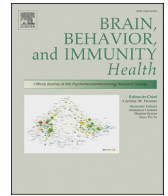


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Review

Neuroinflammation and glial cell activation in mental disorders

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ABSTRACT

Mental disorders (MDs) are highly prevalent and potentially debilitating complex disorders which causes remain elusive. Looking into deeper aspects of etiology or pathophysiology underlying these diseases would be highly beneficial, as the scarce knowledge in mechanistic and molecular pathways certainly represents an important limitation. Association between MDs and inflammation/neuroinflammation has been widely discussed and accepted by many, as high levels of pro-inflammatory cytokines were reported in patients with several MDs, such as schizophrenia (SCZ), bipolar disorder (BD) and major depression disorder (MDD), among others. Correlation of pro-inflammatory markers with symptoms intensity was also reported. However, the mechanisms underlying the inflammatory dysfunctions observed in MDs are not fully understood yet. In this context, microglial dysfunction has recently emerged as a possible pivotal player, as during the neuroinflammatory response, microglia can be over-activated, and excessive production of pro-inflammatory cytokines, which can modify the kynurenine and glutamate signaling, is reported. Moreover, microglial activation also results in increased astrocyte activity and consequent glutamate release, which are both toxic to the Central Nervous System (CNS). Also, as a result of increased microglial activation in MDs, products of the kynurenine pathway were shown to be changed, influencing then the dopaminergic, serotonergic, and glutamatergic signaling pathways. Therefore, in the present review, we aim to discuss how neuroinflammation impacts on glutamate and kynurenine signaling pathways, and how they can consequently influence the monoaminergic signaling. The consequent association with MDs main symptoms is also discussed. As such, this work aims to contribute to the field by providing insights into these alternative pathways and by shedding light on potential targets that could improve the strategies for pharmacological intervention and/or treatment protocols to combat the main pharmacologically unmatched symptoms of MDs, as the SCZ.

1. Introduction

Mental disorders (MDs) are characterized by clinically significant disturbances in cognition and behavior/emotion regulation, which all together reflect a brain dysfunction. MDs are one of the leading global causes of disability, and with recognized high prevalence (Kyū et al.,

2018). Despite the clear genetic predisposition, the etiology of MDs is multifactorial, meaning that there is a combined contribution of genetic and environmental factors (Caspi and Moffitt, 2006). The oldest and most widely accepted hypothesis for the pathophysiology of MDs, such as schizophrenia (SCZ), bipolar disorder (BD), and major depressive disorder (MDD), is based on the dysregulation of the monoaminergic

Abbreviations: MD, mental disorders; SCZ, schizophrenia; BD, bipolar disorder; MDD, major depression disorder; PRRs, pattern recognition receptors; TLRs, toll-like receptors; CLRs, C-type lectin receptors; NF, necrosis factor; IRF, interferon regulatory factor; APCs, antigen presenting cells; IL, interleukin; TNF, tumor necrosis factor; CNS, central nervous system; CCL, C-C motif chemokine ligand; CXCL, X-C motif chemokine ligand; IFN, interferon; TGF, tumor growth factor; CSF, cerebrospinal fluid; IDO, indolamine 2,3-dioxygenase; KYNA, kynurenic acid; NMDA, N-methyl-D-aspartate; AMPA, alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; PPI, prepulse inhibition; NMR, nuclear magnetic resonance; MRI, magnetic resonance imaging; QUIN, quinolinic acid; α -7nAChR, alpha 7 nicotinic acetylcholine receptor; BBB, blood-brain barrier.

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neurotransmission. For instance, the dopaminergic hyperfunction in mesolimbic pathways and hypofunction in mesocortical pathways were shown to explain several of the main symptoms as the general restricted emotional experience, hallucinations, deliriums, and cognitive deficits (Carlsson and Lindqvist, 1963). However, more recent data also indicate an important contribution of the hypofunction of glutamatergic (Kantrowitz and Javitt, 2012) and N-methyl-D-aspartate (NMDA) receptors (Snyder and Gao, 2013).

SCZ is currently among the most impactful and complex MDs, being also recognized as a neurodevelopmental disorder (Jablensky et al., 2017; Murray et al., 2017). Multiple genetic variants are expected to contribute to SCZ, although each one with only small effects (Luo et al., 2019). Together they may represent the potential targets for the environmental risk factors as, for instance, the maternal infections (Weber-Stadlbauer, 2017) or immune activation (Bergdolt and Dunaevsky, 2019), which may be equally capable of affecting the neurodevelopment. Consistently, the neurodevelopmental hypothesis for SCZ conciliates the interaction between the genetic and environmental factors as the possible origin of aberrant process (es) which could affect early brain formation during the embryonic development, potentially explaining the increased risk of developing SCZ (Zwicker et al., 2018; Bergdolt and Dunaevsky, 2019).

SCZ has also been linked to inflammation as the hypothesis of maternal immune activation, since a risk factor for the progeny to develop this disorder in adulthood was previously accepted by many (Mongan et al., 2019). In fact, blood analysis revealed consistent increase of IL-6, IL-12, IL-1 β , tumor necrosis factor alpha (TNF- α), and interferon gamma (IFN- γ) in patients with SCZ, as well as decreased IL-10 in relapsed SCZ inpatients; similarly as observed in the cerebrospinal fluid (CSF) of first onset and acute relapsed patients with SCZ, normalized for the antipsychotic treatment (Miller et al., 2011). In addition, increased levels of midbrain immune-related transcripts observed in SCZ and in murine offspring after maternal immune activation may suggest that immune-related changes in SCZ extend to dopaminergic areas of the midbrain, and that maternal immune activation could be a possible contributing factor underlying these persistent neuroimmune changes, which also includes the activation of microglial and astrocytic cells (Purves-Tyson et al., 2019).

In BD patients, mood fluctuations between episodes of elevation (mania) and depression (recurrent or single), interspersed with periods of euthymia, are classically reported (Grande et al., 2016), and their association with aberrant calcium and glutamate signaling was suggested to explain these phenotypes (Nurnberger et al., 2014). The involvement of the neuroimmune system in the pathological process of BD is also recognized (Niu et al., 2019).

Patients with MDD are characterized by depressed mood and/or loss of interest or pleasure in almost all regular activities, as well as by the presence of other symptoms, including changes in sleep patterns, restlessness, slow motor function, unwanted substantial weight changes, fatigue, loss of energy, feelings of worthlessness or guilt, loss of cognition and memory, among others. The efficacy of medications targeting monoaminergic pathways reinforces the involvement of these systems in the pathophysiology of MDD, although the lack of effectiveness in about one third of the cases may suggest that depression pathophysiology may go beyond monoamines (Chávez-Castillo et al., 2019; Pitsillou et al., 2019). New strategies, such as the use of rapid-action antidepressant ketamine, that targets, among others, the glutamatergic system, indicate the need to modulate other pathways to obtain real responses, especially in the so-called treatment resistant depression (Taylor et al., 2019).

Despite the incomplete knowledge about the pathophysiology of these mental conditions, different studies pointed out to the possible important roles of inflammation in MDs. In fact, a correlation between the peripheral immune modulators and psychiatric symptoms has long been demonstrated in either clinical or animal models (Mosley, 2015; Jeon et al., 2018). For instance, immunological alterations leading to inflammation mediators increases have been observed in several most

common MDs, as a result of differential microglial activation (Barichello et al., 2019). Therefore, further understanding of inflammation in MDs, as well as how it correlates to the known alterations in each disease may open new opportunities for a more suitable and satisfactory treatment/intervention, occasionally with the power to contribute to the prevention as well.

2. Inflammation and neuroinflammation overview

2.1. Inflammation

Inflammation was conceived as the immune system defensive response of an organism against pathogens and/or tissue insults, mainly based on the presence of different pattern recognition receptors (PRRs). PRRs belong to membrane-bound receptor families, which include the Toll-like receptors (TLRs) and C-type lectin receptors (CLRs), in addition to various cytosolic sensors, that once in contact with specific pathogenic proteins, as pathogen- or danger-associated molecular patterns, activate different signaling pathways aiming to isolate, dilute or destroy the noxious agent (Kumar, 2019). For instance, TLRs recognize most bacterial and viral pathogenic threats, and initiate the signaling cascades that culminate in the cell recruitment and production of inflammation mediators, as cytokines and chemokines (Kawasaki and Kawai, 2014). This response initiates with neutrophilic extravasation to the parenchymal tissue, followed by the recruitment of monocytes and lymphocytes (B- and T-cells) (Oo et al., 2010). Dendritic cells, also known as antigen-presenting cells (APCs), present these antigens to naïve T- and B-cells, subsequently, activating them. This process induces the formation of various T-cells subpopulations with different functions in the inflammatory response (Taniuchi, 2018). Once activated, monocytes, lymphocytes, mast cells and macrophages induce further production of inflammatory cytokines as IL-1 β , IL-6, and TNF- α , which are able to modulate the immune response triggered by the infection and/or inflammation, besides regulating the inflammation by itself (Turner et al., 2014). Activated macrophages are specifically divided in two subpopulations, namely M1 and M2, in which M1 macrophages have pro-inflammatory roles (pro-inflammatory cytokine production and pathogen elimination), while M2 macrophages have anti-inflammatory properties (production of IL-10) (Murray and Wynn, 2011).

2.2. Neuroinflammation

Neuroinflammation refers to the inflammation in the CNS, and several evidences have shown a two-way communication between the neuronal and immunological cells, enabling the cooperation at the hybrid junction level, which modulates the synaptic plasticity and neuroimmunity. In fact, inflammatory signaling exists in different ways in accessing the brain. For instance, the cytokines can cross the blood brain barrier (BBB) through circumventricular organs and specific saturable BBB transporters (Limanaqi et al., 2019). After an insult, the CNS activates the glial cells, especially the microglia, and a complex neuro-inflammatory signaling pathway is initiated, leading to the production and release of diverse chemokines and cytokines (Carson et al., 2006).

Microglia are the macrophage-like resident cells in the CNS, where they are responsible for the recognition of pathogen and production of cytokines and chemokines, such as the pro-inflammatory mediators IL-1 β , IL-6, TNF- α , CCL2 (C-C motif chemokine ligand 2), CCL5, CXCL1 (C-X-C motif chemokine ligand 1), nitric oxide (NO) and prostaglandins (DiSabato et al., 2016). Like the peripheral macrophages, in response to different stimuli, microglia can also be activated into two states, namely M1 (pro-inflammatory) and M2 (anti-inflammatory) (Boche et al., 2013).

The M1 activation is a response to the higher levels of IFN- γ and TNF- α , produced by natural killer cells and T-helper 1 lymphocytes, which results in increased release of pro-inflammatory cytokines as IL- β , IL-6, TNF- α , IL-23, and oxygen free radicals, as well as in increased phagocytic activity and antigen presentation. On the other hand, the M2

activation stimulates the release of anti-inflammatory cytokines as IL-10 and tumor growth factor beta (TGF- β), and this activation can inhibit the inflammatory phagocytosis, besides inducing tissue repair/wound healing (Mosser and Edwards, 2008). Microglial activation has a physiological role in cytokine production in early brain development and in adult CNS, as well as it also participates in immunological activities and synapses pruning (Salter and Beggs, 2014). On the other hand, aged microglia are more responsive to stimuli, and produce larger amounts of pro-inflammatory cytokines that can result in persistent neuro-inflammation relevant to diverse neuropathological diseases (Wolf et al., 2017). Apart from aging, stress-associated microglial activation, primarily in the M1 state, could be directly correlated to depressive behaviors and anxiety (Zhu et al., 2019). In this case, the elevated levels of glucocorticoids and catecholamines promote the brain inflammation, with consequent release of IL-1 β and other pro-inflammatory cytokines from the microglia which, in turn, can stimulate the glucocorticoid release by activating the hypothalamic-pituitary-adrenal axis (Sun et al., 2017).

Peripheral increases of inflammatory mediators in MD patients (for instance, in SCZ, BD and MDD) have suggested an association between the chronic inflammation and mental conditions (Niu et al., 2019). In addition, increased levels of pro-inflammatory cytokines and chemokines in the CNS, as a result of microglial activation, can contribute to specific characteristics and symptoms of different MDs (Lurie, 2018), as they may also interfere with relevant signaling pathways in each of these diseases (Haroon et al., 2018; Lanz et al., 2019).

The kynurenine and glutamate signaling pathways, as well as the alterations resulting from the microglial activation, and how they are associated to MDs will be further discussed as follow.

3. Kynurenine pathway

The kynurenine pathway metabolites interact with the glutamatergic and cholinergic receptors, namely NMDA and alpha 7 nicotinic acetylcholine (α 7-nACh) receptors, respectively, and alterations in these glutamatergic/cholinergic pathways were suggested to be relevant to several MDs (Erhardt et al., 2017). In fact, the kynurenine pathway is the main responsible for the production of several neuroactive molecules, including serotonin, quinolinic acid (QUIN), kynurenic acid (KYNA), among others, which are generated by the degradation of the non-essential amino acid tryptophan and other substrates (Ruddick et al., 2006; Carvalho et al., 2017). In the CNS, most cell types, including neurons, microglia and even infiltrating macrophages, produce all enzymes associated with the kynurenine pathway, while the astrocytes and oligodendrocytes do not produce some of these enzymes and, consequently, are unable of synthesizing QUIN (Schwarcz and Stone, 2017).

3.1. Kynurenine pathway and MDs

QUIN is an endogenous NMDA agonist, whereas KYNA is a NMDA

antagonist. Higher concentrations of both were previously reported in the brain during fetal development, which quickly declined after birth, indicating a possible relationship between the kynurenine metabolism and the development of CNS (Notarangelo and Pocivavsek, 2017). Moreover, NMDA receptors were demonstrated to be involved in neuronal migration, synapse formation, neurite outgrowth, and other events related to neuronal plasticity, and therefore, abnormal NMDA activation or inhibition could then compromise its functioning with consequent relevant brain and behavior alterations (Waters and Machaalani, 2004).

Increased pro-inflammatory cytokines, as a result of immune activation, can modulate the kynurenine pathway in the CNS, as IFN- γ stimulates the production of indolamine 2,3-dioxygenases (IDOs) and tryptophan 2,3-dioxygenase (TDOs) (Kim and Jeon, 2018) (Fig. 1). Kynurenine can cross the BBB, and once in the CNS it is absorbed by the glial cells, and subsequently metabolized into KYNA and QUIN (Kennedy et al., 2017), with different consequences, depending on the levels of each metabolite generated. KYNA is often described as neuroprotective in the brain, with sedative and anticonvulsant effects at low concentrations, while QUIN is described to be neurotoxic, inducing apoptosis in Th1 target cells and selectively inhibiting the proliferation of natural killer cells, namely CD4⁺ and CD8⁺ T-lymphocytes, once these cells are in their active state (Chen and Guillemin, 2009).

During the inflammatory response, IDO-1 and IDO-2 are upregulated by pro-inflammatory cytokines (Campbell et al., 2014), resulting in excessive production of KYNA and QUIN, which can both determine the neurotoxic effects mediated by the NMDA receptor (Schwarcz et al., 2012), and also, by indirectly altering other signaling pathways, including the dopaminergic pathway (Erhardt et al., 2017).

SCZ and BD were both previously associated with inflammation and high levels of pro-inflammatory cytokines, and increased levels of KYNA in the CSF and *post-mortem* brain of patients with SCZ and BD have been consistently described (Schwieler et al., 2015; Kindler et al., 2019). However, QUIN levels in SCZ and BD are still a matter of contradiction in the literature. While some studies indicate that the levels of QUIN and its precursor 3-hydroxykynurenine are unaltered in CSF and *post-mortem* brain of BD and SCZ patients (Sathyaikumar et al., 2011; Kegel et al., 2014), some others have suggested elevated peripheral levels of 3-hydroxykynurenine, which were apparently normalized after the treatment with antipsychotic drugs (Oxenkrug et al., 2016). Alterations in the KYN/tryptophan relation, as well as increased level of QUIN and KYNA after a second immune challenge in early adulthood, were observed in animal models for maternal immune activation (Clark et al., 2019).

In addition, in contribution to the theory of NMDA receptor hypofunction in SCZ (Snyder and Gao, 2013), a study with *post-mortem* samples of hippocampus of SCZ patients has demonstrated decreased contents of the NMDA receptor agonist QUIN in microglia, with levels even lower during the psychotic episode (Gos et al., 2014).

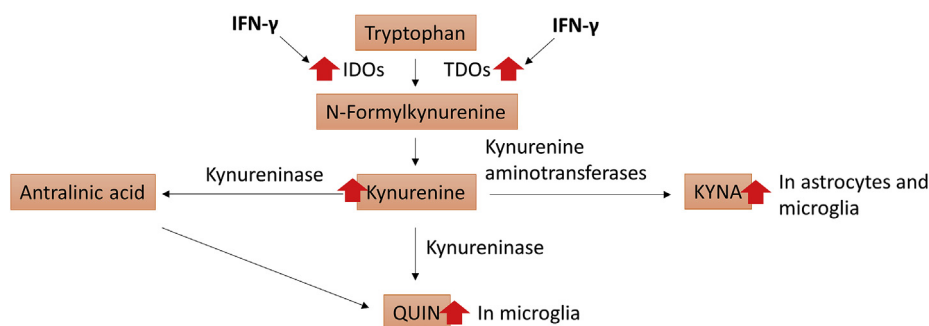


Fig. 1. Schematic representation showing the kynurenine pathway and its neuroactive metabolites QUIN and KYNA. IFN- γ stimulates the activity of IDOs and TDOs in converting tryptophan into kynurenine, which crosses the BBB and is converted into QUIN in the microglia, and KYNA in both microglia and astrocytes.

4. Glutamate pathway

Glutamate is an endogenous amino acid that acts as an excitatory neurotransmitter in the CNS, with a suggested important role in learning and memory, and in neuronal plasticity, by modulating cell elimination and neuronal development (McEntee and Crook, 1993). The neurotransmitter glutamate interacts with different types of receptors as, for instance, the NMDA ligand-gated ion channels and AMPA (alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid) ionotropic receptors (Ribeiro et al., 2017). Its roles in cell proliferation, neuronal migration, synaptic transmission, excitability, plasticity, amongst others, are well established (Moretto et al., 2018), allowing therefore the association of this signaling pathway with a distinct set of disorders, including neurodevelopmental and neurodegenerative psychiatric conditions (Ribeiro et al., 2017; Moretto et al., 2018). This important component to brain health needs a strict signaling regulation, as excessive glutamate can cause cell death in a process referred as "excitotoxicity", which occurs after activation of glutamate receptors in brain cells (Zhou and Danbolt, 2014).

Besides the increased release of pro-inflammatory mediators, microglial activation also results in decreased expression of glutamate transporters and receptors, stimulating increased production and release of glutamate in microglia (Haroon et al., 2017). Moreover, TNF- α also induces glutamate release from microglia and astrocytes (Tasker et al., 2012). Inflammatory mediators, such as chemokines and cytokines, produced either by the activated microglia in the CNS or by the peripheral macrophages in response to an immune response, activate astrocytes and stimulate the production of reactive oxygen species and glutamate release (Haroon and Miller, 2017).

Although the mechanism by which glutamate is correlated to MDs is not completely clear yet, it is known that glutamate is released by astrocytes and neurons in the CNS, in a process involving microglia and oligodendrocytes (Gundersen et al., 2015).

4.1. Glutamatergic signaling and MDs

Various MDs such as MDD, SCZ and post-traumatic stress disorder, are considered to be glutamate-related, especially regarding the interaction with the NMDA receptors (Schwarcz and Stone, 2017; Haroon and Miller, 2017). Interestingly, studies have suggested that the dopaminergic alterations observed in SCZ are mediated by altered glutamatergic signaling, due to a NMDA hypofunction, even considering the assumed increased glutamate release during neuroinflammation (Schwartz et al., 2012). For instance, clinical studies with NMDA antagonists, as phencyclidine and ketamine, have demonstrated the presence of characteristics similar to most common symptoms of SCZ, such as the emotional blunting, thought disorders, hallucinations, cognitive deficits, anhedonia and social retreat (Krystal et al., 2005).

In animal models, NMDA hypofunction (induced either by pharmacological or genetic approaches) resulted in behavioral changes associated with SCZ symptoms, as for instance, the hyperlocomotion in the open field test and the deficits in prepulse inhibition (PPI) of acoustic startle response, which were both reversed by the administration of antipsychotic drugs, such as clozapine and haloperidol (Lee and Zhou, 2019).

The progressive loss of brain tissue reported in SCZ (Chew et al., 2013) could also be correlated to glutamatergic dysregulation during the developmental period, since it could also affect the NMDA roles in synaptic plasticity and neuronal activity (Harrison and Weinberger, 2005). Neonatal administration of NMDAR antagonist phencyclidine in rats induced functional deficits in hippocampal brain network, as denoted by the changes in oscillatory rhythms in the excitation/inhibition balance of the brain (Kjaerby et al., 2017).

Regardless of the possible correlation of glutamate with SCZ, most of data found in the literature refer to the glutamatergic contribution in MDD, in which the association with microglial activation and consequent

neuroinflammation was early demonstrated (Leonard, 2018). In fact, depressive patients presented high glutamate levels in the plasma, CSF and brain (Sanacora et al., 2008). In addition, it is likely that glutamate signaling alterations in the brain are region-dependent, as decreased levels of glutamate in the prefrontal cortex and anterior cingulate cortex were noticed by nuclear magnetic resonance (NMR) spectroscopy studies (Murrough et al., 2017). In addition, the NMDA receptor antagonist ketamine showed antidepressant effects in patients, as suggested by the reduction of sad mood, anhedonia, pessimism and indecision (Williams and Schatzberg, 2016), and also in experimental animal models, as demonstrated by several behavioral tests (Refsgaard et al., 2017).

Clinical evidence also points out to a key role of glutamatergic transmission in the pathophysiology of BD (Blacker et al., 2017). Neurophysiological abnormalities, related to the glutamate-glutamine cycle, membrane turnover, and neuronal integrity, have also been described in BD patients (Kubo et al., 2017). Literature data regarding glutamate/NMDA alterations in BD patients in depressive phase are in accordance with those for depressive patients, mainly concerning the effects of the antidepressant ketamine (Zarate et al., 2012). Studies with BD patients demonstrated links between the genes responsible for coding ionotropic glutamate receptors subunits, the risk for BD, and the response to the treatment with lithium (Le-Niculescu et al., 2009). Lithium has been shown to decrease both glutamatergic and dopaminergic excitatory signaling (Malhi et al., 2013), while proton NMR spectroscopy studies in BD patients revealed elevated prefrontal glutamate across mood states (Smaragdi et al., 2019).

5. Dopamine pathway

Dopamine is a catecholaminergic neurotransmitter that plays an important role in motor function, motivation, cognition, emotion and neuroendocrine secretion. Dopaminergic innervation is the most prominent in the brain, and the four major signaling pathways identified are the nigrostriatal, mesolimbic, mesocortical and tuberoinfundibular (Beaulieu and Gainetdinov, 2011). Dopamine is early expressed in developing brain and it is relevant to the development of neuronal cytoarchitecture, with influence in processes such as the cell proliferation, migration and differentiation (Money and Stanwood, 2013). Therefore, changes in this pathway can lead to altered connectivity and dysfunctional synapses, as demonstrated in animal models with disrupted dopaminergic signaling (Zhang et al., 2010). From the five known dopamine receptors (D1, D2, D3, D4 and D5), three were implied to contribute to SCZ symptoms, with the negative symptoms resulting from reduced D1 activation and possible D3 alterations, and with the positive symptoms resulting from increased D2 expression (Simpson et al., 2014).

5.1. Dopamine system and MDs

The impact of dopaminergic transmission in SCZ is widely studied, since most antipsychotic drugs relieve symptom by interacting directly or indirectly with dopamine receptors D2 (Amato et al., 2018). The dopamine receptors of mesolimbic and mesocortical pathways are the main targets of the antipsychotic drugs currently used for the treatment of several MDs, and increased dopaminergic signaling in the mesolimbic is described to be responsible for the positive symptoms observed in SCZ, while decreased dopaminergic signaling in the cortex is suggested to be responsible for the negative symptoms (McCutcheon et al., 2019).

The mechanisms related to the dopamine neurotransmission and symptoms in BD are similar to that in SCZ (Ashok et al., 2017), in which increased signaling during the mania phase and a decreased signaling during the depressive phase are reported (Tye et al., 2013; Sidor et al., 2015). Moreover, dopamine antagonists and/or partial agonists are often used in the treatment of acute mania, bipolar depression and also for treatment maintenance (Hooshmand et al., 2014). Moreover, dopaminergic transmission seems to be mildly modulated by glutamate and other neuroactive metabolites capable of interacting with NMDA and

AMPA receptors (Felger, 2017).

Deficits in the reward process, motivation and motor function are commonly described in MDs, and a correlation between these symptoms and dopaminergic dysfunction and inflammation is discussed (Felger and Treadway, 2017). Even if the mechanism by which this signaling is altered is still not clear, there is evidence supporting that activation of innate immunity and release of inflammatory cytokines may both play important roles (Harrison et al., 2015), by affecting the dopamine synthesis, release and uptake, in order to reduce the dopamine neurotransmission in the basal ganglia (Felger, 2017). The effect in synthesis is due to increased release of QUIN, KYNA and glutamate, that contribute together to oxidative stress and generation of reactive oxygen species, ultimately leading to decreased production of L-3,4-dihydroxyphenylalanine (L-DOPA) (Vancassel et al., 2018). Besides, KYNA is a NMDA antagonist, and therefore, could negatively modulate the dopamine release (Giménez-Gómez et al., 2018). Increased cytokines could decrease the expression or function of the vesicular monoamines transporter 2 (VMAT2), and can increase the dopaminergic reuptake by the dopamine transporter (DAT), ultimately leading to altered dopaminergic signaling, as it also occurs with serotonin transporters (van Heesch et al., 2013).

6. Conclusion or final remarks

Although there are recommended treatments for MDs, many of them do not show efficacy in all patients or do not necessarily relieve all symptoms, besides showing several undesirable side effects. For this reason, it is important to keep investigating the underlying pathophysiology aiming to discover and characterize new targets for pharmacological intervention.

Microglial activation was shown to modulate the release of glutamate and production of metabolites from the kynurenine pathway, which are

capable of modulating the glutamatergic, serotonergic and dopaminergic signaling, mainly through interaction with NMDA glutamatergic receptors (Fig. 2). These receptors were demonstrated to be altered in a distinct set of MDs, and to contribute to some of reported/observed symptoms. Since it is known that MDs are influenced by environmental factors, besides the importance of the genetic predisposition, the immune activation may emerge as the missing link between these two factors.

Therefore, new treatment possibilities may emerge by interfering not only in the classic monoaminergic signaling pathways, but also targeting upstream in the cascade of events that results in these well-known alterations in the monoaminergic signaling. In this context, early intervention in the production of the inflammatory cytokines, kynurenine pathway metabolites and glutamate, or in their interactions with receptors, may have the power to decrease the effects of other relevant monoaminergic pathways responsible for the MDs symptoms, including the dopaminergic, serotonergic and glutamatergic signaling. We recognize that further experimental data are essential to solidify this suggestion, but this work aimed mainly to contribute to summarize how these altered pathways are correlated to MDs. In addition, we would like to stimulate further studies in the field by shedding light on the potential new possibilities for pharmacological intervention for the treatment of unmet symptoms in MDs.

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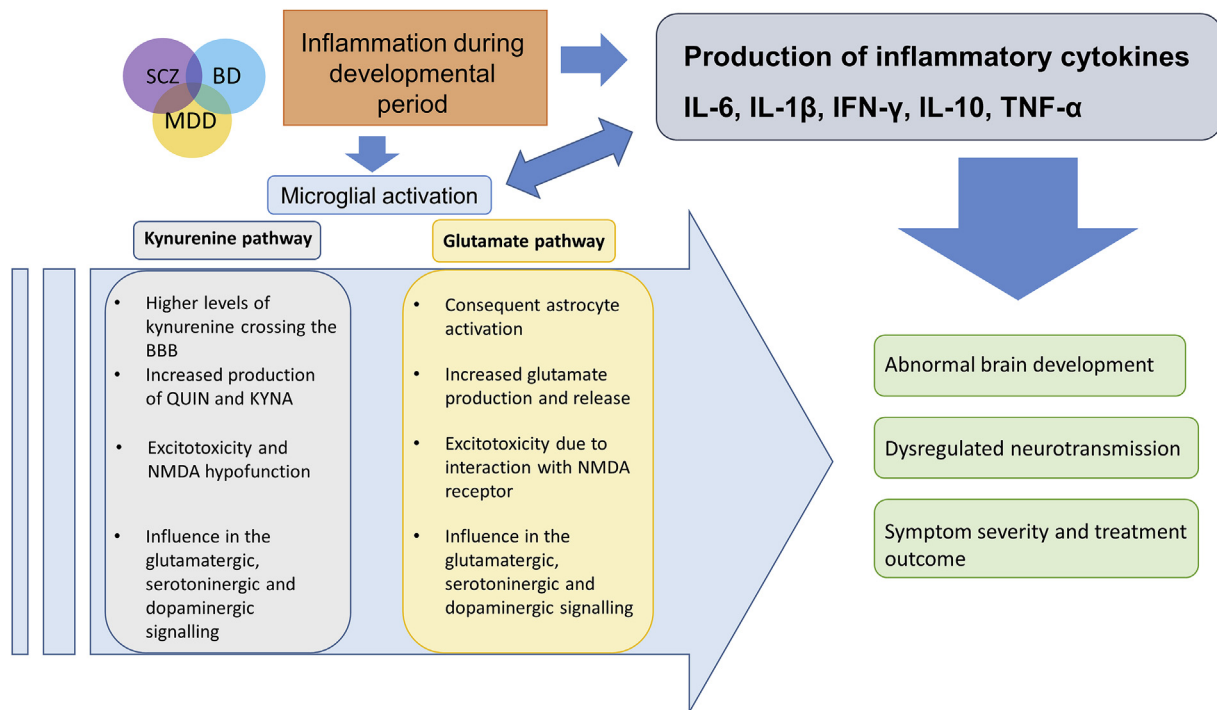


Fig. 2. MDs such as SCZ, BD and MDD, that share symptoms and aspects of their pathophysiology, have been widely associated to immune activation during developmental period. In this case there is an increased production of inflammatory cytokines, both in the periphery and in the CNS, the latter by activated microglia. Microglial activation also results in increased production of the kynurenine pathway neuroactive metabolites QUIN and KYNA, which results in excitotoxicity and NMDA hypofunction. Besides, activated microglia stimulates astrocytic activation, which in turn increases glutamate release with consequent excitotoxicity. Alterations in QUIN, KYNA and glutamate levels can influence the glutamatergic, serotonergic and dopaminergic signaling, and together with the increased inflammatory cytokines, result in abnormal brain development and dysregulation in neurotransmissions, ultimately affecting symptom severity and treatment outcome.

Declaration of competing interest

The authors declare no conflicts of interest.

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References

- Amato, D., Vernon, A.C., Papaleo, F., 2018. Dopamine, the antipsychotic molecule: a perspective on mechanisms underlying antipsychotic response variability. *Neurosci. Biobehav. Rev.* 85, 146–159. <https://doi.org/10.1016/j.neubiorev.2017.09.027>.
- Ashok, A.H., Marques, T.R., Jauhar, S., Nour, M.M., Goodwin, G.M., Young, A.H., Howes, O.D., 2017. The dopamine hypothesis of bipolar affective disorder: the state of the art and implications for treatment. *Mol. Psychiatry* 22 (5), 666–679. <https://doi.org/10.1038/mp.2017.16>.
- Baricello, T., Simoes, L.R., Quevedo, J., Zhang, X.Y., 2019. Microglial activation and psychotic disorders: evidence from pre-clinical and clinical studies. *Curr Top Behav Neurosci.* https://doi.org/10.1007/7854_2018_81 [Epub ahead of print].
- Beaulieu, J.M., Gainetdinov, R.R., 2011. The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacol. Rev.* 63 (1), 182–217. <https://doi.org/10.1124/pr.110.002642>.
- Bergdolt, L., Dunaevsky, A., 2019. Brain changes in a maternal immune activation model of neurodevelopmental brain disorders. *Prog. Neurobiol.* 175, 1–19. <https://doi.org/10.1016/j.pneurobio.2018.12.002>.
- Blacketer, C.J., Lewis, C.P., Frye, M.A., Veldic, M., 2017. Metabotropic glutamate receptors as emerging research targets in bipolar disorder. *Psychiatry Res.* 257, 327–337. <https://doi.org/10.1016/j.psychres.2017.07.059>.
- Boche, D., Perry, V.H., Nicoll, J.A., 2013. Review: activation patterns of microglia and their identification in the human brain. *Neuropathol. Appl. Neurobiol.* 39 (1), 3–18. <https://doi.org/10.1111/nan.12011>.
- Campbell, B.M., Charych, E., Lee, A.W., Möller, T., 2014. Kynurenines in CNS disease: regulation by inflammatory cytokines. *Front. Neurosci.* 8, 12. <https://doi.org/10.3389/fnins.2014.00012>.
- Carlsson, A., Lindqvist, M., 1963. Effect of Chlorpromazine or Haloperidol on formation of 3 methoxyxyramine and normetanephrine in mouse brain. *Acta Pharmacol. Toxicol.* 20, 140–144.
- Carson, M.J., Doose, J.M., Melchior, B., Schmid, C.D., Ploix, C.C., 2006. CNS immune privilege: hiding in plain sight. *Immunol. Rev.* 213, 48–65.
- Carvalho, M.S., Yonamine, C.M., Dal Mas, C., Nunes, D.F.S., Hayashi, M.A.F., 2017. Metabolismo do triptofano em transtornos mentais: um enfoque na esquizofrenia, 29 (2), 44–56. <https://doi.org/10.14295/vitalle.v29i2.6550>.
- Caspi, A., Moffitt, T.E., 2006. Gene-environment interactions in psychiatry: joining forces with neuroscience. *Nat. Rev. Neurosci.* 7 (7), 583–590. <https://doi.org/10.1038/nrn1925>.
- Chávez-Castillo, M., Núñez, V., Nava, M., Ortega, Á., Rojas, M., Bermúdez, V., Rojas-Quintero, J., 2019. Depression as a neuroendocrine disorder: emerging neuropsychopharmacological approaches beyond monoamines. *Adv Pharmacol Sci* 2019, 7943481. <https://doi.org/10.1155/2019/7943481>.
- Chen, Y., Guillemin, G.J., 2009. Kynurenine pathway metabolites in humans: disease and healthy states. *Int. J. Tryptophan Res.* 2, 1–19.
- Chew, L.J., Fusar-Poli, P., Schmitz, T., 2013. Oligodendroglial alterations and the role of microglia in white matter injury: relevance to schizophrenia. *Dev. Neurosci.* 35 (2–3), 102–129. <https://doi.org/10.1159/000346157>.
- Clark, S.M., Notarangelo, F.M., Li, X., Chen, S., Schwarz, R., Tonelli, L.H., 2019. Maternal immune activation in rats blunts brain cytokine and kynurenine pathway responses to a second immune challenge in early adulthood. *Prog Neuropsychopharmacol Biol Psychiatry* 89, 286–294. <https://doi.org/10.1016/j.pnpbp.2018.09.011>.
- DiSabato, D.J., Quan, N., Godbout, J.P., 2016. Neuroinflammation: the devil is in the details. *J. Neurochem.* 139 (2), 136–153. <https://doi.org/10.1111/jnc.13607>.
- Erhardt, S., Schwieler, L., Imbeault, S., Engberg, G., 2017. The kynurenine pathway in schizophrenia and bipolar disorder. *Neuropharmacology* 112 (Pt B), 297–306. <https://doi.org/10.1016/j.neuropharm.2016.05.020>.
- Felger, J.C., 2017. The role of dopamine in inflammation-associated depression: mechanisms and therapeutic implications. *Curr Top Behav Neurosci* 31, 199–219. https://doi.org/10.1007/7854_2016_13.
- Felger, J.C., Treadway, M.T., 2017. Inflammation effects on motivation and motor activity: role of dopamine. *Neuropsychopharmacology* 42 (1), 216–241. <https://doi.org/10.1038/npp.2016.143>.
- Giménez-Gómez, P., Pérez-Hernández, M., Gutiérrez-López, M.D., Vidal, R., Abuin-Martínez, C., O’Shea, E., Colado, M.I., 2018. Increasing kynurenine brain levels reduces ethanol consumption in mice by inhibiting dopamine release in nucleus accumbens. *Neuropharmacology* 135, 581–591. <https://doi.org/10.1016/j.neuropharm.2018.04.016>.
- Gos, T., Myint, A.M., Schiltz, K., Meyer-Lotz, G., Dobrowolny, H., Busse, S., Müller, U.J., Mawrin, C., Bernstein, H.G., Bogerts, B., Steiner, J., 2014. Reduced microglial immunoreactivity for endogenous NMDA receptor agonist quinolinic acid in the hippocampus of schizophrenia patients. *Brain Behav. Immun.* 41, 59–64. <https://doi.org/10.1016/j.bbi.2014.05.012>.
- Grande, I., Berk, M., Birmaher, B., Vieta, E., 2016. Bipolar disorder. *Lancet* 387 (10027), 1561–1572. [https://doi.org/10.1016/S0140-6736\(15\)00241-X](https://doi.org/10.1016/S0140-6736(15)00241-X).
- Gundersen, V., Storm-Mathisen, J., Bergersen, L.H., 2015. Neuroglial transmission. *Physiol. Rev.* 95 (3), 695–726. <https://doi.org/10.1152/physrev.00024.2014>.
- Haroony, E., Chen, X., Li, Z., Patel, T., Woolwine, B.J., Hu, X.P., Felger, J.C., Miller, A.H., 2018. Increased inflammation and brain glutamate define a subtype of depression with decreased regional homogeneity, impaired network integrity, and anhedonia. *Transl. Psychiatry* 8 (1), 189. <https://doi.org/10.1038/s41398-018-0241-4>.
- Haroony, E., Miller, A.H., 2017. Inflammation Effects on glutamate as a pathway to neuroprogression in mood disorders. *Mod Trends Pharmacopsychiatry* 31, 37–55. <https://doi.org/10.1159/000470805>.
- Haroony, E., Miller, A.H., Sanacora, G., 2017. Inflammation, glutamate, and glia: a trio of trouble in mood disorders. *Neuropsychopharmacology* 42 (1), 193–215. <https://doi.org/10.1038/npp.2016.199>.
- Harrison, N.A., Cercignani, M., Voon, V., Critchley, H.D., 2015. Effects of inflammation on hippocampus and substantia nigra responses to novelty in healthy human participants. *Neuropsychopharmacology* 40 (4), 831–838. <https://doi.org/10.1038/npp.2014.222>.
- Harrison, P.J., Weinberger, D.R., 2005. Schizophrenia genes, gene expression, and neuropathology: on the matter of their convergence. *Mol. Psychiatry* 10 (1), 40–68. <https://doi.org/10.1038/sj.mp.4001558>.
- van Heesch, F., Prins, J., Korte-Bouws, G.A., Westphal, K.G., Lemstra, S., Olivier, B., Kraneveld, A.D., Korte, S.M., 2013. Systemic tumor necrosis factor-alpha decreases brain stimulation reward and increases metabolites of serotonin and dopamine in the nucleus accumbens of mice. *Behav. Brain Res.* 253, 191–195. <https://doi.org/10.1016/j.bbr.2013.07.038>.
- Hooshmand, F., Miller, S., Dore, J., Wang, P.W., Hill, S.J., Portillo, N., Ketter, T.A., 2014. Trends in pharmacotherapy in patients referred to a bipolar specialty clinic, 2000–2011. *J. Affect. Disord.* 155, 283–287. <https://doi.org/10.1016/j.jad.2013.10.054>.
- Jablensky, A., McNeil, T.F., Morgan, V.A., 2017. Barbara Fish and a short history of the neurodevelopmental hypothesis of schizophrenia. *Schizophr. Bull.* 43 (6), 1158–1163. <https://doi.org/10.1093/schbul/sbx094>.
- Jeon, Y.W., Han, S.I., Park, E.J., 2018. The relation between immunologic variables and symptom factors in patients with major depressive disorder. *Ann. Gen. Psychiatr.* 17, 32. <https://doi.org/10.1186/s12991-018-0201-7>.
- Kantrowitz, J., Javitt, D.C., 2012. Glutamateergic transmission in schizophrenia: from basic research to clinical practice. *Curr. Opin. Psychiatr.* 25 (2), 96–102. <https://doi.org/10.1097/YCO.0b013e32835035b2>.
- Kawasaki, T., Kawai, T., 2014. Toll-like receptor signaling pathways. *Front. Immunol.* 5, 461. <https://doi.org/10.3389/fimmu.2014.00461>.
- Kegel, M.E., Bhat, M., Skogh, E., Samuelsson, M., Lundberg, K., Dahl, M.L., Sellgren, C., Schwieler, L., Engberg, G., Schuppe-Koistinen, I., Erhardt, S., 2014. Imbalanced kynurenine pathway in schizophrenia. *Int. J. Tryptophan Res.* 7, 15–22. <https://doi.org/10.4137/IJTR.S16800>.
- Kennedy, P.J., Cryan, J.F., Dinan, T.G., Clarke, G., 2017. Kynurenine pathway metabolism and the microbiota-gut-brain axis. *Neuropharmacology* 112 (Pt B), 399–412. <https://doi.org/10.1016/j.neuropharm.2016.07.002>.
- Kim, Y.K., Jeon, S.W., 2018. Neuroinflammation and the immune-kynurenine pathway in anxiety disorders. *Curr. Neuropharmacol.* 16 (5), 574–582. <https://doi.org/10.2174/1570159X15666170913110426>.
- Kindler, J., Lim, C.K., Weickert, C.S., Boerigter, D., Galletly, C., Liu, D., Jacobs, K.R., Balzan, R., Bruggemann, J., O’Donnell, M., Lenroot, R., Guillemin, G.J., Weickert, T.W., 2019. Dysregulation of kynurenine metabolism is related to proinflammatory cytokines, attention, and prefrontal cortex volume in schizophrenia. *Mol. Psychiatry.* <https://doi.org/10.1038/s41380-019-0401-9> [Epub ahead of print].
- Kjaerby, C., Hovelsø, N., Dalby, N.O., Sotty, F., 2017. Phencyclidine administration during neurodevelopment alters network activity in prefrontal cortex and hippocampus in adult rats. *J. Neurophysiol.* 118 (2), 1002–1011. <https://doi.org/10.1152/jn.00081.2017>.
- Krystal, J.H., Perry Jr., E.B., Gueorguieva, R., Belger, A., Madonick, S.H., Abi-Dargham, A., Cooper, T.B., Macdougall, L., Abi-Saab, W., D’Souza, D.C., 2005. Comparative and interactive human psychopharmacological effects of ketamine and amphetamine: implications for glutamatergic and dopaminergic model psychoses and cognitive function. *Arch. Gen. Psychiatr.* 62 (9), 985–994. <https://doi.org/10.1001/archpsyc.62.9.985>.
- Kubo, H., Nakataki, M., Sumitani, S., Iga, J.I., Numata, S., Kameoka, N., Watanabe, S.Y., Umehara, H., Kinoshita, M., Inoshita, M., Tamaru, M., Ohta, M., Nakayama-Yamauchi, C., Funakoshi, Y., Harada, M., Ohmori, T., 2017. ¹H-magnetic resonance spectroscopy study of glutamate-related abnormality in bipolar disorder. *J. Affect. Disord.* 208, 139–144. <https://doi.org/10.1016/j.jad.2016.08.046>.
- Kumar, V., 2019. Toll-like receptors in the pathogenesis of neuroinflammation. *J. Neuroimmunol.* 332, 16–30. <https://doi.org/10.1016/j.jneuroim.2019.03.012>.
- Kyu, H.H., Abate, D., Abate, K.H., Abay, S.M., Abbafati, C., Abbasi, N., Abbastabar, H., Abd-Allah, F., Abdela, J., Abdelalim, A., Abdollahpour, I., 2018. Global, regional, and national disability-adjusted life-years (DALYs) for 359 diseases and injuries and healthy life expectancy (HALE) for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *The Lancet* 392 (10159), 1859–1922.
- Lanz, T.A., Reinhart, V., Sheehan, M.J., Rizzo, S.J.S., Bove, S.E., James, L.C., Volfson, D., Lewis, D.A., Kleiman, R.J., 2019. Postmortem transcriptional profiling reveals widespread increase in inflammation in schizophrenia: a comparison of prefrontal cortex, striatum, and hippocampus among matched tetrads of controls with subjects diagnosed with schizophrenia, bipolar or major depressive disorder. *Transl. Psychiatry* 9 (1), 151. <https://doi.org/10.1038/s41398-019-0492-8>.
- Le-Niculescu, H., Patel, S.D., Bhat, M., Kuczenski, R., Faraone, S.V., Tsuang, M.T., McMahon, F.J., Schork, N.J., Nurnberger Jr., J.I., Niculescu 3rd, A.B., 2009.

- Convergent functional genomics of genome-wide association data for bipolar disorder: comprehensive identification of candidate genes, pathways and mechanisms. *Am J Med Genet B Neuropsychiatr Genet* 150B (2), 155–181. <https://doi.org/10.1002/ajmg.b.30887>.
- Lee, G., Zhou, Y., 2019. NMDAR Hypofunction animal models of schizophrenia. *Front. Mol. Neurosci.* 12, 185. <https://doi.org/10.3389/fnmol.2019.00185>.
- Leonard, B.E., 2018. Inflammation and depression: a causal or coincidental link to the pathophysiology? *Acta Neuropsychiatr.* 30 (1), 1–16. <https://doi.org/10.1017/neu.2016.69>.
- Limanagi, F., Biagioni, F., Gaglione, A., Busceti, C.L., Fornai, F., 2019. A sentinel in the crosstalk between the nervous and immune system: the (immuno)-proteasome. *Front. Immunol.* 10, 628. <https://doi.org/10.3389/fimmu.2019.00628>.
- Luo, Q., Chen, Q., Wang, W., Desrivieres, S., Quinlan, E.B., Jia, T., Macare, C., Robert, G.H., Cui, J., Guedj, M., Palaniyappan, L., Kherif, F., Banaschewski, T., Bokke, A.L.W., Büchel, C., Flor, H., Froin, V., Garavan, H., Gowland, P., Heinz, A., Ittermann, B., Martinot, J.L., Artiges, E., Paillière-Martinot, M.L., Nees, F., Orfanos, D.P., Poustka, L., Fröhner, J.H., Smolka, M.N., Walter, H., Whelan, R., Callicott, J.H., Mattay, V.S., Pausova, Z., Dargatzis, J.F., Tzourio, C., Crivello, F., Berman, K.F., Li, F., Paus, T., Weinberger, D.R., Murray, R.M., Schumann, G., Feng, J., 2019. IMAGEN consortium. Association of a schizophrenia-risk nonsynonymous variant with putamen volume in adolescents: a voxelwise and genome-wide association study. *JAMA Psychiatry* 76 (4), 435–445. <https://doi.org/10.1001/jamapsychiatry.2018.4126>.
- Lurie, D.I., 2018. An integrative approach to neuroinflammation in psychiatric disorders and neuropathic pain. *J. Exp. Neurosci.* 12 <https://doi.org/10.1177/1179069518793639>, 1179069518793639.
- Malhi, G.S., Tanious, M., Das, P., Coulston, C.M., Berk, M., 2013. Potential mechanisms of action of lithium in bipolar disorder. *Curr. understanding. CNS Drugs* 27 (2), 135–153. <https://doi.org/10.1007/s40263-013-0039-0>.
- McCutcheon, R.A., Abi-Dargham, A., Howes, O.D., 2019. Schizophrenia, dopamine and the striatum: from biology to symptoms. *Trends Neurosci.* 42 (3), 205–220. <https://doi.org/10.1016/j.tins.2018.12.004>.
- McEntee, W.J., Crook, T.H., 1993. Glutamate: its role in learning, memory, and the aging brain. *Psychopharmacology (Berlin)* 111 (4), 391–401. <https://doi.org/10.1007/bf02253527>.
- Miller, B.J., Buckley, P., Seabolt, W., Mellor, A., Kirkpatrick, B., 2011. Meta-analysis of cytokine alterations in schizophrenia: clinical status and antipsychotic effects. *Biol. Psychiatry* 70 (7), 663–671. <https://doi.org/10.1016/j.biopsych.2011.04.013>.
- Money, K.M., Stanwood, G.D., 2013. Developmental origins of brain disorders: roles for dopamine. *Front. Cell. Neurosci.* 7, 260. <https://doi.org/10.3389/fncel.2013.00260>.
- Mongan, D., Ramesar, M., Föcking, M., Cannon, M., Cotter, D., 2019. Role of inflammation in the pathogenesis of schizophrenia: a review of the evidence, proposed mechanisms and implications for treatment. *Early Interv Psychiatry*. <https://doi.org/10.1111/eip.12859> [Epub ahead of print].
- Moretto, E., Murru, L., Martano, G., Sassone, J., Passafaro, M., 2018. Glutamatergic synapses in neurodevelopmental disorders. *Prog Neuropsychopharmacol Biol Psychiatry* 84 (Pt B), 328–342. <https://doi.org/10.1016/j.pnpbp.2017.09.014>.
- Mosley, R.L., 2015. Adaptive immunity in neurodegenerative and neuropsychological disorders. *J. Neuroimmune Pharmacol.* 10 (4), 522–527. <https://doi.org/10.1007/s11481-015-9640-y>.
- Mosser, D.M., Edwards, J.P., 2008. Exploring the full spectrum of macrophage activation. *Nat. Rev. Immunol.* 8 (12), 958–969. <https://doi.org/10.1038/nri2448.Review>. Erratum in: *Nat Rev Immunol.*2010;10(6):460.
- Murray, R.M., Bhavsar, V., Tripoli, G., et al., 2017. 30 years on: how the neurodevelopmental hypothesis of schizophrenia morphed into the developmental risk factor model of psychosis. *Schizophr. Bull.* 43 (6), 1190–1196. <https://doi.org/10.1093/schbul/sbx121>.
- Murray, P.J., Wynn, T.A., 2011. Protective and pathogenic functions of macrophage subsets. *Nat. Rev. Immunol.* 11 (11), 723–737. <https://doi.org/10.1038/nri3073>.
- Murrough, J.W., Abdallah, C.G., Mathew, S.J., 2017. Targeting glutamate signalling in depression: progress and prospects. *Nat. Rev. Drug Discov.* 16 (7), 472–486. <https://doi.org/10.1038/nrd.2017.16>.
- Niu, Z., Yang, L., Wu, X., Zhu, Y., Chen, J., Fang, Y., 2019. The relationship between neuroimmunity and bipolar disorder: mechanism and translational application. *Neurosci Bull* 35 (4), 595–607. <https://doi.org/10.1007/s12264-019-00403-7>.
- Notarangelo, F.M., Pociavsek, A., 2017. Elevated kynurenine pathway metabolism during neurodevelopment: implications for brain and behavior. *Neuropharmacology* 112 (Pt B), 275–285. <https://doi.org/10.1016/j.neuropharm.2016.03.001>.
- Nurnberger Jr., J.L., Koller, D.L., Jung, J., Edenberg, H.J., Foroud, T., Guella, I., Vawter, M.P., Kelsoe, J.R., 2014. Identification of pathways for bipolar disorder: a meta-analysis. *JAMA Psychiatry* 71 (6), 657–664. <https://doi.org/10.1001/jamapsychiatry.2014.176>.
- Oo, Y.H., Shetty, S., Adams, D.H., 2010. The role of chemokines in the recruitment of lymphocytes to the liver. *Dig. Dis.* 28 (1), 31–44. <https://doi.org/10.1159/000282062>.
- Oxenkrug, G., van der Hart, M., Roeser, J., Summergrad, P., 2016. Anthranilic Acid: a potential biomarker and treatment target for schizophrenia. *Ann Psychiatry Ment Health* 4 (2), 1059.
- Pitsillou, E., Bresnehan, S.M., Kagarakis, E.A., Wijoyo, S.J., Liang, J., Hung, A., Karagiannis, T.C., 2019. The cellular and molecular basis of major depressive disorder: towards a unified model for understanding clinical depression. *Mol. Biol. Rep.* <https://doi.org/10.1007/s11033-019-05129-3> [Epub ahead of print].
- Purves-Tyson, T.D., Weber-Stadlbauer, U., Ricketto, J., Rothmond, D.A., Labouesse, M.A., Polese, M., Robinson, K., Shannon Weickert, C., Meyer, U., 2019. Increased levels of midbrain immune-related transcripts in schizophrenia and in murine offspring after maternal immune activation. *Mol. Psychiatry*. <https://doi.org/10.1038/s41380-019-0434-0> [Epub ahead of print].
- Réfsgaard, L.K., Pickering, D.S., Andreasen, J.T., 2017. Investigation of antidepressant-like and anxiolytic-like actions and cognitive and motor side effects of four N-methyl-D-aspartate receptor antagonists in mice. *Behav. Pharmacol.* 28 (1), 37–47. <https://doi.org/10.1097/FBP.0000000000000266>.
- Ribeiro, F.M., Vieira, L.B., Pires, R.G., Olmo, R.P., Ferguson, S.S., 2017. Metabotropic glutamate receptors and neurodegenerative diseases. *Pharmacol. Res.* 115, 179–191. <https://doi.org/10.1016/j.phrs.2016.11.013>.
- Ruddick, J.P., Evans, A.K., Nutt, D.J., Lightman, S.L., Rook, G.A., Lowry, C.A., 2006. Tryptophan metabolism in the central nervous system: medical implications. *Expert Rev. Mol. Med.* 8 (20), 1–27. <https://doi.org/10.1017/S1462399406000068>.
- Salter, M.W., Beggs, S., 2014. Sublime microglia: expanding roles for the guardians of the CNS. *Cell* 158 (1), 15–24. <https://doi.org/10.1016/j.cell.2014.06.008>.
- Sanacora, G., Zarate, C.A., Krystal, J.H., Manji, H.K., 2008. Targeting the glutamatergic system to develop novel, improved therapeutics for mood disorders. *Nat. Rev. Drug Discov.* 7 (5), 426–437. <https://doi.org/10.1038/nrd2462>.
- Sathyaikumar, K.V., Stachowski, E.K., Wonodi, I., Roberts, R.C., Rassoulpour, A., McMahon, R.P., Schwarcz, R., 2011. Impaired kynurenine pathway metabolism in the prefrontal cortex of individuals with schizophrenia. *Schizophr. Bull.* 37 (6), 1147–1156. <https://doi.org/10.1093/schbul/sbq112>.
- Schwarcz, R., Stone, T.W., 2017. The kynurenine pathway and the brain: challenges, controversies and promises. *Neuropharmacology* 112 (Pt B), 237–247. <https://doi.org/10.1016/j.neuropharm.2016.08.003>.
- Schwarcz, R., Bruno, J.P., Muchowski, P.J., Wu, H.Q., 2012. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat. Rev. Neurosci.* 13 (7), 465–477. <https://doi.org/10.1038/nrn3257>.
- Schwartz, T.L., Sachdeva, S., Stahl, S.M., 2012. Glutamate neurocircuitry: theoretical underpinnings in schizophrenia. *Front. Pharmacol.* 3, 195. <https://doi.org/10.3389/fphar.2012.00195>.
- Schwieler, L., Larsson, M.K., Skogh, E., Kegel, M.E., Orhan, F., Abdelmoaty, S., Finn, A., Bhat, M., Samuelsson, M., Lundberg, K., Dahl, M.L., Sellgren, C., Schuppe-Koistinen, I., Svensson, C., Erhardt, S., Engberg, G., 2015. Increased levels of IL-6 in the cerebrospinal fluid of patients with chronic schizophrenia—significance for activation of the kynurenine pathway. *J. Psychiatry Neurosci.* 40 (2), 126–133. <https://doi.org/10.1503/jpn.140126>.
- Sidor, M.M., Spencer, S.M., Dzirasa, K., Parekh, P.K., Tye, K.M., Warden, M.R., Arey, R.N., Enwright 3rd, J.F., Jacobsen, J.P., Kumar, S., Remillard, E.M., Caron, M.G., Deisseroth, K., McClung, C.A., 2015. Daytime spikes in dopaminergic activity drive rapid mood-cycling in mice. *Mol. Psychiatry* 20 (11), 1406–1419. <https://doi.org/10.1038/mp.2014.167>.
- Simpson, E.H., Winiger, V., Biezonski, D.K., Haq, I., Kandel, E.R., Kellendonk, C., 2014. Selective overexpression of dopamine D3 receptors in the striatum disrupts motivation but not cognition. *Biol. Psychiatry* 76 (10), 823–831. <https://doi.org/10.1016/j.biopsych.2013.11.023>.
- Smaragdi, A., Chavez, S., Lobaugh, N.J., Meyer, J.H., Kolla, N.J., 2019. Differential levels of prefrontal cortex glutamate + glutamine in adults with antisocial personality disorder and bipolar disorder: a proton magnetic resonance spectroscopy study. *Prog Neuropsychopharmacol Biol Psychiatry* 93, 250–255. <https://doi.org/10.1016/j.pnpbp.2019.04.002>.
- Snyder, M.A., Gao, W.J., 2013. NMDA hypofunction as a convergence point for progression and symptoms of schizophrenia. *Front. Cell. Neurosci.* 7, 31. <https://doi.org/10.3389/fncel.2013.00031>.
- Sun, R., Zhao, Z., Feng, J., Bo, J., Rong, H., Lei, Y., Lu, C., Zhang, X., Hou, B., Sun, Y., Liu, Y., Ma, Z., Gu, X., 2017. Glucocorticoid-potentiated spinal microglia activation contributes to preoperative anxiety-induced postoperative hyperalgesia. *Mol. Neurobiol.* 54 (6), 4316–4328. <https://doi.org/10.1007/s12035-016-9976-1>.
- Taniuchi, I., 2018. CD4 helper and CD8 cytotoxic T cell differentiation. *Annu. Rev. Immunol.* 36, 579–601. <https://doi.org/10.1146/annurev-immunol-042617-053411>.
- Tasker, J.G., Oliet, S.H., Bains, J.S., Brown, C.H., Stern, J.E., 2012. Glial regulation of neuronal function: from synapse to systems physiology. *J. Neuroendocrinol.* 24 (4), 566–576. <https://doi.org/10.1111/j.1365-2826.2011.02259.x>.
- Taylor, R.W., Marwood, L., Greer, B., Strawbridge, R., Cleare, A.J., 2019. Predictors of response to augmentation treatment in patients with treatment-resistant depression: a systematic review. *J. Psychopharmacol.* 33 (11), 1323–1339. <https://doi.org/10.1177/0269881119872194>.
- Turner, M.D., Nedjai, B., Hurst, T., Pennington, D.J., 2014. Cytokines and chemokines: at the crossroads of cell signalling and inflammatory disease. *Biochim. Biophys. Acta* 1843 (11), 2563–2582. <https://doi.org/10.1016/j.bbamcr.2014.05.014>.
- Tye, K.M., Mirzabekov, J.J., Warden, M.R., Ferenczi, E.A., Tsai, H.C., Finkelstein, J., Kim, S.Y., Adhikari, A., Thompson, K.R., Andalman, A.S., Gunaydin, L.A., Witten, I.B., Deisseroth, 2013. Dopamine neurons modulate neural encoding and expression of depression-related behaviour. *Nature* 493 (7433), 537–541. <https://doi.org/10.1038/nature11740>.
- Vancassel, S., Capuron, L., Castanon, N., 2018. Brain kynurenine and BH4 pathways: relevance to the pathophysiology and treatment of inflammation-driven depressive symptoms. *Front. Neurosci.* 12, 499. <https://doi.org/10.3389/fnins.2018.00499>.
- Waters, K.A., Machaalani, R., 2004. NMDA receptors in the developing brain and effects of noxious insults. *Neurosignals* 13 (4), 162–174. <https://doi.org/10.1159/000077523>.
- Weber-Stadlbauer, U., 2017. Epigenetic and transgenerational mechanisms in infection-mediated neurodevelopmental disorders. *Transl. Psychiatry* 7 (5), e1113. <https://doi.org/10.1038/tp.2017.78>.
- Williams, N.R., Schatzberg, A.F., 2016. NMDA antagonist treatment of depression. *Curr. Opin. Neurobiol.* 36, 112–117. <https://doi.org/10.1016/j.conb.2015.11.001>.

- Wolf, S.A., Boddeke, H.W., Kettenmann, H., 2017. Microglia in physiology and disease. *Annu. Rev. Physiol.* 79, 619–643. <https://doi.org/10.1146/annurev-physiol-022516-034406>.
- Zarate Jr., C.A., Brutsche, N.E., Ibrahim, L., Franco-Chaves, J., Diazgranados, N., Cravchik, A., Selter, J., Marquardt, C.A., Liberty, V., Luckenbaugh, D.A., 2012. Replication of ketamine's antidepressant efficacy in bipolar depression: a randomized controlled add-on trial. *Biol. Psychiatry* 71 (11), 939–946. <https://doi.org/10.1016/j.biopsych.2011.12.010>.
- Zhang, X., Bearer, E.L., Boulat, B., Hall, F.S., Uhl, G.R., Jacobs, R.E., 2010. Altered neurocircuitry in the dopamine transporter knockout mouse brain. *PLoS One* 5 (7), e11506. <https://doi.org/10.1371/journal.pone.0011506>.
- Zhou, Y., Danbolt, N.C., 2014. Glutamate as a neurotransmitter in the healthy brain. *J. Neural Transm.* 121 (8), 799–817. <https://doi.org/10.1007/s00702-014-1180-8>.
- Zhu, Y., Klomprens, E.A., Guo, S., Geng, X., 2019. Neuroinflammation caused by mental stress: the effect of chronic restraint stress and acute repeated social defeat stress in mice. *Neurol. Res.* 16, 1–8. <https://doi.org/10.1080/01616412.2019.1615670>.
- Zwicker, A., Denovan-Wright, E.M., Uher, R., 2018. Gene-environment interplay in the etiology of psychosis. *Psychol. Med.* 48 (12), 1925–1936. <https://doi.org/10.1017/S003329171700383X>.