



Article Highly Sensitive and Selective Fluorescence "Turn-On" Detection of Pb (II) Based on Fe₃O₄@Au–FITC Nanocomposite

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Abstract: New nanocomposites, $Fe_3O_4@Au$ –FITC, were prepared and explored to develop a fluorescent detection of Pb^{2+} . The $Fe_3O_4@Au$ -FITC nanocomposites could be etched by Pb^{2+} in the presence of $Na_2S_2O_3$, leading to fluorescence recovery of FITC quenched by $Fe_3O_4@Au$ nanocomposites. With the increase of Pb^{2+} concentration, the fluorescence recovery of $Fe_3O_4@AuNPs$ –FITC increased gradually. Under optimized conditions, a detection limit of 5.2 nmol/L of Pb^{2+} with a linear range of 0.02– $2.0 \ \mu$ mol/L were obtained. The assay demonstrated negligible response to common metal ions. Recoveries of 98.2–106.4% were obtained when this fluorescent method was applied in detecting Pb^{2+} spiked in a lake-water sample. The above results demonstrated the high potential of ion-induced nanomaterial etching in developing robust fluorescent assays.

Keywords: gold nanoparticles; magnetic nanoparticles; FRET; nanomaterial etching; lead ions

1. Introduction

Heavy metal pollution has been a serious long-term threat to human health and ecosystems due to their toxicity. Lead ions (Pb²⁺) are one example of major, ubiquitous, and bio-accumulative heavy metal pollutants, and can cause damage to the kidneys and brain in various ways, even at low concentration [1–3]. The U.S. Environmental Protection Agency (EPA) has set the maximum allowable levels of Pb²⁺ in drinking water at 72 nmol/L (15 ppb) [1].

Conventional analytical methods including atomic absorption spectrometry (AAS), inductively coupled plasma optical emission spectrometry (ICP-OES), inductively coupled plasma mass spectrometry (ICP-MS), etc. are broadly used in central labs for the determination of metal ions [4]. Many methods for the rapid detection of Pb²⁺ were reported including colorimetric [5,6], fluorescent [7,8], and electrochemical methods [9,10]. Among them, the fluorescent method has drawn much attention due to its advantages of high sensitivity, high specificity, and simple operation [11–15].

Nanomaterial-based fluorescent sensors for the detection of Pb²⁺ have attracted a great deal of attention recent years. These methods are mostly related to direct fluorescence quenching or energy transfer processes (i.e., Forster resonance energy transfer (FRET), and nanomaterial surface energy transfer (NSET)). The binding to or recognition of Pb²⁺ mainly includes ion-chelation with ligands [16] or biomolecules (e.g., aptamer, DNAzyme, and antibody) [17] present on the nanomaterial surface.

Another strategy of ion recognition is ion-induced leaching or etching of gold nanoparticles (AuNPs), which have been used to develop non-aggregate AuNPs colorimetric or fluorescent detection for various targets [18–20]. Compared with ion-chelation and biomolecular recognition strategies, very few efforts have been made to develop fluorescent sensing for Pb²⁺ based on the etching of nanomaterials [21].



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). In this work, fluoresce isothiocyanate (FITC) was absorbed onto the surface of Fe₃O₄@Au nanocomposites and then the fluorescence of FITC was quenched. The obtained Fe₃O₄@AuNPs–FITC nanocomposites could be etched by Pb²⁺ in the presence of Na₂S₂O₃, leading to fluorescence recovery of FITC. Based on this principle, a simple, rapid, and highly sensitive fluorescent detection for Pb²⁺ was established. Scheme 1 shows the principle of Fe₃O₄@Au–FITC nanocomposite-based fluorescent detection of Pb²⁺.



Scheme 1. Schematic illustration of Pb²⁺ detection based on Fe₃O₄@Au–FITC nanocomposites.

2. Results

2.1. Characterization of NPs

Figure 1a shows the TEM image of the Fe_3O_4 , which displays a good dispersion with an average diameter of 30 nm. Since the Fe_3O_4 NPs modified with 3-Aminopropyltriethoxysilane (APTES) have abundant amino groups on the surface, the small AuNPs could be absorbed on the Fe_3O_4 NPs by the electrostatic interaction between citric acid and amino groups [22]. As shown in (Figure 1b), the AuNPs was dispersed around spherical Fe_3O_4 particles with an average diameter of 5.0 nm. The nanocomposites can be easily magnetically separated as shown in Figure S1. The nanocomposites can remain stable after storage for 1 month. As shown in the UV-vis spectra (Figure 2), strong characteristic absorption peaks at 498 nm and 520 nm of small AuNPs (5 nm) and AuNPs (15 nm) can be found [23,24].



Figure 1. TEM images of (a) Fe₃O₄ and (b) Fe₃O₄@Au NPs.



Figure 2. UV-vis spectra of AuNPs.

2.2. Etching of Fe₃O₄@AuNPs-FITC by Lead and Thiosulfate Ions

As shown in Figure 3, the fluorescence emission of FITC was almost completely quenched by AuNPs. With the addition of $S_2O_3^{2-}$ and Pb^{2+} ions, the fluorescence intensity of the solution demonstrated slightly increasement. This may be because the above etching of AuNPs (15 nm) only slightly changed the physicochemical property and the etched AuNPs (15 nm) still were big enough to quench FITC.



Figure 3. Fluorescence spectra of the FITC-AuNPs.

Thus, we prepared the Fe₃O₄@AuNPs–FITC to further explore whether the strategy of combing the etching of smaller AuNPs, and the AuNPs separation from FITC together has the potential to develop a sensitive sensor. As shown in Figure 4, very weak fluorescent emission is due to the quenching of FITC by AuNPs. In the presence of Na₂S₂O₃, the fluorescent emission of the solution obviously increased. This may be due to the redox reaction between Na₂S₂O₃ and AuNPs in Fe₃O₄@AuNPs–FITC, forming the Au(S₂O₃)₂³⁺ compound [25,26] and slightly impairing the adsorption of FITC on the surface of AuNPs. With the coexistence of S₂O₃²⁻ and Pb²⁺ ions, the fluorescent emission of the solution increased much more significantly. These results indicate that the etching of AuNPs by ions deserves further investigation for developing a Pb²⁺ sensing method.



Figure 4. Fluorescence spectra of Fe₃O₄@Au-FITC.

2.3. Optimization of the Fluorescent Assay

To achieve a sensitive fluorescent assay for Pb^{2+} , some factors including FITC concentration, incubation time, $Na_2S_2O_3$ concentration, and pH value were investigated. As shown in Figure 5, the fluorescence intensity raises when the FITC concentration is higher than 0.5 µmol/L, which indicates excess FITC appears and separates from the Fe₃O₄@Au NPs (Figure 5). Thus, 0.5 µmol/L FITC was selected for further experiments.



Figure 5. Effect of FITC concentration on Pb²⁺ sensing.

As shown in Figure 6, the recovered fluorescence intensity $(F-F_0)$ increased with the incubation time extension, and nearly leveled off after 10 min. Thus, incubating Fe₃O₄@Au–FITC with etching ions for 10 min was enough for sensing Pb²⁺.



Figure 6. Effect of the incubation time on Pb²⁺ sensing.

As shown in Figure 7, the recovered fluorescence intensity $(F-F_0)$ displayed a gradual increase in the range 0–5.0 mmol/L Na₂S₂O₃ and a higher concentration decreased the fluorescence signal.



Figure 7. Effect of Na₂S₂O₃ concentration on Pb²⁺ sensing.

As shown in Figure 8, the pH value of glycine–NaOH buffer demonstrated big impact on the recovered fluorescence intensity (F-F₀). However, good fluorescence response could be obtained with the glycine–NaOH buffer at the range of pH 8.0–10 and highest signal obtained at pH 9.0.



Figure 8. Effect of the pH value on Pb²⁺ sensing.

2.4. Selectivity of Pb²⁺ Sensing

The selectivity of the assay was investigated by testing some common metal ions $(Cu^{2+}, Mg^{2+}, Ca^{2+}, Co^{2+}, Fe^{3+}, Sr^{2+}, Al^{3+}, Pb^{2+}, Hg^{2+}, Mn^{2+}, Cd^{2+}, Ni^{2+}, Zn^{2+}, Ba^{2+}, and Bi^{2+}, 10 \mu mol/L)$ under the same experimental conditions. As shown in Figure 9, all these metal ions produced negligible fluorescence signal to the Fe₃O₄@Au–FITC-based assay. The results indicated high selectivity toward Pb^{2+} of this Fe₃O₄@Au–FITC-based assay. The high selectivity should be ascribed to two aspects: (1) the magnetic separation and enrichment of Pb^{2+} ions; and (2) the selective etching of AuNPs by Pb^{2+} in the presence of Na₂S₂O₃.



Figure 9. Selectivity of the assay for various metal ions.

2.5. Analytical Performance of Pb²⁺ Sensing

Under optimal conditions, the sensitivity of the fluorescence assay was evaluated. As shown in Figure 10, the fluorescence intensity increased gradually with the increasing Pb²⁺ concentration in the range of 0–10 μ mol/L. A linear relationship between the fluorescence intensity and Pb²⁺ concentration could be achieved in the range 0.02 to 2.0 μ mol/L (Figure 11). The limit of detection (LOD) was 5.2 nmol/L, estimated by a 3-fold signal-to-noise ratio (3S/N). The analytical performance of the Fe₃O₄@Au–FITC-based assay was much more sensitive and rapid than the other reported nanomaterial etching-based methods [25–27] (Table 1). In addition, the LOD of the proposed method was much lower than the maximum allowable levels of Pb²⁺ (72 nmol/L) in drinking water set by the U.S. EPA.



Figure 10. Fluorescence spectra of Fe₃O₄@Au–FITC in the presence of various concentrations of Pb²⁺.



Figure 11. The relationship between the difference value of fluorescence intensities and the different concentrations of Pb^{2+} .

Nanoprobes	Linear Range	LOD	Time	Ref.
CTAB modified AuNPs	1.0~6.0 μmol/L	75 nmol/L	30 min	[25]
AuNPs and graphene oxide	0.1~20 μmol/L	50 nmol/L	20 min	[27]
AuNPs	0.0025~10 µmol/L	0.5 nmol/L	2 h	[26]
Fe3O4@Au-FITC	0.02~2.0 μmol/L	5.2 nmol/L	12 min	This work

Table 1. Comparison of several methods for Pb²⁺ detection.

2.6. Application of Pb²⁺ Sensing

Samples collected from Tai lake (Wuxi, China) were filtered through a microfiltration membrane (0.22 μ m) and spiked with different concentrations of Pb²⁺ (50, 100 and 150 nmol/L). The samples then were measured by the above fluorescent method. As demonstrated in Table 2, the recoveries from 98.2% to 106.4% were obtained with relative standard deviation (RSD) less than 10%. The above results demonstrated the practical potential of this Fe₃O₄@Au–FITC-based fluorescent method for Pb²⁺ in detecting environmental samples.

Table 2. Determination of Pb^{2+} in lake-water samples (n = 3).

Concentration (nmol/L)	Result (nmol/L)	Recovery (%)	RSD (%)
50	53.2	106.4	8.3
100	98.3	98.3	2.5
150	159.0	106.0	3.7

3. Materials and Methods

3.1. Chemicals and Reagents

All reagents were of analytical grade and used without further purification. Sodium borohydride (NaBH₄) and 3-Aminopropyltriethoxysilane (APTES) were purchased from Shanghai Aladdin biochemical technology Co., Ltd. (Shanghai, China) Chloroauric acid hydrate (HAuCl₄·3H₂O) and sodium citrate were purchased from Sigma–Aldrich (Shanghai, China). Fluorescein isothiocyanate (isomeride I) (FITC > 95.0%) was purchased from TCI (Shanghai, China) chemical industry Co., Ltd. Copper, aluminum, nickel, cobalt, barium, cadmium, manganese, iron, zinc, strontium, bismuth, chromium were purchased from Guobiao (Beijing, China) Testing & Certification Co., Ltd. The nitrate, acetate, or chloride salts of other metal ions were purchased from Sinopharm Chemical Reagent Co, Ltd.(Shanghai, China). Ethylene glycol, ethanolamine, ethanol and Poly (ethylene glycol) (PEG2000) were purchased from Sinopharm Chemical Reagent Co, Ltd. (Shanghai, China).

3.2. Apparatus

UV-visible (UV-vis) absorption spectra of nanoparticles were obtained with a microplate reader (Bio-Tek, Elx800, USA) (Winooski, VT, USA). Steady-state fluorescence spectra were measured by a fluorospectrometric photometer (Lengguan F97, Shanghai Lengguan, China). Transmission electron microscopy (TEM) was performed on a JEOL JEM-2100 (Kyoto, Japan) at an accelerating voltage of 200 kV. Energy diffraction X-ray (EDX) spectrum was obtained using the TEM.

3.3. Synthesis of Au Nanoparticles (AuNPs)

AuNPs (5.0 nm) were synthesized by NaBH₄ reduction method [23]. Briefly, HAuCl₄ (0.5 mL, 4.0 g/L) and sodium citrate (2.0 mL, 0.025 mmol/L) were dissolved in 20 mL water under magnetic stirring. Then 0.6 mL NaBH₄ (0.1 mol/L) solution was added and the mixture was stirred thoroughly. The color of the solution changed from light yellow to orange red, which indicated that the AuNPs were successfully synthesized.

3.4. Amine-Functionalization of Fe₃O₄ Nanoparticles Using APTES

Fe₃O₄ nanoparticles were synthesized according to the method with slight modifications [28]. FeCl₃·6H₂O (1.05 g), ethylene glycol (21 mL) and ethanolamine (7 mL) were added to the conical flask under magnetic stirring. Then, sodium acetate trihydrate (NaAc) (2.8 g) and PEG2000 (0.7 g) were added to the solution under vigorous stirring. The reaction was maintained at 200 °C for 8 h. Afterwards, the Fe₃O₄ NPs were obtained after the mixture was washed several times with water and ethanol. The Fe₃O₄ NPs were dried in a vacuum oven at 60 °C for 12 h and Fe₃O₄ NPs powder was obtained.

For APTES modification, the Fe₃O₄ NPs (150 mg) were suspended in a mixture of ethanol (40 mL) and APTES solution (1.0 mL). The mixture was gently stirred at room temperature for 3 h. The Fe₃O₄ NPs were separated using an external magnet, and thoroughly washed with ethanol and water. The amine-functionalization of Fe₃O₄ nanoparticles were finally suspended in 15 mL ethanol and stored at 4 °C for subsequent use.

3.5. Synthesis of Fe₃O₄@AuNCs-FITC

To prepare Fe₃O₄@AuNPs, 3.0 mL of amine-functionalization of Fe₃O₄ NPs was dissolved in 15 mL of water, and then 100 mL of AuNPs (5 nm) was added under stirring for 15 min. The Fe₃O₄@AuNPs were separated using an external magnet, washed with water and finally suspended in 25 mL of water [29]. The Fe₃O₄@AuNP-FITC nanocomposites were synthesized as follows. FITC (50 μ L,50 μ mol/L) onto Fe₃O₄@Au NPs (250 μ L) in 4.7 mL water and stored at 4 °C overnight. FITC–AuNPs was prepared as follows. AuNPs (15 nm) were prepared using trisodium citrate reduction method [24]. Briefly, 100 mL of 0.01% HAuCl₄ solution was slowly stirred, heated to a boil in the conical flask and kept boiling for 5 min. Then, 2 mL of 1% trisodium citrate solution was poured into the flask. The color of mixture changed from pale yellow to gray and finally to wine red. After boiling for 10 min, the obtained AuNPs (3.0 nmol/L) were cooled to room temperature and stored at 4 °C for subsequent use. FITC–AuNPs nanocomposites were synthesized according to the method reported by Wang et al. [30]. FITC solution (5.0 μ L, 1.5 mmol/L) in ethanol was added into 5.0 mL of AuNPs solution and mixed gently. The mixture was stored at 4 °C overnight and then FITC–AuNPs nanocomposites were obtained.

3.6. Fluorescent Detection of Pb²⁺

A total of 10 μ L of Fe₃O₄@AuNP–FITC composites, 40 μ L of H₂O, and 50 μ L of sodium thiosulfate (20 mmol/L) as well as 50 μ L of glycine–NaOH buffer (10 mmol/L, pH 9.0) were mixed gently. After incubation at room temperature for 5.0 min, Pb²⁺ solution was added and incubated at room temperature for 10 min. Then the Fe₃O₄@AuNPs were separated by an external magnet for 2 min. The fluorescence emission spectra of the supernatant were measured by a fluorescence spectrometer with 465 nm excitation.

4. Conclusions

A highly sensitive and selective fluorescent method for Pb^{2+} sensing was developed based on a new nanocomposite, $Fe_3O_4@Au$ –FITC. This method combined the metal leaching by the $Pb^{2+}-S_2O_3^{2-}$ system with magnetic separation together. Considering that ioninduced etching of metal NPs has been applied in the development of analytical methods for the detection of metal ions, anions, small molecules, and proteins [31,32], the strategy proposed here is expected to be further explored in some challenging situations.

Supplementary Materials: The following are available online, Figure S1: Magnetic seperation of the Fe₃O₄@Au–FITC nanocomposites.

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