

Revascularization of Transplanted Islets

Can It Be Improved?

Marcela Brissova¹ and Alvin C. Powers^{1,2,3}

Pancreatic islets are highly vascularized, which is important in their ability to quickly secrete insulin in response to changes in blood glucose. Although pancreatic islets comprise only 1–2% of pancreatic mass, they receive 5–10% of pancreatic blood flow. Blood vessels within pancreatic islets are of a greater density than those in surrounding exocrine tissue and are lined with fenestrated endothelial cells. These specialized features are responsible for the greater partial pressure of oxygen in islets compared with acinar tissue and other organs, which is likely important for normal islet cell function. Islet production of angiogenic factors such as vascular endothelial growth factor-A (VEGF-A) and angiopoietin-1 is critical for creating this highly vascularized state (1,2). During embryonic development, reciprocal endothelial-endocrine cell signaling and the formation of functional blood vessels appear to instruct pancreatic differentiation and morphogenesis (3–5). Development of the islet vasculature is coordinated with islet formation, but blood flow to endocrine cells precedes their final assembly into a mature islet (2).

Pancreatic islet isolation severs the connections between the islet vasculature and the systemic circulation. In contrast with whole-organ transplantation, where organ perfusion is quickly reestablished by reconnection of arterial and venous vessels, the reestablishment of blood flow to transplanted islets requires several days and involves angiogenesis and possibly vasculogenesis. Not only are islets avascular for several days following transplantation, they are less vascularized and have a lower oxygen tension than islets in the pancreas when revascularization is complete (6,7). The death of significant numbers of islets in the days following transplantation results from several factors, but ischemia and inadequate blood supply are likely contributors to islet death in the immediate posttransplant period and may impair islet survival and function long term. Thus, improvements in the revascularization of transplanted islets may enhance islet survival and the outcomes of islet transplantation.

Efforts to improve the revascularization of transplanted

islets are hindered because the responsible ligands, receptors, cells, and mechanisms are not well defined. Recent evidence indicates that the endothelial cells creating new capillaries or vessels within the islet graft arise from three sources (Fig. 1). The first source is the endothelial cells from the transplant recipient, which are recruited into the islet graft. A second source is intraislet endothelial cells, which exist in large numbers in isolated islets and may account for up to 40% of the endothelial cells lining capillaries within a revascularized graft (8,9). Interestingly, functional vessels within a revascularized graft are often chimeric, consisting of both endothelial cells from the transplant recipient and donor-derived, intraislet endothelial cells. Bone marrow–derived cells are a third, but likely minor, source of endothelial cells (10,11). The factors produced by transplanted islet cells that stimulate or recruit endothelial cells from the three potential sources in the graft include VEGF-A (2), but other pro- or antiangiogenic molecules could also play a role (Fig. 1). The formation of new vessels also requires vascular remodeling involving the basement membrane, vascular supporting cells such as pericytes, and the extracellular matrix. Little is known about these processes in islet revascularization.

The revascularization of transplanted islets might be enhanced or accelerated by several types of interventions. One approach would be to increase the action of proangiogenic factors or to inhibit antiangiogenic factors and thus stimulate the proliferation, migration, and maturation of endothelial cells into functional vessels. This approach has had some hints of success (12–15), but it is likely that the optimal formation of mature, fully functional islet vasculature will require precise control of the timing, dose, and duration of angiogenic factor action in the posttransplant period. A second approach could directly target endothelial cells or enhance their ability to form mature, functional vessels and might involve the addition of preactivated endothelial cells or some type of endothelial progenitor cell population. These two approaches should be applicable to isolated islets before transplantation or, also, could be used to prepare the transplantation site before transplantation of isolated islets.

In this issue of *Diabetes*, Johansson et al. (16) propose a new approach using tissue engineering to enhance islet revascularization. These investigators provide evidence that the coculture of mesenchymal stem cells (MSCs) and endothelial cells with human islets in vitro before transplantation initiates formation of vessel-like structures that may promote islet engraftment after transplantation. MSCs, multipotent cells usually isolated from bone marrow but also present in other tissues, exhibit a wide range of properties in other settings, properties that might enhance islet survival (17–19). For example, MSCs positively modulate inflammation, tissue regeneration, and

From the ¹Department of Medicine, Division of Diabetes, Endocrinology, and Metabolism, Vanderbilt University, Nashville, Tennessee; the ²Department of Molecular Physiology and Biophysics, Vanderbilt University, Nashville, Tennessee; and the ³VA Tennessee Valley Healthcare System, Nashville, Tennessee.

Corresponding author: Alvin C. Powers, al.powers@vanderbilt.edu.

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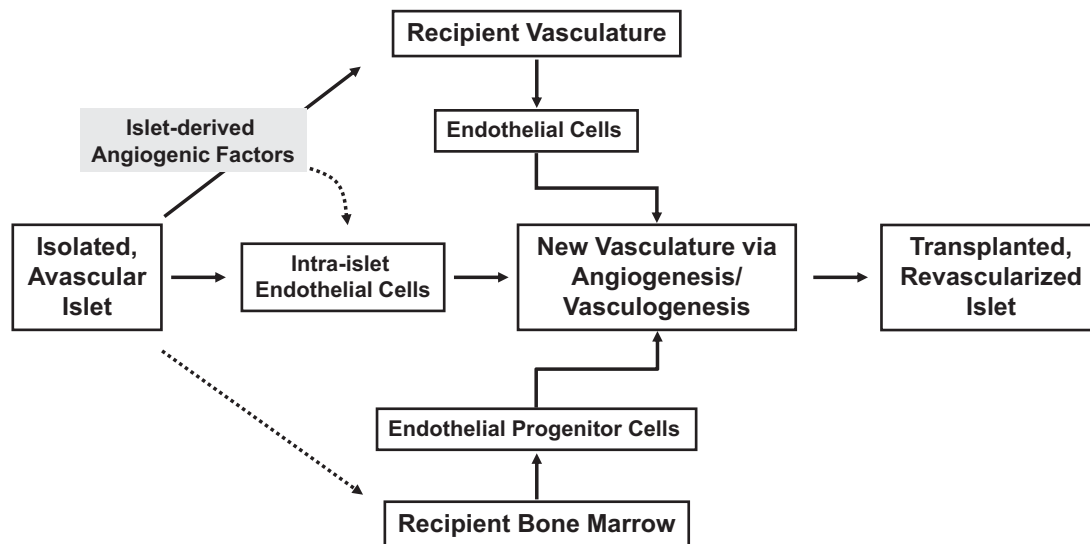


FIG. 1. Model of islet revascularization after transplantation. Endothelial cells from three sources (recipient endothelial cells at the site of transplant, intra-islet endothelial cells, and endothelial progenitor cells from the bone marrow) contribute to revascularization. The dotted line refers to processes or factors not yet defined.

immune attack either through cell-to-cell contact, differentiation into other cell types, or by the local production of factors such as platelet-derived growth factor. Johansson et al. purified MSCs from normal human bone marrow using cell-surface markers and found that MSCs or factors produced by these cells promoted endothelial cell proliferation and migration and the “coating” of cultured islets with endothelial cells (16). Using an in vitro system to study angiogenesis, these investigators demonstrated that this mixture of MSCs, endothelial cells, and islets promoted the migration of exogenous endothelial cells into the cultured islets; the formation of chimeric, vessel-like structures between the endogenous intra-islet endothelial cells and the endothelial cells added to the islet culture; and the formation of new vessel “sprouts” from islets. A unique and possibly critical component in these studies was the microvascular endothelial cells harvested from human dermis, which are likely more receptive to remodeling signals from the MSCs. Such MSCs and microvascular endothelial cells could likely be harvested and expanded from the bone marrow or adipose tissue of humans selected to receive an islet transplant.

So, how did MSCs promote these changes in endothelial cells and promote the formation of new intraislet vascular-like structures, and how might this be translated to islet transplantation? Additional work is needed to define the ligands, receptors, and mechanisms responsible for these effects, but Johansson et al. speculate that proteases from MSCs may degrade the islet extracellular matrix and thus allow the migration of endothelial cells that have been stimulated by growth factors such as VEGF-A produced by MSCs. Identification of these factors should allow one to test whether addition of these factors to cultured islets could substitute for the MSCs. As one considers how to extend these in vitro findings, demonstration that this coculture approach improves the function and survival of transplanted islets using in vivo models is the critical next step. Likewise, the long-term fate and growth potential of MSCs must be determined and may be dependent on the transplantation site microenvironment. Hopefully, as we better understand the molecular events of islet revascularization, delivery of angiogenic factors at the optimal dose, time, and duration combined with tissue engineering ap-

proaches such as that described by Johansson et al. will accelerate and enhance the revascularization of transplanted islets and lead to improved islet function and survival.

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REFERENCES

- Lammert E, Gu G, McLaughlin M, Brown D, Brekken R, Murtaugh LC, Gerber HP, Ferrara N, Melton DA: Role of VEGF-A in vascularization of pancreatic islets. *Curr Biol* 13:1070–1074, 2003
- Brissova M, Shostak A, Shiota M, Wiebe PO, Poffenberger G, Kantz J, Chen Z, Carr C, Jerome WG, Chen J, Baldwin HS, Nicholson W, Bader DM, Jetton T, Gannon M, Powers AC: Pancreatic islet production of vascular endothelial growth factor—a is essential for islet vascularization, revascularization, and function. *Diabetes* 55:2974–2985, 2006
- Lammert E, Cleaver O, Melton D: Induction of pancreatic differentiation by signals from blood vessels. *Science* 294:564–567, 2001
- Edsbacke J, Johansson JK, Esni F, Luo Y, Radice GL, Semb H: Vascular function and sphingosine-1-phosphate regulate development of the dorsal pancreatic mesenchyme. *Development* 132:1085–1092, 2005
- Jacquemin P, Yoshitomi H, Kashima Y, Rousseau GG, Lemaigre FP, Zaret KS: An endothelial-mesenchymal relay pathway regulates early phases of pancreas development. *Dev Biol* 290:189–199, 2006
- Carlsson PO, Palm F, Andersson A, Liss P: Markedly decreased oxygen tension in transplanted rat pancreatic islets irrespective of the implantation site. *Diabetes* 50:489–495, 2001
- Mattsson G, Jansson L, Carlsson PO: Decreased vascular density in mouse pancreatic islets after transplantation. *Diabetes* 51:1362–1366, 2002
- Brissova M, Fowler MJ, Wiebe P, Shostak A, Shiota M, Radhika A, Lin PC, Gannon M, Powers AC: Intra-islet endothelial cells contribute to revascularization of transplanted pancreatic islets. *Diabetes* 53:1318–1325, 2004
- Nyqvist D, Kohler M, Wahlstedt H, Berggren PO: Donor islet endothelial cells participate in formation of functional vessels within pancreatic islet grafts. *Diabetes* 54:2287–2293, 2005
- Contreras JL, Smyth CA, Eckstein C, Bilbao G, Thompson JA, Young CJ, Eckhoff DE: Peripheral mobilization of recipient bone marrow-derived endo-

- thelial progenitor cells enhances pancreatic islet revascularization and engraftment after intraportal transplantation. *Surgery* 134:390–398, 2003
11. Miller R, Cirulli V, Diaferia GR, Ninniri S, Hardiman G, Torbett BE, Benezra R, Crisa L: Switching-on survival and repair response programs in islet transplants by bone marrow-derived vasculogenic cells. *Diabetes* 57:2402–2412, 2008
 12. Lai Y, Schneider D, Kiszun A, Hauck-Schmalenberger I, Breier G, Brandhorst D, Brandhorst H, Iken M, Brendel MD, Bretzel RG, Linn T: Vascular endothelial growth factor increases functional beta-cell mass by improvement of angiogenesis of isolated human and murine pancreatic islets. *Transplantation* 79:1530–1536, 2005
 13. Olerud J, Johansson M, Lawler J, Welsh N, Carlsson PO: Improved vascular engraftment and graft function following inhibition of the angiostatic factor thrombospondin-1 in mouse pancreatic islets. *Diabetes* 57:1870–1877, 2008
 14. Su D, Zhang N, He J, Qu S, Slusher S, Bottino R, Bertera S, Bromberg J, Dong HH: Angiopoietin-1 production in islets improves islet engraftment and protects islets from cytokine-induced apoptosis. *Diabetes* 56:2274–2283, 2007
 15. Zhang N, Richter A, Suriawinata J, Harbaran S, Altomonte J, Cong L, Zhang H, Song K, Meseck M, Bromberg J, Dong H: Elevated vascular endothelial growth factor production in islets improves islet graft vascularization. *Diabetes* 53:963–970, 2004
 16. Johansson U, Rasmuson I, Niclou SP, Forslund N, Gustavsson L, Nilsson B, Korsgren O, Magnusson PU: Formation of composite endothelial cell-mesenchymal stem cell-islets; a novel approach to promote islet revascularization. *Diabetes* 57:2393–2401, 2008
 17. Ball SG, Shuttleworth CA, Kieley CM: Mesenchymal stem cells and neovascularization: role of platelet-derived growth factor receptors. *J Cell Mol Med* 11:1012–1030, 2007
 18. Brooke G, Cook M, Blair C, Han R, Heazlewood C, Jones B, Kambouris M, Kollar K, McTaggart S, Pelekanos R, Rice A, Rossetti T, Atkinson K: Therapeutic applications of mesenchymal stromal cells. *Semin Cell Dev Biol* 18:846–858, 2007
 19. Kumar S, Chanda D, Ponnazhagan S: Therapeutic potential of genetically modified mesenchymal stem cells. *Gene Ther* 15:711–715, 2008