



## The Key Role of TNF-TNFR2 Interactions in the Modulation of Allergic Inflammation: A Review

Suhana Ahmad<sup>1</sup>, Nor Azrini Azid<sup>1</sup>, Jennifer C. Boer<sup>2</sup>, JitKang Lim<sup>3</sup>, Xin Chen<sup>4</sup>, Magdalena Plebanski<sup>5</sup> and Rohimah Mohamud<sup>1,6\*</sup>

<sup>1</sup> Department of Immunology, School of Medical Sciences, Universiti Sains Malaysia, Kelantan, Malaysia, <sup>2</sup> Department of Immunology and Pathology, Monash University, Melbourne, VIC, Australia, <sup>3</sup> School of Chemical Engineering, Universiti Sains Malaysia, Pulau Pinang, Malaysia, <sup>4</sup> State Key Laboratory of Quality Research in Chinese Medicine, Institute of Chinese Medical Sciences, University of Macau, Taipa, China, <sup>5</sup> School of Health and Biomedical Sciences, RMIT, Melbourne, VIC, Australia, <sup>6</sup> Hospital Universiti Sains Malaysia, Universiti Sains Malaysia, Kelantan, Malaysia

Tumor necrosis factor-alpha (TNF) is a pleiotropic cytokine, which is thought to play a major role in the pathogenesis of inflammatory diseases, including allergy. TNF is produced at the early stage of allergen sensitization, and then continues to promote the inflammation cascade in the effector phase of allergic reactions. Consequently, anti-TNF treatment has been proposed as a potential therapeutic option. However, recent studies reveal anti-intuitive effects of TNF in the activation and proliferative expansion of immunosuppressive Tregs, tolerogenic DCs and MDSCs. This immunosuppressive effect of TNF is mediated by TNFR2, which is preferentially expressed by immunosuppressive cells. These findings redefine the role of TNF in allergic reaction, and suggest that targeting TNF-TNFR2 interaction itself may represent a novel strategy in the treatment of allergy.

### HIGHLIGHTS

- Pleiotropic function of TNF in allergy is likely mediated by its two receptors, TNFR1 and TNFR2
- Activation by TNFR1 results in the allergic inflammatory responses while TNFR2 plays a role in the immune tolerance to allergens
- TNFR2 is preferentially expressed by highly suppressive and replicating Tregs and TNFR2 signaling leads to the activation and proliferation of Tregs
- Targeting of TNFR2 to boost Treg activity may represent a novel strategy for treating patients with allergy.

Keywords: allergy, TNF, TNFR2, regulatory T cells, tolerogenic dendritic cells

## INTRODUCTION

Allergy is an immune-mediated hypersensitivity to allergens. Exposure of allergens through inhalation, ingestion or skin contact leads to diseases such as asthma, allergic rhinitis, food allergy, and atopic dermatitis. Allergy is a complex disease, and both genetic and environmental factors contribute to its pathogenesis. It affects 30–35% of the population at some point in their life,

### OPEN ACCESS

### Edited by:

Luis Graca, Universidade de Lisboa, Portugal

#### Reviewed by:

Olga Ucar, Independent Researcher, Manchester, United Kingdom Marie-Claude Gaudreau, Bristol Myers Squibb, United States

> \*Correspondence: Rohimah Mohamud rohimahm@usm.my

#### Specialty section:

This article was submitted to Immunological Tolerance and Regulation, a section of the journal Frontiers in Immunology

Received: 06 July 2018 Accepted: 18 October 2018 Published: 09 November 2018

#### Citation:

Ahmad S, Azid NA, Boer JC, Lim JK, Chen X, Plebanski M and Mohamud R (2018) The Key Role of TNF-TNFR2 Interactions in the Modulation of Allergic Inflammation: A Review. Front. Immunol. 9:2572. doi: 10.3389/fimmu.2018.02572

1

with incidence continuing to rapidly grow each year. The past decades have exhibited large-scale anthropogenic changes which are currently considered the leading causes of the increasing burden of allergic diseases.

## EFFECTOR MECHANISMS IN ALLERGIC REACTIONS

The development of allergy can be divided into two phases: (1) the sensitization and memory phase, and (2) the effector phase, which can be further staged into immediate and late responses (1) (Figure 1). Sensitization occurs upon the first encounter with allergen that leads to the production of proinflammatory cytokines [Interleukin (IL) 33 (IL-33), thymic stromal lymphopoietin (TSLP), tumor necrosis factor (TNF), IL-1 $\beta$ ] by epithelial cells. This allergic sensitization is determined by both genetic polymorphisms (2) and environmental risk factors (3). Studies have identified several risk alleles, including cadherin-related protein 3 (CDHR3) and protocadherin 1 (PCDH1), which are both thought to be involved in facilitating allergic sensitization (4). The ubiquitous presence of lipopolysaccharide (LPS) in the environment may also contribute to the exacerbation of allergic responses. Exposure to LPS triggers signaling of toll-like receptor 4 (TLR4) on epithelial cells and can either promote or suppress the sensitization to an allergen in a dose-dependent manner (5, 6). Allergy is primarily a T helper 2 (Th2)-driven disease (7) in which dendritic cells (DCs) stimulated with cytokines released by sensitized epithelial cells have the capacity to induce Th2 responses. This cascade of events drives IgE synthesis and promotes the generation of memory allergen-specific T and B cells (8).

The effector phase of allergic responses is initiated when the allergen cross-links IgE-FceRI complexes on sensitized mast cells. Subsequently, mediators of the allergic responses such as histamine, leukotrienes, cytokines, chemokines and proteases, largely responsible for type 1 hypersensitivity are released (9). The release of these mediators causes the acute signs and symptoms of allergy, such as vasodilation and airway constriction. While the continuous exposure to allergen activates T cells and consequently triggers the late phase response, it is the allergen-specific Th2 cells that produce IL-4, IL-5, IL-9, and IL-13, which are responsible for the maintenance of allergenspecific IgE levels and activation of tissue eosinophilia, mucus overproduction, and tissue remodeling (10–13).

Furthermore, Th17 cells were shown to be associated with a more severe asthma, which is less responsive to corticosteroid (14), induces neutrophils recruitment (15) and increases airway inflammation and remodeling (16). In addition to Th1, Th2,

and Th17 cells, other cell types are also involved in allergy. For example, Th9 and Th22 cells have been shown to play a crucial role in both early and chronic allergic inflammation. The Th9 cells are vital for the recruitment and activation of mast cells during early allergic response and their increased number in allergic patients is correlated with elevated IgE levels (17). On the other hand, Th22 cells are increased in children with asthma and atopic dermatitis. These cells act by increasing the recruitment of leukocytes and disrupting the epithelial integrity on the skin and in the lungs (18, 19).

Another subset of CD4T cells, which play an indispensable role in the induction and maintenance of tolerance to allergens, are the immunosuppressive CD4<sup>+</sup>Foxp3<sup>+</sup> regulatory T cells (Tregs) (20). In one study, the CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> Treg numbers were found to be normal but the expression of FoxP3 protein, a critical marker for Tregs (21) was diminished. However, the authors did not investigate whether there are functional consequences related to a reduced FoxP3 expression in these cells (22). In another study, Fontenot et al., found that FoxP3 displayed reduced expression in patients with allergic rhinitis (23). Because in general, patients with FoxP3 mutations exhibit excessive autoimmunity with high levels of IgE, peripheral eosinophilia and Th2 skewing, the reduced FoxP3 expression may be a contributing factor to the development of allergic diseases in humans (24). Furthermore, in addition to Th2 responses, other types of effector T cells (Teff) are also attributable to the pathogenesis of allergy (25). In the stage of allergy sensitization, Th1 responses are inhibited through reduction of IL-12. However, in the later stage of allergic response, Th1 cells have been shown to coexist with Th2 cells. Such Th1 cells can exacerbate the symptoms and lead to severe allergy manifestation (26).

Overall a proper maturity and homeostasis of immune system of neonates during the first years of life is fundamental to minimize allergic development. Maternal allergy correlates well with impaired frequency and function of Tregs in neonates and consequently the increased susceptibility for the development of allergy in early childhood (27–29). A study found that Th2 cells were increased in newborns' cord blood with maternal allergy, accompanied by a decreased Tregs/Th2 ratio, indicative of an increased risk for developing atopic dermatitis (29). Furthermore, prenatal environmental exposure such as smoking and the usage of harsh chemicals like disinfectants were found to be associated with reduced Tregs number in newborns' cord blood, which further increases the risk of development of allergy later in life (28). Therefore, as shown by these and other extensive studies [reviewed by (30)], Tregs play a fundamental role in the pathogenesis of allergy and their modulation may harness great potential for treatment of allergic diseases.

### **GENERAL BIOLOGY OF TNF**

TNF is a pleiotropic cytokine that plays important dual roles in maintaining immune homeostasis and in promoting the development of diseases. TNF is required for host defense against pathogens, immune surveillance against

**Abbreviations:** CDHR3, Cadherin-related protein 3; DCs, Dendritic cells; Interleukin, IL; IL-17RD, Interleukin 17 receptor D; MAPK, Mitogen-activated protein kinases; MDSC, Myeloid-derived suppressor cell; NFκB, Nuclear factor kappa-light-chain-enhancer of activated B cells; NK, Natural killer cells; NO, nitric oxide; PCDH1, Protocadherin 1; RCT, Randomized controlled trial; SIT, Specific immunotherapy; Teff, Effector T cells; Th, T helper cells; TNF, Tumor necrosis factor; TNFR, Tumor necrosis factor receptor; TRADD, TNFR-associated death domain protein; TRAF, TNFR-associated factor; Tregs, Regulatory T cells; TSLP, Thymic stromal lymphopoietin.

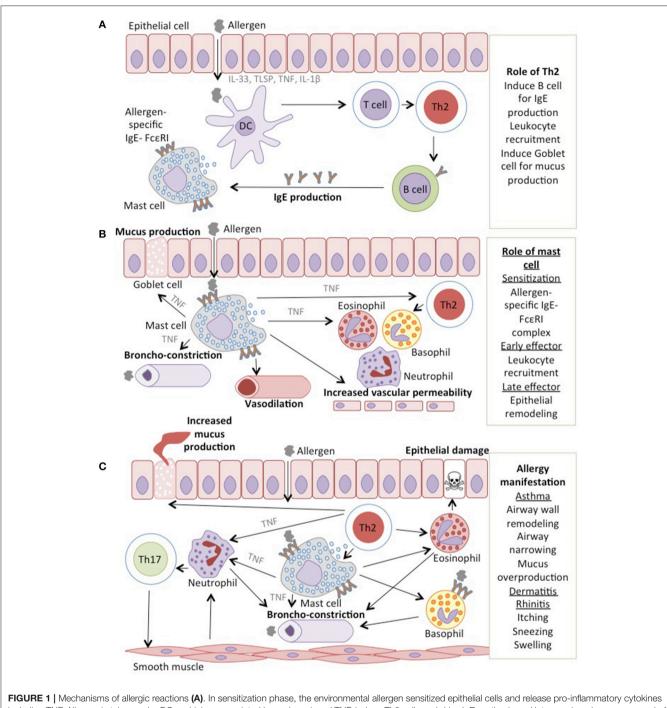


FIGURE 1 [Mechanisms of allergic reactions (A). In sensitization phase, the environmental allergen sensitized epithelial cells and release pro-inflammatory cytokines including TNF. Allergen is taken up by DCs, which are regulated by and produced TNF, induce Th2 cells and drive IgE synthesis and later produced a memory pool of allergen-specific T and B cells (B). Next encounter with the allergen induces assembly with antigen-specific IgE-Fc<sub>2</sub>RI complex on mast cells that secretes TNF and later recruits leukocytes. The leukocytes; basophils, neutrophils, eosinophils and mast cells interact with each other to produce more mucus (C). In the late effector phase, the epithelial remodeling will exhibit the allergic manifestation as in asthmatic and rhinitis patients, the airway wall are narrowed and mucus overproduced while in dermatitis patients, the vasodilation resulted in the itching and swelling of the skin.

malignancies, as well as cell proliferation and survival (31). This cytokine is widely considered as an important inflammatory mediator of several diseases such as autoimmune diseases, cancer, hypernociception, cardiovascular disease and fibrosis (32).

Dual biological functions of TNF are likely to be transduced through its two distinct receptors, TNFR1 and TNFR2. TNF and its receptors have both a membrane-bound form and a soluble form. TNFR1 shows high affinity toward both forms of TNF, while TNFR2 is only fully activated by membrane-bound TNF (33). Once activated, TNF elicits its biological functions through the activation of two major signaling pathways, nuclear factor kappa-light-chain-enhancer of activated B cells (NF $\kappa$ B) and mitogen-activated protein kinases (MAPK). These two signaling pathways mediate various cellular activities including proliferation and survival as well as apoptosis and cell death, depending on which receptor is bound and activated by TNF (32).

## PATHOGENIC ROLE OF TNF IN ALLERGIC MANIFESTATION

TNF is reported to play significant role in the pathogenesis of allergy and contributes to both early and late stages of allergy development. This is evident when allergy manifestations are inhibited in TNF-knockout (KO) mice as well as with anti-TNF treatments (34–37). Furthermore, upon allergen exposure, TNF is produced by sensitized epithelial barriers and immune cells (such as macrophages, mast cells, DCs). It has the capacity to promote Th2 responses (38), resulting in high levels of IL-4, IL-5, and IL-13, which further activate eosinophils, mast cells and basophils (7). In allergic rhinitis, TNF is essential for the recruitment of eosinophils to the site of allergic inflammation through the induction of adhesion molecules (34). In asthma, TNF is shown to synergize with IL-17 by promoting neutrophil recruitment (36), whereas in atopic dermatitis, TNF is responsible for the production of other cytokines, including IL-32 which induces keratinocyte apoptosis (39). Furthermore, TNF, both induced and are secreted by Th2 cells which promote the production of antigen-specific IgE isoforms from B cells (34). A meta-analysis study suggested that TNF polymorphisms were significantly associated with asthma susceptibility (40). Elevated TNF levels in severe allergy have been shown to contribute to the epithelial barrier dysfunction by upregulating the adhesion molecules (p120, E-cadherin) and increasing the endothelial permeability to allergens (41).

# Proinflammatory TNF-TNFR1 Signaling in Allergy

Although binding of TNF to its distinct receptors generally activates the same major signaling pathways (NF $\kappa$ B and MAPK), the distinct structure and motifs of TNFR1 and TNFR2 result in entirely different functional consequences. TNFR1 bears the death domain, which recruits several death signaling proteins such as TNFR1-associated death domain protein (TRADD), Fas associated protein with death domain (FADD) and the TNFR-associated factor (TRAF)-1 to induce inflammation and apoptosis (42).

In the context of apoptosis, TNF-TNFR1 axis signals through caspase 3 and 8, and is mainly responsible for the host defense of pathogen (43) and anti-tumoral activities (44). The impaired regulation of this TNF receptor can cause autoimmune diseases, cancer, chronic infections and allergy (45). In allergic diseases, elevated levels of TNF reportedly trigger inflammatory cascade through TNFR1 (46, 47). For example, Maillet et al. have shown that in a murine model, soluble TNF is a primary driver of

allergic airway inflammation, which can be effectively attenuated by neutralization of soluble TNF (48). Increased expression of TNFR1 is shown to play a crucial role in allergic inflammation through the recruitment of eosinophils, neutrophils and other lymphocytes (47). Surprisingly, one study demonstrates antiapoptotic effects of TNFR1 by enhanced eosinophil survival in asthma, which is contradictory to its known regulation, given the existing cross talk between NFkb and another pathway called c-jun-N-terminal kinase (JNK) pathway (49).

## Immunosuppressive TNF-TNFR2 Signaling in Allergy

Unlike TNFR1, TNFR2 lacks the death domain where the activation of TNFR2 recruits TNFR-associated factor 2 (TRAF2) that mainly promotes cell proliferation and survival (50). In allergy, impaired TNF-TNFR2 signaling promotes the polarization of Th2 and Th17 cells, thus aggravating the allergy manifestations (51). This notion was further supported by a study showing that TNFR2-KO mice displayed an increased eosinophilic inflammation, one of the most common allergic manifestations, in comparison with wild type mice, whereas TNFR1-KO mice had a weaker response (47). Nevertheless, as TNFR2 promotes cell survival, this receptor signaling also protect inflammatory cells in diseases including eosinophils (52), thus maintain disease progression. Furthermore, TNFR2 signaling on natural killer (NK) cells help to induce Th2 sensitization toward inhaled allergen (53). To add, in certain inflammatory conditions, TNFR2 can also induce apoptosis when it cross-talks with TNFR1 (54). In the cross-talk, TNFR2 induces TRAF2 to deplete cIAP1/2, the apoptosis inhibitor, thus accelerating the TNFR1-dependent apoptosis (54). In addition, with Fas and Fas ligand, TNFR2 signaling induces apoptosis in IFN-y-Th1 cells, resulting in a predominance of Th2 in atopic individuals (55). Under certain conditions including prolonged cell stress in disease condition, shift of TNFR2 to TNFR1 apoptotic signaling can occur, leading to opposite known function of TNFR2.

Distinct functions of TNF and its receptor led to the prospective of selectively targeting TNFR1 to inhibit apoptosis, and TNFR2 to induce cell survival. This approach intends to achieve homeostasis in various autoimmune and inflammatory diseases including allergy. These disorders are primarily associated with defects in TNF signaling through TNFR2. Therefore, this particular receptor is of interest and important roles of TNF-TNFR2 interaction on various cell types are further discussed in the next section (**Figure 2, Table 1**).

### **TNF-TNFR2** on Tregs

Due to its important role in allergy manifestations, TNF has been evaluated as a target for therapy and findings have led to the discovery of a more prominent role of TNFR2 in the pathogenesis of allergy. Only a few effects of the exclusive TNF signaling via TNFR2 have been characterized, since its expression is limited to certain cell populations and is only fully activated by membranebound TNF. In T cell biology, TNFR2 is directly associated with proliferation and maintenance of function, both in Tregs (67, 68) and Teff (69, 70). What is more interesting is the restricted

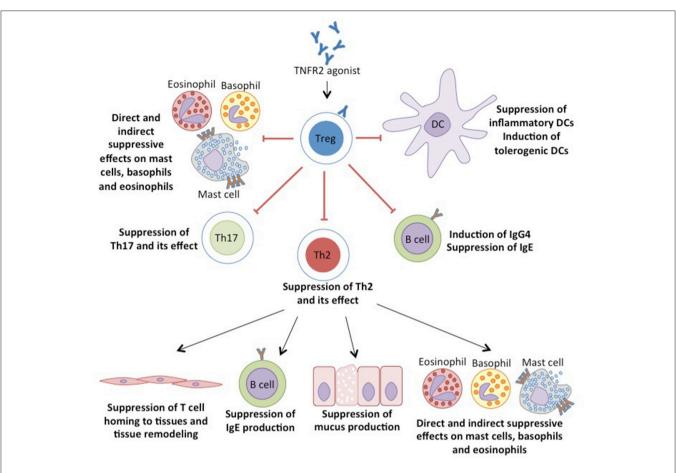


FIGURE 2 | TNFR2 agonist target to enhance the proliferation and suppressive capacity of Tregs. The suppressive Tregs inhibits the inflammation by suppressing the inflammatory cells and induces the tolerogenic DCs by the suppressive MDSCs, hence enabling the suppression of allergic manifestations.

**TABLE 1** | Regulation of TNF on Tregs, DCs and MDSCs for protective immune response against allergic reactions.

Target cells	Protective mechanisms	References	
Tregs	Increased suppressive capacity	(56, 57)	
	Increased proliferative response	(58, 59)	
	Inhibited excessive Th2 and Th17 polarization via inhibition on NF-kB signaling	(51)	
DCs	Regulate maturation and survival	(60, 61)	
	Tolerogenic DCs	(62–64)	
MDSCs	Inhibit activation of Th2 cells	(65, 66)	

abundance of TNFR2 on Tregs, leading to strong activity of this receptor on the mediator of tolerance.

TNFR2, which is preferentially expressed on human and mouse Tregs, is associated with both phenotypic and functional properties of Tregs (67). Tregs are shown to inhibit inflammatory responses by TNF through TNFR2 signaling (71). In addition, in diseases, downregulation of Foxp3 has been associated with TNF- TNFR2 interaction and is later restored by blockade of TNF with TNF antagonist (59, 72, 73). Unlike Foxp3 of which the forced expression can convert CD4<sup>+</sup>CD25<sup>-</sup> Teff into functional Tregs (74), TNFR2, does not induce CD4<sup>+</sup>Foxp3<sup>-</sup> T cells to become suppressive (70). Instead, TNFR2 expressing CD4<sup>+</sup>Foxp3<sup>-</sup> T cells show greater resistance to suppression by TNFR2- Tregs. In addition, TNFR2 is also shown to be responsible for a more potent suppressive capacity of Tregs when TNFR2<sup>+</sup> Tregs preferentially accumulated intratumorally than in periphery (68). Previously, Tregs were described as CD4<sup>+</sup> T cells expressing CD25, the IL-2R  $\alpha$  chain, and CD45RB with Foxp3 as functional transcription marker (75). Co-expression of TNFR2 with CD25 has been suggested to identify more functional suppressive Foxp3<sup>+</sup> Tregs in human (76). Although TNFR2 induces proliferation of both Tregs and Teff, TNFR2<sup>+</sup> Tregs are able to overcome the inhibition of suppression of TNFR2<sup>+</sup> Teff (70). A reduction of Tregs in TNFR2 deficient mice indicates the role of TNFR2 in promoting the generation and homeostasis of this cell subtype (56). Both in vivo and ex vivo expansions of Tregs while maintaining their suppressive capacity have been demonstrated by utilizing TNFR2 signaling (77, 78).

Furthermore, a subset of Treg, CD8<sup>+</sup> Tregs, which can be rapidly generated in the presence of IL-4 and IL-12, can both

block activation of naïve or Teff and suppress IgG/IgE antibody response (79). Expansion of this Treg subset is induced in the presence of activated CD8<sup>+</sup> T cells via TNFR2 signaling in which TNFR2 is usually identified on a more potent subpopulation (80, 81). However, there are studies that experimentally demonstrate the inhibition of Tregs suppressive function through TNF-TNFR2 axis (57, 73, 82). These inhibitory effects of TNF-TNFR2 axis in Tregs have been associated with several mechanisms including activation of NFkB cascade, preferentially activated by pro-inflammatory TNFR1, (82) and activation of Akt with Smad3 that reduced Foxp3 transcription (57). This discrepancy of TNFR2 signaling on the function of Tregs is hypothesized to be related with the crosstalk of TNFR2 with TNFR1. Under certain conditions including prolonged cell stress in disease condition, shift of TNFR2 to TNFR1 apoptotic signaling can occur, leading to opposite primary function of TNFR2 (50).

#### **TNF-TNFR2** on DCs

The biological function of DCs, the professional APCs, can be regulated by TNF. Conversely, TNF is produced by DCs upon exposure to exogenous antigen or allergen. Subsequently, depending on the cytokine milieu including TNF, DCs would activate distinct T cell responses, which in allergy corresponds primarily to a Th2 response (83). Furthermore, TNF differentially regulates maturation and survival of DCs through interaction with its two receptors, TNFR1 and TNFR2. (60). Interaction between TNFR2 and membrane-bound TNF expressed by tolerogenic DCs induces generation of suppressive Tregs (84). In addition, TNFR2 also regulates survival of DCs, which is crucial in maintaining immunity (60, 61). These studies showed that TNFR2-deficient mice failed to develop matured myeloid cells and DCs thus reducing T cell activation including Tregs. However, the maturation state of DCs when regulating the immune tolerance has been subject of debates (85). Previously, tolerance in terms of T cell anergy and deletion is induced by immature DCs (86). Currently, mature DCs which are characterized by the typical phenotypic expression of CD103, are capable of promoting antigen specific Tregs (87). CD103<sup>+</sup> DCs have been shown to selectively prime Th2 responses to inhaled allergens and impaired allergic airway inflammation in mice lacking CD103<sup>+</sup> DCs (88). Alternately, CD103<sup>+</sup> DCs in lymphoid organs have the capacity to promote generation of Foxp3<sup>+</sup> Tregs by metabolizing dietary vitamin A (89). Also, CD103<sup>+</sup> DCs in the airway has been shown to restrain airway inflammation, for example through the induction of IL-10 (62) and production of IL-12 that control Th1 (90). Previously, it was shown that TNFR2 maintains adequate IL-12 production by DCs in inflammatory responses by regulating endogenous TNF level (91). In addition, selective expansion of maximally suppressive TNFR2<sup>+</sup>Foxp3<sup>+</sup> Tregs in pulmonary inflammation can be induced by engineered nanoparticles through the activation of CD103<sup>+</sup> DCs (92).

#### **TNF-TNFR2** on MDSCs

Furthermore, TNFR2 also acts as a co-stimulatory receptor, crucial for the development and regulation of myeloid suppressor cells (61). Myeloid-derived suppressor cells (MDSCs) are an innate heterogeneous cell population that plays a crucial role

in dampening inflammatory responses. The accumulation and expansion of MDSCs has been observed in several diseases like tumors, infectious diseases, trauma, autoimmune diseases and asthma as well. The suppressive MDSCs regulate the adaptive immunity by inhibiting the activation of T cells, especially Th2 cells in allergy (65, 66), although the upregulation of MDSCs, synergistically with Th2 polarization in asthma has also been observed (65). Similar to Tregs, the TNF-TNFR2 axis also promotes the activation and suppression of MDSCs in tumor progression by several mechanisms such as promoting secretion of nitric oxide (NO), IL-10 and TGF-B as well as enhancing the inhibition of lymphocyte proliferation (93, 94). However, it is unclear whether the interaction of TNFR2 with Tregs and MDSCs have the same signaling pathway of the suppressive mechanism in tumor development and immune evasion. For example the activation of TNFR2 on Tregs enhances immune suppression in vivo and stimulates proliferation in vitro (61) while MDSCs rely on TNFR2 activation for their maturation and to reach their optimal suppressive function (95).

## THERAPEUTIC TARGETING OF TNF-TNFR2 IN ALLERGY

Currently, treatment for allergy such as antihistamine and glucocorticoid only temporarily relieve symptoms in terms of inflammation, hence a disease-modifying treatment is fundamental. In allergic models, TNF antagonist have been shown to induce anti-allergic effects by reducing IgE level, Th2 cytokines, and eosinophils infiltration (35, 48, 96). Moreover, preferential expression of TNFR2 on Tregs makes it a more attractive molecular target for drug development. Studies showing the efficacy of targeting TNF-TNFR2 in allergies such as atopic dermatitis (97) and asthma (46, 59) have made this as a promising treatment option. Although such molecules are capable of neutralizing TNF, their affinity and avidity toward both soluble and transmembrane TNF is highly variable as well as the effects they exert on TNF-producing cells. The big differences can specifically be found in their distinct pharmalogical properties, which explains their variable efficacy in allergy treatment modalities (98) (Table 2).

In clinical settings, five TNF antagonists (three human monoclonal antibodies; infliximab, adalimumab, and golimumab, a TNFR2 receptor etanercept and certolizumab, the PEGylated Fab antibodies) have been established as therapeutic options in inflammatory diseases. Etanercept, a genetically engineered recombinant protein that comprises of TNFR2 and Fc portion of human IgG1, specifically binds to TNF and blocks its interaction with cell surface receptors (99). Studies have shown that in an allergy model, etanercept attenuates allergic lung inflammation (48) while in allergic asthma it can even reverse the inhibitory activity of TNF onto Tregs (59). However, in a randomized, phase II controlled trial (RCT), in comparison to placebo, etanercept only showed to be a well-tolerated therapy in moderate-to-severe asthma but with no significant clinical efficacy (100). Targeting TNF with etanercept in mild-tomoderate allergic asthma increased the TNFR2 levels but failed to attenuate disease pathologies (101). Moreover, treatment

TABLE 2	TNF antagonist and its efficacy in a	llergy as well as their effects on Tregs.
	The analysing and to onloady in a	norgy do won do thon oncoto on nogo.

Biologic name (trade name)	Type of agent	Approved indication	Efficacy in allergy		Effects on Tregs	References
			in vitro	in vivo		
Etanercept (Enbrel)	Human TNFR2-Fc fusion protein	RA, JIA, PA, AS, PP	+++	+	Decrease of Foxp3+ cells in vitro.	(66, 96–100)
Infliximab (Remicade)	Chimeric anti-TNF mAb	RA, CD, UC, AS, PS, PP	+++	+	Expansion of Tregs in RA	(34, 70, 93, 94, 101)
Adalimumab (Humira)	Human anti-TNF mAb	RA, PA, CD, PP	nd	Nd	Expansion of Tregs in RA	(99, 102)
Golimumab (Simponi)	Human anti-TNF mAb	RA, AS, PA	nd	-	nd	(103, 104)
Certolizumab Pegol (Cimzia)	Human PEGylated Fab anti-TNF mAb	RA, CD, PA, AS	nd	nd	nd	(105)

RA, rheumatoid arthritis; AS, ankylosing spondylitis; CD, Crohn's disease; PP, plaque psoriasis; PA, psoriatic arthritis; UC, ulcerative colitis; JIA, juvenile idiopathic arthritis; + + +, very strong; +, weak; -, no effect; nd; no data.

with etanercept in severe atopic dermatitis only showed modest effects (102). Infliximab, the chimeric monoclonal anti-TNF, showed significantly, reduced pathological inflammation in allergy model (35, 96). In addition, treatment with infliximab improved clinical outcome in both moderate and severe atopic dermatitis, but failed to respond in the maintenance therapy (97). Another anti-TNF monoclonal antibodies, golimumab, demonstrated to be unsuitable for treatment in severe asthma when it shows unfavorable risk-benefit profile in a RCT (103). Although it is well established that TNF plays a prominent role in establishment and maintenance of allergy, treatment with its antagonist in allergy population shows to be inefficient although they successfully attenuate the symptoms in allergy model. The severe side effects (immunosuppression, risk of infection, hematological malignancies, demyelinating events and neuropathies, impact on cardiovascular), and unsuccessful trials, are a limitation to their use as a general treatment in allergy (42).

Unlike in allergy, TNF-TNFR2 axis in autoimmune diseases and cancer has been well established. Adalimumab has shown to expand functional Tregs through TNF-TNFR2 axis on monocytes in rheumatoid arthritis (104). Another study in a rheumatoid arthritis model exhibited selective blockade of TNFR1 while sparing TNFR2 ameliorated inflammation and enhanced number and function of Tregs (105). Nguyen and Ehrenstein (104) have demonstrated a mechanism where TNF antagonism selectively neutralized the pro-inflammatory soluble TNF while it enhanced the immunosuppressive function of membrane TNF (104). The aforementioned studies may suggest that targeting TNF through TNFR2 not only neutralizes its proinflammatory activities but also induces tolerance by activation and expansion of Tregs (Table 2). In allergy perspective, although TNF antagonism does increase the TNFR2 levels (101), but other study demonstrated a functional insufficiency by TNF via TNFR2 signaling pathway (59). Interestingly, effectiveness toward anti-TNF treatment has also been associated with polymorphism of TNF receptor superfamily member 1B that code for TNFR2 protein (106). This may explain the effectiveness of TNF antagonism in only certain allergy population.

Due to its restricted cellular expression and prominent role in immune regulation, TNFR2 is still a more attractive target for treatment in diseases, including allergy. Faustman and Davis (50) suggested both short and long-term strategies to refine TNFR2 as a target in therapy. These include a better TNFR2 agonist by mean to improve specificity, binding duration, and affinity, as well as TNF inducers, modulation in NFkB pathway and CD3-specific antibodies (107). Furthermore, utilization of nanoparticles to modulate immune response has been widely investigated (108). Nanoparticles, with their various immunological effects in the lung (109), can be utilized to selectively block TNFR1 and/or activate TNFR2. A synthetic nanoparticle has been used to imprint innate immunity when pre-exposed mice preferentially expanded TNFR2+Foxp3+ Tregs after allergen challenge, partly via the activation of CD103<sup>+</sup> DCs (92, 110). This non-toxic engineered nanoparticle evidenced the selective modulation of Tregs homeostasis through mechanisms such as maintenance of TNF-TNFR2 interaction, targeting CD103<sup>+</sup> DCs, and expanding the proliferative rate of Tregs, thus decreasing the susceptibility to allergic disease. This strategy to use engineered targeting can also be adopted in specific immunotherapy (SIT). It can be considered as an achievable long-term cure for allergy as basic principle in SIT, which involves inducing immune tolerance toward allergens by specifically and repeatedly administrating the causative allergen (111).

### **CONCLUSIVE REMARKS**

In allergy, TNF may have dual pro-inflammatory and antiinflammatory activities, which are likely mediated by its two distinct receptors. TNFR2 signaling is attributable to the immunosuppressive effects of TNF and thus is protective against allergy. Consequently, targeting TNF-TNFR2 pathway may represent future direction to develop new therapies in allergy. However, as a pleiotropic cytokine, TNF and its effects on TNFR signaling is diverse and can either regulate allergic manifestations or to control disease pathogenesis. Implications of TNFR2 in health and diseases requires additional investigation to further elucidate their exact mechanisms and provide more insight for future strategies in manipulating TNFR2 for therapeutics.

### **AUTHOR CONTRIBUTIONS**

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

### REFERENCES

- Larche M, Akdis CA, Valenta R. Immunological mechanisms of allergen-specific immunotherapy. Nat Rev Immunol. (2006) 6:761–71. doi: 10.1038/nri1934
- Portelli MA, Hodge E, Sayers I. Genetic risk factors for the development of allergic disease identified by genome-wide association. *Clin Exp Allergy* (2015) 45:21–31. doi: 10.1111/cea.12327
- Stein MM, Hrusch CL, Gozdz J, Igartua C, Pivniouk V, Murray SE, et al. Innate immunity and asthma risk in amish and hutterite farm children. N Engl J Med. (2016) 375:411–21. doi: 10.1056/NEJMoa1508749
- Lambrecht BN, Hammad H. Allergens and the airway epithelium response: gateway to allergic sensitization. *J Allergy Clin Immunol.* (2014) 134:499–507. doi: 10.1016/j.jaci.2014.06.036
- Arora M, Poe SL, Ray A, Ray P. LPS-induced CD11b+Gr1(int)F4/80+ regulatory myeloid cells suppress allergen-induced airway inflammation. *Int Immunopharmacol.* (2011) 11:827–32. doi: 10.1016/j.intimp.2011.01.034
- Eisenbarth SC, Piggott DA, Huleatt JW, Visintin I, Herrick CA, Bottomly K. Lipopolysaccharide-enhanced, toll-like receptor 4-dependent T helper cell type 2 responses to inhaled antigen. *J Exp Med.* (2002) 196:1645–51. doi: 10.1084/jem.20021340
- Paul WE, Zhu J. How are TH2-type immune responses initiated and amplified? *Nat Rev Immunol.* (2010) 10:225–35. doi: 10.1038/nri2735
- Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation. Nature (2008) 454:445–54. doi: 10.1038/nature07204
- Amin K. The role of mast cells in allergic inflammation. *Respir Med.* (2012) 106:9–14. doi: 10.1016/j.rmed.2011.09.007
- Kuhn R, Rajewsky K, Muller W. Generation and analysis of interleukin-4 deficient mice. *Science* (1991) 254:707–10. doi: 10.1126/science.1948049
- Bousquet J, Chanez P, Lacoste JY, Barneon G, Ghavanian N, Enander I, et al. Eosinophilic inflammation in asthma. N Engl J Med. (1990) 323:1033–9. doi: 10.1056/NEJM199010113231505
- 12. Sutton BJ, Gould HJ. The human IgE network. *Nature* (1993) 366:421–8. doi: 10.1038/366421a0
- McKenzie GJ, Emson CL, Bell SE, Anderson S, Fallon P, Zurawski G, et al. Impaired development of Th2 cells in IL-13-deficient mice. *Immunity* (1998) 9:423–32. doi: 10.1016/s1074-7613(00)80625-1
- Wong CK, Ho CY, Ko FWS, Chan CHS, Ho ASS, Hui DSC, et al. Proinflammatory cytokines (IL-17, IL-6, IL-18 and IL-12) and Th cytokines (IFN-gamma, IL-4, IL-10 and IL-13) in patients with allergic asthma. *Clin Exp Immunol.* (2001) 125:177–83. doi: 10.1046/j.1365-2249.2001.01602.x
- Liu F, Yu J, Bai L, Xue Z, Zhang X. Pingchuan formula improves asthma via restoration of the Th17/Treg balance in a mouse model. *BMC Complement Altern Med.* (2015) 15:234. doi: 10.1186/s12906-015-0755-8
- Newcomb DC, Peebles RS, Jr. Th17-mediated inflammation in asthma. *Curr* Opin Immunol. (2013) 25:755–60. doi: 10.1016/j.coi.2013.08.002
- Jones CP, Gregory LG, Causton B, Campbell GA, Lloyd CM. Activin A and TGF-beta promote T(H)9 cell-mediated pulmonary allergic pathology. J Allergy Clin Immunol. (2012) 129:1000–10 e3. doi: 10.1016/j.jaci.2011.12.965
- Farfariello V, Amantini C, Nabissi M, Morelli MB, Aperio C, Caprodossi S, et al. IL-22 mRNA in peripheral blood mononuclear cells from allergic rhinitic and asthmatic pediatric patients. *Pediatr Allergy Immunol.* (2011) 22:419–23. doi: 10.1111/j.1399-3038.2010.01116.x
- Ro ADB, Simpson MR, Ro TB, Storro O, Johnsen R, Videm V, et al. Reduced Th22 cell proportion and prevention of atopic dermatitis in infants following maternal probiotic supplementation. *Clin Exp Allergy* (2017) 47:1014–21. doi: 10.1111/cea.12930
- 20. Pellerin L, Jenks JA, Begin P, Bacchetta R, Nadeau KC. Regulatory T cells and their roles in immune dysregulation and allergy.

### FUNDING

This work was supported by the Fundamental Research Grant Scheme (FRGS) by Ministry of Higher Education (MOHE) Malaysia (203/PPSP/6171198) and Universiti Sains Malaysia (USM) Research Grant (1001/PPSP/ 8012294).

*Immunol Res.* (2014) 58:358–68. doi: 10.1007/s12026-014-8512-5

- Fontenot JD, Rasmussen JP, Williams LM, Dooley JL, Farr AG, Rudensky AY. Regulatory T cell lineage specification by the forkhead transcription factor foxp3. *Immunity* (2005) 22:329–41. doi: 10.1016/j.immuni.2005.01.016
- 22. Provoost S, Maes T, van Durme YM, Gevaert P, Bachert C, Schmidt-Weber CB, et al. Decreased FOXP3 protein expression in patients with asthma. *Allergy* (2009) 64:1539–46. doi: 10.1111/j.1398-9995.2009.02056.x
- Sogut A, Yilmaz O, Kirmaz C, Ozbilgin K, Onur E, Celik O, et al. Regulatory-T, T-Helper 1, and T-Helper 2 cell differentiation in nasal mucosa of allergic rhinitis with olive pollen sensitivity. *Int Arch Allergy Immunol.* (2012) 157:349–53. doi: 10.1159/000329159
- 24. Lee SM, Gao B, Dahl M, Calhoun K, Fang D. Decreased FoxP3 gene expression in the nasal secretions from patients with allergic rhinitis. *Otolaryngol Head Neck Surg.* (2009) 140:197–201. doi: 10.1016/j.otohns.2008.08.016
- Alnahas S, Hagner S, Raifer H, Kilic A, Gasteiger G, Mutters R, et al. IL-17 and TNF-alpha are key mediators of moraxella catarrhalis triggered exacerbation of allergic airway inflammation. *Front Immunol.* (2017) 8:1562. doi: 10.3389/fimmu.2017.01562
- Rogala B, Bozek A, Gluck J, Jarzab J. Prevalence of IgE-mediated allergy and evaluation of Th1/Th2 cytokine profiles in patients with severe bronchial asthma. *Postepy Dermatol Alergol.* (2015) 32:274–80. doi: 10.5114/pdia.2015.53323
- Meng SS, Gao R, Yan BD, Ren J, Wu F, Chen P, et al. Maternal allergic disease history affects childhood allergy development through impairment of neonatal regulatory T-cells. *Respir Res.* (2016) 17:114. doi: 10.1186/s12931-016-0430-8
- Hinz D, Bauer M, Roder S, Olek S, Huehn J, Sack U, et al. Cord blood Tregs with stable FOXP3 expression are influenced by prenatal environment and associated with atopic dermatitis at the age of one year. *Allergy* (2012) 67:380–9. doi: 10.1111/j.1398-9995.2011.02767.x
- Fu Y, Lou H, Wang C, Lou W, Wang Y, Zheng T, et al. T cell subsets in cord blood are influenced by maternal allergy and associated with atopic dermatitis. *Pediatr Allergy Immunol.* (2013) 24:178–86. doi: 10.1111/pai.12050
- Zhang HY, Kong H, Zeng XN, Guo LY, Sun XY, He SH. Subsets of regulatory T cells and their roles in allergy. J Transl Med. (2014) 12:125. doi: 10.1186/1479-5876-12-125
- Aggarwal BB. Signalling pathways of the TNF superfamily: a double-edged sword. Nat Rev Immunol. (2003) 3:745–56. doi: 10.1038/nri1184
- Kalliolias GD, Ivashkiv LB. TNF biology, pathogenic mechanisms and emerging therapeutic strategies. *Nat Rev Rheumatol.* (2016) 12:49–62. doi: 10.1038/nrrheum.2015.169
- 33. Grell M, Douni E, Wajant H, Lohden M, Clauss M, Maxeiner B, et al. The transmembrane form of tumor-necrosis-factor is the prime activating ligand of the 80 Kda tumor-necrosis-factor receptor. *Cell* (1995) 83:793–802. doi: 10.1016/0092-8674(95)90192-2
- Iwasaki M, Saito K, Takemura M, Sekikawa K, Fujii H, Yamada Y, et al. TNFalpha contributes to the development of allergic rhinitis in mice. J Allergy Clin Immunol. (2003) 112:134–40. doi: 10.1067/mai.2003.1554
- Mo JH, Kang EK, Quan SH, Rhee CS, Lee CH, Kim DY. Anti-tumor necrosis factor-alpha treatment reduces allergic responses in an allergic rhinitis mouse model. *Allergy* (2011) 66:279–86. doi: 10.1111/j.1398-9995.2010.02476.x
- Manni ML, Trudeau JB, Scheller EV, Mandalapu S, Elloso MM, Kolls JK, et al. The complex relationship between inflammation and lung function in severe asthma. *Mucosal Immunol.* (2014) 7:1186–98. doi: 10.1038/mi.2014.8
- 37. Lee HS, Park HW, Song WJ, Jeon EY, Bang B, Shim EJ, et al. TNF-alpha enhance Th2 and Th17 immune responses regulating by

IL23 during sensitization in asthma model. Cytokine (2016) 79:23-30. doi: 10.1016/j.cyto.2015.12.001

- Choi JP, Kim YS, Kim OY, Kim YM, Jeon SG, Roh TY, et al. TNF-alpha is a key mediator in the development of Th2 cell response to inhaled allergens induced by a viral PAMP double-stranded RNA. *Allergy* (2012) 67:1138–48. doi: 10.1111/j.1398-9995.2012.02871.x
- Meyer N, Zimmermann M. IL-32 is expressed by human primary keratinocytes and modulates keratinocyte apoptosis in atopic dermatitis. J Allergy Clin Immunol. (2010) 125:858–65.e10. doi: 10.1016/j.jaci.2010.01.016
- Huang H, Nie W, Qian J, Zang Y, Chen J, Lai G, et al. Effects of TNF-alpha polymorphisms on asthma risk: a systematic review and meta-analysis. J Investig Allergol Clin Immunol. (2014) 24:406–17.
- Hardyman MA, Wilkinson E, Martin E, Jayasekera NP, Blume C, Swindle EJ, et al. TNF-α-mediated bronchial barrier disruption and regulation by src-family kinase activation. *J Allergy Clin Immunol.* (2013) 132:665–75.e8. doi: 10.1016/j.jaci.2013.03.005
- Sedger LM, McDermott MF. TNF and TNF-receptors: from mediators of cell death and inflammation to therapeutic giants - past, present and future. *Cytokine Growth Factor Rev.* (2014) 25:453–72. doi: 10.1016/j.cytogfr.2014.07.016
- Dreschers S, Gille C, Haas M, Seubert F, Platen C, Orlikowsky TW. Reduced internalization of TNF-a/TNFR1 down-regulates caspase dependent phagocytosis induced cell death (PICD) in neonatal monocytes. *PLoS ONE* (2017) 12:e0182415. doi: 10.1371/journal.pone.0182415
- MacEwan DJ. TNF ligands and receptors-a matter of life and death. Br J Pharmacol. (2002) 135:855-75. doi: 10.1038/sj.bjp.0704549
- Puimege L, Libert C, Hauwermeiren FV. Regulation and dysregulation of tumor necrosis factor receptor-1. *Cytokine Growth Factor Rev.* (2014) 25:285–300. doi: 10.1016/j.cytogfr.2014.03.004
- 46. Berry MA, Hargadon B, Shelley M, Parker D, Shaw DE, Green RH, et al. Evidence of a role of tumor necrosis factor alpha in refractory asthma. N Engl J Med. (2006) 354:697–708. doi: 10.1056/NEJMoa050580
- Whitehead GS, Thomas SY, Shalaby KH, Nakano K, Moran TP, Ward JM, et al. TNF is required for TLR ligand-mediated but not proteasemediated allergic airway inflammation. J Clin Invest. (2017) 127:3313–26. doi: 10.1172/JCI90890
- Maillet I, Schnyder-Candrian S, Couillin I, Quesniaux VF, Erard F, Moser R, et al. Allergic lung inflammation is mediated by soluble tumor necrosis factor (TNF) and attenuated by dominant-negative TNF biologics. *Am J Respir Cell Mol Biol.* (2011) 45:731–9. doi: 10.1165/rcmb.2010-0512OC
- 49. Kankaanranta H, Ilmarinen P, Zhang X, Adcock IM, Lahti A, Barnes PJ, et al. Tumour necrosis factor-alpha regulates human eosinophil apoptosis via ligation of TNF-receptor 1 and balance between NF-kappaB and AP-1. *PLoS ONE* (2014) 9:e90298. doi: 10.1371/journal.pone.0090298
- 50. Faustman D, Davis M. TNF receptor 2 pathway: drug target for autoimmune diseases. *Nat Rev Drug Discov*. (2010) 9:482–93. doi: 10.1038/nrd3030
- Li XM, Chen X, Gu W, Guo YJ, Cheng Y, Peng J, et al. Impaired TNF/TNFR2 signaling enhances Th2 and Th17 polarization and aggravates allergic airway inflammation. *Am J Physiol Lung Cell Mol Physiol.* (2017) 313:L592–601. doi: 10.1152/ajplung.00409.2016
- Yang S, Wang Y, Mei K, Zhang S, Sun X, Ren F, et al. Tumor necrosis factor receptor 2 (TNFR2).interleukin-17 receptor D (IL-17RD) heteromerization reveals a novel mechanism for NF-kappaB activation. *J Biol Chem.* (2015) 290:861–71. doi: 10.1074/jbc.M114.586560
- Choi JP, Kim YM, Choi HI, Choi SJ, Park HT, Lee WH, et al. An important role of tumor necrosis factor receptor-2 on natural killer T cells on the development of dsRNA-enhanced Th2 cell response to inhaled allergens. *Allergy* (2014) 69:186–98. doi: 10.1111/all.12301
- Fotin-Mleczek M, Henkler F, Samel D, Reichwein M, Hausser A, Parmryd I, et al. Apoptotic crosstalk of TNF receptors: TNF-R2-induces depletion of TRAF2 and IAP proteins and accelerates TNF-R1-dependent activation of caspase-8. J Cell Sci. (2002) 115:2757–70.
- 55. Akkoc T, de Koning PJ, Ruckert B, Barlan I, Akdis M, Akdis CA. Increased activation-induced cell death of high IFN-gamma-producing T(H)1 cells as a mechanism of T(H)2 predominance in atopic diseases. J Allergy Clin Immunol. (2008) 121:652–8e1. doi: 10.1016/j.jaci.2007.12.1171
- 56. Chen X, Wu X, Zhou Q, Howard OM, Netea MG, Oppenheim JJ. TNFR2 is critical for the stabilization of the CD4+Foxp3+ regulatory T. cell

phenotype in the inflammatory environment. J Immunol. (2013) 190:1076–84. doi: 10.4049/jimmunol.1202659

- Zhang Q, Cui F, Fang L, Hong J, Zheng B, Zhang JZ. TNF-alpha impairs differentiation and function of TGF-beta-induced Treg cells in autoimmune diseases through Akt and Smad3 signaling pathway. *J Mol Cell Biol.* (2013) 5:85–98. doi: 10.1093/jmcb/mjs063
- Verhagen J, Akdis M, Traidl-Hoffmann C, Schmid-Grendelmeier P, Hijnen D, Knol EF, et al. Absence of T-regulatory cell expression and function in atopic dermatitis skin. J Allergy Clin Immunol. (2006) 117:176–83. doi: 10.1016/j.jaci.2005.10.040
- Lin YL, Shieh CC, Wang JY. The functional insufficiency of human CD4+CD25 high T-regulatory cells in allergic asthma is subjected to TNF-alpha modulation. *Allergy* (2008) 63:67–74. doi: 10.1111/j.1398-9995.2007.01526.x
- Maney NJ, Reynolds G, Krippner-Heidenreich A, Hilkens CMU. Dendritic cell maturation and survival are differentially regulated by TNFR1 and TNFR2. J Immunol. (2014) 193:4914–23. doi: 10.4049/jimmunol.1302929
- Schmid T, Falter L, Weber S, Muller N, Molitor K, Zeller D, et al. Chronic inflammation increases the sensitivity of mouse treg for TNFR2 costimulation. *Front Immunol.* (2017) 8:1471. doi: 10.3389/fimmu.2017.01471
- Engler DB, Reuter S, van Wijck Y, Urban S, Kyburz A, Maxeiner J, et al. Effective treatment of allergic airway inflammation with Helicobacter pylori immunomodulators requires BATF3-dependent dendritic cells and IL-10. *Proc Natl Acad Sci USA*. (2014) 111:11810–5. doi: 10.1073/pnas.1410579111
- Bernatchez E, Gold MJ, Langlois A, Lemay AM, Brassard J, Flamand N, et al. Pulmonary CD103 expression regulates airway inflammation in asthma. *Am J Physiol Lung Cell Mol Physiol.* (2015) 308:816–26. doi: 10.1152/ajplung.00319.2014
- Conejero L, Khouili SC, Martinez-Cano S, Izquierdo HM, Brandi P, Sancho D. Lung CD103+ dendritic cells restrain allergic airway inflammation through IL-12 production. *JCI Insight* (2017). doi: 10.1172/jci.insight.90420. [Epub ahead of print].
- 65. Wu Y, Yan Y, Su Z, Bie Q, Wu J, Wang S, et al. Enhanced circulating ILC2s accompany by upregulated MDSCs in patients with asthma. *Int J Clin Exp Pathol.* (2015) 8:3568–79.
- Song C, Yuan Y, Wang XM, Li D, Zhang GM, Huang B, et al. Passive transfer of tumour-derived MDSCs inhibits asthma-related airway inflammation. *Scand J Immunol.* (2014) 79:98–104. doi: 10.1111/sji.12140
- Hamano R, Huang J, Yoshimura T, Oppenheim JJ, Chen X. TNF optimally activatives regulatory T cells by inducing TNF receptor superfamily members TNFR2, 4-1BB and OX40. *Eur J Immunol.* (2011) 41:2010–20. doi: 10.1002/eji.201041205
- Chen X, Subleski JJ, Kopf H, Howard OMZ, Mannel DN, Oppenheim JJ. Expression of TNFR2 defines a maximally suppressive subset of mouse CD4(+)CD25(+)FoxP3(+) T regulatory cells: applicability to tumor-infiltrating T regulatory cells. J Immunol. (2008) 180:6467–71. doi: 10.4049/jimmunol.180.10.6467
- 69. Chen X, Nie Y, Xiao H, Bian Z, Scarzello AJ, Song NY, et al. TNFR2 expression by CD4 effector T cells is required to induce full-fledged experimental colitis. *Sci Rep.* (2016) 6:32834. doi: 10.1038/srep32834
- Chen X, Hamano R, Subleski JJ, Hurwitz AA, Howard OM, Oppenheim JJ. Expression of costimulatory TNFR2 induces resistance of CD4+FoxP3conventional T cells to suppression by CD4+FoxP3+ regulatory T cells. J Immunol. (2010) 185:174–82. doi: 10.4049/jimmunol.0903548
- van Mierlo GJD, Scherer HU, Hameetman M, Morgan ME, Flierman R, Huizinga TWJ, et al. Cutting edge: TNFR-shedding by CD4+CD25+ regulatory T cells inhibits the induction of inflammatory mediators. J Immunol. (2008) 180:2747–51. doi: 10.4049/jimmunol.180.5.2747
- Nie H, Zheng Y, Li R, Guo TB, He D, Fang L, et al. Phosphorylation of FOXP3 controls regulatory T cells function and is inhibited by TNF-a in rheumatoid arthritis. *Nat Med.* (2013) 19:322–8. doi: 10.1038/nm/3085
- Valencia X, Stephens G, Goldbach-Mansky R, Wilson M, Shevach EM, Lipsky PE. TNF downmodulates the function of human CD4+CD25hi T-regulatory cells. *Blood* (2006) 108:253–61. doi: 10.1182/blood-2005-11-4567
- Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol.* (2003) 4:330-6. doi: 10.1038/ni904

- Rivas MN, Chatila TA. Regulatory T cells in allergic diseases. J Allergy Clin Immun. (2016) 138:639–52. doi: 10.1016/j.jaci.2016.06.003
- Chen X, Subleski JJ, Hamano R, Howard OM, Wiltrout RH, Oppenheim JJ. Co-expression of TNFR2 and CD25 identifies more of the functional CD4+FOXP3+ regulatory T cells in human peripheral blood. *Eur J Immunol.* (2010) 40:1099–106. doi: 10.1002/eji.200940022
- Okubo Y, Torrey H, Butterworth J, Zheng H, Faustman DL. Treg activation defect in type 1 diabetes: correction with TNFR2 agonism. *Clin Transl Immunol.* (2016) 5:e56. doi: 10.1038/cti.2015.43
- He X, Landman S, Bauland SC, van den Dolder J, Koenen HJ, Joosten I. A TNFR2-agonist facilitates high purity expansion of human low purity treg cells. *PLoS ONE* (2016) 11:e0156311. doi: 10.1371/journal.pone.0156311
- Siegmund K, Ruckert B, Ouaked N, Burgler S, Speiser A, Akdis CA, et al. Unique phenotype of human tonsillar and in vitro-induced FOXP3+CD8+ T cells. *J Immunol.* (2009) 182:2124–30. doi: 10.4049/jimmunol.0802271
- Joedicke JJ, Myers L, Carmody AB, Messer RJ, Wajant H, Lang KS, et al. Activated CD8+ T cells induce expansion of Vbeta5+ regulatory T cells via TNFR2 signaling. J Immunol. (2014) 193:2952–60. doi: 10.4049/jimmunol.1400649
- Ablamunits V, Bisikirska B, Herold KC. Acquisition of regulatory function by human CD8(+) T cells treated with anti-CD3 antibody requires TNF. *Eur J Immunol.* (2010) 40:2891–901. doi: 10.1002/eji.2010 40485
- Nagar M, Jacob-Hirsch J, Vernitsky H, Berkun Y, Ben-Horin S, Amariglio N, et al. TNF activates a NF-kappaB-regulated cellular program in human CD45RA- regulatory T cells that modulates their suppressive function. J Immunol. (2010) 184:3570–81. doi: 10.4049/jimmunol.0902070
- Na H, Cho M, Chung Y. Regulation of Th2 cell immunity by dendritic cells. *Immune Netw.* (2016) 16:1–12. doi: 10.4110/in.2016.16.1.1
- Kleijwegt FS, Laban S, Duinkerken G, Joosten AM, Zaldumbide A, Nikolic T, et al. Critical role for TNF in the induction of human antigen-specific regulatory T cells by tolerogenic dendritic cells. *J Immunol.* (2010) 185:1412– 8. doi: 10.4049/jimmunol.1000560
- Schmidt SV, Nino-Castro AC, Schultze JL. Regulatory dendritic cells: there is more than just immune activation. *Front Immunol.* (2012) 3:274. doi: 10.3389/fimmu.2012.00274
- Manicassamy S, Pulendran B. Dendritic cell control of tolerogenic responses. *Immunol Rev.* (2011) 241:206–27. doi: 10.1111/j.1600-065X.2011.01015.x
- Mildner A, Jung S. Development and function of dendritic cell subsets. *Immunity* (2014) 40:642–56. doi: 10.1016/j.immuni.2014.04.016
- Nakano H, Free ME, Whitehead GS, Maruoka S, Wilson RH, Nakano K, et al. Pulmonary CD103(+) dendritic cells prime Th2 responses to inhaled allergens. *Mucosal Immunol.* (2012) 5:53–65. doi: 10.1038/mi.2011.47
- Reynolds LA, Finlay BB. Early life factors that affect allergy development. Nat Rev Immunol. (2017) 17:518–28. doi: 10.1038/nri.2017.39
- Hamza T, Barnett JB, Li B. Interleukin 12 a key immunoregulatory cytokine in infection applications. *Int J Mol Sci.* (2010) 11:789–806. doi: 10.3390/ijms11030789
- Martin EM, Remke A, Pfeifer E, Polz J, Pietryga-Krieger A, Steffens-Weber D, et al. TNFR2 maintains adequate IL-12 production by dendritic cells in inflammatory responses by regulating endogenous TNF levels. *Innate Immun.* (2014) 20:712–20. doi: 10.1177/1753425913506949
- Mohamud R, LeMasurier JS, Boer JC, Sieow JL, Rolland JM, O'Hehir RE, et al. Synthetic nanoparticles that promote tumor necrosis factor receptor 2 expressing regulatory T cells in the lung and resistance to allergic airways inflammation. *Front Immunol.* (2017) 8:1812. doi: 10.3389/fimmu.2017.01812
- Polz J, Remke A, Weber S, Schmidt D, Weber-Steffens D, Pietryga-Krieger A, et al. Myeloid suppressor cells require membrane TNFR2 expression for suppressive activity. *Immun Inflammat Dis.* (2014) 2:121–30. doi: 10.1002/iid3.19
- 94. Hu X, Li B, Li X, Zhao X, Wan L, Lin G, et al. Transmembrane TNFalpha promotes suppressive activities of myeloid-derived suppressor cells via TNFR2. *J Immunol.* (2014) 192:1320–31. doi: 10.4049/jimmunol.12 03195
- Zhao X, Rong L, Zhao X, Li X, Liu X, Deng J, et al. TNF signaling drives myeloid-derived suppressor cell accumulation. *J Clin Invest.* (2012) 122:4094–104. doi: 10.1172/JCI64115

- 96. Wei-Xu H, Wen-Yun Z, Xi-Ling Z, Zhu W, Li-Hua W, Xiao-Mu W, et al. Anti-interleukin-1 beta/tumor necrosis factor-alpha IgY antibodies reduce pathological allergic responses in Guinea pigs with allergic rhinitis. *Mediat Inflamm*. (2016) 2016:3128182. doi: 10.1155/2016/3128182
- Jacobi A, Antoni C, Manger B, Schuler G, Hertl M. Infliximab in the treatment of moderate to severe atopic dermatitis. J Am Acad Dermatol. (2005) 52:522–6. doi: 10.1016/j.jaad.2004.11.022
- Benucci M, Saviola G, Manfredi M, Sarzi-Puttini P, Atzeni F. Tumor necrosis factors blocking agents: analogies and differences. *Acta Biomed.* (2012) 83:72–80.
- 99. Scott LJ. Etanercept: a review of its use in autoimmune inflammatory diseases. Drugs (2014) 74:1379–410. doi: 10.1007/s40265-014-0258-9
- 100. Holgate ST, Noonan M, Chanez P, Busse W, Dupont L, Pavord I, et al. Efficacy and safety of etanercept in moderate-to-severe asthma: a randomised, controlled trial. *Eur Respir J.* (2011) 37:1352–9. doi: 10.1183/09031936.00063510
- 101. Rouhani FN, Meitin CA, Kaler M, Miskinis-Hilligoss D, Stylianou M, Levine SJ. Effect of tumor necrosis factor antagonism on allergenmediated asthmatic airway inflammation. *Respir Med.* (2005) 99:1175–82. doi: 10.1016/j.rmed.2005.02.031
- Buka RL, Resh B, Roberts B, Cunningham BB, Friedlander S. Etanercept is minimally effective in 2 children with atopic dermatitis. *J Am Acad Dermatol.* (2005) 53:358–9. doi: 10.1016/j.jaad.2005.03.013
- 103. Wenzel SE, Barnes PJ, Bleecker ER, Bousquet J, Busse W, Dahlen SE, et al. A randomized, double-blind, placebo-controlled study of tumor necrosis factor-alpha blockade in severe persistent asthma. *Am J Respir Crit Care Med.* (2009) 179:549–58. doi: 10.1164/rccm.200809-1512OC
- Nguyen DX, Ehrenstein MR. Anti-TNF drives regulatory T cell expansion by paradoxically promoting membrane TNF-TNF-RII binding in rheumatoid arthritis. J Exp Med. (2016) 213:1241–53. doi: 10.1084/jem.20151255
- 105. McCann FE, Perocheau DP, Ruspi G, Blazek K, Davies ML, Feldmann M, et al. Selective tumor necrosis factor receptor I blockade is antiinflammatory and reveals immunoregulatory role of tumor necrosis factor receptor II in collagen-induced arthritis. *Arthritis Rheumatol.* (2014) 66:2728–38. doi: 10.1002/art.38755
- 106. Chen W, Xu H, Wang X, Gu J, Xiong H, Shi Y. The tumor necrosis factor receptor superfamily member 1B polymorphisms predict response to anti-TNF therapy in patients with autoimmune disease: a meta-analysis. *Int Immunopharmacol.* (2015) 28:146–53. doi: 10.1016/j.intimp.2015.05.049
- 107. Faustman DL, Davis M. TNF receptor 2 and disease: autoimmunity and regenerative medicine. *Front Immunol.* (2013) 4:478. doi: 10.3389/fimmu.2013.00478
- Ahmad S, Zamry AA, Tan HT, Wong KK, Lim J, Mohamud R. Targeting dendritic cells through gold nanoparticles: a review on the cellular uptake and subsequent immunological properties. *Mol Immunol.* (2017) 91:123–33. doi: 10.1016/j.molimm.2017.09.001
- 109. Mohamud R, Xiang SD, Selomulya C, Rolland JM, O'Hehir RE, Hardy CL, et al. The effects of engineered nanoparticles on pulmonary immune homeostasis. *Drug Metab Rev.* (2014) 46:176–90. doi: 10.3109/03602532.2013.859688
- 110. Hardy CL, Lemasurier JS, Mohamud R, Yao J, Xiang SD, Rolland JM, et al. Differential uptake of nanoparticles and microparticles by pulmonary APC subsets induces discrete immunological imprints. *J Immunol.* (2013) 191:5278–90. doi: 10.4049/jimmunol.1203131
- Akdis CA. Therapies for allergic inflammation: refining strategies to induce tolerance. Nat Med. (2012) 18:736–49. doi: 10.1038/nm.2754

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Ahmad, Azid, Boer, Lim, Chen, Plebanski and Mohamud. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.