



# Airway remodeling in asthma

Carolyn Damilola Ekpruke<sup>^</sup>, Patricia Silveyra<sup>^</sup>

Department of Environmental and Occupational Health, Indiana University Bloomington, School of Public Health, Bloomington, IN, USA

*Correspondence to:* Patricia Silveyra. Department of Environmental and Occupational Health, Indiana University Bloomington, School of Public Health, Bloomington, IN 47405, USA. Email: psilveyr@iu.edu.

*Comment on:* Huang Y, Qiu C. Research advances in airway remodeling in asthma: a narrative review. *Ann Transl Med* 2022;10:1023.

Submitted Oct 13, 2022. Accepted for publication Oct 21, 2022.

doi: 10.21037/atm-22-5059

**View this article at:** <https://dx.doi.org/10.21037/atm-22-5059>

Asthma is a heterogeneous lung disease affecting approximately 300 million patients worldwide. Asthma is generally characterized by anatomical alterations of the airway. Such alterations are collectively referred to as airway remodeling. In airway remodeling, both small and large airways are affected (1). Airway remodeling comprises the thickening of airway smooth muscle layers due to increased cell proliferation and inflammation (1). In allergic asthma, pattern recognition receptors in epithelial cells help secrete chemokines and cytokines to recruit dendritic cells; the reason why epithelial cells are referred to as airway remodeling initiators (2). Immune cells release interleukin-13, platelet-derived growth factor, and transforming growth factor- $\beta$  which help in the amplification of the airway remodeling magnitude in asthma (3). These cells are also referred to as airway remodeling mediators through the activation of fibroblasts in the submucosa (4).

Several scientists have reported the pathological features of airway remodeling, including thickening of the airway wall and subepithelial reticular basement membrane, damage to epithelial cells, alterations in airway smooth muscle cells, mucus gland hyperplasia and hypertrophy, and angiogenesis and vascular remodeling. In this issue of the journal, Huang and Qiu reviewed the most recent research advances in airway remodeling in asthma, from the literature available between 2001–2022. The authors addressed this topic by discussing the various pathological features of airway remodeling, including the interaction of different cell types within the airway smooth muscle layer and the submucosa (5). They indicated that airway remodeling can be either pathological or physiological and either prevent or contribute to bronchoconstriction.

For example, bronchoconstriction in asthma can result from the degradation of the cartilage thereby contributing to the stiffness of the airway (6). Similarly, goblet cell hyperplasia causes an increase in the production of sputum, consequently leading to the narrowing and thickening of the airway walls (5). However, some studies suggest that the thickening of the airway walls in airway remodeling may also prevent bronchoconstriction (6). During airway remodeling, the deposition of the matrix to the subepithelial layer can make the airway stiff and therefore prevent its narrowing (7). Similarly, the migration and increased contractility of the airway smooth muscle cells may help prevent obstruction of airflow in the airway (8).

The study by Huang and Qiu looked at the various ways of evaluating airway remodeling either by directly or indirectly assessing the airway tissues using invasive or non-invasive tools. The authors, clearly discuss the direct methods and stated the limitations of these methods as reported by the American Thoracic Society guidelines. Direct methods of evaluating airway remodeling include carrying out endobronchial biopsy (9), transbronchial biopsy, and trans-endobronchial cryobiopsy (9), though these methods have limitations. Indirectly, airway remodeling has been evaluated by collecting and analyzing the bronchoalveolar lavage fluid (BALF). While BALF is a well-recognized method for quantifying the number of cells released during the asthma allergic cascade, the dilution factor used by different investigators is usually inconsistent, which makes the method less accurate. Other relatively non-invasive, indirect methods used to assess airway remodeling are sputum analysis (10), exhaled breath condensate,

<sup>^</sup> ORCID: Carolyn Damilola Ekpruke, 0000-0002-9045-4457; Patricia Silveyra, 0000-0001-7083-8915.

and blood and urine analysis (11). These methods are easier than the direct methods, but their accuracy remain controversial. Alternative tools use to assess air remodeling in asthma include computed tomography (12), optical coherence tomography (13), endobronchial ultrasound (14), position emission tomography (15), hyperpolarized magnetic resonance imaging and Birefringence microscopy. These methods are very efficient but also costly. Similarly, high resolution non-linear optical microscopy has replaced the normal tissue staining method by scientists evaluating airway remodeling in asthma (16). The ratio of forced expiratory volume in one second and forced vital capacity values obtained during lung function testing can also be used to characterize this major asthma component (17).

The authors also discussed different ways of treating airway remodeling in asthma. These methods include the use of glucocorticoids, anti-IgE, anti-TNF antibodies and vitamin D therapies. They also made mention of a non-pharmacological, safe, and efficient method, bronchial thermoplasty and the development of individualized treatment protocols. Some researchers reported that glucocorticoid drugs may not be effective for the treatment of airway remodeling as they observed that it can only partially reduce lung function and airway thickness (18), while others reported that the use of these sets of drugs induced epithelial cell apoptosis and hindered their migration (19). The effectiveness of monoclonal antibodies, include varieties of anti-IL-5, IL-4/IL-13 and IL-33 in the treatment of airway remodeling that occurs in asthma has also been investigated (20). Recently, the use of bronchial thermoplasty was reported as a new therapy for the treatment of severe refractory asthma (21). This procedure consists of the introduction of radiofrequency into the airway wall through bronchoscopy and has proven to reduce airway remodeling. All these methods and their limitations are described in the review.

This study has several limitations. First, it concentrates on clinical evidence and did not discuss information from experimental models used to understand mechanisms of airway remodeling in the past few years. Such models range from *in vitro* to *in vivo* and have been specifically developed to study the pathogenesis of asthma and airway remodeling. Examples of models that were recently used for this purpose include the epithelial-mesenchymal trophic unit model of the bronchial mucosa (22), and porcine airway smooth muscle strips to assess airway contractility (23). Second, sex differences in airway remodeling and asthma treatment were not discussed. There is well-established evidence that sex differences occur in asthma across the life span, and many researchers are advocating for a sex-specific treatment

of the disease. Finally, the authors do not mention potential ways of improving the present methods of treating asthma to make them affordable and accessible to more patients. Recently, some researchers have studied the role of nutrition in the prevention and treatment of asthma (24) as there is a general belief that nutrition or diet possesses immunomodulatory effect. This may help to complement the existing methods for treatment lung disease.

In short, this review summarizes the different pathological features of airway remodeling and the existing different ways of treating asthma. The summary provided by the authors might help provide information leading to development of better strategies for evaluating airway remodeling mechanisms both in animal models and clinical studies. For instance, sex-differences in airway remodeling that occur in asthma need to be investigated, and sex-specific responses to treatment reviewed. Future review studies should focus on integrating evidence of airway remodeling in both clinical and animal studies and providing therapeutic options that account for sex differences in disease pathogenesis and presentation, as well as consideration of more affordable therapeutic options such as nutritional interventions.

## Acknowledgments

*Funding:* This study was supported by the NIH funding (No. R01HL159764).

## Footnote

*Provenance and Peer Review:* This article was commissioned by the editorial office, *Annals of Translational Medicine*. The article did not undergo external peer review.

*Conflicts of Interest:* The authors have completed the ICMJE uniform disclosure form (available at <https://atm.amegroups.com/article/view/10.21037/atm-22-5059/coif>). The authors have no conflicts of interest to declare.

*Ethical Statement:* Both authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-

commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

1. Elliot JG, Jones RL, Abramson MJ, et al. Distribution of airway smooth muscle remodelling in asthma: relation to airway inflammation. *Respirology* 2015;20:66-72.
2. Nathan AT, Peterson EA, Chakir J, et al. Innate immune responses of airway epithelium to house dust mite are mediated through beta-glucan-dependent pathways. *J Allergy Clin Immunol* 2009;123:612-8.
3. Holgate ST. A look at the pathogenesis of asthma: the need for a change in direction. *Discov Med* 2010;9:439-47.
4. Zhang S, Smartt H, Holgate ST, et al. Growth factors secreted by bronchial epithelial cells control myofibroblast proliferation: an in vitro co-culture model of airway remodeling in asthma. *Lab Invest* 1999;79:395-405.
5. Hough KP, Curtiss ML, Blain TJ, et al. Airway Remodeling in Asthma. *Front Med (Lausanne)* 2020;7:191.
6. Haraguchi M, Shimura S, Shirato K. Morphometric analysis of bronchial cartilage in chronic obstructive pulmonary disease and bronchial asthma. *Am J Respir Crit Care Med* 1999;159:1005-13.
7. Noble PB, Turner DJ, Mitchell HW. Relationship of airway narrowing, compliance, and cartilage in isolated bronchial segments. *J Appl Physiol* (1985) 2002;92:1119-24.
8. Joubert P, Lajoie-Kadoch S, Labonté I, et al. CCR3 expression and function in asthmatic airway smooth muscle cells. *J Immunol* 2005;175:2702-8.
9. Pepe C, Foley S, Shannon J, et al. Differences in airway remodeling between subjects with severe and moderate asthma. *J Allergy Clin Immunol* 2005;116:544-9.
10. Suzuki R, Kato T, Miyazaki Y, et al. Matrix metalloproteinases and tissue inhibitors of matrix metalloproteinases in sputum from patients with bronchial asthma. *J Asthma* 2001;38:477-84.
11. Belleguic C, Corbel M, Germain N, et al. Increased release of matrix metalloproteinase-9 in the plasma of acute severe asthmatic patients. *Clin Exp Allergy* 2002;32:217-23.
12. Fuso L, Macis G, Condoluci C, et al. Impulse oscillometry and nitrogen washout test in the assessment of small airway dysfunction in asthma: Correlation with quantitative computed tomography. *J Asthma* 2019;56:323-31.
13. Williamson JP, McLaughlin RA, Noffsinger WJ, et al. Elastic properties of the central airways in obstructive lung diseases measured using anatomical optical coherence tomography. *Am J Respir Crit Care Med* 2011;183:612-9.
14. Bullone M, Beauchamp G, Godbout M, et al. Endobronchial Ultrasound Reliably Quantifies Airway Smooth Muscle Remodeling in an Equine Asthma Model. *PLoS One* 2015;10:e0136284.
15. Trivedi A, Hall C, Hoffinan EA, et al. Using imaging as a biomarker for asthma. *J Allergy Clin Immunol* 2017;139:1-10.
16. Mostaço-Guidolin LB, Osei ET, Ullah J, et al. Defective Fibrillar Collagen Organization by Fibroblasts Contributes to Airway Remodeling in Asthma. *Am J Respir Crit Care Med* 2019;200:431-43.
17. Ward C, Johns DP, Bish R, et al. Reduced airway distensibility, fixed airflow limitation, and airway wall remodeling in asthma. *Am J Respir Crit Care Med* 2001;164:1718-21.
18. Chakir J, Shannon J, Molet S, et al. Airway remodeling-associated mediators in moderate to severe asthma: effect of steroids on TGF-beta, IL-11, IL-17, and type I and type III collagen expression. *J Allergy Clin Immunol* 2003;111:1293-8.
19. Royce SG, Li X, Tortorella S, et al. Mechanistic insights into the contribution of epithelial damage to airway remodeling. Novel therapeutic targets for asthma. *Am J Respir Cell Mol Biol* 2014;50:180-92.
20. Brightling CE, Chaney P, Leigh R, et al. Efficacy and safety of tralokinumab in patients with severe uncontrolled asthma: a randomised, double-blind, placebo-controlled, phase 2b trial. *Lancet Respir Med* 2015;3:692-701.
21. Facciolo N, Bonacini M, Galeone C, et al. Bronchial thermoplasty in severe asthma: a real-world study on efficacy and gene profiling. *Allergy Asthma Clin Immunol* 2022;18:39.
22. Bucchieri F, Pitruzzella A, Fucarino A, et al. Functional characterization of a novel 3D model of the epithelial-mesenchymal trophic unit. *Exp Lung Res* 2017;43:82-92.
23. Dogan M, Han YS, Delmotte P, et al. TNF $\alpha$  enhances force generation in airway smooth muscle. *Am J Physiol Lung Cell Mol Physiol* 2017;312:L994-L1002.
24. Alwarith J, Kahleova H, Crosby L, et al. The role of nutrition in asthma prevention and treatment. *Nutr Rev* 2020;78:928-38.

**Cite this article as:** Ekpruke CD, Silveyra P. Airway remodeling in asthma. *Ann Transl Med* 2022;10(22):1189. doi: 10.21037/atm-22-5059