



Interleukin 15 Pharmacokinetics and Consumption by a Dynamic Cytokine Sink

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Interleukin-15 (IL-15) is crucial for the proliferation and survival of NK and CD8⁺ T memory cells, and of significant interest in immuno-oncology. Immune cell expansion requires continuous IL-15 exposure above a threshold concentration for an extended period. However, the short $t_{1/2}$ of IL-15 makes this impossible to achieve after a single injection without a high C_{max} and toxicities. The most effective way to deliver IL-15 is continuous intra-venous infusion, but this administration mode is impractical. Efforts have been devoted to developing IL-15 agonists which after a single injection maintain the cytokine in a narrow therapeutic window for a long period. Enigmatically, although the half-life extension technologies used often extend the half-life of a protein to 1 or more weeks, the modified IL-15 agonists studied usually have systemic elimination half-lives — common to all circulating IL-15 agonists that accompanies target immune cell proliferation. What is needed is an IL-15 agonist that is as effective as continuous intravenous infusion, but with the convenience and acceptance of single injections at 1-week or longer intervals.

Keywords: cytokine, immuno-oncology, NK cells, CD8⁺ T cells, target-mediated drug disposition, interleukin-15, pharmacokinetics

INTRODUCTION

Interleukin 15 (IL-15) is a ~14 kDa four- α -helix protein belonging to a family of six interleukins that use a common cytokine-receptor γ -chain (1). The cytokine is crucial in the proliferation, maintenance, and survival of NK and CD8⁺ T memory cells, and is of major interest in immuno-oncology (2). IL-15 stimulates immune cell responses through the same dimeric IL-2/15R β , γ receptor complex as IL-2 (**Figure 1A**), but the two cytokines exhibit functionally distinct activities due to their private α -receptor subunits. As a consequence, either IL-15 or IL-2 stimulates NK and CD8⁺ T cells, but only IL-2 binds to and stimulates regulatory T cells that possess the IL-2R α in a trimeric R α , β , γ complex. IL-15 is expressed in association with its high affinity IL-15R α on the surface of IL-15-producing cells and is trans-presented to immune cells that express dimeric IL-2/15R β , γ subunits (3). Once bound to target NK and memory CD8⁺ T cells, IL-15 stimulates their proliferation, and supports their survival (4, 5). Despite dramatic augmentation of NK cells and CD8⁺ T cells, IL-15 has minimal anticancer activity as a single agent. However, in combination with other immuno-oncology agents, it shows significant efficacy and it is in this setting that IL-15 will likely find success (6).

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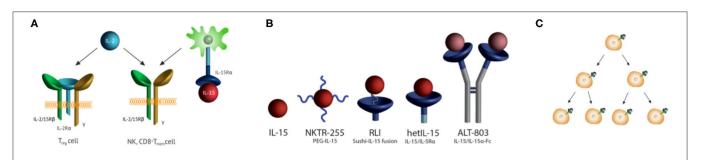


FIGURE 1 | IL-15 receptor, receptor agonists, half-lives, and immune cell proliferation. (A) IL-15 and IL-2 receptor specificities. IL-15 is trans-presented in association with IL-15R α to R β , γ subunits also used by IL-2. Trans-presentation of IL-15/IL-15R α to responding cells leads to the activation of JAK1/3 which phosphorylate STAT3/5 to prompt transcription of IL-15-modulated genes. IL-2 binds to cells with the same R β , γ subunits, as well as the high-affinity trimeric IL-2 R α , β , γ receptor on T_{Reg} cells. (B) IL-15R agonists in human clinical trials. (I) NKTR-255 is a PEGylated IL-15 (7), (ii) RLI is a fusion of the C-terminus of the IL-15R α Sushi domain to the N-terminus of IL-15 via a 20 AA linker (8), (iii) HetIL-15 (aka NIZ985) is a heterodimeric complex of IL-15 and the extracellular R α (9), and (iv) ALT-803 (aka N-803) has IL-15N72D bound to a Fc –IL-15R α Sushi domain fusion (10, 11). (C) Receptor-mediated cell proliferation and increased cytokine consumption. The cytokine binds its extracellular receptor, signals to cause cell proliferation, and is endocytosed with variable degrees of destruction. With proliferating cells consuming more of the cytokine, clearance increases as target immune cells expand.

The pharmacokinetics and pharmacodynamics of IL-15 agonists are profoundly impacted by target-mediated drug disposition (TMDD) wherein the cytokine is consumed and cleared by target immune cells (12-14)-a "cytokine sink." At high initial IL-15 concentrations, target cells are saturated with cytokine-(i.e., capacity-limited TMDD)-and clearance of the agonist is dominated by renal and/or metabolic elimination. IL-15 subsequently causes a time-dependent proliferation of target immune cells and commensurate time-dependent increase of cytokine consumption and clearance; once expanding target cells exceed the cytokine concentration and K_d of the IL-15- $R\beta$, γ the pharmacokinetics manifest as TMDD. The purposes of this review are to consolidate and scrutinize dispersed reports of the pharmacokinetics of IL-15 agonists, and to evaluate the evidence for generation of a dynamic cytokine sink that expands concomitantly with the proliferation of IL-15 target immune cells.

Target Immune Cell Proliferation and Maintenance

NK and memory T cell proliferation requires continuous IL-15 exposure at a level above a threshold concentration for an extended period (4, 5). However, with a $t_{1/2}$ of only ~2.5 h in the human it is impossible to achieve long exposure of IL-15 after a single IV injection without a very high C_{max} and resultant toxicities (15). For example, a bolus IV dose of 3 µg/kg IL-15 is necessary to maintain serum IL-15 at ≥ 1 pM – ~10-fold lower than the K_d of IL-15 for IL-2/15R β , γ – for a 24h period but gives a C_{max} of ~3,000 pM, some 30-fold higher than the 100 pM C_{max} with a 0.3 µg/kg MTD (15). A possible and popular solution to this problem is to simply extend the $t_{1/2}$ of the cytokine. This would concurrently flatten the *C* vs. *t* plot, enable prolonged exposure to optimize efficacy, and reduce the C_{max} peak responsible for toxicities.

Continuous IV (CIV) infusions of the rapeutics are often safer than bolus injections since the flat $C\,{\rm vs.}\,t$ profile is absent the high $C_{\rm max}$ peak often associated with toxicity. Indeed, CIV infusion is

the most efficient delivery method for IL-15, requiring low doses and resulting in large increases of target immune cells (16). In humans, 2 µg IL-15/kg/day of CIV infusion for 10 days led to a 5-fold increase in circulating CD8⁺ T cells, a 38-fold increase in the total NK cells, and a massive 358-fold increase in CD56^{bright} NK cells. Usually, infusing a drug at a constant rate that is balanced with its rate of elimination results in maintenance of a constant serum level. However, with CIV infusion of IL-15 serum levels decreased over the time of infusion—starting at ${\sim}360~\text{pM}$ and lowering to \sim 70 pM by the end of the infusion period indicating a time dependent increase in IL-15 clearance. This is in accord with an expanding cytokine sink where the initial lower clearance rate reflects capacity-limited TMDD (12, 14) but which after target expansion to about 10^{-9} M presents a large receptacle exceeding both IL-15 concentration and the K_d of the IL-15- $R\beta$, γ complex; under saturating sink conditions, the clearance of IL-15 would be maximal. An interesting transition of the IL-15 dose-exposure relationship occurs between infusion of 0.5- and $1 \,\mu g/kg/day$ whereby a doubling in dose results in an >45-fold increase in serum IL-15. This further supports a conversion from capacity-limited TMDD to a clearance mechanism whereby the IL-15 exceeds the capacity of the cytokine sink. Notwithstanding, CIV infusion of IL-15 for 10 days is an inconvenient, expensive mode of long-term drug administration and is unacceptable to both patients and physicians. It would be desirable to have an IL-15 agonist that could be administered as single injections at intervals of 1 or more weeks.

Pharmacokinetics of IL-15 Agonists

There have been extensive efforts to develop potent, long-acting IL-15 receptor agonists—referred to as "super-agonists" (17)— which after a single injection maintains the agonist in a narrow therapeutic window for a long period. **Figure 1B** depicts IL-15R agonists in clinical trials that have reported pharmacokinetic data, and **Table 1** presents their reported $t_{1/2}$ values in different species with different routes of administration. The approaches used to increase the $t_{1/2}$ of IL-15 generally involve increasing

| Agonist | Route | t _{1/2} , hr | | |
|----------|-------|-----------------------|------------------|---------------------------|
| | | Mouse | NHP | Human |
| IL-15 | IV | 0.64 ^b | 1.1° | 2.5 ^d |
| | SC | 0.67 ^e | 2.7 ^f | $\sim 4^{g}$ |
| | IP | 0.50 ^h | | |
| RLI | IP | 3 ^h | | |
| hetlL-15 | IV | | 1.5 ^j | |
| | SC | | 12 ^j | |
| | IP | 4 ⁱ | | |
| ALT-803 | IV | 7.5 ^k | 7.5 ¹ | 0.75- to 5 ^{m,n} |
| | SC | | | 30 ^{m,n} |
| NKTR-255 | IV | 14° | 30° | |

^aOmissions indicate data is not available.

^bHan et al. (18).

^cWaldmann et al. (19). ^dConlon et al. (15). ^eZhao et al. (20). ^fSneller et al. (21). ^gMiller et al. (22). ^hBessard et al. (23). ⁱChertova et al. (23). ⁱBergamaschi et al. (24). ^kLiu et al. (11). ^mMargolin et al. (25). ⁿRomee et al. (26).

^oKuo (7)

the molecular size to reduce renal elimination; they include incorporation of part of the IL-15R α and/or attachment to PEG or Fc to the cytokine. There are also current investigations not covered here—of IL-2R β -"biased" IL-2 constructs that bind poorly to IL-2R α in the trimeric receptor of regulatory T cells, and specifically bind to IL-2/15R β , γ mimicking the specificity of IL-15 (27, 28) which are reviewed in Klein et al. (29).

Significantly, and enigmatically, although the half-life extension technologies used often lengthen the $t_{1/2}$ of a protein to a week or longer in humans, the IL-15 agonists studied have systemic elimination $t_{1/2}$ s of not much longer than 1 day and usually only several hours (Table 1). Indeed, the only current IL-15 agonists that might be sufficient for QWk injections are the SC administered ALT-803 and the IV-administered PEGylated IL-15, NKTR-255. Interestingly, IV administration of the "long-acting" ALT-803 agonist in humans has a dose-dependent elimination $t_{1/2}$ of ~ 0.75 to 5 h (26), which is not significantly different than the 2.5 h $t_{1/2}$ of IV-administered native IL-15 (15). The dose-dependent elimination rate is likely due to a significant proportion of the drug being consumed by target cells-[i.e., TMDD-as observed with NKTR-255 (13)]. Thus, the longer apparent t_{1/2} of 30 h for SC administered ALT-803 is not due to the retardation of systemic elimination intended by its large size, but rather due to slow absorption from the injection site. Regardless, even with the 30 h $t_{1/2}$ for SC ALT-803 in humans, the level of exposure needed for proliferation and survival of target cells is sustainable for only a few days, and >90% of the drug will be eliminated in 4 days.

Although slow SC absorption provides a simple, practical approach toward achieving half-life extension, it may have unintended shortcomings. For example, SC administration of proteins may or may not have adequate bioavailability. For native IL-15, the SC $t_{1/2}$ in humans is close to that of the IV injection $(\sim 4 h \text{ for SC vs. } 2.5 h \text{ for IV})$, and the bioavailability is estimated from reported data to be near 100% (15, 22). Hence, most of the SC-administered IL-15 is rapidly and completely transferred to the systemic circulation. In contrast, in humans SC ALT-803 shows a long $t_{1/2}$ of \sim 30 h but a bioavailability of only about 3% (26). Interestingly, the 40 pM C_{max} of the agonist administered SC is some 100-fold lower than that of the same amount injected IV, yet it is significantly more effective at NK and CD8⁺ T cell expansion; clearly, the superior pharmacodynamic effects of SC vs IV administered ALT-803 are due to its longer effective $t_{1/2}$. The very low bioavailability of SC ALT-803 reveals that most $(\sim 97\%)$ of the administered drug never reaches the systemic circulation and may activate and be consumed by immune cells near the injection site. Hence, the substantial local toxicity of SCinjected ALT-803-which is not observed with IL-15-may be due to the very high level of the super-agonist deposited at the site of injection and its slow departure. Enigmatically, the problem cannot be solved by simply changing the mode of administration of ALT-803 from SC to IV or intraperitoneal (30) injections since the $t_{1/2}$ of ALT-803 via these routes are so much lower. Whether the other IL-15 super-agonists - such as hetIL-15 and RLI - will face the same conundrum in humans is not known since relevant pharmacokinetic parameters have not yet been reported.

IL-15 Consumption by a Cytokine Sink

Why do the IL-15 agonists have shorter than expected t1/2s? There is abundant evidence for an IL-15-induced "cytokine sink" that increases with proliferation of target immune cells (Figure 1C) and causes commensurate increases in the consumption/clearance of IL-15 (Table 1)—a prototypical example of "drug-induced" or "dynamic" TMDD (14). First, CIV infusion of IL-15 in monkeys and in man initially results in the expected steady state level of serum IL-15 which then decreases \sim 4- to 5-fold over time (16, 21); this unusual effect can best be explained by a time dependent increase of clearance of IL-15. Second, sequential SC administration of the same doses of the super-agonist hetIL-15 results in decreasing Cmax and Cmin upon repeated injections (24); but, when the dose is doubled in each injection by a "doubling step dose" a constant Cmax and Cmin can be maintained. This shows that increased hetIL-15 dosing is necessary to satiate the increased appetite for consumption by expanded target cells. Third, pharmacokinetic modeling of cergutuzomab amunaleukin-a CEA targeted immunocytokine containing an IL-15-like IL-2Rβ biased IL-2 mutein-revealed expansion of a drug-induced peripheral sink that increases clearance (27). Fourth, the IL-15/ILRa Fc-fusion XmAb24306 shows low potency but a long $t_{1/2}$, which is in accord with the inverse relationship between cytokine-induced expansion of a peripheral sink and cytokine $t_{1/2}$ (31). Finally, several IL-15

fusions show unexpectedly high rates of systemic elimination that are best explained by a large peripheral sink. The ~114 kDa ALT-803—expected to have a slow systemic elimination rate because of its large size—has an average elimination t_{1/2} of only ~3 h in humans when injected IV; and, connection of RLI to the carboxy terminus of anti-CD20 rituximab—which has a t_{1/2} of about 100 h in mice—results in a large fusion protein with >10-fold lower t_{1/2} than the antibody carrier itself (32). Hence, expansion of target cells by IL-15 agonists is accompanied by a dynamic increased consumption, increased clearance, and decreased lifetime of the agonists.

Summary

The efficacy of IL-15 agonists in stimulating target immune cell proliferation is paradoxically coupled to its more rapid consumption and clearance. By far, the most efficacious way to deliver IL-15 is by CIV infusion, but this mode of administration is generally impractical. The available IL-15 super-agonists are administered as single injections at intervals of several days

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to 1 week and are more practical to administer than CIV infusion; however, because of the expanding consumptive sink that accompanies target cell proliferation, they have relatively short systemic half-lives and do not achieve the level of immune cell proliferation and maintenance as CIV infusion. Ideally, what is needed is an IL-15 agonist that is as effective as CIV infusion, but with the convenience and acceptance of single injections at one-week or longer intervals. In a forthcoming report, we will describe our efforts at developing a very slow-releasing hydrogel depot of IL-15 that attempts to achieve this goal.

AUTHOR CONTRIBUTIONS

JH wrote and edited the manuscript. DS conceived, wrote, and edited the manuscript. TW provided critical comments, concepts, and insights. All authors read and agreed to the content of this work prior to submission. All authors contributed to the article and approved the submitted version.

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Conflict of Interest: JH and DS are employees and shareholders of ProLynx.

The remaining author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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