Plastic wrap combined with alcohol wiping is an effective method of preventing bacterial colonization on mobile phones

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Abstract

Introduction: Using mobile phones for communication in emergency departments is a common practice; however, several studies have demonstrated that they may act as vectors for bacteria and viruses. This study evaluated the effectiveness of plastic wrapping in decreasing bacterial contamination on mobile phone surfaces.

Method: We used culture dishes and a luminometer to detect bacterial colonies and contamination on the phone surfaces.

Result: Our experiment showed that bacterial colonies exist on mobile phones before and after work. We found that wiping with 75% alcohol sanitizers effectively reduces the number of colonies on either a mobile phone or a temporary plastic covering. In addition, we found that bacterial colonies do not contaminate or adhere to plastic wrap any easier than to mobile phones.

Conclusion: These results demonstrated the effectiveness of plastic wrap for protecting mobile phone surfaces against bacterial colonization. In addition, applying a layer of plastic wrap protects the phone from potential damage due to the alcohol.

Abbreviation: COVID-19 = coronavirus disease 2019.

Keywords: alcohol, bacteria, infection prevention, mobile phone, plastic wrap

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The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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1. Introduction

Nosocomial infection prevention is a major challenge for emergency department. Patients in the emergency room may not only be infected while receiving treatment, but may infect other patients or hospital staff due to their own diseases. Bacteria and viruses can be transmitted by airborne droplet nuclei, large particle droplets, or direct contact with patients and their immediate environment.^[1]

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There are many recommendations for the control and prevention of hospital infections, including environmental hygiene, hand hygiene, antibiotic stewardship, and vaccinations, etc.^[2,3] Among them, hand hygiene is the simplest method, and many studies have confirmed that frequent washing can effectively control and prevent infections.^[4,5]

Using mobile phones for communication is a common practice in emergency departments; however, several studies have demonstrated that they may act as vectors for bacteria and viruses.^[6–9] Therefore, some hospital staff members began wrapping their mobile phones in plastic because they thought this would decrease the chances of contamination. This study evaluated the effectiveness of plastic wrapping in decreasing bacterial contamination on mobile phone surfaces.

2. Method

We utilized the mobile phones of 6 people working in the emergency department with 2 people per group. When the first 2 people worked together, they randomly decided who would use a phone with a plastic wrap. When the next 2 people worked together for the next phase of the experiment, the mobile phone



was replaced and enclosed in a new plastic wrap package. A total of 4 phases were conducted for each group, that is, twice with plastic wraps and twice without.

We recorded the bacterial colonization on the mobile phone surfaces at various time points (Fig. 1). We used cotton swabs to collect the bacteria on the phone after each 6-hour work shift. We also used sterilized cotton swabs to scratch the smart phone surfaces and used these to create agar plates. After a 48-hour incubation period in a 37 °C aerobic environment, we counted the bacterial colonies growing on the surface of the Mueller-Hinton agars. To reduce the chance of bias, the same person collected the swab and wrapped the mobile phone. The practical steps for this procedure are shown in Figure 1.

After using culture dishes to detect the cultured bacterial colonies, we used a 3 M Clean-Trace NG luminometer (Taipei, Taiwan) to detect colonies on the phone surfaces. This luminometer is used with Clean-Trace adenosine triphosphate surface tests and water tests to determine the level of contamination in a sample, and results are available in seconds. Adenosine triphosphate is a standard measurement of biological residue and an effective marker to assess the hygienic status of an environmental surface. The 3 M Clean-Trace Luminometer cell counts are measured by levels of adenosine triphosphate.^[10] The steps and times of the experiment are identical to the culture dishes, as is the number of experiments.

The statistical methods employed included Wilcoxon signed rank tests and Mann–Whitney *U* tests to analyze whether there were notable differences before and after alcohol disinfection. After a work shift, the differences in the degree of bacterial colonies or contamination on the surface of the mobile phones before and after wrapping them in plastic were recorded. This research does not involve the collection of any medical records and body fluids related to the human body. It is only to detect the bacteria colony on the mobile phone. The institutional review board of the Kaohsiung Medical University Hospital waived the need for ethical approval for this study.

3. Results

The results collected from the culture dishes and luminometer recordings are shown in the appendix file. In this experiment, the initial time point represented the condition of the mobile phone prior to work. It was clearly shown that bacteria on the surface of the mobile phones were brought into the hospital from home.

When we compared the first and second time points by means of Wilcoxon signed rank tests, we found that 75% alcohol sanitizers effectively reduced the bacterial colonies on mobile phone surfaces (P < .05; Table 1). We compared the fourth and fifth time points and found the same result. In other words, with or without the plastic wraps, a 75% alcohol solution effectively reduced the colonies (P < .05; Table 1). At the third time point of the experiment, the recently placed plastic phone wraps were almost sterile, and they were made even cleaner after disinfection with the alcohol sanitizer.

We then compared the data of the mobile phones with no plastic wrap at the sixth time point to the fourth time point with

Table 1

The	effect of alcohol	disinfection	on the	surface	of mobile	phone
and	plastic wrap.					

	Before alcohol	After alcohol	P value			
Time point I and II (n = 24) (The mobile phone surface)						
Culture dish	11.46±17.118	0.21 ± 0.658 (24)	P<.0005			
Luminometer	828.42±720.093	29.37 ± 14.331 (24)	P<.0005			
Time point IV and V	(n=12) (The plastic wraction of the plastic wractic wraction of the plastic wractic wractic wractic	ap on the surface of the r	mobile phone)			
Culture dish	7.75±7.956	0.17 ± 0.389	P=.003			
Luminometer	425.25 ± 335.726	22.83 ± 8.601	P=.002			

Table 2

Comparison of the number of bacterial colonies and contamination on mobile phones with and without plastic wrap.

	Experimental group with plastic wrap (n=12)	Control group without plastic wrap ($n = 12$)	P value
Time point I (Before work)			
Culture dish	10.75 ± 14.937	12.17 ± 19.711	P = .817
Luminometer	827.17 ± 792.252	829.67±675.679	P = .817
Time point II (Before work	post alcohol disinfection)		
Culture dish	0.25 ± 0.866	0.17 ± 0.389	P = .615
Luminometer	29.25 ± 13.778	29.50 ± 15.477	P = .773
Time point IV and VI (After	r work)		
Culture dish	7.75 ± 7.956	8.42±17.686	P = .383
Luminometer	425.25 ± 335.726	617.50 ± 472.748	P = .248
Time point VI (After work,	remove plastic wrap)		
Culture dish	0.08 ± 0.289	8.42±17.686	P<.0005
Luminometer	33.67±17.778	617.50 ± 472.748	P<.0005

plastic wrap using a Mann–Whitney *U* test. After the work shift, we found that there were colonies attached to the surfaces of both the mobile phones and the plastic wraps. There were no obvious differences between the 2 groups (P > .05; Table 2). Finally, we compared the mobile phones with and without plastic wraps at the sixth time point. We found that the plastic wraps significantly reduced the number of bacterial colonies on the mobile phone surfaces (P < .05; Table 2).

4. Discussion

Bacteria and viruses can be spread in three main ways: droplets, contact exposure and airborne. The common pathogens transmitted by droplets include *Hemophilus influenzae*, Group A *Streptococcus*, *Bordetella pertussis*, respiratory viruses, seasonal influenza and meningococcal disease. As part of standard precautions, wearing a surgical mask can provide adequate droplet protection for the health care professional.^[11]

As for contact exposure-related transmission, according to previous studies, patients infected or colonized with multidrugresistant organisms can be found in the emergency department. The pathogens transmitted in this manner include methicillinresistant *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Enterobacteriaceae*, vancomycin-resistant *Enterococcus*, vancomycin-resistant *Enterococcus*, *Acinetobacter baumannii* and other enteric pathogens. Contact prevention measures require the use of protective clothing and gloves during patient care to prevent healthcare personnel from being infected by these pathogens and spreading them to other patients.

As for airborne transmission, the pathogens which can potentially be transmitted in the emergency department such as tuberculosis, measles, varicella, influenza, and even severe acute respiratory distress syndrome. Appropriate protection to prevent droplet being infected by nuclei in the air requires the use of N95 or electric purifying air respirators. After the coronavirus disease 2019 (COVID-19) broke out in Wuhan, China, in December 2019, it quickly spread worldwide.^[12] The first COVID-19 case in Taiwan was confirmed on January 21, 2020.^[13] Several strategies for infection prevention in hospitals were suggested, such as limiting the number of simultaneous visitors, drive-through pharmacies for long-term medication control patients, new equipment and methods for testing patients suspected of being infected with COVID-19, and denying hospital admission when a patient had either previous contact or travel history and was presenting signs of fever or upper respiratory infection. Hand hygiene using alcohol-based hand sanitizers is practiced worldwide to prevent COVID-19 cross-transmission, and it has been one of the most effective, simplest and cheapest procedures.^[14]

In today's world, electronic devices like mobile phones, laptops, and desktop computers are essential parts of daily life. Although people are taught to wash their hands frequently and to maintain good overall hygiene, they may not clean these devices (e.g., touch screens, keyboards) as often as they should. According to previous research, bacteria may not only collect and live on keyboards^[6] but also on the surfaces of mobile phones.^[7] The most common pathogens found on mobile phones include Coagulase-negative Staphylococcus, S. aureus, Micrococcus species, Enterobacter species, Escherichia coli, Klebsiella species and Enterococcus species. This includes not only pathogens but also bacteria that are resistant to many drugs such as methicillin-resistant S. aureus.^[8] In addition, viruses may also be present on mobile phones. Viruses previously found on mobile phones include influenza viruses, noroviruses, rotaviruses, and respiratory syncytial viruses.^[15,16] The transmission of infections via mobile phones is not limited to medical care institutions because phones are ubiquitous in every community and have proven to be a possible vehicle for transmission.^[17]

Although previous research has found that 70% alcohol wipes can effectively reduce the amount of colonies and further reduce the risk of nosocomial infections,^[18] alcohol can cause damage to their phones, which may account for people being reluctant to adopt this method.

Based on the results of our experiment, bacterial colonies may be present on mobile phones before and after medical personnel use them at work. Wiping with 75% alcohol sanitizers can effectively reduce the number of colonies on either a mobile phone or its temporary plastic covering. Plastic wrap is cheap and easy to replace. A mobile phone wrapped with plastic can be fully disinfected with alcohol while avoiding having the alcohol contact the phone itself. In addition, we found that bacterial colonies do not contaminate or adhere to plastic wrap any easier than to mobile phones.

The results of the 2 experimental methods (luminometer and culture dish) demonstrated the effectiveness of plastic wrap for protecting mobile phones. The 2 methods have advantages and disadvantages. Culture dishes are very cost effective, but they require incubation and take a long time to produce results. Luminometers produce results much faster but are more expensive.

5. Limitations

Our research has several limitations, such as the small sample, the difference in personal hygiene habits between participants, the differences in workload and the amount of contact with patients, and the size of mobile phone brands among other possible factors that may affect results. To mitigate potential bias, the sampling, plastic wrapping and calculation of the number of colonies on wraps and phones were all conducted by the same person.

6. Conclusion

To summarize the results of our experiment, we believe that encasing mobile phones in plastic wrap is an effective method of preventing them from becoming vectors for bacterial transmission.

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