



## Complete Genome Sequence of the Secondary Alcohol-Utilizing Methanogen *Methanospirillum hungatei* Strain GP1

Leslie A. Day,<sup>a</sup> <sup>(D)</sup>Kyle C. Costa<sup>a</sup>

<sup>a</sup>Department of Plant and Microbial Biology, University of Minnesota, St. Paul, Minnesota, USA

**ABSTRACT** We report the complete genome sequence of *Methanospirillum hungatei* strain GP1 (DSM 1101). Strain GP1 oxidizes  $H_2$ , formate, and secondary alcohols as the substrates for methanogenesis. Members of the genus are model organisms used to study syntrophic growth with bacterial partners, but secondary alcohol metabolism remains poorly studied.

*Phatomatrix in the direction of ketone reduction at high partial pressures of H*<sub>2</sub> (6). While the mechanisms of H<sub>2</sub>- and formate-dependent growth are well understood for hydrogenotrophic methanogens (7), the basis of alcohol utilization is underexplored. We sequenced the genome of *M. hungatei* strain GP1 to better understand how electrons from secondary alcohols feed into methanogenes from secondary alcohols are output to be the mechanisms of H<sub>2</sub> and formate-dependent growth are well understood for hydrogenotrophic methanogens (7), the basis of alcohol utilization is underexplored. We sequenced the genome of *M. hungatei* strain GP1 to better understand how electrons from secondary alcohols feed into methanogens.

A liquid culture of *M. hungatei* strain GP1 was obtained from Deutsche Sammlung von Mikroorganismen und Zellkulturen (DSMZ). The material was transferred and propagated in liquid medium under an atmosphere of H<sub>2</sub>:CO<sub>2</sub> (80:20, 140 kPa) in DSMZ medium 119 with sludge fluid omitted and 0.4% tryptone, 0.4% peptone, and 0.2% Casamino Acids added. Genomic DNA was isolated from two separate cultures using the Qiagen blood and tissue kit and submitted to the Microbial Genome Sequencing Center (https://www.migscenter.com/). One DNA extraction was used for Nanopore sequencing (ligation sequencing kit; Oxford Nanopore), and a separate DNA extraction was used for Illumina library preparation (DNA prep kit; Illumina, Inc.) and sequencing. Illumina NextSeg 2000 sequencing (151-bp paired-end reads) generated 3,715,804 reads, totaling 541,571,161 bp. The Illumina reads were trimmed using Trim Galore v. 0.6.6 (https://github.com/FelixKrueger/TrimGalore) to remove adapters and for quality control. Nanopore sequencing was performed on a MinION R9 flow cell and generated 128,156 reads, totaling 640,042,040 bp, with a read  $N_{50}$  value of 23,336 bp. Guppy v. 4.2.2 (Oxford Nanopore) was used for base calling. De novo hybrid assembly and polishing of Illumina and Nanopore reads were performed using Unicycler v. 4.9 and resulted in a single circular chromosome of 3,393,136 bp with  $170 \times$  read coverage (8). The genomic G+C content is 42.2%. The assembly annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline v. 5.2 (9–11). All programs were run with default parameters.

The strain GP1 genome contains 3,139 protein coding genes and 70 RNA coding genes (genes for 55 tRNAs, 5 5S rRNAs, 4 16S rRNAs, 4 23S rRNAs, and 2 noncoding RNAs [ncRNAs]). There is a single putative coenzyme  $F_{420}$ -dependent secondary alcohol dehydrogenase that shares 79% amino acid identity with the alcohol dehydrogenase of *Methanoculleus* thermophilus strain TCI (4). Recent data suggest that methanogens from the order

Citation Day LA, Costa KC. 2021. Complete genome sequence of the secondary alcoholutilizing methanogen *Methanospirillum hungatei* strain GP1. Microbiol Resour Announc 10:e00708-21. https://doi.org/10.1128/MRA .00708-21.

Editor David A. Baltrus, University of Arizona Copyright © 2021 Day and Costa. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Kyle C. Costa, kcosta@umn.edu.

Received 16 July 2021 Accepted 26 July 2021 Published 12 August 2021 *Methanomicrobiales* can oxidize  $F_{420}H_2$  and produce formate through the action of a reversible  $F_{420}$ -dependent formate dehydrogenase (12). It may be that alcohol oxidation ultimately leads to formate production before  $CO_2$  reduction occurs. The strain GP1 genome encodes six putative formate dehydrogenases and several hydrogenases, including  $F_{420}$ reducing hydrogenase, 5,10-methenyltetrahydromethanopterin hydrogenase (Hmd), and membrane-associated, energy-converting hydrogenases. To our knowledge, strain GP1 is the only member of the genus *Methanospirillum* known to possess Hmd.

**Data availability.** The GenBank accession number for the complete genome sequence is CP077107.1. The raw sequence reads are available in the Sequence Read Archive under the accession numbers SRX11342833 for the Nanopore reads and SRX11342832 for the Illumina paired-end reads.

## ACKNOWLEDGMENT

This work was supported by the U.S. Department of Energy, Office of Science, Basic Energy Sciences, under grant number DE-SC0019148.

## REFERENCES

- Morris BEL, Henneberger R, Huber H, Moissl-Eichinger C. 2013. Microbial syntrophy: interaction for the common good. FEMS Microbiol Rev 37: 384–406. https://doi.org/10.1111/1574-6976.12019.
- Patel GB, Roth LA, van den Berg L, Clark DS. 1976. Characterization of a strain of *Methanospirillum hungatii*. Can J Microbiol 22:1404–1410. https://doi.org/10 .1139/m76-208.
- Widdel F, Rouvière PE, Wolfe RS. 1988. Classification of secondary alcoholutilizing methanogens including a new thermophilic isolate. Arch Microbiol 150:477–481. https://doi.org/10.1007/BF00422290.
- Aufhammer SW, Warkentin E, Berk H, Shima S, Thauer RK, Ermler U. 2004. Coenzyme binding in F<sub>420</sub>-dependent secondary alcohol dehydrogenase, a member of the bacterial luciferase family. Structure 12:361–370. https:// doi.org/10.1016/j.str.2004.02.010.
- Bleicher K, Winter J. 1991. Purification and properties of F<sub>420<sup>-</sup></sub> and NADP<sup>+</sup>dependent alcohol dehydrogenases of *Methanogenium liminatans* and *Methanobacterium palustre*, specific for secondary alcohols. Eur J Biochem 200:43–51. https://doi.org/10.1111/j.1432-1033.1991.tb21046.x.
- Bleicher K, Zellner G, Winter J. 1989. Growth of methanogens on cyclopentanol/CO<sub>2</sub> and specificity of alcohol dehydrogenase. FEMS Microbiol Lett 59: 307–312. https://doi.org/10.1111/j.1574-6968.1989.tb03130.x.
- Costa KC, Leigh JA. 2014. Metabolic versatility in methanogens. Curr Opin Biotechnol 29:70–75. https://doi.org/10.1016/j.copbio.2014.02.012.

- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. Nucleic Acids Res 49: D1020–D1028. https://doi.org/10.1093/nar/gkaa1105.
- Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. Nucleic Acids Res 46:D851–D860. https://doi.org/10.1093/ nar/gkx1068.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/ nar/gkw569.
- 12. Abdul Halim MF, Day LA, Costa KC. 2021. Formate dependent heterodisulfide reduction in a *Methanomicrobiales* archaeon. Appl Environ Microbiol 87:e02698-20. https://doi.org/10.1128/AEM.02698-20.