# Methylenetetrahydrofolate Reductase (MTHFR) Polymorphisms and Susceptibility for Cervical Lesions: A Meta-Analysis

## Shuyu Long<sup>19</sup>, Xingliang Yang<sup>29</sup>, Xiaojiao Liu<sup>1</sup>, Pei Yang<sup>1\*</sup>

1 Department of Gynecology and Obstetrics, West China Second Hospital, Sichuan University, Chengdu, Sichuan, China, 2 Department of Urology, Second Affiliated Hospital, Third Military Medical University, Chongqing, China

## Abstract

**Background:** The association between the methylenetetrahydrofolate reductase (MTHFR) C677T/A1298C polymorphisms and the susceptibility to cervical lesions was unclear. This study was designed to investigate their precise association using a large-scale meta-analysis.

*Methods:* The previous 16 studies were identified by searching PubMed, Embase and CBM databases. The crude odds ratios and their corresponding 95% confidence intervals (Cls) were used to estimate the association between the MTHFR C677T/ A1298C polymorphisms and the susceptibility to the cervical lesions. The subgroup analyses were made on the following: pathological history, geographic region, ethnicity, source of controls and source of DNA for genotyping.

**Results:** Neither of the polymorphisms had a significant association with the susceptibility to the cervical lesions in all genetic models. Similar results were found in the subgroup analyses. No association was found between the MTHFR C677T polymorphism and the cervical lesions in the Asia or the America populations though a significant inverse association was found in the Europe population (additive model: P = 0.006, OR = 0.83, 95% CI = 0.72-0.95; CT vs. CC: P = 0.05, OR = 0.83, 95% CI = 0.69-1.00; TT vs. CC: P = 0.05, OR = 0.73, 95% CI = 0.53-1.00). Interestingly, women with the MTHFR A1298C polymorphisms had a marginally increased susceptibility to invasive cancer (ICC) when compared with no carriers but no statistically significant difference in the dominant model (P = 0.06, OR = 1.21, 95% CI = 0.99-1.49) and AC vs. AA (P = 0.09, OR = 1.21, 95% CI = 0.97-1.51).

*Conclusions:* The MTHFR C677T and A1298C polymorphisms may not increase the susceptibility to cervical lesions. However, the meta-analysis reveals a negative association between the MTHFR C677T polymorphisms and the cervical lesions, especially in the European populations. The marginal association between the MTHFR A1298C polymorphisms and the susceptibility to cervical cancer requires a further study.

Citation: Long S, Yang X, Liu X, Yang P (2012) Methylenetetrahydrofolate Reductase (MTHFR) Polymorphisms and Susceptibility for Cervical Lesions: A Meta-Analysis. PLoS ONE 7(12): e52381. doi:10.1371/journal.pone.0052381

Editor: Surinder K. Batra, University of Nebraska Medical Center, United States of America

Received August 3, 2012; Accepted November 12, 2012; Published December 21, 2012

**Copyright:** © 2012 Long et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The authors have no support or funding to report.

Competing Interests: The authors have declared that no competing interests exist.

\* E-mail: YangP790@126.com

9 These authors contributed equally to this work.

## Introduction

Cervical cancer is the third most frequently encountered cancer and the fourth leading cause of the women's cancer death in the world, accounting for 9% (529,800) of the total newly-diagnosed cancer cases and 8% (275,100) of the total cancer deaths among females in 2008 [1]. However, cervical cancer is considered a preventable disease because of its relatively long period of precancerous lesions, including cervical intraepithelial neoplasia (CIN). The virological, molecular, clinical and epidemiological studies have provided evidence that cervical cancer is in fact a sequel to a long-term unresolved infection of certain genotypes of the Human Papilloma Virus (HPV) [2,3]. High-risk HPVs are known to infect cervical epithelium, with a subset of these being associated with preneoblastic lesions that can progress to cervical cancer. Nevertheless, despite the extremely high rate of infection by these viruses, the rate of cervical cancer, even in the prescreening area, has been less than one tenth that of exposure [4,5]. Thus, other factors are important for cervical lesion development and progression such as a long-term use of hormonal contraceptives, multiparty, smoking, and some nutritional factors [6-8].

Association between micronutrient depletion, particularly folate deficiency, and cervical lesions has been studied for a long time. Folate deficiency, as a potential risk for cervical cancer, was first reported by some cytopathologists in the 1960s, who had found that the cervical epithelial cells from folate-deficient women had some similarity to the dysplastic cervical cells in cytology [9]. Later on, Whitehead et al. demonstrated that macrocytic changes in the cervical cells of the oral contraceptive users could be reversed with folic acid supplementation [10]. However, conflicting results still

existed in the conclusion of the association between the folate deficiency and the cervical dysplasia [11–13]. Furthermore, various clinical epidemiological studies have shown that low-level folate was not directly increase risk of cervical dysplasia but enhance HPV infection instead [14–16]. Therefore, despite the lack of a statistically significant association between folate status and cervical dysplasia, these trials indicated that folate may involve along with HPV to induce cervical carcinogenesis.

The apparent role of folate in carcinogenesis in cervical tissue has stimulated investigations of polymorphisms in the folate metabolizing enzymes. As we know, Methylenetetrahydrofolate reductase (MTHFR) is a crucial enzyme that can regulate the metabolism of folate and methionine, both of which are important in DNA methylation and synthesis [17]. This occurs through the conversion of 5, 10-methyltetrahydrofolate to 5-methyltetrahydrofolate (1-carbon metabolism), which is a dominant circulating form of folate. The MTHFR gene is located on the short arm of chromosome 1 (1p36.3) and has several well-described single nucleotide polymorphisms (SNPs). Two common SNPs are known to affect enzyme function and have been shown to have clinical significance. The most common mutation is a C-to-T transition at nucleotide 677 (rs1801133, C677T) in exon 4, resulting in a substitution of alanine with valine that affects the catalytic domain of the enzyme, leading to the enzyme activity reduction [18]. Another common variant is an A-to-C transversion at position 1298 in exon 7 (rs1801131, A1298C), resulting in a substitution of glutamate with alanine at codon 429. This polymorphism also reduces the enzyme activity to a lesser extent [19].

Several studies had been designed to evaluate associations between MTHFR genotypes and cervical lesions, including cervical cancer, but the results were inconsistent because of different stages of cervical lesions and the combinatorial effects of other risk factors. Precancerous cervical lesions are classified according to the degree of cellular abnormality. The lowest grade of abnormality is CIN1, and CIN2 and CIN3 describe the progressive epithelial dysplasia leading to invasive cancer. Preinvasive lesions have also been classified in terms of squamous intraepithelial lesions (SILs) included low-grade squamous intraepithelial lesions (LSIL, including HPV infection and CIN1) and high-grade squamous intraepithelial lesions (HSIL, including CIN2 and CIN3). The majority of the case-control genetic studies revealed no association between cervical lesions and MTHFR SNPs [20-25]. But some evidences indicated that the MTHFR variants are positively associated with the cervical cancer risk [26-31]; some other evidence indicated that the MTHFR variants are inversely associated with the cervical cancer risk [32-35]. For example, one study reported that the MTHFR variant genotype may increase CIN and cervical cancer risk in women who had low-level folate status [26]. Another study suggested women with MTHFR polymorphism and low riboflavin status were significantly less likely to have HSIL than women without the polymorphism and high riboflavin status [33].

These inconclusive results may due to limited sample size, because any single study may be underpowered to detect the precise effects. In addition, there also may be the causes of different characteristics among studies, such as ethnicity, pathological history, sources of controls, and source of DNA for genotyping. Therefore, we have done a meta-analysis on association between MTHFR polymorphisms and cervical lesions using data obtained from the published case-control genetic studies. Our aim was to identify whether the MTHFR polymorphisms affect the susceptibility to SIL or cervical cancer by means of a large-scale meta-analysis. Furthermore, we wanted to summarize the effect size of the polymorphism associated with the susceptibility to the cervical lesions.

## **Materials and Methods**

## Search Strategy and Selection Criteria

The computer-based search strategy was comprehensively used to find eligible studies for this meta-analysis. Two investigators (Long, Yang) searched in the PubMed and Emase independently from inception to July 22, 2012, for the studies on the association between the MTHFR C677T polymorphism (rs1801133) and A1289C polymorphism (rs1801131) and the cervical lesions. Following Medical Subject Heading (MeSH) terms and/or text words were used in our search, such as for methylenetetrahydrofolate reductase ("MTHFR" or "methylenetetrahydrofolate reductase" or Methylenetetrahydrofolate Reductase AND (NADPH2)) with terms for genetic variations ("polymorphism" or "variation" or "mutation" or "Single Nucleotide Polymorphism" or Polymorphism, Single Nucleotide" or "SNPs" ) and terms for cervical lesions("Uterine Cervical Cancer" or "Neoplasms, Cervix" or "Neoplasms, Cervical" or "Cervix Neoplasms" or "Cervix Cancer" or "Cervical Neoplasms" or "Cancer of the Uterine Cervix" or "Cancer of the Cervix" or "Cancer of Cervix" or "Uterine Cervical Neoplasms" or "Uterine Cervical Neoplasms" or "Uterine Cervical Dysplasia" or "Neoplasia, Cervical Intraepithelia" or "Intraepithelial Neoplasia, Cervical" or "Cervical Intraepithelial Neoplasms" or "Cervical Intraepithelial Neoplasia" or "cin" ). Meanwhile, China Biological Medicine Database (CBM) was also searched for the eligible studies. Full articles published in English or Chinese were considered to be eligible for our study. In addition, reference list of the original research articles and reviews were also manually searched.

The eligible studies must meet the following inclusion criteria: (1) Exploration of associations between the MTHFR genetic polymorphisms (including C677T or A1298C or both) and the susceptibility to cervical cancer or SIL; (2) A case-control study; (3) Provision of information on genotype frequencies of the MTHFR C677T and/or A1298C polymorphism(s) or sufficient data for the calculation. The exclusion criteria were as follows: (1) A review, case report, editorial, or comment; (2) A duplicated study; (3) Laboratory molecular or animal studies. If studies contained overlapping cases and/or controls, the largest study with extractable data was preferred.

Because the data included in this study was taken from literatures, written consent given by the patients and ethical approval acquired by certain committee were not needed in our meta-analysis.

#### Data Extraction

According to the inclusion and exclusion criteria, extraction from each study was conducted independently by two authors (Long, Yang) and the consensus was achieved for all the data, which were as follows: the first author's name, year of publication, source of controls, source of DNA for genotyping, country, ethnicity, goodness-in-fitness of Hardy-Weinberg Equilibrium (HWE) in the control group, histological stage of cervical lesions, numbers of cases/patients and controls, and distribution of genotypes in the case and control groups. The patients were recruited into the study regardless of whether they had a firstdegree relative with cervical lesions. The controls were recruited regardless of whether they had other diseases, e.g., hysteromyoma. For studies with inadequate information, authors of those studies were contacted for further information by E-mail if possible.

#### Statistical Analysis

Meta-analysis was performed and reported as described previously [36,37]. Crude ORs with 95% CIs were computed to assess the strength of the correlation between the MTHFR C677T/A1298C polymorphisms and the susceptibility to cervical lesions. The pooled ORs were performed for the dominant model (aa+Aa vs. AA), recessive model (aa vs. Aa+AA) and additive model (A vs. a). Moreover, the pooled estimates were also calculated for the pair-wise comparisons (allele Aa vs. AA, and allele aa vs. AA). The above-mentioned A and a represented the major and the minor allele respectively. Taking consideration of possible between-study heterogeneity, a statistical test for heterogeneity was performed by the  $\chi^2$  test or Fisher exact test when appropriate. P < 0.10 or  $I^2 > 50\%$  indicated an obvious of the between-study heterogeneity, and OR (95% CI) was calculated by the random-effects model using the DerSimonian and Laird method; otherwise, the fixed-effects model was used by the Mantel-Haenszel method [38,39]. Subgroup analyses were mainly conducted using the corresponding pathological history (ICC, SIL), geographic region (Asia, Europe, United States), ethnicity (Asian, Caucasian, mixed), source of controls (healthy persons, patients), and source of DNA for genotyping (blood, cervical cells or tissue sample), all of which were used to explore and explain the heterogeneity between the different studies.

The allele frequencies, at which the MTHFR C677T/A1298C polymorphisms occurred in each respective study, were determined by the allele-counting method. A chi-square test was used to determine whether the observed frequencies of the genotypes in the controls conformed to Hardy Weinberg-Equilibrium (HWE) expectations if genotype data were available. Sensitivity analyses were performed on stability of the results, namely, one case-control study omitted each time to reflect the influence of the individual data set on the pooled OR. Several methods were used to detect any probable publication bias. Asymmetry of the funnel plot indicated the possible publication bias. In addition, the Egger and Begg quantitative tests were also used, and P < 0.05 was considered a statistical significance [40,41].

All analyses were performed using the RevMan 5.0 program (Cochrane Library, UK) and the STATA package version 11.0 program (Stata Corporation, College Station, Texas, USA). All *P* values were two-sided. To ensure the reliability of data, two reviewers (Long, Yang) independently performed the data analysis using the statistics programs for the same results.

### Results

## Characteristics of Eligible Studies

Detailed information for selecting eligible studies was showed in Figure 1. After comprehensively searching, 67 potentially-relevant publications were identified, and none of them were selected from the reference lists of the identified articles. After the careful selection, 16 eligible studies were finally included in our metaanalysis. Among them, 16 studies investigated the MTHFR C677T polymorphism with 3498 cases and 3594 controls and 5 studies investigated the MTHFR A1298C polymorphism with 1087 cases and 1202 controls. General characteristics of the included studies were evaluated for the association between variants and cervical lesions (Table 1, Table 2). For C677T, 11 studies recruited the controls from healthy persons; 1 study from hospital patients and 4 studies from both. 9 studies were performed in Asia; 4 studies performed in Europe; 3 studies performed in America. 5 studies talked about ICC; 3 studies talked about SIL and 8 studies talked about both. For A1298C, all 5 studies performed in Asian; 4 studies recruited controls from healthy

persons and 1 study from both healthy persons and hospital patients. 1 study talked about ICC and 4 studies talked about both ICC and SIL. 14 of the studies presented NS (not significant) were conformed to Hardy Weinberg-Equilibrium (HWE) expectations (P>0.05). However, two of the studies [27,35] presented NA (not available) were because we could not perform the HWE test for the subjects (either cases or controls) in those studies, for only the total number of the combined genotypes (CT/TT vs. CC or AC/CC vs. AA) were available. Therefore, this study was included in the analysis on the dominant model, not on other genetic models. Furthermore, the allele and genotype frequencies, at which the MTHFR C677T and the A1298C polymorphisms occurred in case and controls in each of the studies, were also summarized (Table 1, Table 2).

#### Quantitative Synthesis

Association between the MTHFR C677T polymorphisms and cervical lesions. As for the C677T polymorphism, no association was found between the polymorphism and the susceptibility to cervical lesions in all the genetic models (Table 3, dominant model: OR = 0.99, 95% CI = 0.78-1.26, Figure 2A; recessive model: OR = 1.05, 95% CI = 0.80-1.38; additive model: OR = 0.97, 95% CI = 0.80-1.18,; CT vs. CC: OR = 0.97, 95% CI = 0.78–1.20, Figure 2B; TT vs. CC: OR = 1.06, 95% CI = 0.76 - 1.48, Figure 2C). The heterogeneity was significant in all the genetic models ( $P \le 0.05$ ) and the random-effects model was used in the meta-analysis. The subgroup analysis of the C677T polymorphisms in the histological stages of the cervical lesions also revealed that the polymorphism was not associated with the risk of ICC or SIL in all the genetic models (Table 3). Although the subgroup analysis of C677T in the geographic regions revealed that no association was found between the C677T polymorphism and the cervical lesions in either the Asia or the America populations, the Europe population showed a significant inverse association in some genetic models (additive model: P = 0.006, OR = 0.83, 95% CI = 0.72–0.95; CT vs. CC: P = 0.05, OR = 0.83,95% CI = 0.69–1.00; TT vs. CC: P = 0.05, OR = 0.73, 95% CI = 0.53 - 1.00). The heterogeneity was significantly reduced in the Europe populations in the recessive, additive, C/T vs. C/C, and T/T vs. C/C models.

In the sensitivity analysis, the overall association between the MTHFR C677T genotype and the cervical lesions was unchanged after an exclusion of the individual study, including two studies [27,35], which lacked enough data to calculate if it conformed to HWE among the control group. Similar results were found in the sensitivity analyses on the association between the MTHFR C677T genotype and ICC or SIL, indicating that our results were statistically robust. No obvious publication bias was detected according to the shapes of the funnel plots for the C677T polymorphism in all the genetic models (Figure 3). Consistent results of the Egger's and the Begg's tests were also obtained in all the genetic models (Table 3). Moreover, neither the funnel plots nor the Begg's or Egger's test detected any obvious evidence for the publication bias in the subgroup analyses on all the genetic models (data not shown).

Association between the MTHFR A1298C polymorphisms and cervical lesions. As for the A1298C polymorphism, no association was found between the polymorphism and the cervical lesions in all the genetic models (Table 4, dominant model: OR = 1.21, 95% CI = 0.87-1.690, Figure 4A; recessive model: OR = 0.81, 95% CI = 0.54-1.23; additive model: OR = 0.98, 95% CI = 0.85-1.14; AC vs. AA: OR = 1.02, 95% CI = 0.85-1.24, Figure 4B; CC vs. AA: OR = 0.80, 95% CI = 0.52-1.24, Figure 4C). The heterogeneity was significant in the dominant



Figure 1. Flow diagram of the study selection process. doi:10.1371/journal.pone.0052381.g001

And    Construction    C    T    C    C    T    C    C    T    C	First author [reference] Y	fear	Source of control	Source of DNA	Country	Ethnicity	HWE	listology	Samp size	<u>a</u>	Case					C	Contro	-				
The state of									case	control	υ	⊢	ប	Ե	E		U		0	F	5	E ±
Mostowska [1] 2011 Healthy persons Blood korea and N S SL 14 18 19 7 5 6 5 2 10 5 2 10 5 10 5 10 10 10 10 10 10 10 10 10 10 10 10 10	Prasad [20] 2	2011	Mixed	Blood	India	Asian	NS IC	0	62	125	119	5	57	5	2		240	1	16 8	-	6	
Transitional problem of the second of	Mostowska [21] 2	2011	Healthy persons	Blood	Poland	Caucasian	NS IC	Ŋ	124	168	194	77	56	59	9	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	219	117 6	.8 6	18	66	
HSI    HSI <td>Tong [26] 2</td> <td>2011</td> <td>Healthy persons</td> <td>Blood</td> <td>Korea</td> <td>Asian</td> <td>NS</td> <td>SIL</td> <td>159</td> <td>427</td> <td>186</td> <td>132</td> <td>52</td> <td>82</td> <td>25 1</td> <td>07</td> <td>502</td> <td>352 1</td> <td>52 19</td> <td>98 77</td> <td>27</td> <td>'n</td>	Tong [26] 2	2011	Healthy persons	Blood	Korea	Asian	NS	SIL	159	427	186	132	52	82	25 1	07	502	352 1	52 19	98 77	27	'n
ICC    ICC <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>Τ</td> <td>SIL</td> <td>160</td> <td>427</td> <td>182</td> <td>138</td> <td>54</td> <td>74</td> <td>32 1</td> <td>90</td> <td>502</td> <td>352 1</td> <td>52 19</td> <td>98 77</td> <td>27</td> <td>Ŀ0</td>							Τ	SIL	160	427	182	138	54	74	32 1	90	502	352 1	52 19	98 77	27	Ŀ0
Kohar (22)    200    Haitypersons    Tisue or cell    India    Afan    NS    HSL    23    G7    11    28    11    29    31      Shekar [22]    2008    Haitypersons    Blood    India    Afan    NS    ICC    164    231    27    13    47    4    51      Shekar [23]    2008    Haitypersons    Blood    India    Afan    NS    ICC    200    206    71    26    71    26    20 <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>⊻</td> <td>Ŋ</td> <td>146</td> <td>427</td> <td>171</td> <td>121</td> <td>53</td> <td>65</td> <td>28</td> <td>с. С</td> <td>502</td> <td>352 1</td> <td>52 19</td> <td>98 77</td> <td>27</td> <td>'n</td>							⊻	Ŋ	146	427	171	121	53	65	28	с. С	502	352 1	52 19	98 77	27	'n
Heat    IC    16    21    23    5    13    47    4    51    33      Shefat    200    Heathy persons    Blood    India    Aian    NS    ICC    200    36    32    17    26    36    31      Nucher [23]    2008    Heathy persons    Blood    India    Aian    NS    ICC    200    36    32    17    36    NA    46    30    30      Pythikk    300    Mixed    Blood    USA    Mixed    Blood    USA    MS    HSL    26    30    30    30    30    30      Pythikk    300    Mixed    Blood    USA    Mixed    Blood    No    No    No    30	Kohaar [22] 2	2010	Healthy persons	Tissue or cell	India	Asian	NS H	SIL	39	231	67	1	28	11	-	-	387	75 1	61 6	5	70	
Shekar [32]    2008    Healthypersons    Blood    India    Asian    NS    ICC    200    36    37    70    28    3    30    30      Nuchar [37]    2008    Healthypersons    Blood    India    Asian    NA    Si    70    Asia    NA    70    Asia    Asia    Asia    NA    Si    71    NA    70    Asia    Asia    NA      Phythike [33]    2007    Nixed    Blood    USA    Mixed    Si    71    NA    70 <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>⊻</td> <td>Ŋ</td> <td>164</td> <td>231</td> <td>273</td> <td>55</td> <td>113</td> <td>47</td> <td>5</td> <td>-</td> <td>387</td> <td>75 1</td> <td>61 6</td> <td>5</td> <td>70</td> <td></td>							⊻	Ŋ	164	231	273	55	113	47	5	-	387	75 1	61 6	5	70	
Number (27)    2008    Heithypersons    Biood    India    Sile    N	Shekari [32] 2	2008	Healthy persons	Blood	India	Asian	NS IC	Ŋ	200	200	368	32	170	28	3	0	318 8	32 1	25 68	8 7	75	
Index    Index <th< td=""><td>Nandan [27] 2</td><td>2008</td><td>Healthy persons</td><td>Blood</td><td>India</td><td>Asian</td><td>NA S</td><td>_</td><td>80</td><td>77</td><td>NA</td><td>NA</td><td>34</td><td>NA</td><td>NA 4</td><td>9</td><td>AN I</td><td>NA 5</td><td>с N</td><td>A N/</td><td>1 24</td><td></td></th<>	Nandan [27] 2	2008	Healthy persons	Blood	India	Asian	NA S	_	80	77	NA	NA	34	NA	NA 4	9	AN I	NA 5	с N	A N/	1 24	
Pythtlake [3]    207    Mixed    Bood    Usd    Need    NS    HSL    SS    35    15    50    16    5    17    203    313    201    Mixed    32    32    36    37    30    313    30      Zoodsma [34]    2005    Mixed    Bood    Netherlands    Caucasian    NS    HSL    26    32    16    17    20    32    30<							⊻	Ŋ	62	77	NA	NA	36	NA	VA 2	9	A N	VA 5	E N	A N/	1 24	
Zoodsma [34]    Zoodsma [34]    Mixed    Blood    Netherlands    Caucasian    NS    HSL    Zood    Mise	Piyathilake [33]	2007	Mixed	Blood	USA	Mixed	NS	SIL	80	355	134	26	59	16	2	-	562	48 2	23 1	16 16	13	2
ICC    635    592    944    337    337    337    337    33    337    33    337	Zoodsma [34] 2	2005	Mixed	Blood	Netherlands	Caucasian	NS	SIL	264	592	362	166	121	120	23 1	43	308	376 2	73 20	52 57	31	6
Kang [23]    2005    Heathypersons    Blood    Korean    Asian    NS    ICC    79    74    86    72    27    20    22    20    22    20    20    20      Sull [28]    Jodd    Heathypersons    Blood    Korean    Asian    NS    LSIL    45    45    38    10    23    8    20    52    38    30    52    30							⊻	Ŋ	636	592	944	328	357	230 4	t9 2	79	308	376 2	73 20	52 57	31	6
Sull (28)    2004    Healthy persons    Biod    Koran    Asian    NS    LSIL    46    45    38    10    22    8    30    537      Ambropoulos (24)    2003    Healthy persons    Tissue or cell    Greece    Gucasian    NS    LSIL    74    190    162    70    76    73    713    73    713    73    713    73    713    73    713    73    713    73    713    73    713    73	Kang [23] 2	2005	Healthy persons	Blood	Korean	Asian	NS IC	Ŋ	79	74	86	72	27	32	20 5	5	92	56 3	0 3.	2 12	44	
HSIL  75  45  19  16  50  36  126  57    Indroboulos [24]  203  Healthy persons  Tissue or cell  Greece  Caucasian  NS  LSI  26  26  37  115  58  173  52    Indroboulos [24]  203  Healthy persons  Tissue or cell  Greece  Caucasian  NS  LSI  64  91  68  38  73 <	Sull [28] 2	2004	Healthy persons	Blood	Korean	Asian	NS	SIL	40	454	42	38	10	22 8	m m	0	527	381 1	53 2.	21 80	30	F
Indepondenci (24)    Zord (28)    Heading (24)    Zord (28)    Heading (28)    Tissue or cell (28)    Greece (28)    NS    EV    Zord (28)    Zord (28) <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>Т</td> <td>SIL</td> <td>176</td> <td>454</td> <td>190</td> <td>162</td> <td>50</td> <td>6</td> <td>36 1</td> <td>26</td> <td>527</td> <td>381 1</td> <td>53 2:</td> <td>21 80</td> <td>30</td> <td>-</td>							Т	SIL	176	454	190	162	50	6	36 1	26	527	381 1	53 2:	21 80	30	-
Lambropoulos [24]    2003    Healthy persons    Tissue or cell    Greece    Caucasian    NS    LSL    53    91    68    38    20    28    5    33    121      Ambropoulos [24]    No    Healthy persons    Tissue or cell    Greece    Gaucasian    NS    LSL    64    91    83    27    29    8    73    121      Goodman [29]    2001    Hospital patients    Blood    USA    Mixed    NS    SIL    150    179    213    87    73    67    10    121      Pyathilake [30]    2000    Healthy persons    Tissue or cell    USA    Mixed    NS    LL    25    11    87    26    19    73    64    44      Pyathilake [30]    2000    Healthy persons    Tissue or cell    USA    Mixed    NS    LL    25    26    13    73    26    14    44    44    44    44    44    14    14							⊻	2	246	454	261	231	73	115	58 1	73	527	381 1	53 23	21 80	30	E
HSIL  64  91  83  45  27  29  8  37  121    Goodman [29]  2001  Hospital patients  Blood  USA  Mixed  NS  SIL  150  19  20  10  27  29  8  37  121    Piyathilake [30]  2001  Hospital patients  Blood  USA  Mixed  NS  SIL  150  179  213  87  73  67  10  73  261    Piyathilake [30]  2000  Healthy persons  Tissue or cell  USA  Mixed  NS  LSL  25  31  27  26  10  73  26    Piyathilake [30]  2010  Healthy persons  Tissue or cell  USA  Mixed  NS  LSL  25  31  27  26  28  24  28  24  28  24  28	Lambropoulos [24] 2	2003	Healthy persons	Tissue or cell	Greece	Caucasian	NS	SIL	53	91	68	38	20	28	ŝ	e e	121 (	51 4	2 3	7 12	49	_
Image: Section 129  2001  Hospital patients  Blood  USA  Mixed  NS  SlL  15  17  8  2  10  121    Pyathilake [30]  2001  Hospital patients  Blood  USA  Mixed  NS  SlL  150  179  213  87  73  67  10  77  261    Pyathilake [30]  2000  Healthy persons  Tissue or cell  USA  Mixed  NS  LSIL  25  31  12  23  67  10  77  261    Pyathilake [30]  2010  Healthy persons  Tissue or cell  USA  Mixed  NS  LSIL  25  31  12  23  44    Agodi [35]  2010  Healthy persons  Cell  tay  N  NI  N  NI							Т	SIL	64	91	83	45	27	29 8	e e	2	121 6	51 4	2	7 12	49	
Goodman [29]    2001    Hospital patients    Blood    USA    Mixed    NS    SIL    150    173    87    73    67    10    77    261      Pyathilake [30]    2000    Healthy persons    Tissue or cell    USA    Mixed    NS    LIL    25    31    25    6    13    67    19    74    261      Pyathilake [30]    2000    Healthy persons    Tissue or cell    USA    Mixed    NS    LIL    25    31    15    23    44    44      Adodi [35]    2010    Healthy persons    Cell    tap    SIL    123    66    NA    NA    5    NA    5    74      Adodi [35]    2011    Mixed    Blood    China    Asian    NS    SIL    33    32    66    NA    NA    5    73    5    5    50    73    5    50    50    50    50    50    50    50    50							⊻	Ŋ	21	91	30	12	11	∞	1	0	121 (	51 4	.2	7 12	49	
Plyathilake [30]    2000    Healthy persons    Tissue or cell    USA    Mixed    NS    LSIL    25    31    25    6    13    6    19    44      Apodi [35]    2010    Healthy persons    Cell    Italy    Gaucasian    NA    SIL    123    66    NA    11    23    5    28    44      Agodi [35]    2010    Healthy persons    Cell    Italy    Gaucasian    NA    SIL    123    66    NA    NA    78    70    78    70      Vang [25]    2011    Mixed    Blood    China    Asian    NS    SIL    38    382    60    16    17    15    50    70    50 <td>Goodman [29]</td> <td>2001</td> <td>Hospital patients</td> <td>Blood</td> <td>USA</td> <td>Mixed</td> <td>NS S</td> <td>_</td> <td>150</td> <td>179</td> <td>213</td> <td>87</td> <td>73</td> <td>. 29</td> <td>0</td> <td>-</td> <td>261 9</td> <td>9 76</td> <td>3 7</td> <td>11</td> <td>86</td> <td></td>	Goodman [29]	2001	Hospital patients	Blood	USA	Mixed	NS S	_	150	179	213	87	73	. 29	0	-	261 9	9 76	3 7	11	86	
Agodi [35]  2010  Healthy persons  Cell  Italy  Caucasian  NA  SIL  13  65  NA  NA  5  28  44    Yang [25]  2011  Mixed  Blood  China  Asian  NS  SIL  38  382  60  16  17  15  50  Van    Vang [25]  2011  Mixed  Blood  China  Asian  NS  SIL  38  382  60  16  17  15  530    Yang [25]  2011  Mixed  Blood  China  Asian  NS  SIL  38  382  60  16  17  15  530	Piyathilake [30]	2000	Healthy persons	Tissue or cell	USA	Mixed	NS	SIL	25	31	25	25	9	13	-	6	4	8	6 1:	5	15	
Agodi [35]    2010    Healthy persons    Cell    Italy    Caucasian    NA    SIL    123    66    NA    NA    SE    NA    5    NA      Yang [25]    2011    Mixed    Blood    China    Asian    NS    SIL    38    382    60    16    23    14    15    530      Yang [25]    2011    Mixed    Blood    China    Asian    NS    SIL    38    382    60    16    14    15    530							т	SIL	39	31	45	33	11	23	2	~	44	8	6 1:	3	15	
Yang [25]    2011    Mixed    Blood    China    Asian    NS    SIL    38    382    60    16    23    14    1    15    530      Yang [25]    2011    Mixed    Blood    China    Asian    NS    SIL    38    382    60    16    23    14    1    15    530      IC    157    382    229    85    77    75    5    80    530	Agodi [35] 2	2010	Healthy persons	Cell	Italy	Caucasian	NA S	_	123	66	NA	NA	118	NA	VA 5		A N	VA 5	5	A N/	11	
ICC 157 382 229 85 77 75 5 80 530	Yang [25] 2	2011	Mixed	Blood	China	Asian	NS S	_	38	382	60	16	23	14	-	5	530	234 1	82 1(	56 34	20	0
							⊻	Ŋ	157	382	229	85	77	75	80	0	230	234 1	82 1(	56 34	20	0
Ma [31] 2006 Hospital patients Blood China Asian NS ICC 111 111 93 129 20 53 38 91 126	Ma [31] 2	2006	Hospital patients	Blood	China	Asian	NS	0	111	111	93	129	20	23	38	-	126	90 90	9 0(	0 18	78	

ŝ
Ë
.0
ě
Ξ
ß
·>
ē
0
.⊆
٦
S
Ē
<u>d</u>
2
ξ
-fi
ă
υ
õ
5
2
4
Æ
Ī
F
2
e
÷
ç
0
es
ö
Ξ
S
0
Ę
E C
Ä
ψ
as
0
b
ð
<u>n</u>
2
.=
he
Ŧ
ę
S
Ľ.
ist
ē
せ
Гa
Ja
Ū
<b>_</b> i
e
ą
L0

First author [reference]	Year	Source of control	Source of DNA	Country	Ethnicity	HWE	Histology	, Sample	size	Case					Cont	lo				
								case	control	A	U	AA	AC AC	₹ U	C+CC A	υ	AA	AC	ម	AC+CC
Tong [26]	2011	Healthy persons	Blood	Korea	Asian	NS	SIL	160	428	260	60	107	46 7	5	3 688	168	278	132	18	150
							HSIL	160	428	273	47	117	39 4	4	688	168	278	132	18	150
								148	428	235	61	68	57 2	Ň	9 688	168	278	132	18	150
Kohaar [22]	2010	Healthy persons	Tissue or cell	India	Asian	NS	HSIL	39	231	50	28	15	20 4	2	4 289	173	85	119	27	146
							IJ	164	231	199	129	28	33 2	-	06 289	173	85	119	27	146
Nandan [27]	2008	Healthy persons	Blood	India	Asian	NA	SIL	80	77	NA	NA	14	AA P	IA 6	5 NA	ΝA	37	NA	NA	40
							S	62	77	NA	NA	20	AA P	IA 4	2 NA	ΝA	37	NA	NA	40
Kang [23]	2005	Healthy persons	Blood	Korea	Asian	NS	S	79	84	132	26	55	22 2	2	4 141	27	58	25	-	26
Yang [25]	2011	Mixed	Blood	China	Asian	NS	SIL	38	382	62	14	24	14	Ē	4 606	158	237	132	13	145
							ICC	157	382	245	69	68	57 1	9	3 606	158	237	132	13	145
Abbreviations:   intra-epithelia    doi:10.1371/jou	HWE: H lesion. rnal.po	lardy-Weinberg Eq me.0052381.t002	luilibrium; NA, not av	ailable; NS, no	ot significant	;; LSIL, lo	w-grade squ	ui snomer	traepithelia	al lesion; H	ISIL, high-	grade sq	ramous	intraep	ithelial lesior	ı; ICC, inva	sive cer	vical car	icer; SIL,	squamous

model ( $I^2 = 68\%$ , P = 0.01) and the random-effects model was performed. However, there was no significant heterogeneity for the comparison of other genetic models (P > 0.1) and the fixedeffects method was performed for our investigation. In the subgroup analysis, no association was found between the A1298C polymorphism and SIL. Interestingly, the investigation on the women with A1298C polymorphisms vs. no carriers showed a marginally increased susceptibility to ICC but no statistically significant difference in dominant model (P = 0.06, OR = 1.21, 95% CI = 0.99–1.49) and AC vs. AA (P = 0.09, OR = 1.21, 95% CI = 0.97–1.51).

In the sensitivity analyses, the overall association between the MTHFR A1298C genotype and the cervical lesions was changed after an exclusion of one study [27] which lacked enough data to calculate if it conformed to HWE among the control group. However, the results of the sensitivity analysis on the cervical lesions were virtually unchanged after an exclusion of any other individual study (Figure 5). The shape of the funnel plots was symmetrical, which showed that no evidence was found for the publication bias among the studies (Figure 6). No publication bias was also detected according to the results of Egger's and Begg's tests (Table 4). Furthermore, neither the funnel plots nor the Begg's and Egger's tests found any obvious evidence for the publication bias in the subgroup analysis on all genetic models (data not shown).

#### Discussion

As we know, HPV infection may be necessary but is not sufficient to cause cervical cancer. Other factors may play some important roles in this cancer development. For example, the nutritional factors may affect the persistence of HPV infection and thereby influence progression of early precancerous lesions to invasive cancer. Specifically, folate plays a key role in DNA synthesis, repair, and methylation, and this forms the basis of mechanistic explanations for a putative role for folate in cancer prevention. However, the effect of folate in these processes may be modulated by the genotype for the common C677T or A1298C variants of MTHFR, the homozygosity of which is associated with a lower level of the enzyme activity, lower plasma and red blood cell folate, and elevated plasma homocysteine [42,43]. Several studies investigated the association between the MTHFR polymorphisms and the preinvasive cervical lesions or cervical cancer, but the results were not consistent. Thus, our meta-analysis could better evaluate association between the MTHFR C677T/ A1298C polymorphisms and the susceptibility to cervical lesions. Our findings demonstrate that there was no association between them. To our knowledge, this is the first meta-analysis on association between MTHFR C677T/A1298C polymorphisms and susceptibility to cervical lesions, and the largest-scale metaanalysis examining the risk of cervical cancer.

As for the MTHFR C677T, most evidence points to decrease in the susceptibility to colorectal cancer and an increase in the susceptibility to esophagus and gastric cancer [44–48], but the effect on the cervical cancer susceptibility was not consistent. In our meta-analysis, no statistically significant difference was found in the frequency of the MTHFR C677T polymorphism in the patients with cervical lesions when compared with the controls. This finding was consistent with that of one previous meta-analysis [49]. However, 9 new studies [20–22,25–27,32,33,35] have been published since 2006 and all recruited in our study dramatically increased the case number of cervical lesion and controls with genetic information, which indicated that our results could be more reliable. In addition, multiple subgroup analyses made our

Α	Experim	ental	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% CI	M-H, Random, 95% Cl
Agodi 2010	5	123	11	66	3.1%	0.21 [0.07, 0.64]	
Goodman 2001	77	150	86	179	7.0%	1.14 [0.74, 1.76]	
Kang 2005	52	79	44	74	5.4%	1.31 [0.68, 2.53]	
Kohaar 2010	62	203	70	231	7.2%	1.01 [0.67, 1.52]	<b>T</b>
Lambropoulos 2003	80	138	49	91	6.3%	1.18 [0.69, 2.01]	
Ma 2006	91	111	78	111	5.5%	1.93 [1.02, 3.62]	
Mostowska 2011	68	124	99	168	6.7%	0.85 [0.53, 1.35]	
Nandan 2008 Divethilelie 2000	12	142	24		5.9%	2.27 [1.27, 4.07]	
Piyathilake 2000	47	04	122	266	4.070	2.90 [1.40, 0.22]	
Proceed 2011	- 21	62	132	126	2.2%	1 1 2 (0 26 2 62)	
Shekari 2009	30	200	75	200	2.370	0.20 (0.10, 0.33)	
Sull 2004	329	462	301	454	81%	1 26 (0.95 1.66)	-
Tong 2011	306	465	275	427	8.1%	1.06 (0.81, 1.40)	+
Yang 2011	95	195	200	382	7.7%	0.86 [0.61, 1.22]	
Zoodsma 2005	422	900	319	592	8.5%	0.76 [0.61, 0.93]	-
		2400		2504	100.0%	0.0010.70.4.261	
Total (95% CI)	1760	3498	1002	3594	100.0%	0.99 [0.78, 1.26]	Ť
Heterogeneity: Tau <sup>2</sup> = (	1702 116:Chi≧=	66 87	df = 15.02		0001) <sup>.</sup> I <sup>2</sup> =	. 78%	
Test for overall effect: Z	C = 0.06 (P)	= 0.95)	ui - 10 (i	. 0.0			0.01 0.1 1 10 100
-			~ .			F	avours experimental Favours control
B	Experim	ental	Contr	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	lotal	Events	lotal	weight	M-H, Random, 95% Cl	M-H, Random, 95% Cl
Goodman 2001	67	140	75	168	7.8%	1.14 [0.73, 1.78]	
Kang 2005	32	59	32	62	5.2%	1.11 [0.54, 2.27]	
Kohaar 2010	58	199	65	226	8.1%	1.02 [0.67, 1.55]	
Lambropoulos 2003	65	123	37	79	6.5%	1.27 [0.72, 2.24]	
Ma 2006	53	73	60	93	5.6%	1.46 [0.75, 2.84]	<b>—</b>
Mostowska 2011	59	115	81	150	7.4%	0.90 [0.55, 1.46]	
Piyathilake 2000	36	53	24	56	4.6%	2.82 [1.29, 6.18]	
Piyathilake 2007	16	75	116	339	6.2%	0.52 (0.29, 0.95)	
Prasad 2011	5	62	8	124	2.7%	1.27 [0.40, 4.06]	
Shekari 2008	28	198	68	193	7.3%	0.30 (0.18, 0.50)	
Sull 2004	227	360	221	374	9.6%	1.18 [0.88, 1.59]	<b>+</b> -
Tong 2011	221	380	198	350	9.7%	1.07 [0.80, 1.43]	+
Yang 2011	89	189	166	348	8.9%	0.98 (0.68, 1.39)	+
Zoodsma 2005	350	828	262	535	10.5%	0.76 [0.61, 0.95]	-
Total (95% CI)		2854		3097	100.0%	0.97 [0.78, 1.20]	•
Total events	1306	2001	1413		1001070		1
Heterogeneity: Tau <sup>2</sup> = (	1.10: Chi²=	= 41.92	df = 13.0	P < 0.0	$001$ ); $I^2 = I$	69%	
Test for overall effect: Z	(= 0.32 (P	= 0.75)					
<u>^</u>	E		Contr	-1		h Odda Datia	avours experimental Favours control
C Stucky or Subgroup	Experime	Total	Evente	Total	Moight	M H Pandom 95% CL	M H Bandom 95% Cl
Goodman 2004	10	00	11	104	6.0%	1 16 /0 47 0 00	
Kong 2005	20	03	10	104	0.3%	1.10 [0.47, 2.00]	
Kang 2005 Kohaar 2010	20	145	5	166	1 1 96	0.01 [0.70, 4.45]	
Lambropoulos 2003	15	73	12	54	7.4%	0.31 [0.24, 3.47]	
Ma 2006	38	58	18	51	8.0%	3 48 [1 58 7 67]	
Mostowska 2011	g	65	18	87	7.2%	0.62 [0.26 1.48]	<b>_</b> _
Pivathilake 2000	11	28	6	38	5.3%	3 45 [1 09 10 96]	
Piyathilake 2007	5	64	16	239	6.0%	1.18 [0.42, 3.36]	
Prasad 2011	ň	57	1	117	1.0%	0.68.00.03.16.84	
Shekari 2008	2	172	7	132	3.4%	0.21 [0.04, 1.03]	
Sull 2004	102	235	80	233	12.2%	1.47 [1.01, 2.13]	<b>-</b>
Tong 2011	85	244	77	229	12.1%	1.06 [0.72, 1.54]	+
Yang 2011	6	106	34	216	7,0%	0.32 [0.13, 0.79]	<b>_</b> _
Zoodsma 2005	72	550	57	330	12.1%	0.72 [0.49, 1.05]	
Total (95% CI)		1027		2030	100.0%	1 06 10 76 1 401	▲
Total evente	270	1921	264	2030	100.0%	1.00 [0.70, 1.48]	Ť
Heterogeneity Tau? - 0	379 120:Ωbi≧−	33 71	004 df=12/9		11): E – 64	1%	
Test for overall effect: Z	(= 0.35 (P =	= 0.73)	ar⊐ 13 (r	- 0.00			0.01 0.1 1 10 100
		,				F	avours experimental Favours control

**Figure 2. Forest plot describing the association between the C677T polymorphism and the risk of cervical lesions.** (A) Meta-analysis in a random-effects model for CT+TT vs. CC (dominant model). (B) Meta-analysis in a random-effects model for CT vs. CC. (C) Meta-analysis in a random-effects model for TT vs. CC. Each study is shown by the point estimate of the OR (the size of the square is proportional to the weight of each study) and 95% CI for the OR (extending lines). doi:10.1371/journal.pone.0052381.g002

Genetic model	Number of study	Sample	Size	Analysis	l² (%)	Ph	Test o	f Association	P(Publication	bias test)
		Case	Control	Model	-		Р	OR(95%CI)	Begg's test	Egger's test
Total										
Dominant model	16	3498	3594	R	78	0.00	0.95	0.99 [0.78, 1.26]	0.558	0.626
Recessive model	14	3233	3451	R	51	0.01	0.75	1.05 [0.80, 1.38]	0.827	0.956
Additive model	14	6177	6902	R	79	0.00	0.79	0.97 [0.80, 1.18]	1.000	0.659
CT vs. CC	14	2854	3097	R	69	0.00	0.75	0.97 [0.78, 1.20]	0.443	0.490
TT vs. CC	14	1927	2038	R	61	0.00	0.73	1.06 [0.76, 1.48]	0.913	0.614
Pathological type										
ICC										
Dominant model	12	2008	2932	R	73	0.00	0.62	0.94 [0.72, 1.21]		
Dominant model*	11	1946	2855	R	73	0.00	0.44	0.90 [0.69, 1.18]		
Recessive model	11	1946	2855	R	59	0.00	0.96	1.01 [0.70, 1.45]		
Additive model	11	3915	5710	R	80	0.00	0.51	0.92 [0.73, 1.17]		
CT vs. CC	11	1731	2534	R	64	0.00	0.29	0.88 [0.69, 1.12]		
TT vs. CC	11	1229	1657	R	65	0.00	0.84	0.96 [0.62, 1.47]		
SIL										
Dominant model	11	1490	2916	R	71	0.00	0.54	1.09 [0.82, 1.45]		
Dominant model	9	1287	2773	R	52	0.04	0.51	1.08 [0.86, 1.35]		
Recessive model	9	1287	2773	F	0	0.79	0.80	1.03 [0.83, 1.27]		
Additive model	9	2574	5546	R	43	0.08	0.59	1.04 [0.90, 1.21]		
CT vs. CC	9	1123	2475	R	47	0.06	0.27	1.09 [0.94, 1.26]		
TT vs. CC	9	698	1609	F	0	0.45	0.36	1.11 [0.88, 1.40]		
Geographic area										
Asian										
Dominant model	9	1919	2081	R	80	0.00	0.71	1.07 [0.76, 1.49]		
Recessive model	8	1777	2004	R	65	0.00	0.74	1.08 [0.70, 1.66]		
Additive model	8	3242	4008	R	83	0.00	0.82	0.97 [0.71, 1.31]		
	8	1520	1770	R	72	0.00	0.72	0.95 [0.70, 1.28]		
TT vs. CC	8	1064	1186	R	69	0.00	0.77	1.08 [0.65, 1.80]		
European										
Dominant model	4	1285	917	R	62	0.05	0.18	0.77 [0.52.1.13]		
Recessive model	3	1162	851	F	0	0.89	0.13	0.79 [0.58.1.07]		
Additive model	3	2347	1702	F	0	0.42	0.006	0.83 [0.72.0.95]		
	3	1066	764	F	30	0.24	0.05	0.83 [0.69.1.00]		
	3	688	471	F	0	0.82	0.05	0.73 [0.53 1.00]		
USA	-	000		,	v	0.02	0.00			
Dominant model	3	294	596	R	83	0.00	0.62	1.22 [0.56, 2.65]		
Becessive model	3	294	596	F	0	0.72	0.25	1 39 [0 79 2 45]		
Additive model	3	588	1192	R	76	0.02	0.57	1 16 [0 70 1 93]		
	3	268	563	R	83	0.02	0.74	1 15 [0 50 2 63]		
	2	175	201	E	20	0.00	0.12	1 56 [0.99 2.77]		

Table 3. Pooled Analysis on Association between the MTHFR C677T polymorphism and the cervical lesion risk.

Dominant model: CT+TT vs. CC; Recessive model: TT vs. CC+CT; Additive model: T vs. C; R, Random-effects model; F, fixed-effects model; ICC: invasive cervical cancer; SIL, squamous intra-epithelial lesion; Dominant model\*: one study [27] omitted; Dominant model<sup>2</sup>: two studies [27,35] omitted. doi:10.1371/journal.pone.0052381.t003



**Figure 3. Funnel plot analysis on the detection of the publication bias for the C677T polymorphism.** (A) Meta-analysis in a randomeffects model for CT+TT vs. CC (dominant model). (B) Meta-analysis in a random-effects model for CT vs. CC. (C) Meta-analysis in a random-effects model for TT vs. CC. Each point represents an individual study for the indicated association. LogOR, natural logarithm of OR. Perpendicular line denotes the mean effect size. doi:10.1371/journal.pone.0052381.q003

meta-analysis more convincing too. We meta-analyzed the eligible case-control studies for C677T by geographic regions. No association was found between the C677T polymorphism and the cervical lesions in either in the Asian or in the American populations. However, a significant inverse association was found in the European population. Different genetic backgrounds or environmental conditions could explain the discrepancy. The meta-analysis also stratified by histological stages of cervical lesions showed that there was no association between the MTHFR C677T variants and cervical lesion development. To assess the effect of individual study on the overall meta-analysis estimate, we excluded one study at a time, and the exclusion of any single report did not change the significance of the final conclusion,

which indicated that the outcomes were robust. Taken together, we could make a conclusion that cervical lesion were not primarily caused by genetically-determined enzymatic defects in the folate metabolic pathway, which might be different from the pathways supposed for colorectal or gastric carcinogenesis. The effect of those polymorphisms on the cervical cancer susceptibility seems to be further modulated by other cofactors such as infection with the HPV and smoking.

As for MTHFR A1298C, some studies reported a positive association with cervical lesions, which had only borderline significance [25]. More recent studies have revealed no association between the MTHFR A1298C and the cervical lesions [22,23,26,27]. Our meta-analysis confirmed that there is no

A	Experime	ental	Contro	ol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Random, 95% C	M-H, Random, 95% Cl
Kang 2005	24	79	26	84	13.9%	0.97 [0.50, 1.90]	1
Kohaar 2010	130	203	146	231	21.7%	1.04 [0.70, 1.53]	」
Nandan 2008	108	142	40	77	15.8%	2.94 [1.63, 5.30]	] –––
Tong 2011	155	468	150	428	25.5%	0.92 [0.70, 1.21]	] 4
Yang 2011	82	195	145	382	23.1%	1.19 (0.83, 1.69)	」
Total (95% CI)		1087		1202	100.0%	1.21 [0.87, 1.69]	। ♦
Total events	499		507				
Heterogeneity: Tau <sup>2</sup> =	0.09; Chi <sup>2</sup>	= 12.66	, df = 4 (F	P = 0.01	); l <sup>z</sup> = 68%	6	
Test for overall effect:	Z=1.12 (P	e 0.26)					0.01 0.1 1 10 100
							Favours experimental Favours control
В	Experim	ental	Cont	rol		Odds Ratio	Odds Ratio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H, Fixed, 95% Cl	M-H, Fixed, 95% Cl
Kang 2005	22	77	25	83	8.0%	0.93 [0.47, 1.83]	
Kohaar 2010	103	176	119	204	21.3%	1.01 [0.67, 1.52]	<u>+</u>
Tong 2011	142	455	132	410	44.4%	0.96 [0.72, 1.27]	<b>+</b>
Yang 2011	81	204	132	369	26.3%	1.18 [0.83, 1.68]	
Total (95% CI)		912		1066	100.0%	1.02 [0.85, 1.24]	+
Total events	348		408				
Heterogeneity: Chi <sup>2</sup> =	0.95, df =	3 (P = 0	.81); I <sup>2</sup> =	0%			
Test for overall effect:	Z=0.25 (	P = 0.80	))			F	avours experimental Favours control
C	Exnerim	ental	Cont	rol		Odds Ratio	Odds Batio
Study or Subgroup	Events	Total	Events	Total	Weight	M-H. Fixed, 95% Cl	M-H. Fixed, 95% Cl
Kang 2005	2	57	1	59	2.1%	2 11 10 19 23 921	
Kohaar 2010	27	100	27	112	40.7%	1 16 [0 63 2 16]	
Tong 2011	13	326	18	296	39.6%	0.64 [0.31, 1.33]	
Yang 2011	1	114	13	250	17.7%	0.16 (0.02, 1.25)	<b>e</b>
Tung 2011				200		0.10 [0.02] 1.20]	
Total (95% CI)		597		717	100.0%	0.80 [0.52, 1.24]	•
Total events	43		59				
Heterogeneity: Chi <sup>2</sup> =	4.73, df=	3 (P = 0	.19); I <sup>2</sup> =	37%			
Toet for overall effect:							

**Figure 4. Forest plot describing the association between the A1298C polymorphism and the risk of cervical lesions.** (A) Meta-analysis in a random-effects model for AC+CC vs. AA (dominant model). (B) Meta-analysis in a random-effects model for AC vs. AA. (C) Meta-analysis in a random-effects model for CC vs. AA. Each study is shown by the point estimate of the OR (the size of the square is proportional to the weight of each study) and 95% CI for the OR (extending lines). doi:10.1371/journal.pone.0052381.g004

PLOS ONE | www.plosone.org

Table 4. Pooled Analysis on Association between the MTHFR A1298C polymorphism and the cervical lesion risk.

Genetic model	Number of study	Sample	Size	Analysis	l <sup>2</sup> (%)	Ph	Test o	of Association	P(Publication	bias test)
		Case	Control	Model	-		Р	OR(95%CI)	Begg's test	Egger's test
Total										
Dominant model	5	1087	1202	R	68	0.01	0.26	1.21[0.87, 1.69]	0.462	0.290
Recessive model	4	945	1125	F	42	0.16	0.33	0.81[0.54, 1.23]	1.000	0.992
Additive model	4	1890	2250	F	0	0.81	0.82	0.98[0.85, 1.14]	1.000	0.587
AC vs. AA	4	912	1066	F	0	0.81	0.80	1.02[0.85, 1.24]	1.000	0.930
CC vs. AA	4	597	717	F	37	0.19	0.31	0.80[0.52, 1.24]	1.000	0.971
Pathological type										
ICC										
Dominant model	5	610	1202	F	0	0.63	0.06	1.21[0.99, 1.49]		
Recessive model	4	548	1125	R	51	0.10	0.46	0.67[0.24, 1.93]		
Additive model	4	1096	2250	F	0	1.00	0.43	1.07[0.90, 1.27]		
AC vs. AA	4	520	1066	F	0	0.62	0.09	1.21[0.97, 1.51]		
CC vs. AA	4	319	717	F	43	0.15	0.46	0.82[0.49, 1.38]		
SIL										
Dominant model	4	477	1118	R	83	0.00	0.49	1.28[0.63, 2.60]		
Recessive model	3	397	1041	F	0	0.85	0.43	0.78[0.42, 1.44]		
Additive model	3	794	2082	F	0	0.90	0.14	0.85[0.68, 1.06]		
AC vs. AA	3	382	983	F	0	0.75	0.25	0.85[0.65, 1.12]		
CC vs. AA	3	278	658	F	0	0.86	0.34	0.74[0.40, 1.38]		

Dominant model: CC+AC vs. AA; Recessive model: CC vs. AC+AA; Additive model: C vs. A; R, Random-effects model; F, fixed-effects model; ICC, invasive cervical cancer; ICC: invasive cervical cancer; SIL, squamous intra-epithelial lesion.

doi:10.1371/journal.pone.0052381.t004



Figure 5. Influence analysis of the summary odds ratio coefficients on the association between the A1298C polymorphism and cervical cancer in dominant model. The results were computed by omitting each study (left column) in turn. Bars, 95% Cls. doi:10.1371/journal.pone.0052381.g005



**Figure 6. Funnel plot analysis on the detection of the publication bias for the A1298C polymorphism.** (A) Meta-analysis in a randomeffects model for AC+CC vs. AA (dominant model). (B) Meta-analysis in a random-effects model for AC vs. AA. (C) Meta-analysis in a random-effects model for CC vs. AA. Each point represents an individual study for the indicated association. LogOR, natural logarithm of OR. Perpendicular line denotes the mean effect size. doi:10.1371/journal.pone.0052381.q006

association between the A1298C polymorphism and cervical lesions, similar to that found by the subgroup analysis on the ethnic groups and the histological stages of cervical lesions. No association was found between the A1298C polymorphism and SIL, but the ICC showed a marginally positive association though with no statistically significant difference. This result suggested that a probably higher risk for cervical cancer was linked to the A1298C variants, implying their important role in later stages of cervical carcinogenesis but not in SILs. Sensitivity analyses revealed that the overall association between the MTHFR A1298C genotype and cervical lesions could be changed after excluding one study [27] which lacked sufficient data to calculate whether it conformed to HWE among or not in the control group. In contrast, the results were virtually unchanged after the exclusion of any other individual study. To sum up, it is possibly indicated that the study by Nandan et al. could be the main source of the observed heterogeneity across the studies in this metaanalysis. Alternatively, the study may had limitations or because of other unknown factors.

To some extent, several limitations of this meta-analysis should be addressed. One limitation of the present study was that the sample size of A1298C mutation involved is not big enough. We neen more original researches to make our conclusions more reliable and accurate. The studies on the A1298C variant had reported only 5 articles, and their participants were entirely Asians with no population variation in minor allele frequency. So, the subgroup meta-analysis on this gene polymorphism was not possible by race. Another limitation was that significant heterogeneity in the studies was mainly present in overall analyses and subgroup analyses. Though several possible sources of the between-study heterogeneity were investigated, including patho-

#### References

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, et al. (2011) Global cancer statistics. CA Cancer J Clin 61: 69–90.
- Bosch F, Lorincz A, Munoz N, Meijer C, Shah K (2002) The causal relation between human papillomavirus and cervical cancer. J Clin Pathol 55: 244–265.
- Walboomers JMM, Jacobs MV, Manos MM, Bosch FX, Kummer JA, et al. (1999) Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. J Pathol 189: 12–19.
- Elfgren K, Kalantari M, Moberger B, Hagmar B, Dillner J (2000) A populationbased five-year follow-up study of cervical human papillomavirus infection. Am J Obstet Gynecol 183: 561–567.
- Insinga RP, Dasbach EJ, Elbasha EH (2009) Epidemiologic natural history and clinical management of Human Papillomavirus (HPV) disease: a critical and systematic review of the literature in the development of an HPV dynamic transmission model. BMC Infect Dis 9: 119–119.
- Castellsagué X, Muñoz N (2003) Cofactors in human papillomavirus carcinogenesis–role of parity, oral contraceptives, and tobacco smoking. JNCI Monographs 2003: 20–28.
- Castellsague X, Bosch FX, Munoz N (2002) Environmental co-factors in HPV carcinogenesis. Virus Res 89: 191–199.
- García-Closas R, Castellsagué X, Bosch X, González CA (2005) The role of diet and nutrition in cervical carcinogenesis: a review of recent evidence. Int J Cancer 117: 629–637.
- Van Niekerk W (1966) Cervical cytological abnormalities caused by folic acid deficiency. Acta Cytol 10: 67–73.
- Whitehead N, Reyner F, Lindenbaum J (1973) Megaloblastic changes in the cervical epithelium: association with oral contraceptive therapy and reversal with folic acid. JAMA 226: 1421–1424.
- VanEenwyk J, Davis FG, Colman N (1992) Folate, vitamin C, and cervical intraepithelial neoplasia. Cancer Epidemiol Biomarkers Prev 1: 119–124.
- Butterworth Jr C, Hatch KD, Macaluso M, Cole P, Sauberlich HE, et al. (1992) Folate deficiency and cervical dysplasia. JAMA 267: 528–533.

logical history, geographic region, ethnicity, source of controls, and source of DNA for genotyping ethnicity (data not shown), none of them could sufficiently explain the heterogeneity. The effect estimates might depend on some unidentified sources of heterogeneity. Besides, part of the exposure information was still lacking in the available studies, E.g., HPV infection status, smoking status or nutritional status (particularly folate intake or level). Therefore, effects of environment exposure or lifestyle on association between MTHFR variants and cervical lesions could not be determined by this meta-analysis.

In summary, despite the above-mentioned limitations, the present study provides evidence that the MTHFR C677T and A1298C polymorphisms may not increase the susceptibility to cervical cancer development. However, our meta-analysis reveals a negative association between the MTHFR C677T mutations and cervical lesions, especially in the European populations. The marginal association between the MTHFR A1298C polymorphisms and the susceptibility for cervical cancer need to be further studied.

#### **Supporting Information**

Table S1PRISMA checklist.(DOC)

#### **Author Contributions**

Conceived and designed the experiments: SL XY PY. Performed the experiments: SL XY XL. Analyzed the data: SL XY XL. Contributed reagents/materials/analysis tools: SL XY XL. Wrote the paper: SL XY. Helped edit the manuscript: XL PY.

- Potischman N, Brinton LA, Laiming VA, Reeves WC, Brenes MM, et al. (1991) A case-control study of serum folate levels and invasive cervical cancer. Cancer Res 51: 4785–4789.
- Sedjo RL, Inserra P, Abrahamsen M, Harris RB, Roe DJ, et al. (2002) Human papillomavirus persistence and nutrients involved in the methylation pathway among a cohort of young women. Cancer Epidemiol Biomarkers Prev 11: 353– 359.
- Piyathilake CJ, Henao OL, Macaluso M, Cornwell PE, Meleth S, et al. (2004) Folate is associated with the natural history of high-risk human papillomaviruses. Cancer Res 64: 8788–8793.
- Pillai M, Chacko P, Kesari L, Jayaprakash P, Jayaram H, et al. (2003) Expression of folate receptors and heterogeneous nuclear ribonucleoprotein E1 in women with human papillomavirus mediated transformation of cervical tissue to cancer. J Clin Pathol 56: 569–574.
- Blount BC, Mack MM, Wehr CM, MacGregor JT, Hiatt RA, et al. (1997) Folate deficiency causes uracil misincorporation into human DNA and chromosome breakage: implications for cancer and neuronal damage. Proc Natl Acad Sci USA 94: 3290–3295.
- Yamada K, Chen Z, Rozen R, Matthews RG (2001) Effects of common polymorphisms on the properties of recombinant human methylenetetrahydrofolate reductase. Proc Nati Acad Sci USA 98: 14853–14858.
- Prasad V, Wilkhoo H (2011) Association of the Functional Polymorphism C677T in the Methylenetetrahydrofolate Reductase Gene with Colorectal, Thyroid, Breast, Ovarian, and Cervical Cancers. Onkologie 34: 422–426.
- Mostowska A, Myka M, Lianeri M, Roszak A, Jagodzinski PP (2011) Folate and choline metabolism gene variants and development of uterine cervical carcinoma. Clin Biochem 44: 596–600.

- Kohaar I, Kumar J, Thakur N, Hussain S, Niyaz MK, et al. (2010) Homocysteine levels are associated with cervical cancer independent of methylene tetrahydrofolate reductase gene (MTHFR) polymorphisms in Indian population. Biomarkers 15: 61–68.
- Kang S, Kim JW, Kang GH, Park NH, Song YS, et al. (2005) Polymorphism in folate-and methionine-metabolizing enzyme and aberrant CpG island hypermethylation in uterine cervical cancer. Gynecol Oncol 96: 173–180.
- Lambropoulos A, Agorastos T, Foka Z, Chrisafi S, Constantinidis T, et al. (2003) Methylenetetrahydrofolate reductase polymorphism C677T is not associated to the risk of cervical dysplasia. Cancer Lett 191: 187–191.
- Yang F, Zhou Y, Jiang Y, Fan Y, Li J (2010) Study on the correlation between polymorphism of MTHFR gene and the pathogenesis of cervical cancer. J China Maternal Child Health 23: 4122–4124.
- 26. Tong S, Kim MK, Lee JK, Lee JM, Choi SW, et al. (2011) Common polymorphisms in methylenetetrahydrofolate reductase gene are associated with risks of cervical intraepithelial neoplasia and cervical cancer in women with low serum folate and vitamin B12. Cancer Causes Control 22: 63–72.
- Nandan NK, Wajid S, Biswas S, Juneja SS, Rizvi M, et al. (2008) Allelic variations in 5, 10-methylenetetrahydrofolate reductase gene and susceptibility to cervical cancer in Indian women. Drug Metabolism Lett 2: 18–22.
- Sull JW, Jee SH, Yi S, Lee JE, Park JS, et al. (2004) The effect of methylenetetrahydrofolate reductase polymorphism C677T on cervical cancer in Korean women. Gynecol Oncol 95: 557–563.
- Goodman MT, McDuffie K, Hernandez B, Wilkens LR, Bertram CC, et al. (2001) Association of methylenetetrahydrofolate reductase polymorphism C677T and dietary folate with the risk of cervical dysplasia. Cancer Epidemiol Biomarkers Prev 10: 1275–1280.
- Piyathilake C, Macaluso M, Johanning G, Whiteside M, Heimburger D, et al. (2000) Methylenetetrahydrofolate reductase (MTHFR) polymorphism increases the risk of cervical intraepithelial neoplasia. Anticancer Res 20: 1751–1757.
- Ma X, Wang J, Zhou Q, Ding L, Cheng Y, et al. (2006) Relationship between Methylenetetrahydrofolate reductase polymorphism and cervical cancer susceptibility. China Public Health 22: 1247–1248.
- Shekari M, Sobti RC, Kordi Tamandani DM, Suri V (2008) Impact of methylenetetrahydrofolate reductase (MTHFR) codon (677) and methionine synthase (MS) codon (2756) on risk of cervical carcinogenesis in North Indian population. Arch Gynecol Obstet 278: 517–524.
- Piyathilake CJ, Azrad M, Macaluso M, Johanning GL, Cornwell PE, et al. (2007) Protective association of MTHFR polymorphism on cervical intraepithelial neoplasia is modified by riboflavin status. Nutrition 23: 229–235.
- Zoodsma M, Nolte IM, Schipper M, Oosterom E, van der Steege G, et al. (2005) Methylenetetrahydrofolate reductase (MTHFR) and susceptibility for (pre)neoplastic cervical disease. Hum Genet 116: 247–254.

- Agodi A, Barchitta M, Cipresso R, Marzagalli R, La Rosa N, et al. (2010) Distribution of p53, GST, and MTHFR polymorphisms and risk of cervical intraepithelial lesions in sicily. Int J Gynecol Cancer 20: 141–146.
- Collin SM, Metcalfe C, Zuccolo L, Lewis SJ, Chen L, et al. (2009) Association of folate-pathway gene polymorphisms with the risk of prostate cancer: a population-based nested case-control study, systematic review, and metaanalysis. Cancer Epidemiol Biomarkers Prev 18: 2528–2539.
- Moher D, Liberati A, Tetzlaff J, Altman DG, The PRISMA Group (2009) Preferred reporting items for systematic reviews and meta-analyses: The PRISMA Statement. PLoS Med 6(6): e1000097. doi:10.1371/journal.pmed1000097.
- Higgins J, Thompson SG (2002) Quantifying heterogeneity in a meta-analysis. Stat Med 21: 1539–1558.
- DerSimonian R, Laird N (1986) Meta-analysis in clinical trials. Control Clini Trials 7: 177–188.
- Begg CB, Mazumdar M (1994) Operating characteristics of a rank correlation test for publication bias. Biometrics 50: 1088–1101.
- Egger M, Smith GD, Schneider M, Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. BMJ 315: 629–634.
- Jacques PF, Bostom AG, Williams RR, Ellison RC, Eckfeldt JH, et al. (1996) Relation between folate status, a common mutation in methylenetetrahydrofolate reductase, and plasma homocysteine concentrations. Circulation 93: 7–9.
- 43. Ashfield-Watt PAL, Pullin CH, Whiting JM, Clark ZE, Moat SJ, et al. (2002) Methylenetetrahydrofolate reductase 677C>T genotype modulates homocysteine responses to a folate-rich diet or a low-dose folic acid supplement: a randomized controlled trial. Am J Clin Nutrition 76: 180–186.
- Hubner RA, Houlston RS (2007) MTHFR C677T and colorectal cancer risk: A meta-analysis of 25 populations. Int J Cancer 120: 1027–1035.
- Huang Y, Han S, Li Y, Mao Y, Xie Y (2007) Different roles of MTHFR C677T and A1298C polymorphisms in colorectal adenoma and colorectal cancer: a meta-analysis. J Hum Genet 52: 73–85.
- Larsson SC, Giovannucci E, Wolk A (2006) Folate Intake, MTHFR Polymorphisms, and Risk of Esophageal, Gastric, and Pancreatic Cancer: A Meta-analysis. Gastroenterology 131: 1271–1283.
- Langevin S, Lin D, Matsuo K, Gao C, Takezaki T, et al. (2009) Review and pooled analysis of studies on MTHFR C677T polymorphism and esophageal cancer. Toxicol Lett 184: 73–80.
- Zintzaras E (2006) Association of methylenetetrahydrofolate reductase (MTHFR) polymorphisms with genetic susceptibility to gastric cancer: a metaanalysis. J Hum Genet 51: 618–624.
- Zacho J, Yazdanyar S, Bojesen SE, Tybjærg-Hansen A, Nordestgaard BG (2011) Hyperhomocysteinemia, methylenetetrahydrofolate reductase 677C>T polymorphism and risk of cancer: Cross-sectional and prospective studies and meta-analyses of 75,000 cases and 93,000 controls. Int J Cancer 128: 644–652.