



Draft Genome Sequences of Four *Rhodobacter sphaeroides* Strains Isolated from a Marine Ecosystem

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ABSTRACT *Rhodobacter sphaeroides* is an alphaproteobacterium found in freshwater and marine ecosystems. To better understand the metabolic diversity within this species, we isolated and sequenced four *R. sphaeroides* isolates obtained from Trunk River in Woods Hole, Massachusetts. Here, we report the draft genome sequences of *R. sphaeroides* AB24, AB25, AB27, and AB29.

Rhodobacter sphaeroides is a model purple nonsulfur bacterium for studying microbial metabolism and bioenergetics (1, 2). It is exceptionally metabolically versatile, being capable of photoheterotrophy, photoautotrophy, chemoheterotrophy, chemoautotrophy, and fermentation. *R. sphaeroides* has also been investigated for its biotechnological (3, 4) and bioremediation (5–8) potential. To date, only 13 *R. sphaeroides* genome sequences have been deposited in the GenBank database. To examine the genetic diversity of natural isolates within this species, we isolated and sequenced the genomes of four *R. sphaeroides* strains from Trunk River in Woods Hole, Massachusetts. We determined that these isolates are 99% similar to *R. sphaeroides* KD131 (9) based on 16S rRNA gene sequence analysis.

Seawater was sampled from Trunk River, and 500 μ l was used as an inoculum into Pfenning bottles containing anoxic artificial seawater medium (10) supplemented with 20 mM acetate. Enrichments were cultivated with \sim 850-nm light at 30°C and passaged six times in anoxic artificial seawater medium, followed by streaking oxically 6 times on Bacto agar with Difco marine broth 2216 (BD Diagnostic Systems, Sparks, MD, USA). Genomic DNA was isolated with the DNeasy blood and tissue kit according to the manufacturer's recommendations (Qiagen, Dusseldorf, Germany) from single colonies cultivated in marine broth to mid-log phase. Paired-end 250-bp Illumina sequencing libraries were prepared using the Nextera sample prep kit (San Diego, CA) and sequenced on a MiSeq instrument using v2 chemistry (Illumina, Inc.) to 300 \times (AB24), 38 \times (AB25), 36 \times (AB27), or 33 \times (AB29) coverage. Reads were trimmed with Trimmomatic version 0.38 with the program's default parameters for paired-end reads (11). The trimmed reads were *de novo* assembled with SPAdes version 3.13.0 using the program's default parameters (12). Contigs were extended using the reference-guided scaffolder MeDuSa version 1.6 with the complete genome of *R. sphaeroides* KD131, using the program's default parameters (13). Alignment of the scaffolded genomes of AB24, AB25, AB27, and AB29 was performed with LASTZ version 1.02.00 to examine plasmid and chromosomal synteny between the isolates and reference genomes (14). Sequences were submitted for annotation to the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (15). The identity of each strain was determined by analyzing the full-length 16S rRNA gene sequence predicted by PGAP in the genome assemblies. NCBI BLASTN analysis was performed on these sequences to determine the identity using the program's default parameters (16). Phylogenetic analysis was performed with the BLASTN alignments using the BLAST Tree View widget, with the program's default parameters (<http://blast.ncbi.nlm.nih.gov/>).

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TABLE 1 Genome statistics and accession numbers

Strain	No. of reads	Assembly size (Mb)	No. of scaffolds	SRA accession no.	GenBank accession no.
AB24	2,899,454	3.26	4	SRR8107695	CP033434 – CP033437
AB25	676,996	3.23	4	SRR8107697	CP033442 – CP033445
AB27	637,638	3.23	4	SRR8107696	CP033446 – CP033449
AB29	569,874	3.24	4	SRR8107698	CP033450 – CP033453

The genomes of AB24, AB25, AB27, and AB29 have a total assembly length of ~3.2 Mb (Table 1) and a GC content of 69.1%. Genome scaffolding produced four sequences for each strain (Table 1) that mapped to chromosomes I (3.2 Mb) and II (1 Mb), as well as plasmids A (0.1 Mb) and B (0.2 Mb), of *R. sphaeroides* KD131 and *R. sphaeroides* 2.4.1, as determined by synteny analysis (17). Neither contigs nor sequencing reads mapped to plasmid C, D, or E of *R. sphaeroides* 2.4.1, similar to *R. sphaeroides* KD131. NCBI PGAP predicted 4,280 (AB24), 4,271 (AB25), 4,086 (AB27), and 4,169 (AB29) open reading frames. The isolates contain multiple copies of genes involved in DNA replication, amino acid metabolism, motility and chemotaxis, photosynthetic light harvesting, and central carbon metabolism, as has been characterized in *R. sphaeroides* 2.4.1 (1, 17). The isolates encode proteins involved in lithotrophic metabolism (Ni-Fe uptake hydrogenase and CO dehydrogenase), nitrogen fixation (Fe-Mo nitrogenase), and denitrification (nitrous oxide and nitric oxide reductase) (5, 18–20). These genomes provide opportunities for future studies into the metabolic potential of *R. sphaeroides* in marine ecosystems.

Data availability. These whole-genome shotgun (WGS) projects and the raw sequencing reads have been deposited in GenBank and the NCBI Sequence Read Archive (SRA), respectively, under the accession numbers listed in Table 1.

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REFERENCES

- Mackenzie C, Choudhary M, Larimer FW, Predki PF, Stilwagen S, Armitage JP, Barber RD, Donohue TJ, Hosler JP, Newman JE, Shapleigh JP, Sockett RE, Zeilstra-Ryalls J, Kaplan S. 2001. The home stretch, a first analysis of the nearly completed genome of *Rhodobacter sphaeroides* 2.4.1. *Photosynth Res* 70:19–41. <https://doi.org/10.1023/A:1013831823701>.
- Imam S, Yilmaz S, Sohmen U, Gorzalski AS, Reed JL, Noguera DR, Donohue TJ. 2011. iRsp1095: a genome-scale reconstruction of the *Rhodobacter sphaeroides* metabolic network. *BMC Syst Biol* 5:116. <https://doi.org/10.1186/1752-0509-5-116>.
- Koku H, Eroglu I, Gunduz U, Yucel M, Turker L. 2002. Aspects of the metabolism of hydrogen production by *Rhodobacter sphaeroides*. *Int J Hydrogen Energy* 27:1315–1329. [https://doi.org/10.1016/S0360-3199\(02\)00127-1](https://doi.org/10.1016/S0360-3199(02)00127-1).
- Lee I-H, Park J, Kho D, Kim M-S, Lee J. 2002. Reductive effect of H₂ uptake and poly-β-hydroxybutyrate formation on nitrogenase-mediated H₂ accumulation of *Rhodobacter sphaeroides* according to light intensity. *Appl Microbiol Biotechnol* 60:147–153. <https://doi.org/10.1007/s00253-002-1097-2>.
- Moore MD, Kaplan S. 1992. Identification of intrinsic high-level resistance to rare-earth oxides and oxyanions in members of the class *Proteobacteria*: characterization of tellurite, selenite, and rhodium sesquioxide reduction in *Rhodobacter sphaeroides*. *J Bacteriol* 174:1505–1514. <https://doi.org/10.1128/jb.174.5.1505-1514.1992>.
- O’Gara JP, Gomelsky M, Kaplan S. 1997. Identification and molecular genetic analysis of multiple loci contributing to high-level tellurite resistance in *Rhodobacter sphaeroides* 2.4.1. *Appl Environ Microbiol* 63:4713–4720.
- Nepple B, Kessi J, Bachofen R. 2000. Chromate reduction by *Rhodobacter sphaeroides*. *J Ind Microbiol Biotechnol* 25:198–203. <https://doi.org/10.1038/sj.jim.7000049>.
- Giotta L, Agostiano A, Italiano F, Milano F, Trotta M. 2006. Heavy metal ion influence on the photosynthetic growth of *Rhodobacter sphaeroides*. *Chemosphere* 62:1490–1499. <https://doi.org/10.1016/j.chemosphere.2005.06.014>.
- Lim S-K, Kim SJ, Cha SH, Oh Y-K, Rhee H-J, Kim M-S, Lee JK. 2009. Complete genome sequence of *Rhodobacter sphaeroides* KD131. *J Bacteriol* 191:1118–1119. <https://doi.org/10.1128/JB.01565-08>.
- Straub KL, Rainey FA, Widdel F. 1999. *Rhodovulum iodolum* sp. nov. and *Rhodovulum robiginosum* sp. nov., two new marine phototrophic ferrous-iron-oxidizing purple bacteria. *Int J Syst Evol Microbiol* 49:729–735. <https://doi.org/10.1099/00207713-49-2-729>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for

- Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
12. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
 13. Bosi E, Donati B, Galardini M, Brunetti S, Sagot M-F, Lió P, Crescenzi P, Fani R, Fondi M. 2015. MeDuSa: a multi-draft based scaffolder. *Bioinformatics* 31:2443–2451. <https://doi.org/10.1093/bioinformatics/btv171>.
 14. Harris RS. 2007. Improved pairwise alignment of genomic DNA. PhD thesis. Pennsylvania State University, State College, PA.
 15. Tatusova T, DiCuccio M, Badretdin A, Chetvermin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
 16. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. *J Mol Biol* 215:403–410. [https://doi.org/10.1016/S0022-2836\(05\)80360-2](https://doi.org/10.1016/S0022-2836(05)80360-2).
 17. Kontur WS, Schackwitz WS, Ivanova N, Martin J, Labutti K, Deshpande S, Tice HN, Pennacchio C, Sodergren E, Weinstock GM, Noguera DR, Donohue TJ. 2012. Revised sequence and annotation of the *Rhodobacter sphaeroides* 2.4.1 genome. *J Bacteriol* 194:7016–7017. <https://doi.org/10.1128/JB.01214-12>.
 18. Tosques IE, Shi J, Shapleigh JP. 1996. Cloning and characterization of *nnrR*, whose product is required for the expression of proteins involved in nitric oxide metabolism in *Rhodobacter sphaeroides* 2.4.3. *J Bacteriol* 178:4958–4964. <https://doi.org/10.1128/jb.178.16.4958-4964.1996>.
 19. Bartnikas TB, Tosques IE, Laratta WP, Shi J, Shapleigh JP. 1997. Characterization of the nitric oxide reductase-encoding region in *Rhodobacter sphaeroides* 2.4.3. *J Bacteriol* 179:3534–3540. <https://doi.org/10.1128/jb.179.11.3534-3540.1997>.
 20. Meijer WG, Tabita FR. 1992. Isolation and characterization of the *nifUSVW-rpoN* gene cluster from *Rhodobacter sphaeroides*. *J Bacteriol* 174:3855–3866. <https://doi.org/10.1128/jb.174.12.3855-3866.1992>.