of specific microRNAs in colorectal neoplasia. *Mol. Cancer Res.* **2003**, *1*, 882–891. [[PubMed](#)]
7. Zavoral, M.; Suchanek, S.; Zavada, F.; Dusek, L.; Muzik, J.; Seifert, B.; Fric, P. Colorectal cancer screening in Europe. *World J. Gastroenterol.* **2009**, *15*, 5907–5915. [[CrossRef](#)] [[PubMed](#)]
8. Altobelli, E.; Lattanzi, A.; Paduano, R.; Varassi, G.; di Orio, F. Colorectal cancer prevention in Europe: Burden of disease and status of screening programs. *Prev. Med.* **2014**, *62*, 132–141. [[CrossRef](#)] [[PubMed](#)]
9. Sovich, J.L.; Sartor, Z.; Misra, S. Developments in screening tests and strategies for colorectal cancer. *Biomed. Res. Int.* **2015**, *2015*, 326728. [[CrossRef](#)] [[PubMed](#)]
10. Van Rijn, J.C.; Reitsma, J.B.; Stoker, J.; Bossuyt, P.M.; van Deventer, S.J.; Dekker, E. Polyp miss rate determined by tandem colonoscopy: A systematic review. *Am. J. Gastroenterol.* **2006**, *101*, 343–350. [[CrossRef](#)] [[PubMed](#)]
11. Leufkens, A.M.; van Oijen, M.G.; Vleggaar, F.P.; Siersema, P.D. Factors influencing the miss rate of polyps in a back-to-back colonoscopy study. *Endoscopy* **2012**, *44*, 470–475. [[CrossRef](#)] [[PubMed](#)]
12. Dong, Y.; Wu, W.K.; Wu, C.W.; Sung, J.J.; Yu, J.; Ng, S.S. MicroRNA dysregulation in colorectal cancer: A clinical perspective. *Br. J. Cancer* **2011**, *104*, 893–898. [[CrossRef](#)] [[PubMed](#)]
13. Ren, A.; Dong, Y.; Tsoi, H.; Yu, J. Detection of miRNA as non-invasive biomarkers of colorectal cancer. *Int. J. Mol. Sci.* **2015**, *16*, 2810–2823. [[CrossRef](#)] [[PubMed](#)]
14. Akao, Y.; Nakagawa, Y.; Naoe, T. MicroRNAs 143 and 145 are possible common onco-microRNAs in human cancers. *Oncol. Rep.* **2006**, *16*, 845–850. [[CrossRef](#)] [[PubMed](#)]
15. Schee, K.; Boye, K.; Abrahamsen, T.W.; Fodstad, O.; Flatmark, K. Clinical relevance of microRNA miR-21, miR-31, miR-92a, miR-101, miR-106a, and miR-145 in colorectal cancer. *BMC Cancer* **2012**, *12*, 505. [[CrossRef](#)] [[PubMed](#)]
16. Xuan, Y.; Yang, H.; Zhao, L.; Lau, W.B.; Lau, B.; Ren, N.; Hu, Y.; Yi, T.; Zhao, X.; Zhou, S.; et al. MicroRNAs in colorectal cancer: Small molecules with big functions. *Cancer Lett.* **2015**, *360*, 89–105. [[CrossRef](#)] [[PubMed](#)]
17. Shen, W.W.; Zeng, Z.; Zhu, W.X.; Fu, G.H. MiR-142-3p functions as a tumor suppressor by targeting CD133, ABCG2, and LGR5 in colon cancer cells. *J. Mol. Med.* **2013**, *91*, 989–1000. [[CrossRef](#)] [[PubMed](#)]
18. Schetter, A.J.; Okayama, H.; Harris, C.C. The role of microRNAs in colorectal cancer. *Cancer J.* **2012**, *18*, 244–252. [[CrossRef](#)] [[PubMed](#)]
19. Yang, I.P.; Tsai, H.L.; Miao, Z.F.; Huang, C.W.; Kuo, C.H.; Wu, J.Y.; Wang, W.M.; Juo, S.H.; Wang, J.Y. Development of a deregulating microRNA panel for the detection of early relapse in postoperative colorectal cancer patients. *J. Transl. Med.* **2016**, *14*, 108. [[CrossRef](#)] [[PubMed](#)]



20. Bovell, L.C.; Shanmugam, C.; Putcha, B.D.; Katkooori, V.R.; Zhang, B.; Bae, S.; Singh, K.P.; Grizzle, W.E.; Manne, U. The prognostic value of microRNAs varies with patient race/ethnicity and stage of colorectal cancer. *Clin. Cancer Res.* **2013**, *19*, 3955–3965. [[CrossRef](#)] [[PubMed](#)]
21. Cheng, D.; Zhao, S.; Tang, H.; Zhang, D.; Sun, H.; Yu, F.; Jiang, W.; Yue, B.; Wang, J.; Zhang, M.; et al. MicroRNA-20a-5p promotes colorectal cancer invasion and metastasis by downregulating SMAD4. *Oncotarget* **2016**, *7*, 45199–45213. [[CrossRef](#)] [[PubMed](#)]
22. Schetter, A.J.; Leung, S.Y.; Sohn, J.J.; Zanetti, K.A.; Bowman, E.D.; Yanaihara, N.; Yuen, S.T.; Chan, T.L.; Kwong, D.L.; Au, G.K.; et al. MicroRNA expression profiles associated with prognosis and therapeutic outcome in colon adenocarcinoma. *JAMA* **2008**, *299*, 425–436. [[CrossRef](#)] [[PubMed](#)]
23. Mima, K.; Nishihara, R.; Yang, J.; Dou, R.; Masugi, Y.; Shi, Y.; da Silva, A.; Cao, Y.; Song, M.; Nowak, J.; et al. MicroRNA miR21 (miR-21) and PTGS2 expression in colorectal cancer and patient survival. *Clin. Cancer Res.* **2016**, *22*, 3841–3848. [[CrossRef](#)] [[PubMed](#)]
24. Peng, Q.; Zhang, X.; Min, M.; Zou, L.; Shen, P.; Zhu, Y. The clinical role of microRNA-21 as a promising biomarker in the diagnosis and prognosis of colorectal cancer: A systematic review and meta-analysis. *Oncotarget* **2017**, *8*, 44893–44909. [[CrossRef](#)] [[PubMed](#)]
25. Chen, Z.; Liu, H.; Jin, W.; Ding, Z.; Zheng, S.; Yu, Y. Tissue microRNA-21 expression predicted recurrence and poor survival in patients with colorectal cancer—A meta-analysis. *Onco. Targets Ther.* **2016**, *9*, 2615–2624. [[PubMed](#)]
26. Xia, S.S.; Zhang, G.J.; Liu, Z.L.; Tian, H.P.; He, Y.; Meng, C.Y.; Li, L.F.; Wang, Z.W.; Zhou, T. MicroRNA-22 suppresses the growth, migration and invasion of colorectal cancer cells through a sp1 negative feedback loop. *Oncotarget* **2017**, *8*, 36266–36278. [[CrossRef](#)] [[PubMed](#)]
27. Ke, T.W.; Wei, P.L.; Yeh, K.T.; Chen, W.T.; Cheng, Y.W. MiR-92a promotes cell metastasis of colorectal cancer through PTEN-mediated PI3K/AKT pathway. *Ann. Surg. Oncol.* **2015**, *22*, 2649–2655. [[CrossRef](#)] [[PubMed](#)]
28. Shibuya, H.; Iinuma, H.; Shimada, R.; Horiuchi, A.; Watanabe, T. Clinicopathological and prognostic value of microRNA-21 and microRNA-155 in colorectal cancer. *Oncology* **2010**, *79*, 313–320. [[CrossRef](#)] [[PubMed](#)]
29. Han, H.B.; Gu, J.; Zuo, H.J.; Chen, Z.G.; Zhao, W.; Li, M.; Ji, D.B.; Lu, Y.Y.; Zhang, Z.Q. Let-7c functions as a metastasis suppressor by targeting MMP11 and PBX3 in colorectal cancer. *J. Pathol.* **2012**, *226*, 544–555. [[CrossRef](#)] [[PubMed](#)]
30. Hansen, T.F.; Kjaer-Frifeldt, S.; Morgenthaler, S.; Blondal, T.; Lindebjerg, J.; Jakobsen, A.; Sorensen, F.B. The prognostic value of microRNA-126 and microvessel density in patients with stage ii colon cancer: Results from a population cohort. *J. Transl. Med.* **2014**, *12*, 254. [[CrossRef](#)] [[PubMed](#)]
31. Colangelo, T.; Fucci, A.; Votino, C.; Sabatino, L.; Pancione, M.; Laudanna, C.; Binaschi, M.; Bigioni, M.; Maggi, C.A.; Parente, D.; et al. MicroRNA-130b promotes tumor development and is associated with poor prognosis in colorectal cancer. *Neoplasia* **2013**, *15*, 1086–1099. [[CrossRef](#)] [[PubMed](#)]
32. Mokutani, Y.; Uemura, M.; Munakata, K.; Okuzaki, D.; Haraguchi, N.; Takahashi, H.; Nishimura, J.; Hata, T.; Murata, K.; Takemasa, I.; et al. Down-regulation of microRNA-132 is associated with poor prognosis of colorectal cancer. *Ann. Surg. Oncol.* **2016**, *23*, 599–608. [[CrossRef](#)] [[PubMed](#)]
33. Liu, X.; Duan, B.; Dong, Y.; He, C.; Zhou, H.; Sheng, H.; Gao, H.; Zhang, X. MicroRNA-139-3p indicates a poor prognosis of colon cancer. *Int. J. Clin. Exp. Pathol.* **2014**, *7*, 8046–8052. [[PubMed](#)]
34. Xiao, G.; Tang, H.; Wei, W.; Li, J.; Ji, L.; Ge, J. Aberrant expression of microRNA-15a and microRNA-16 synergistically associates with tumor progression and prognosis in patients with colorectal cancer. *Gastroenterol. Res. Pract.* **2014**, *2014*, 364549. [[CrossRef](#)] [[PubMed](#)]
35. Zhou, T.; Zhang, G.J.; Zhou, H.; Xiao, H.X.; Li, Y. Overexpression of microRNA-183 in human colorectal cancer and its clinical significance. *Eur. J. Gastroenterol. Hepatol.* **2014**, *26*, 229–233. [[CrossRef](#)] [[PubMed](#)]
36. Wang, X.; Wang, J.; Ma, H.; Zhang, J.; Zhou, X. Downregulation of miR-195 correlates with lymph node metastasis and poor prognosis in colorectal cancer. *Med. Oncol.* **2012**, *29*, 919–927. [[CrossRef](#)] [[PubMed](#)]
37. Ge, J.; Chen, Z.; Li, R.; Lu, T.; Xiao, G. Upregulation of microRNA-196a and microRNA-196b cooperatively correlate with aggressive progression and unfavorable prognosis in patients with colorectal cancer. *Cancer Cell. Int.* **2014**, *14*, 128. [[CrossRef](#)] [[PubMed](#)]
38. Liang, Y.; Yang, W.; Zhu, Y.; Yuan, Y. Prognostic role of microRNA-203 in various carcinomas: Evidence from a meta-analysis involving 13 studies. *Springerplus* **2016**, *5*, 1538. [[CrossRef](#)] [[PubMed](#)]

39. Karaayvaz, M.; Pal, T.; Song, B.; Zhang, C.; Georgakopoulos, P.; Mehmood, S.; Burke, S.; Shroyer, K.; Ju, J. Prognostic significance of miR-215 in colon cancer. *Clin. Colorectal Cancer* **2011**, *10*, 340–347. [[CrossRef](#)] [[PubMed](#)]
40. Liao, W.T.; Li, T.T.; Wang, Z.G.; Wang, S.Y.; He, M.R.; Ye, Y.P.; Qi, L.; Cui, Y.M.; Wu, P.; Jiao, H.L.; et al. MicroRNA-224 promotes cell proliferation and tumor growth in human colorectal cancer by repressing PHLPP1 and PHLPP2. *Clin. Cancer Res.* **2013**, *19*, 4662–4672. [[CrossRef](#)] [[PubMed](#)]
41. Takeyama, H.; Yamamoto, H.; Yamashita, S.; Wu, X.; Takahashi, H.; Nishimura, J.; Haraguchi, N.; Miyake, Y.; Suzuki, R.; Murata, K.; et al. Decreased miR-340 expression in bone marrow is associated with liver metastasis of colorectal cancer. *Mol. Cancer Ther.* **2014**, *13*, 976–985. [[CrossRef](#)] [[PubMed](#)]
42. Gao, J.; Li, N.; Dong, Y.; Li, S.; Xu, L.; Li, X.; Li, Y.; Li, Z.; Ng, S.S.; Sung, J.J.; et al. MiR-34a-5p suppresses colorectal cancer metastasis and predicts recurrence in patients with stage II/III colorectal cancer. *Oncogene* **2015**, *34*, 4142–4152. [[CrossRef](#)] [[PubMed](#)]
43. Perez-Carbonell, L.; Sinicrope, F.A.; Alberts, S.R.; Oberg, A.L.; Balaguer, F.; Castells, A.; Boland, C.R.; Goel, A. MiR-320e is a novel prognostic biomarker in colorectal cancer. *Br. J. Cancer.* **2015**, *113*, 83–90. [[CrossRef](#)] [[PubMed](#)]
44. Diaz, R.; Silva, J.; Garcia, J.M.; Lorenzo, Y.; Garcia, V.; Pena, C.; Rodriguez, R.; Munoz, C.; Garcia, F.; Bonilla, F.; et al. Deregulated expression of miR-106a predicts survival in human colon cancer patients. *Genes Chromosomes Cancer* **2008**, *47*, 794–802. [[CrossRef](#)] [[PubMed](#)]
45. Ma, Y.; Zhang, P.; Wang, F.; Zhang, H.; Yang, J.; Peng, J.; Liu, W.; Qin, H. MiR-150 as a potential biomarker associated with prognosis and therapeutic outcome in colorectal cancer. *Gut* **2012**, *61*, 1447–1453. [[CrossRef](#)] [[PubMed](#)]
46. Weissmann-Brenner, A.; Kushnir, M.; Lithwick Yanai, G.; Aharonov, R.; Gibori, H.; Purim, O.; Kundel, Y.; Morgenstern, S.; Halperin, M.; Niv, Y.; et al. Tumor microRNA-29a expression and the risk of recurrence in stage II colon cancer. *Int. J. Oncol.* **2012**, *40*, 2097–2103. [[PubMed](#)]
47. Yamashita, S.; Yamamoto, H.; Mimori, K.; Nishida, N.; Takahashi, H.; Haraguchi, N.; Tanaka, F.; Shibata, K.; Sekimoto, M.; Ishii, H.; et al. MicroRNA-372 is associated with poor prognosis in colorectal cancer. *Oncology* **2012**, *82*, 205–212. [[CrossRef](#)] [[PubMed](#)]
48. Iwaya, T.; Yokobori, T.; Nishida, N.; Kogo, R.; Sudo, T.; Tanaka, F.; Shibata, K.; Sawada, G.; Takahashi, Y.; Ishibashi, M.; et al. Downregulation of miR-144 is associated with colorectal cancer progression via activation of mTOR signaling pathway. *Carcinogenesis* **2012**, *33*, 2391–2397. [[CrossRef](#)] [[PubMed](#)]
49. Nishimura, J.; Handa, R.; Yamamoto, H.; Tanaka, F.; Shibata, K.; Mimori, K.; Takemasa, I.; Mizushima, T.; Ikeda, M.; Sekimoto, M.; et al. MicroRNA-181a is associated with poor prognosis of colorectal cancer. *Oncol. Rep.* **2012**, *28*, 2221–2226. [[CrossRef](#)] [[PubMed](#)]
50. Menendez, P.; Padilla, D.; Villarejo, P.; Palomino, T.; Nieto, P.; Menendez, J.M.; Rodriguez-Montes, J.A. Prognostic implications of serum microRNA-21 in colorectal cancer. *J. Surg. Oncol.* **2013**, *108*, 369–373. [[CrossRef](#)] [[PubMed](#)]
51. Sun, Y.; Liu, Y.; Cogdell, D.; Calin, G.A.; Sun, B.; Kopetz, S.; Hamilton, S.R.; Zhang, W. Examining plasma microRNA markers for colorectal cancer at different stages. *Oncotarget* **2016**, *7*, 11434–11449. [[CrossRef](#)] [[PubMed](#)]
52. Jinushi, T.; Shibayama, Y.; Kinoshita, I.; Oizumi, S.; Jinushi, M.; Aota, T.; Takahashi, T.; Horita, S.; Dosaka-Akita, H.; Iseki, K. Low expression levels of microRNA-124-5p correlated with poor prognosis in colorectal cancer via targeting of SMC4. *Cancer Med.* **2014**, *3*, 1544–1552. [[CrossRef](#)] [[PubMed](#)]
53. Cheng, H.; Zhang, L.; Cogdell, D.E.; Zheng, H.; Schetter, A.J.; Nykter, M.; Harris, C.C.; Chen, K.; Hamilton, S.R.; Zhang, W. Circulating plasma miR-141 is a novel biomarker for metastatic colon cancer and predicts poor prognosis. *PLoS ONE* **2011**, *6*, e17745. [[CrossRef](#)] [[PubMed](#)]
54. Lv, Z.C.; Fan, Y.S.; Chen, H.B.; Zhao, D.W. Investigation of microRNA-155 as a serum diagnostic and prognostic biomarker for colorectal cancer. *Tumour. Biol.* **2015**, *36*, 1619–1625. [[CrossRef](#)] [[PubMed](#)]
55. Toiyama, Y.; Hur, K.; Tanaka, K.; Inoue, Y.; Kusunoki, M.; Boland, C.R.; Goel, A. Serum mir-200c is a novel prognostic and metastasis-predictive biomarker in patients with colorectal cancer. *Ann. Surg.* **2014**, *259*, 735–743. [[CrossRef](#)] [[PubMed](#)]
56. Yu, H.; Gao, G.; Jiang, L.; Guo, L.; Lin, M.; Jiao, X.; Jia, W.; Huang, J. Decreased expression of miR-218 is associated with poor prognosis in patients with colorectal cancer. *Int. J. Clin. Exp. Pathol.* **2013**, *6*, 2904–2911. [[PubMed](#)]

57. Pu, X.X.; Huang, G.L.; Guo, H.Q.; Guo, C.C.; Li, H.; Ye, S.; Ling, S.; Jiang, L.; Tian, Y.; Lin, T.Y. Circulating miR-221 directly amplified from plasma is a potential diagnostic and prognostic marker of colorectal cancer and is correlated with p53 expression. *J. Gastroenterol. Hepatol.* **2010**, *25*, 1674–1680. [[CrossRef](#)] [[PubMed](#)]
58. Tsai, H.L.; Yang, I.P.; Huang, C.W.; Ma, C.J.; Kuo, C.H.; Lu, C.Y.; Juo, S.H.; Wang, J.Y. Clinical significance of microRNA-148a in patients with early relapse of stage II stage and III colorectal cancer after curative resection. *Transl. Res.* **2013**, *162*, 258–268. [[CrossRef](#)] [[PubMed](#)]
59. Yuan, D.; Li, K.; Zhu, K.; Yan, R.; Dang, C. Plasma miR-183 predicts recurrence and prognosis in patients with colorectal cancer. *Cancer Biol. Ther.* **2015**, *16*, 268–275. [[CrossRef](#)] [[PubMed](#)]
60. Schou, J.V.; Rossi, S.; Jensen, B.V.; Nielsen, D.L.; Pfeiffer, P.; Hogdall, E.; Yilmaz, M.; Tejpar, S.; Delorenzi, M.; Kruhoffer, M.; et al. MiR-345 in metastatic colorectal cancer: A non-invasive biomarker for clinical outcome in non-kras mutant patients treated with 3rd line cetuximab and irinotecan. *PLoS ONE* **2014**, *9*, e99886. [[CrossRef](#)] [[PubMed](#)]
61. Matsumura, T.; Sugimachi, K.; Iinuma, H.; Takahashi, Y.; Kurashige, J.; Sawada, G.; Ueda, M.; Uchi, R.; Ueo, H.; Takano, Y.; et al. Exosomal microRNA in serum is a novel biomarker of recurrence in human colorectal cancer. *Br. J. Cancer* **2015**, *113*, 275–281. [[CrossRef](#)] [[PubMed](#)]
62. Hur, K.; Toiyama, Y.; Okugawa, Y.; Ide, S.; Imaoka, H.; Boland, C.R.; Goel, A. Circulating microRNA-203 predicts prognosis and metastasis in human colorectal cancer. *Gut* **2017**, *66*, 654–665. [[CrossRef](#)] [[PubMed](#)]
63. Takano, Y.; Masuda, T.; Iinuma, H.; Yamaguchi, R.; Sato, K.; Tobo, T.; Hirata, H.; Kuroda, Y.; Nambara, S.; Hayashi, N.; et al. Circulating exosomal microRNA-203 is associated with metastasis possibly via inducing tumor-associated macrophages in colorectal cancer. *Oncotarget* **2017**. [[CrossRef](#)]
64. Lee, J.K.; Liles, E.G.; Bent, S.; Levin, T.R.; Corley, D.A. Accuracy of fecal immunochemical tests for colorectal cancer: Systematic review and meta-analysis. *Ann. Intern. Med.* **2014**, *160*, 171. [[CrossRef](#)] [[PubMed](#)]
65. Ahmed, F.E.; Jeffries, C.D.; Vos, P.W.; Flake, G.; Nuovo, G.J.; Sinar, D.R.; Naziri, W.; Marcuard, S.P. Diagnostic microRNA markers for screening sporadic human colon cancer and active ulcerative colitis in stool and tissue. *Cancer Genom. Proteom.* **2009**, *6*, 281–295.
66. Ahmed, F.E.; Ahmed, N.C.; Vos, P.W.; Bonnerup, C.; Atkins, J.N.; Casey, M.; Nuovo, G.J.; Naziri, W.; Wiley, J.E.; Mota, H.; et al. Diagnostic microRNA markers to screen for sporadic human colon cancer in stool: I. Proof of principle. *Cancer Genom. Proteom.* **2013**, *10*, 93–113.
67. Li, J.M.; Zhao, R.H.; Li, S.T.; Xie, C.X.; Jiang, H.H.; Ding, W.J.; Du, P.; Chen, W.; Yang, M.; Cui, L. Down-regulation of fecal miR-143 and miR-145 as potential markers for colorectal cancer. *Saudi Med. J.* **2012**, *33*, 24–29. [[PubMed](#)]
68. Link, A.; Balaguer, F.; Shen, Y.; Nagasaka, T.; Lozano, J.J.; Boland, C.R.; Goel, A. Fecal microRNAs as novel biomarkers for colon cancer screening. *Cancer Epidemiol. Biomarkers Prev.* **2010**, *19*, 1766–1774. [[CrossRef](#)] [[PubMed](#)]
69. Wu, C.W.; Ng, S.S.; Dong, Y.J.; Ng, S.C.; Leung, W.W.; Lee, C.W.; Wong, Y.N.; Chan, F.K.; Yu, J.; Sung, J.J. Detection of miR-92a and miR-21 in stool samples as potential screening biomarkers for colorectal cancer and polyps. *Gut* **2012**, *61*, 739–745. [[CrossRef](#)] [[PubMed](#)]
70. Wu, C.W.; Ng, S.C.; Dong, Y.; Tian, L.; Ng, S.S.; Leung, W.W.; Law, W.T.; Yau, T.O.; Chan, F.K.; Sung, J.J.; et al. Identification of microRNA-135b in stool as a potential noninvasive biomarker for colorectal cancer and adenoma. *Clin. Cancer Res.* **2014**, *20*, 2994–3002. [[CrossRef](#)] [[PubMed](#)]
71. Zhu, Y.; Xu, A.; Li, J.; Fu, J.; Wang, G.; Yang, Y.; Cui, L.; Sun, J. Fecal miR-29a and miR-224 as the noninvasive biomarkers for colorectal cancer. *Cancer Biomark.* **2016**, *16*, 259–264. [[CrossRef](#)] [[PubMed](#)]
72. Koga, Y.; Yasunaga, M.; Takahashi, A.; Kuroda, J.; Moriya, Y.; Akasu, T.; Fujita, S.; Yamamoto, S.; Baba, H.; Matsumura, Y. MicroRNA expression profiling of exfoliated colonocytes isolated from feces for colorectal cancer screening. *Cancer Prev. Res.* **2010**, *3*, 1435–1442. [[CrossRef](#)] [[PubMed](#)]
73. Yau, T.O.; Wu, C.W.; Dong, Y.; Tang, C.M.; Ng, S.S.; Chan, F.K.; Sung, J.J.; Yu, J. MicroRNA-221 and microRNA-18a identification in stool as potential biomarkers for the non-invasive diagnosis of colorectal carcinoma. *Br. J. Cancer* **2014**, *111*, 1765–1771. [[CrossRef](#)] [[PubMed](#)]
74. Giraldez, M.D.; Lozano, J.J.; Ramirez, G.; Hijona, E.; Bujanda, L.; Castells, A.; Gironella, M. Circulating microRNAs as biomarkers of colorectal cancer: Results from a genome-wide profiling and validation study. *Clin. Gastroenterol. Hepatol.* **2013**, *11*, 681–688. [[CrossRef](#)] [[PubMed](#)]
75. Luo, X.; Stock, C.; Burwinkel, B.; Brenner, H. Identification and evaluation of plasma microRNAs for early detection of colorectal cancer. *PLoS ONE* **2013**, *8*, e62880. [[CrossRef](#)] [[PubMed](#)]

76. Jiang, J.X.; Zhang, N.; Liu, Z.M.; Wang, Y.Y. Detection of microRNA-21 expression as a potential screening biomarker for colorectal cancer: A meta-analysis. *Asian Pac. J. Cancer Prev.* **2014**, *15*, 7583–7588. [[CrossRef](#)] [[PubMed](#)]
77. Yu, W.; Wang, Z.; Shen, L.I.; Wei, Q. Circulating microRNA-21 as a potential diagnostic marker for colorectal cancer: A meta-analysis. *Mol. Clin. Oncol.* **2016**, *4*, 237–244. [[CrossRef](#)] [[PubMed](#)]
78. Zhu, M.; Huang, Z.; Zhu, D.; Zhou, X.; Shan, X.; Qi, L.W.; Wu, L.; Cheng, W.; Zhu, J.; Zhang, L.; et al. A panel of microRNA signature in serum for colorectal cancer diagnosis. *Oncotarget* **2017**, *8*, 17081–17091. [[CrossRef](#)] [[PubMed](#)]
79. Fang, Z.; Tang, J.; Bai, Y.; Lin, H.; You, H.; Jin, H.; Lin, L.; You, P.; Li, J.; Dai, Z.; et al. Plasma levels of microRNA-24, microRNA-320a, and microRNA-423–5p are potential biomarkers for colorectal carcinoma. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 86. [[CrossRef](#)] [[PubMed](#)]
80. Huang, Z.; Huang, D.; Ni, S.; Peng, Z.; Sheng, W.; Du, X. Plasma microRNAs are promising novel biomarkers for early detection of colorectal cancer. *Int. J. Cancer* **2010**, *127*, 118–126. [[CrossRef](#)] [[PubMed](#)]
81. Nugent, M.; Miller, N.; Kerin, M.J. Circulating miR-34a levels are reduced in colorectal cancer. *J. Surg. Oncol.* **2012**, *106*, 947–952. [[CrossRef](#)] [[PubMed](#)]
82. Xu, L.; Li, M.; Wang, M.; Yan, D.; Feng, G.; An, G. The expression of microRNA-375 in plasma and tissue is matched in human colorectal cancer. *BMC Cancer* **2014**, *14*, 714. [[CrossRef](#)] [[PubMed](#)]
83. Wang, S.; Xiang, J.; Li, Z.; Lu, S.; Hu, J.; Gao, X.; Yu, L.; Wang, L.; Wang, J.; Wu, Y.; et al. A plasma microRNA panel for early detection of colorectal cancer. *Int. J. Cancer* **2015**, *136*, 152–161. [[CrossRef](#)] [[PubMed](#)]
84. Ng, E.K.; Chong, W.W.; Jin, H.; Lam, E.K.; Shin, V.Y.; Yu, J.; Poon, T.C.; Ng, S.S.; Sung, J.J. Differential expression of microRNAs in plasma of patients with colorectal cancer: A potential marker for colorectal cancer screening. *Gut* **2009**, *58*, 1375–1381. [[CrossRef](#)] [[PubMed](#)]
85. Wang, J.; Huang, S.K.; Zhao, M.; Yang, M.; Zhong, J.L.; Gu, Y.Y.; Peng, H.; Che, Y.Q.; Huang, C.Z. Identification of a circulating microRNA signature for colorectal cancer detection. *PLoS ONE* **2014**, *9*, e87451. [[CrossRef](#)] [[PubMed](#)]
86. Zanutto, S.; Pizzamiglio, S.; Ghilotti, M.; Bertan, C.; Ravagnani, F.; Perrone, F.; Leo, E.; Pilotti, S.; Verderio, P.; Gariboldi, M.; et al. Circulating miR-378 in plasma: A reliable, haemolysis-independent biomarker for colorectal cancer. *Br. J. Cancer* **2014**, *110*, 1001–1007. [[CrossRef](#)] [[PubMed](#)]
87. Wang, Q.; Huang, Z.; Ni, S.; Xiao, X.; Xu, Q.; Wang, L.; Huang, D.; Tan, C.; Sheng, W.; Du, X. Plasma miR-601 and miR-760 are novel biomarkers for the early detection of colorectal cancer. *PLoS ONE* **2012**, *7*, e44398. [[CrossRef](#)] [[PubMed](#)]
88. Zheng, G.; Du, L.; Yang, X.; Zhang, X.; Wang, L.; Yang, Y.; Li, J.; Wang, C. Serum microRNA panel as biomarkers for early diagnosis of colorectal adenocarcinoma. *Br. J. Cancer* **2014**, *111*, 1985–1992. [[CrossRef](#)] [[PubMed](#)]
89. Chang, P.Y.; Chen, C.C.; Chang, Y.S.; Tsai, W.S.; You, J.F.; Lin, G.P.; Chen, T.W.; Chen, J.S.; Chan, E.C. MicroRNA-223 and microRNA-92a in stool and plasma samples act as complementary biomarkers to increase colorectal cancer detection. *Oncotarget* **2016**, *7*, 10663–10675. [[CrossRef](#)] [[PubMed](#)]
90. Yau, T.O.; Wu, C.W.; Tang, C.M.; Chen, Y.; Fang, J.; Dong, Y.; Liang, Q.; Ng, S.S.; Chan, F.K.; Sung, J.J.; et al. MicroRNA-20a in human faeces as a non-invasive biomarker for colorectal cancer. *Oncotarget* **2016**, *7*, 1559–1568. [[CrossRef](#)] [[PubMed](#)]
91. Kalimutho, M.; Del Vecchio Blanco, G.; Di Cecilia, S.; Sileri, P.; Cretella, M.; Pallone, F.; Federici, G.; Bernardini, S. Differential expression of miR-144\* as a novel fecal-based diagnostic marker for colorectal cancer. *J. Gastroenterol.* **2011**, *46*, 1391–1402. [[CrossRef](#)] [[PubMed](#)]
92. Hunter, M.P.; Ismail, N.; Zhang, X.; Aguda, B.D.; Lee, E.J.; Yu, L.; Xiao, T.; Schafer, J.; Lee, M.L.; Schmittgen, T.D.; et al. Detection of microRNA expression in human peripheral blood microvesicles. *PLoS ONE* **2008**, *3*, e3694. [[CrossRef](#)] [[PubMed](#)]
93. Chen, X.; Ba, Y.; Ma, L.; Cai, X.; Yin, Y.; Wang, K.; Guo, J.; Zhang, Y.; Chen, J.; Guo, X.; et al. Characterization of microRNAs in serum: A novel class of biomarkers for diagnosis of cancer and other diseases. *Cell. Res.* **2008**, *18*, 997–1006. [[CrossRef](#)] [[PubMed](#)]
94. Colombo, M.; Raposo, G.; Thery, C. Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. *Annu. Rev. Cell. Dev. Biol.* **2014**, *30*, 255–289. [[CrossRef](#)] [[PubMed](#)]
95. Mirzaei, H.; Sahebkar, A.; Jaafari, M.R.; Goodarzi, M.; Mirzaei, H.R. Diagnostic and therapeutic potential of exosomes in cancer: The beginning of a new tale? *J. Cell. Physiol.* **2016**, *232*, 3251–3260. [[CrossRef](#)] [[PubMed](#)]

96. Silva, J.; Garcia, V.; Rodriguez, M.; Compte, M.; Cisneros, E.; Veguillas, P.; Garcia, J.M.; Dominguez, G.; Campos-Martin, Y.; Cuevas, J.; et al. Analysis of exosome release and its prognostic value in human colorectal cancer. *Genes Chromosomes Cancer*. **2012**, *51*, 409–418. [[CrossRef](#)] [[PubMed](#)]
97. Hollis, M.; Nair, K.; Vyas, A.; Chaturvedi, L.S.; Gambhir, S.; Vyas, D. MicroRNAs potential utility in colon cancer: Early detection, prognosis, and chemosensitivity. *World J. Gastroenterol.* **2015**, *21*, 8284–8292. [[CrossRef](#)] [[PubMed](#)]
98. Merhautova, J.; Demlova, R.; Slaby, O. MicroRNA-based therapy in animal models of selected gastrointestinal cancers. *Front. Pharmacol.* **2016**, *7*, 329. [[CrossRef](#)] [[PubMed](#)]
99. Zhu, L.; Fang, J. The structure and clinical roles of microRNA in colorectal cancer. *Gastroenterol. Res. Pract.* **2016**, *2016*, 1360348. [[CrossRef](#)] [[PubMed](#)]
100. Xie, T.; Huang, M.; Wang, Y.; Wang, L.; Chen, C.; Chu, X. MicroRNAs as regulators, biomarkers and therapeutic targets in the drug resistance of colorectal cancer. *Cell. Physiol. Biochem.* **2016**, *40*, 62–76. [[CrossRef](#)] [[PubMed](#)]
101. Hansen, T.F.; Andersen, C.L.; Nielsen, B.S.; Spindler, K.L.; Sorensen, F.B.; Lindebjerg, J.; Brandslund, I.; Jakobsen, A. Elevated microRNA-126 is associated with high vascular endothelial growth factor receptor 2 expression levels and high microvessel density in colorectal cancer. *Oncol. Lett.* **2011**, *2*, 1101–1106. [[CrossRef](#)] [[PubMed](#)]
102. Suto, T.; Yokobori, T.; Yajima, R.; Morita, H.; Fujii, T.; Yamaguchi, S.; Altan, B.; Tsutsumi, S.; Asao, T.; Kuwano, H. MicroRNA-7 expression in colorectal cancer is associated with poor prognosis and regulates cetuximab sensitivity via EGFR regulation. *Carcinogenesis* **2015**, *36*, 338–345. [[CrossRef](#)] [[PubMed](#)]
103. Igarashi, H.; Kurihara, H.; Mitsunashi, K.; Ito, M.; Okuda, H.; Kanno, S.; Naito, T.; Yoshii, S.; Takahashi, H.; Kusumi, T.; et al. Association of microRNA-31–5p with clinical efficacy of anti-EGFR therapy in patients with metastatic colorectal cancer. *Ann. Surg. Oncol.* **2015**, *22*, 2640–2648. [[CrossRef](#)] [[PubMed](#)]
104. Cappuzzo, F.; Sacconi, A.; Landi, L.; Ludovini, V.; Biagioni, F.; D’Incecco, A.; Capodanno, A.; Salvini, J.; Corgna, E.; Cupini, S.; et al. MicroRNA signature in metastatic colorectal cancer patients treated with anti-EGFR monoclonal antibodies. *Clin. Colorectal Cancer* **2014**, *13*, 37–45. [[CrossRef](#)] [[PubMed](#)]
105. Fang, L.; Li, H.; Wang, L.; Hu, J.; Jin, T.; Wang, J.; Yang, B.B. MicroRNA-17–5p promotes chemotherapeutic drug resistance and tumour metastasis of colorectal cancer by repressing PTEN expression. *Oncotarget* **2014**, *5*, 2974–2987. [[CrossRef](#)] [[PubMed](#)]
106. Oue, N.; Anami, K.; Schetter, A.J.; Moehler, M.; Okayama, H.; Khan, M.A.; Bowman, E.D.; Mueller, A.; Schad, A.; Shimomura, M.; et al. High miR-21 expression from FFPE tissues is associated with poor survival and response to adjuvant chemotherapy in colon cancer. *Int. J. Cancer* **2014**, *134*, 1926–1934. [[CrossRef](#)] [[PubMed](#)]
107. Qian, X.; Yu, J.; Yin, Y.; He, J.; Wang, L.; Li, Q.; Zhang, L.Q.; Li, C.Y.; Shi, Z.M.; Xu, Q.; et al. MicroRNA-143 inhibits tumor growth and angiogenesis and sensitizes chemosensitivity to oxaliplatin in colorectal cancers. *Cell. Cycle* **2013**, *12*, 1385–1394. [[CrossRef](#)] [[PubMed](#)]
108. Takahashi, M.; Cuatrecasas, M.; Balaguer, F.; Hur, K.; Toiyama, Y.; Castells, A.; Boland, C.R.; Goel, A. The clinical significance of miR-148a as a predictive biomarker in patients with advanced colorectal cancer. *PLoS ONE* **2012**, *7*, e46684. [[CrossRef](#)] [[PubMed](#)]
109. Song, Y.; Xu, Y.; Wang, Z.; Chen, Y.; Yue, Z.; Gao, P.; Xing, C.; Xu, H. MicroRNA-148b suppresses cell growth by targeting cholecystokinin-2 receptor in colorectal cancer. *Int. J. Cancer* **2012**, *131*, 1042–1051. [[CrossRef](#)] [[PubMed](#)]
110. Bhangu, A.; Wood, G.; Brown, G.; Darzi, A.; Tekkis, P.; Goldin, R. The role of epithelial mesenchymal transition and resistance to neoadjuvant therapy in locally advanced rectal cancer. *Colorectal Dis.* **2014**, *16*, O133–O143. [[CrossRef](#)] [[PubMed](#)]
111. Wan, L.Y.; Deng, J.; Xiang, X.J.; Zhang, L.; Yu, F.; Chen, J.; Sun, Z.; Feng, M.; Xiong, J.P. MiR-320 enhances the sensitivity of human colon cancer cells to chemoradiotherapy in vitro by targeting foxm1. *Biochem. Biophys. Res. Commun.* **2015**, *457*, 125–132. [[CrossRef](#)] [[PubMed](#)]
112. Rasmussen, M.H.; Jensen, N.F.; Tarpgaard, L.S.; Qvortrup, C.; Romer, M.U.; Stenvang, J.; Hansen, T.P.; Christensen, L.L.; Lindebjerg, J.; Hansen, F.; et al. High expression of microRNA-625–3p is associated with poor response to first-line oxaliplatin based treatment of metastatic colorectal cancer. *Mol. Oncol.* **2013**, *7*, 637–646. [[CrossRef](#)] [[PubMed](#)]

113. Boisen, M.K.; Dehlendorff, C.; Linnemann, D.; Nielsen, B.S.; Larsen, J.S.; Osterlind, K.; Nielsen, S.E.; Tarpgaard, L.S.; Qvortrup, C.; Pfeiffer, P.; et al. Tissue microRNAs as predictors of outcome in patients with metastatic colorectal cancer treated with first line capecitabine and oxaliplatin with or without bevacizumab. *PLoS ONE* **2014**, *9*, e109430. [[CrossRef](#)] [[PubMed](#)]
114. Hansen, T.F.; Carlsen, A.L.; Heegaard, N.H.; Sorensen, F.B.; Jakobsen, A. Changes in circulating microRNA-126 during treatment with chemotherapy and bevacizumab predicts treatment response in patients with metastatic colorectal cancer. *Br. J. Cancer* **2015**, *112*, 624–629. [[CrossRef](#)] [[PubMed](#)]
115. Hu, J.; Cai, G.; Xu, Y.; Cai, S. The plasma microRNA miR-1914\* and -1915 suppresses chemoresistant in colorectal cancer patients by down-regulating NFIX. *Curr. Mol. Med.* **2016**, *16*, 70–82. [[CrossRef](#)] [[PubMed](#)]
116. Svoronos, A.A.; Engelman, D.M.; Slack, F.J. OncomiR or tumor suppressor? The duplicity of microRNAs in cancer. *Cancer Res.* **2016**, *76*, 3666–3670. [[CrossRef](#)] [[PubMed](#)]
117. Bao, Y.; Guo, Y.; Li, Z.; Fang, W.; Yang, Y.; Li, X.; Li, Z.; Xiong, B.; Chen, Z.; Wang, J.; et al. MicroRNA profiling in MUC2 knockout mice of colitis-associated cancer model reveals epigenetic alterations during chronic colitis malignant transformation. *PLoS ONE* **2014**, *9*, e99132. [[CrossRef](#)] [[PubMed](#)]
118. Zhu, L.; Wang, Z.; Fan, Q.; Wang, R.; Sun, Y. MicroRNA-27a functions as a tumor suppressor in esophageal squamous cell carcinoma by targeting Kras. *Oncol. Rep.* **2014**, *31*, 280–286. [[CrossRef](#)] [[PubMed](#)]
119. Scheibner, K.A.; Teaboldt, B.; Hauer, M.C.; Chen, X.; Cherukuri, S.; Guo, Y.; Kelley, S.M.; Liu, Z.; Baer, M.R.; Heimfeld, S.; et al. MiR-27a functions as a tumor suppressor in acute leukemia by regulating 14–3-3theta. *PLoS ONE* **2012**, *7*, e50895. [[CrossRef](#)] [[PubMed](#)]
120. Chintharlapalli, S.; Papineni, S.; Abdelrahim, M.; Abudayyeh, A.; Jutooru, I.; Chadalapaka, G.; Wu, F.; Mertens-Talcott, S.; Vanderlaag, K.; Cho, S.D.; et al. Oncogenic microRNA-27a is a target for anticancer agent methyl 2-cyano-3,11-dioxo-18beta-olean-1,12-dien-30-oate in colon cancer cells. *Int. J. Cancer* **2009**, *125*, 1965–1974. [[CrossRef](#)] [[PubMed](#)]
121. Xiang, X.; Zhuang, X.; Ju, S.; Zhang, S.; Jiang, H.; Mu, J.; Zhang, L.; Miller, D.; Grizzle, W.; Zhang, H.G. MiR-155 promotes macroscopic tumor formation yet inhibits tumor dissemination from mammary fat pads to the lung by preventing EMT. *Oncogene* **2011**, *30*, 3440–3453. [[CrossRef](#)] [[PubMed](#)]
122. Shi, J. Considering exosomal miR-21 as a biomarker for cancer. *J. Clin. Med.* **2016**, *5*, 42. [[CrossRef](#)] [[PubMed](#)]
123. Shen, J.; Liu, Z.; Todd, N.W.; Zhang, H.; Liao, J.; Yu, L.; Guarnera, M.A.; Li, R.; Cai, L.; Zhan, M.; et al. Diagnosis of lung cancer in individuals with solitary pulmonary nodules by plasma microRNA biomarkers. *BMC Cancer* **2011**, *11*, 374. [[CrossRef](#)] [[PubMed](#)]
124. Liu, J.; Gao, J.; Du, Y.; Li, Z.; Ren, Y.; Gu, J.; Wang, X.; Gong, Y.; Wang, W.; Kong, X. Combination of plasma microRNAs with serum CA19–9 for early detection of pancreatic cancer. *Int. J. Cancer* **2012**, *131*, 683–691. [[CrossRef](#)] [[PubMed](#)]
125. Zhou, J.; Yu, L.; Gao, X.; Hu, J.; Wang, J.; Dai, Z.; Wang, J.F.; Zhang, Z.; Lu, S.; Huang, X.; et al. Plasma microRNA panel to diagnose hepatitis b virus-related hepatocellular carcinoma. *J. Clin. Oncol.* **2011**, *29*, 4781–4788. [[CrossRef](#)] [[PubMed](#)]
126. Chen, W.; Cai, F.; Zhang, B.; Barekati, Z.; Zhong, X.Y. The level of circulating miRNA-10b and miRNA-373 in detecting lymph node metastasis of breast cancer: Potential biomarkers. *Tumour. Biol.* **2013**, *34*, 455–462. [[CrossRef](#)] [[PubMed](#)]
127. Shimomura, A.; Shiino, S.; Kawauchi, J.; Takizawa, S.; Sakamoto, H.; Matsuzaki, J.; Ono, M.; Takeshita, F.; Niida, S.; Shimizu, C.; et al. Novel combination of serum microRNA for detecting breast cancer in the early stage. *Cancer. Sci.* **2016**, *107*, 326–334. [[CrossRef](#)] [[PubMed](#)]
128. Ogata-Kawata, H.; Izumiya, M.; Kurioka, D.; Honma, Y.; Yamada, Y.; Furuta, K.; Gunji, T.; Ohta, H.; Okamoto, H.; Sonoda, H.; et al. Circulating exosomal microRNAs as biomarkers of colon cancer. *PLoS ONE* **2014**, *9*, e92921. [[CrossRef](#)] [[PubMed](#)]
129. Xiang, M.; Zeng, Y.; Yang, R.; Xu, H.; Chen, Z.; Zhong, J.; Xie, H.; Xu, Y.; Zeng, X. U6 is not a suitable endogenous control for the quantification of circulating microRNAs. *Biochem. Biophys. Res. Commun.* **2014**, *454*, 210–214. [[CrossRef](#)] [[PubMed](#)]
130. Yu, H.W.; Sze, D.M.; Cho, W.C. MicroRNAs involved in anti-tumour immunity. *Int. J. Mol. Sci.* **2013**, *14*, 5587–5607. [[CrossRef](#)] [[PubMed](#)]
131. Lodish, H.F.; Zhou, B.; Liu, G.; Chen, C.Z. Micromanagement of the immune system by microRNAs. *Nat. Rev. Immunol.* **2008**, *8*, 120–130. [[CrossRef](#)] [[PubMed](#)]

132. Xiao, C.; Rajewsky, K. MicroRNA control in the immune system: Basic principles. *Cell* **2009**, *136*, 26–36. [[CrossRef](#)] [[PubMed](#)]
133. Li, X.; Nie, J.; Mei, Q.; Han, W.D. MicroRNAs: Novel immunotherapeutic targets in colorectal carcinoma. *World J. Gastroenterol.* **2016**, *22*, 5317–5331. [[CrossRef](#)] [[PubMed](#)]



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