



Successful Immunotherapy for Pancreatic Cancer in a Patient With TSC2 and SMAD4 Mutations: A Case Report

Yanghui Ye^{1,2} and Song Zheng^{2,3,4*}

¹ The Fourth School of Clinical Medicine, Zhejiang Chinese Medical University, Hangzhou, China, ² Department of Oncology, Affiliated Hangzhou First People's Hospital, Zhejiang University School of Medicine, Hangzhou, China, ³ Department of Oncology, Affiliated Hangzhou Cancer Hospital, Zhejiang University School of Medicine, Hangzhou, China, ⁴ Laboratory of Clinical Cancer Pharmacology and Toxicology Research of Zhejiang Province, Hangzhou, China

OPEN ACCESS

Edited by:

Liangrong Shi, Central South University, China

Reviewed by:

Sid P. Kerkar, EXUMA Biotech, United States Philippe Rochigneux, Institut Paoli-Calmettes (IPC), France

> *Correspondence: Song Zheng tztree@126.com

Specialty section:

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

Received: 29 September 2021 Accepted: 01 November 2021 Published: 22 November 2021

Citation:

Ye Y and Zheng S (2021) Successful Immunotherapy for Pancreatic Cancer in a Patient With TSC2 and SMAD4 Mutations: A Case Report. Front. Immunol. 12:785400. doi: 10.3389/fimmu.2021.785400 **Background:** Pancreatic cancer has a poor prognosis, and it is traditionally treated with chemotherapy. Fortunately, immunotherapy has rapidly changed the landscape of solid tumor treatment, and improving the survival of cancer patients. However, pancreatic cancer is non-immunogenic, and single agent immunotherapies are unfavorable to its prognosis.

Case Presentation: Here, we report a case of stage IV pancreatic cancer in a patient with TSC2 and SMAD4 mutations treated with immunotherapy when the disease progressed after multi-line chemotherapy. Next generation sequencing (NGS) confirmed the presence of TSC2 and SMAD4 mutations and microsatellite stability (MSS). When the disease progressed after chemotherapy, a combination strategy was devised consisting of chemotherapy (S-1) and sintilimab. The patient had a partial response to therapy with this regimen, the lesions were significantly reduced and nearly disappeared. In metastatic pancreatic cancer, responses of this magnitude are rarely seen.

Conclusions: This outcome reveals that this combination can be effective in treating metastatic pancreatic cancer, especially in pancreatic cancer patients with SMAD4 and TSC2 mutations. This may help increase the use of this therapy in large-scale clinical research.

Keywords: pancreatic cancer, immunotherapy, chemotherapy, next generation sequencing (NGS), SMAD4, TSC2

Abbreviations: MRI, magnetic resonance imaging; PET, positron emission tomography; CT, computed tomography; NGS, next generation sequencing; TGF- β , transforming growth factor beta; TMB, tumor mutational burden; MSS, microsatellite stable; MSI, microsatellite instability; MSI-H, high levels of microsatellite instability; ICI, immune checkpoint inhibitor; ORR, objective response rate; PFS, progression-free survival; OS, overall survival.

INTRODUCTION

Pancreatic cancer has a high incidence and mortality, and its special structure can protect pancreatic cancer cells from chemotherapeutic agents (1, 2). However, pancreatic cancer is non-immunogenic and single agent immunotherapies are unfavorable to the prognosis of patients. Several clinical trials showed that single agent immunotherapies are ineffective for the treatment of advanced pancreatic cancer (3–5). Moreover, there was a clinical trial confirmed that objective response rate (ORR) was 0 for patients receiving single agent immunotherapies (5).

However, the effect of combination therapy is also not optimistic. As for chemotherapy plus immunotherapy, some clinical trials confirmed that the safety profile of combination therapy at standard doses in advanced pancreatic cancer was manageable (6, 7), but there was no significant improvement in progression-free survival (PFS) and overall survival (OS) (6, 8).

Though, the pancreatic cancer has little response rate to immunotherapy, it may be effective for specific patient. For example, several gene mutations can improve the effective of immunotherapy (9), including high levels of microsatellite instability (MSI-H), POLE, POLD1, et al.

CASE PRESENTATION

We present the case of a 56-year-old Chinese man who had been smoking and drinking for decades. He was hospitalized for six months with abdominal pain. In September 2019, magnetic resonance imaging (MRI) of the upper abdomen identified a pancreatic head mass, multiple retroperitoneal enlarged lymph nodes, and abnormal enhancement near the inferior vena cava in right lobe of the liver. Ultrasound guided biopsy of the pancreas was performed. Pathology diagnosis was pancreatic ductal adenocarcinoma (stage IV) (Figures 1A, B). Next generation sequencing (NGS) confirmed the tumor was microsatellite stability (MSS), and a total of 6 gene mutations, TSC2, CREBBP, HIST1H3I, MAP2K4, SMAD4, and STK11 were detected. Additionally, frame shift mutation occurred in exon 17 of TSC2 gene (32.13%); missense mutation occurred in exon 9 (7.47%) and nonsense mutation occurred in exon 5 (9.49%) of SMAD4 gene. The NGS confirmed that there were no targeted drug-related gene mutations and the tumor mutational burden (TMB) was 7.1 mut/Mb.

In October 2019, the patient's primary oncologist started him on AG (gemcitabine and nab-paclitaxel). The gemcitabine (1000mg/m² over 30 minutes, weekly for 2 weeks, every 21 days) and nab-paclitaxel (125mg/m², weekly for 2 weeks, every 21 days) were administered intravenously. However, in the first three cycles and the fifth cycle of chemotherapy, for several reasons (such as COVID-19), he only received chemotherapeutic agents (AG) on the first day. In March 2020, the repeated MRI showed progressive disease. In late March 2020, the patient developed back pain and underwent endoscopic retrograde cholangiopancreatography (ERCP) and endoscopic metal biliary endoprosthesis (EMBE) due to obstructive jaundice. In April 2020, positron emission tomography/computed tomography (PET/CT) revealed a 6.0 cm \times 6.0 cm mass in pancreatic head, multiple enlarged lymph nodes in different sizes were found in pancreatic head and retroperitoneum, and multiple low-density shadows in the liver (**Figures 1C-F**).

Based on these findings, the patient was treated with FOLFIRINOX (oxaliplatin, Irinotecan, calcium folinate and 5-Fluorouracil). However, considering that the patient had just undergone surgery, the patient was asked by his doctor to eat S-1 for 14 days (from April 2, 2020 to April 16, 2020) before he received FOLFIRINOX treatment on April 25, 2020. In June 2020, after 4 cycles of chemotherapy, upper-abdomen enhanced CT revealed that the size of pancreatic head lesion was significantly decreased to 2.3 cm \times 2.5 cm (**Figure 2A**), more that 50% decrease, and multiple low-density shadows in the liver. The patient was repeated CT about every two months (**Figures 2B, C**). Until October 2020, the CT only revealed a mass in the pancreatic head, and the lesion was slightly low density with peripheral lymph node metastasis.

In October 2020, the patient underwent one cycle of Olaparib by himself, although he did not have the targeted drug-related gene mutation. On October 16, his abdomen enhanced CT scan revealed multiple liver metastases (**Figure 2F**), and the largest measured 2.3 cm. We changed the chemotherapy regimen to AG again (gemcitabine and nab-paclitaxel). In November 2020, the abdomen enhanced CT scan revealed progressive disease (**Figure 2D**), and the size of the pancreatic head lesion was 6.4 cm.

The patient has undergone multi-line chemotherapy, chemotherapy alone could not inhibit the progression of the disease, and there was no standard treatment after two or more lines of systemic chemotherapy. In addition, his Eastern Cooperative Oncology Group Performance Status (ECOG-PS) was 2, he suffered from severe cancer pain all over the body (OxyContin 30mg, every 12 hours) and weight loss, the aspartate transaminase and alanine transaminase were at normal level, and alkaline phosphatase increased slightly (not higher than 2 times the normal value). Thus, we decided to use chemotherapy (S-1) combined with immunotherapy (sintilimab). Considering the economic status of patient, we decided to choose sintilimab (more economical). The sintilimab is a fully human IgG4 monoclonal antibody; it can bind to PD-1, block the interaction of PD-1 with its ligands, and help recover the anti-tumor response of T-cells, and it has been approved to treat relapsed or refractory classical Hodgkin lymphoma, advanced non-small cell lung cancer and metastatic hepatocellular carcinoma. The anti-tumor effect of sintilimab is similar to that of other anti-PD-1/PD-L1 antibodies (10). Sintilimab was administered intravenously (200mg, every 3 weeks); and S-1was orally (40mg, twice a day orally for 2 weeks, every 21 days). The patient started to receive S-1 on November 24, 2020, and sintilimab on November 27, 2020.

The patient did not come to our hospital again until June 2021 (**Figure 3**). He said he was treated at a local hospital during these seven months, and received a total of 10 cycles of sintilimab combined with S-1. The performance status was better than before (his PS improved to ECOG-PS 1); his severe cancer pain was well controlled (OxyContin 10mg, every 12 hours). The abdomen enhanced CT scan



FIGURE 1 | (A, B) pancreatic ductal adenocarcinoma. (A) 200X, (B) 400X. The pancreatic head puncture smear showed significant atypical epithelial mass, irregular nuclei, which was consistent with the changes of adenocarcinoma. (C–F) PET/CT images in April 2020. (C) showed a 6.0cm *6.0 cm mass in pancreatic head (arrow); (D) confirmed enlarged lymph nodes in pancreatic head (arrow), and low-density shadows in liver (within cycle); panels (E, F) revealed enlarged multiple lymph nodes in different sizes in retroperitoneum (arrow).





showed that the intrahepatic metastases (**Figure 2G**) and pancreatic head lesion (**Figure 2E**) were significantly reduced (the size of the pancreatic head lesion was 1 cm). By standard Response Evaluation Criteria in Solid Tumors (RECIST) criteria, version 1.1, the patient had a partial response (more than 80% decrease) to therapy with this regimen. Moreover, throughout his treatment, the CA 19-9 level was always normal.

At the latest follow-up (August 2021), the patient still had a partial response to this regimen (S-1 combined with sintilimab), and the duration of partial response was 8 months.

DISCUSSION

Chemotherapy has been the mainstay of treatment for many malignant tumors, including pancreatic cancer. Due to the lack of effective screening tools, most patients lose the opportunity to undergo surgery, which makes chemotherapy the standard treatment. However, chemotherapy has a poor therapeutic effect on pancreatic cancer due to its aggressive nature. Pancreatic cancer is composed of malignant cells and desmoplastic stroma (1). Desmoplastic stoma serves as a





physical barrier that protects pancreatic cancer cells and prevent the effective delivery of chemotherapeutic agents (2).

In this case, the chemotherapy was effective for this patient at first. Additionally, after the disease progressed, the immunotherapy combined with chemotherapy had a significant effect. In the past 5 years, immunotherapy has rapidly changed the landscape of solid tumor treatment. What is more, tumor testing can help patients to get a better treatment. The results of a tumor testing making a patient eligible for treatment with immunotherapy, whose disease (advanced pancreatic cancer) progressed after neoadjuvant chemotherapy and adjuvant chemotherapy (11). Recent studies have identified several positive predictive markers for immune checkpoint inhibitors (ICIs), such as high levels of MSI-H, PD-L1 overexpression, high TMB, and gene mutations (9).

TSC2 is a tumor suppressor gene; it negatively regulates the cellular signaling networks that control cellular growth and proliferation (12). The TSC2 protein forms a complex and functions as a tumor suppressor by inhibiting mTORC1 kinase (13). It has been shown that, in TSC2-deficient tumors, the single-agent PD-1 or CTLA-4 blockade, or a combination of them, can inhibit the growth of tumors (14). Additionally, the combination of PD-1 and CTLA-4 antibody treatment or single-

agent treatment can increase CD8+ T-cell infiltration in TSC2deficient human tumors (14), and the level of infiltration is correlated with the degree of response to therapy.

Transforming growth factor beta (TGF- β) is an immune regulator; it can suppress the immune response via many different mechanisms (15). Moreover, it can inhibit tumor growth at the early stages of disease and promote tumor development at the later stages (16). However, the tumorsuppressive role of TGF- β is only effective when the TGF- β signaling pathway is not defective (17). SMAD4 serves as the central mediator of the TGF- β signaling pathway (18), and it is the only common mediator. The TGF-B/SMAD4 signaling pathway plays a tumor suppressive role in early stages of disease, mainly by inducing cell cycle arrest and apoptosis. TGF- β can stimulate regulatory T-cells, which inhibit the function of other lymphocytes (19). PD-1 is highly expressed on tumor infiltrating lymphocytes; it has been shown that human PD-1 expression may under direct transcriptional control by TGF- β , and TGF- β can enhance the expression of PD-1, suppressing anti-tumor immunity (20). TGF-β inhibits CD8+ T-cell effector function through TGF- β signaling pathway (21). Pancreatic cancer cells have lost their tumor-suppressive roles, but they possess tumor-promoting effects induced by increased

TGF- β (22). In a tumor microenvironment, TGF- β expression is very high.

In pancreatic cancer, alterations of TGF- β signaling occur through the mutation of the genes involved in the pathway (including SMAD4); this activity is present in 47% of pancreatic cancer patients (23). The loss of SMAD4 will abrogate the canonical TGF- β /SMAD4 signaling pathway (24), and it may make pancreatic cancer more aggressive (25). It has been shown that SMAD4-deleted pancreatic ductal adenocarcinoma cells are sensitive to agents modulating the cell cycle (26). The loss of SMAD4 counteracted TGF-\beta-induced cell cycle arrest and apoptosis (27). Furthermore, it has been reported that the loss of SMAD4 expression is significantly associated with better survival after resection (28). The inhibition of TGF- β has been reported to have a variety of antitumor effects (29). A TGF-B blockade can reverse the suppressive effects of apoptotic cells on inflammation and adaptive immunity (30). In T-cell excluded mouse models, immune checkpoint-resistant MSS colorectal cancers and liver tumors were rendered susceptible to anti-PD-1/PD-L1 therapy with a TGF- β blockade (31).

Blockade of immune checkpoints by anti-CTLA-4 or anti-PD-1/anti-PD-L1 agents leads to T-cell activation, and it provides an effective approach for tumor immunotherapy (32). And the high PD-L1 expression may have a better clinical benefit. There was a case report showed that blocking the PD-L1 pathway combined with chemotherapy was effective for pancreatic squamous cell carcinoma patients with high PD-L1 expression (33).

Pancreatic cancer is intrinsically non-immunogenic (34). Single agent immunotherapies are unlikely to be successful in treating this type of cancer (35), but immunotherapy combined with chemotherapy has a synergistic effect (36). Chemotherapeutic agents could promote the release of tumor antigens from the cancer cells and reactivate an anti-cancer immune response to suppress tumor growth (37). Besides, according to several ongoing clinical trials, there are other regimens of combination therapy for the treatment of pancreatic cancer, such as BL-8040 (chemokine receptor type 4 inhibitors) and pembrolizumab combined with chemotherapy (NCT02826486), olaparib plus pembrolizumab (NCT04666740 and NCT04548752), olaparib or selumetinib plus durvalumab (NCT04348045). Additionally, the COMBAT/ KEYNOTE-202 Trial (NCT02826486) revealed that the ORR was 21.1%, and the triple combination of BL-8040, pembrolizumab, and chemotherapy was safe and well tolerated, but no significant improvement in PFS and OS (38).

Other ICIs can also improve the effect of therapy. TMB is the total number of mutations per coding area of a tumor gene,

REFERENCES

- Neesse A, Algül H, Tuveson DA, Gress TM. Stromal Biology and Therapy in Pancreatic Cancer: A Changing Paradigm. *Gut* (2015) 64(9):1476–84. doi: 10.1136/gutjnl-2015-309304
- Torphy RJ, Zhu Y, Schulick RD. Immunotherapy for Pancreatic Cancer: Barriers and Breakthroughs. Ann Gastroenterol Surg (2018) 2(4):274–81. doi: 10.1002/ags3.12176

which can increase the sensitivity to immunotherapy (39). Generally, we defined TMB \geq 20 mutations/Mb as high TMB, TMB \leq 10 mutations/Mb as low TMB. Patients with a high TMB also have a better prognosis with immunotherapy. For example, a higher TMB was associated with better response in non-small cell lung cancer patients receiving pembrolizumab (40). A higher TMB had a clinical benefit in malignant melanoma patients receiving either ipilimumab or tremelimumab (41). Additionally, a case report revealed that combined antiangiogenic therapy and immunotherapy is effective for pancreatic cancer with high TMB (42). However, patients with pancreatic cancer generally have a low TMB in comparison to patients with other malignancies (43). Moreover, pancreatic cancer is a tumor with low immunogenicity, which is attributed to low TMB (36).

CONCLUSION

In this case, the effect of immunotherapy combined with chemotherapy seems to be very effective. We also established a hypothesis that the SMAD4 and TSC2 mutations improved the efficacy of immunotherapy, prolonging the survival of patients. However, very few studies have investigated the relationship between SMAD4 mutation and immunotherapy in pancreatic cancer. Thus, more studies and clinical trials are needed.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

AUTHOR CONTRIBUTIONS

SZ is the guarantor. YY wrote the manuscript. All authors read, provided feedback, and approved the final version.

FUNDING

The National Natural Science Foundation of China (81372660), Zhejiang Province Public Welfare Technology Application Research Project (2017C33200), and Laboratory of Clinical Cancer Pharmacology and Toxicology Research of Zhejiang Province.

 Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and Activity of Anti-PD-L1 Antibody in Patients With Advanced Cancer. N Engl J Med (2012) 366(26):2455–65. doi: 10.1056/NEJMoa 1200694

Royal RE, Levy C, Turner K, Mathur A, Hughes M, Kammula US, et al. Phase 2 Trial of Single Agent Ipilimumab (Anti-CTLA-4) for Locally Advanced or Metastatic Pancreatic Adenocarcinoma. *J Immunother* (2010) 33(8):828–33. doi: 10.1097/CJI.0b013e3181eec14c

- O'Reilly EM, Oh DY, Dhani N, Renouf DJ, Lee MA, Sun W, et al. Durvalumab With or Without Tremelimumab for Patients With Metastatic Pancreatic Ductal Adenocarcinoma: A Phase 2 Randomized Clinical Trial. *JAMA Oncol* (2019) 5(10):1431–8. doi: 10.1001/jamaoncol.2019.1588
- Wainberg ZA, Hochster HS, Kim EJ, George B, Kaylan A, Chiorean EG, et al. Open-Label, Phase I Study of Nivolumab Combined With Nab-Paclitaxel Plus Gemcitabine in Advanced Pancreatic Cancer. *Clin Cancer Res* (2020) 26 (18):4814–22. doi: 10.1158/1078-0432.CCR-20-0099
- O'Hara MH, O'Reilly EM, Varadhachary G, Wolff RA, Wainberg ZA, Ko AH, et al. CD40 Agonistic Monoclonal Antibody APX005M (Sotigalimab) and Chemotherapy, With or Without Nivolumab, for the Treatment of Metastatic Pancreatic Adenocarcinoma: An Open-Label, Multicentre, Phase 1b Study. *Lancet Oncol* (2021) 22(1):118–31. doi: 10.1016/S1470-2045(20)30532-5
- Renouf DJ, Knox JJ, Kavan P, Jonker D, Welch S, Couture F, et al. The Canadian Cancer Trials Group PA.7 Trial: Results of a Randomized Phase II Study of Gemcitabine (GEM) and Nab-Paclitaxel (Nab-P) vs. GEM, Nab-P, Durvalumb (D) and Tremelimumab (T) as First Line Therapy in Metastatic Pancreatic Ductal Adenocarcinoma (Mpdac) [Conference Presentation]. *ESMO 2020 Virtual* (2020) 31(Supplement 4):S1195. doi: 10.1016/j.annonc.2020.08.2300
- Samstein RM, Lee CH, Shoushtari AN, Hellmann MD, Shen R, Janjigian Y, et al. Tumor Mutational Load Predicts Survival After Immunotherapy Across Multiple Cancer Types. *Nat Genet* (2019) 51(2):202–6. doi: 10.1038/s41588-018-0312-8
- Zhang L, Mai W, Jiang W, Geng Q. Sintilimab: A Promising Anti-Tumor Pd-1 Antibody. Front Oncol (2020) 10:594558. doi: 10.3389/fonc.2020.594558
- Ngo P, Shanshal M, Rojan A. Immunotherapy in Pancreatic Cancer and the Importance of Tumour Testing. *BMJ Case Rep* (2020) 13(7):e235774. doi: 10.1136/bcr-2020-235774
- Dang HX, White BS, Foltz SM, Miller CA, Luo J, Fields RC, et al. Clonevol: Clonal Ordering and Visualization in Cancer Sequencing. *Ann Oncol* (2017) 28(12):3076–82. doi: 10.1093/annonc/mdx517
- Cingarlini S, Bonomi M, Corbo V, Scarpa A, Tortora G. Profiling Mtor Pathway in Neuroendocrine Tumors. *Target Oncol* (2012) 7(3):183–8. doi: 10.1007/s11523-012-0226-9
- Liu HJ, Lizotte PH, Du H, Speranza MC, Lam HC, Vaughan S, et al. TSC2-Deficient Tumors Have Evidence of T Cell Exhaustion and Respond to Anti-PD-1/Anti-CTLA-4 Immunotherapy. JCI Insight (2018) 3(8):e98674. doi: 10.1172/jci.insight.98674
- Shull MM, Ormsby I, Kier AB, Pawlowski S, Diebold RJ, Yin M, et al. Targeted Disruption of the Mouse Transforming Growth Factor-Beta 1 Gene Results in Multifocal Inflammatory Disease. *Nature* (1992) 359(6397):693–9. doi: 10.1038/359693a0
- Wang Y, Chu J, Yi P, Dong W, Saultz J, Wang Y, et al. SMAD4 Promotes TGF-β-Independent NK Cell Homeostasis and Maturation and Antitumor Immunity. J Clin Invest (2018) 128(11):5123–36. doi: 10.1172/JCI121227
- Principe DR, Doll JA, Bauer J, Jung B, Munshi HG, Bartholin L, et al. Tgf-β: Duality of Function Between Tumor Prevention and Carcinogenesis. J Natl Cancer Inst (2014) 106(2):djt369. doi: 10.1093/jnci/djt369
- Massagué J. How Cells Read TGF-Beta Signals. Nat Rev Mol Cell Biol (2000) 1 (3):169–78. doi: 10.1038/35043051
- Dardare J, Witz A, Merlin JL, Gilson P, Harlé A. SMAD4 and the Tgfβ Pathway in Patients With Pancreatic Ductal Adenocarcinoma. Int J Mol Sci (2020) 21(10):3534. doi: 10.3390/ijms21103534
- Park BV, Freeman ZT, Ghasemzadeh A, Chattergoon MA, Rutebemberwa A, Steigner J, et al. Tgfβ1-Mediated SMAD3 Enhances PD-1 Expression on Antigen-Specific T Cells in Cancer. *Cancer Discov* (2016) 6(12):1366–81. doi: 10.1158/2159-8290.CD-15-1347
- Thomas DA, Massagué J. TGF-Beta Directly Targets Cytotoxic T Cell Functions During Tumor Evasion of Immune Surveillance. *Cancer Cell* (2005) 8(5):369–80. doi: 10.1016/j.ccr.2005.10.012
- 22. Wagner M, Kleeff J, Lopez ME, Bockman I, Massaqué J, Korc M. Transfection of the Type I TGF-Beta Receptor Restores TGF-Beta Responsiveness in Pancreatic Cancer. Int J Cancer (1998) 78(2):255–60. doi: 10.1002/(sici) 1097-0215(19981005)78:2<255::aid-ijc21>3.0.co;2-8
- Bailey P, Chang DK, Nones K, Johns AL, Patch AM, Gingras MC, et al. Genomic Analyses Identify Molecular Subtypes of Pancreatic Cancer. *Nature* (2016) 531(7592):47–52. doi: 10.1038/nature16965
- 24. Derynck R, Jarrett JA, Chen EY, Eaton DH, Bell JR, Assoian RK, et al. Human Transforming Growth Factor-Beta Complementary DNA Sequence and

Expression in Normal and Transformed Cells. Nature (1985) 316 (6030):701-5. doi: 10.1038/316701a0

- Yamada S, Fujii T, Shimoyama Y, Kanda M, Nakayama G, Sugimoto H, et al. SMAD4 Expression Predicts Local Spread and Treatment Failure in Resected Pancreatic Cancer. *Pancreas* (2015) 44(4):660–4. doi: 10.1097/MPA. 0000000000000315
- Hsieh YY, Liu TP, Chou CJ, Chen HY, Lee KH, Yang PM. Integration of Bioinformatics Resources Reveals the Therapeutic Benefits of Gemcitabine and Cell Cycle Intervention in SMAD4-Deleted Pancreatic Ductal Adenocarcinoma. *Genes (Basel)* (2019) 10(10):766. doi: 10.3390/genes10100766
- 27. Duda DG, Sunamura M, Lefter LP, Furukawa T, Yokoyama T, Yatsuoka T, et al. Restoration of SMAD4 by Gene Therapy Reverses the Invasive Phenotype in Pancreatic Adenocarcinoma Cells. *Oncogene* (2003) 22 (44):6857–64. doi: 10.1038/sj.onc.1206751
- Biankin AV, Morey AL, Lee CS, Kench JG, Biankin SA, Hook HC, et al. DPC4/ Smad4 Expression and Outcome in Pancreatic Ductal Adenocarcinoma. J Clin Oncol (2002) 20(23):4531–42. doi: 10.1200/JCO.2002.12.063
- Ungefroren H. Blockade of TGF-β Signaling: A Potential Target for Cancer Immunotherapy? *Expert Opin Ther Targets* (2019) 23(8):679–93. doi: 10.1080/ 14728222.2019.1636034
- Larson C, Oronsky B, Carter CA, Oronsky A, Knox SJ, Sher D, et al. TGF-Beta: A Master Immune Regulator. *Expert Opin Ther Targets* (2020) 24 (5):427–38. doi: 10.1080/14728222.2020.1744568
- Tauriello DVF, Palomo-Ponce S, Stork D, Berenguer-Llergo A, Badia-Ramentol J, Iglesias M, et al. Tgfβ Drives Immune Evasion in Genetically Reconstituted Colon Cancer Metastasis. *Nature* (2018) 554(7693):538–43. doi: 10.1038/nature25492
- 32. Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T, Minato N. Involvement of PD-L1 on Tumor Cells in the Escape From Host Immune System and Tumor Immunotherapy by PD-L1 Blockade. *Proc Natl Acad Sci U S A* (2002) 99 (19):12293–7. doi: 10.1073/pnas.192461099
- 33. Yang B, Ren H, Yu G. Case Report: Squamous Cell Carcinoma of Pancreas With High PD-L1 Expression: A Rare Presentation. *Front Oncol* (2021) 11:680398. doi: 10.3389/fonc.2021.680398
- Pico de Coaña Y, Choudhury A, Kiessling R. Checkpoint Blockade for Cancer Therapy: Revitalizing a Suppressed Immune System. *Trends Mol Med* (2015) 21(8):482–91. doi: 10.1016/j.molmed.2015.05.005
- Sideras K, Braat H, Kwekkeboom J, van Eijck CH, Peppelenbosch MP, Sleijfer S, et al. Role of the Immune System in Pancreatic Cancer Progression and Immune Modulating Treatment Strategies. *Cancer Treat Rev* (2014) 40 (4):513–22. doi: 10.1016/j.ctrv.2013.11.005
- Schizas D, Charalampakis N, Kole C, Economopoulou P, Koustas E, Gkotsis E, et al. Immunotherapy for Pancreatic Cancer: A 2020 Update. *Cancer Treat Rev* (2020) 86:102016. doi: 10.1016/j.ctrv.2020.102016
- Zitvogel L, Galluzzi L, Smyth MJ, Kroemer G. Mechanism of Action of Conventional and Targeted Anticancer Therapies: Reinstating Immunosurveillance. *Immunity* (2013) 39(1):74-88. doi: 10.1016/ j.immuni.2013.06.014
- Bockorny B, Macarulla T, Semenisty V, Borazanci E, Feliu J, Ponz-Sarvise M, et al. Motixafortide and Pembrolizumab Combined to Nanoliposomal Irinotecan, Fluorouracil, and Folinic Acid in Metastatic Pancreatic Cancer: The COMBAT/KEYNOTE-202 Trial. *Clin Cancer Res* (2021) 27(18):5020–7. doi: 10.1158/1078-0432.CCR-21-0929
- Champiat S, Ferté C, Lebel-Binay S, Eggermont A, Soria JC. Exomics and Immunogenics: Bridging Mutational Load and Immune Checkpoints Efficacy. Oncoimmunology (2014) 3(1):e27817. doi: 10.4161/onci.27817
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer Immunology. Mutational Landscape Determines Sensitivity to PD-1 Blockade in Non-Small Cell Lung Cancer. *Science* (2015) 348(6230):124–8. doi: 10.1126/science.aaa1348
- Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, et al. Genetic Basis for Clinical Response to CTLA-4 Blockade in Melanoma. N Engl J Med (2014) 371(23):2189–99. doi: 10.1056/NEJMoa1406498
- 42. Chen M, Yang S, Fan L, Wu L, Chen R, Chang J, et al. Combined Antiangiogenic Therapy and Immunotherapy Is Effective for Pancreatic Cancer With Mismatch Repair Proficiency But High Tumor Mutation Burden: A Case Report. *Pancreas* (2019) 48(9):1232–6. doi: 10.1097/ MPA.000000000001398

 Chalmers ZR, Connelly CF, Fabrizio D, Gay L, Ali SM, Ennis R, et al. Analysis of 100,000 Human Cancer Genomes Reveals the Landscape of Tumor Mutational Burden. *Genome Med* (2017) 9(1):34. doi: 10.1186/s13073-017-0424-2

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Ye and Zheng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.