

## Review Article

# LncRNAs in Stem Cells

**Shanshan Hu and Ge Shan**

*School of Life Sciences and CAS Key Laboratory of Brain Function and Disease, University of Science and Technology of China, Hefei, Anhui 230027, China*

Correspondence should be addressed to Ge Shan; [shange@ustc.edu.cn](mailto:shange@ustc.edu.cn)

Received 5 September 2015; Accepted 3 December 2015

Academic Editor: Yujing Li

Copyright © 2016 S. Hu and G. Shan. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Noncoding RNAs are critical regulatory factors in essentially all forms of life. Stem cells occupy a special position in cell biology and Biomedicine, and emerging results show that multiple ncRNAs play essential roles in stem cells. We discuss some of the known ncRNAs in stem cells such as embryonic stem cells, induced pluripotent stem cells, mesenchymal stem cells, adult stem cells, and cancer stem cells with a focus on long ncRNAs. Roles and functional mechanisms of these lncRNAs are summarized, and insights into current and future studies are presented.

## 1. Introduction

Less than 2% of human genome is composed of protein-coding genes, but transcription is widespread and corresponding to more than ~80% of the genome [1, 2]. With the development of deep sequencing and bioinformatics, a large number of noncoding RNAs (ncRNAs) are identified in eukaryotic cells [3, 4]. ncRNAs greater than 200 nucleotides are grouped as long noncoding RNAs. Majority of lncRNAs are intergenic transcripts and sense or antisense to other transcripts. Similar to mRNAs, most lncRNAs are Pol II transcripts which have a poly-A tail and 5' cap. But compared to protein-coding genes, lncRNAs show lower evolutionary conservation and lower expression level. lncRNAs are predominantly localized in nucleus [5–7]. lncRNAs have been associated with significant roles in diverse biological events such as chromatin modifications, transcriptional regulation, and posttranscriptional regulation [8–10].

Stem cells are undifferentiated cells that can either self-renew or differentiate into specialized cells. Stem cells are of great interest to both basic and biomedical research due to their unique properties in cell biology [11–13]. In recent years, researchers are paying more and more attention to roles of ncRNAs in stem cells.

In this review, we will discuss some of the known roles of ncRNAs in stem cells, including embryonic stem cells (ESCs), induced pluripotent stem cells (iPSCs), mesenchymal stem

cells (MSCs), adult stem cells, and cancer stem cells. Table 1 gives a brief summary to key properties of each stem cell type. The focus is on lncRNAs, and for roles of small ncRNAs such as microRNAs and piRNAs in stem cells, readers are referred to other articles [14–17].

## 2. LncRNAs in ESCs

Mohamed et al. performed a genome-wide screen to identify lncRNAs transcriptionally regulated by Oct4 and Nanog in mouse ESCs (mESCs) by combining the mESC transcriptome with the chromatin immunoprecipitation genomic location maps of Oct4 and Nanog [18]. Two conserved lncRNAs AK028326 and AK141205 were further confirmed to be directly activated by Oct4 or suppressed by Nanog, respectively. In fact, AK028326 might be a coactivator of Oct4. Later, a systematic survey for lincRNAs, which are a subclass of lncRNAs transcribed from intergenic regions of coding genes, was conducted with several murine cell lines including ESCs [19]. More than a thousand lincRNAs were identified, and many of them might possess putative functions in the pluripotency or proliferation of ESCs. Transcription of some lincRNAs was also regulated by key transcription factors such as Sox2, Oct4, and Nanog in ESCs.

In human ESCs (hESCs), multiple ESC specific lncRNAs were identified with microarray assays [20]. Some of these

TABLE 1: Key properties of each stem cell type.

Stem cell	Key properties
Embryonic stem cell	Derived from the blastocyst stage early mammalian embryos and has the ability to differentiate into any cell type and propagate
Induced pluripotent stem cell	A type of pluripotent stem cell that can be generated directly from adult cells; classic four specific genes encoding transcription factors (Oct4, Sox2, cMyc, and Klf) could convert adult cells into iPSCs, holding great promise in the field of regenerative medicine because of indefinite propagation
Mesenchymal stem cell	Multipotent stromal cells that can differentiate into a variety of cell types including osteoblasts, chondrocytes, myocytes, and adipocytes
Adult stem cell	Undifferentiated cells, which are found throughout the body after development, can divide or self-renew indefinitely, and generate all the cell types of the organ from which they originate
Cancer stem cell	Cancer cells that possess the ability to give rise to all cell types found in particular cancer samples and can generate tumors through the stem cell processes of self-renewal and differentiation into multiple cell types

lncRNAs were involved in pluripotency maintenance and interacted with SOX2 and SUZ12 (a subunit of PRC2 complex). A subset of the identified lncRNAs was involved in neurogenesis of ESCs, and these lncRNAs potentially played neurogenic roles via physical interaction with important epigenetic factors such as REST and SUZ12. One of the lncRNAs identified in this study, rhabdomyosarcoma 2-associated transcript (RMST), was further investigated in a later study [21, 22]. RMST interacted directly with and served as a transcriptional coregulator of SOX2 to modulate the transcription of a number of SOX2 downstream genes. Recently, the long terminal repeats of human specific endogenous retrovirus subfamily H (HERVH), which is transposable elements expressed preferentially in hESCs, was found to function as nuclear lncRNAs with associations with OCT4, coactivators such as p300 and Mediator subunits [23].

A recent report showed that pluripotency-associated transcription factor SOX2 has an overlapping long non-coding transcript SOX2OT, which was highly expressed in embryonic stem cells and downregulated upon the induction of differentiation [24]. SOX2OT had a potential role in modulating pluripotency through the regulation of SOX2 expression.

Another lncRNA, linc-RoR might function as microRNA sponge to prevent mRNA of some key transcription factors in hESCs from microRNA mediated regulation [25]. Thus, this lncRNA played a role in ESC maintenance and differentiation in the cytoplasm.

These results implicated that lncRNAs could modulate the pluripotency and differentiation of ESCs via intervening the transcriptional and epigenetic regulatory networks in the nucleus or tuning the microRNA functions in the cytoplasm. Other cytoplasmic and nuclear roles of lncRNAs in ESCs are waiting for further investigations.

### 3. LncRNAs in iPSCs

The conversion of lineage-committed cells to iPSCs is a process called reprogramming. In a 2010 paper, Rinn Lab characterized the transcriptional reorganization of lncRNAs and identified a vast number of lncRNAs associating

with reprogramming of human iPSCs (hiPSCs) [26]. These lncRNAs were overexpressed in both hESCs and hiPSCs but showed an elevated level in iPSCs compared to ESCs, suggesting that their activation might promote the iPSCs. One of the lncRNAs named lincRNA-RoR for “regulator of reprogramming” was regulated by pluripotency transcription factors such as OCT4, SOX2, and NANOG. Knockdown of lincRNA-RoR resulted in an inhibition in reprogramming. Interestingly, linc-RoR was later found to function as a competing endogenous RNA in hESCs as already discussed.

By analyzing single-cell transcriptomes during somatic cell reprogramming, more than 400 hundred lncRNAs were characterized that were dynamically expressed at defined stages of reprogramming [27]. During reprogramming, sets of relevant lncRNAs were activated to modulate signaling pathways, repress lineage-specific genes, and regulate metabolic gene expression.

The reprogramming process is accompanied by global epigenetic remodeling [28]. The lncRNA *Xist* responsible for X chromosome inactivation (XCI) has been investigated as a classic epigenetic regulator in the context of reprogramming [29, 30]. Female mESCs have low *Xist* expression and two active X chromosomes. Successfully reprogrammed female murine iPSCs (miPSCs) have undergone X chromosome reactivation and have the same feature in *Xist* expression as mESCs. Upon differentiation, female mESCs and miPSCs initiate *Xist* expression and XCI. Surprisingly, hESCs and hiPSCs can be divided into three classes based on their different states of *Xist* expression. Class I hESCs and hiPSCs behave just like mESCs and miPSCs. Class II human pluripotent stem cells express *Xist* and exhibit random XCI even before differentiation. Class III cells have lost *XIST* expression but have already initiated XCI, regardless of whether they are in the undifferentiated or differentiating conditions [31].

It seems that lncRNAs play essential roles in the reprogramming process. A better understanding in lncRNAs and more broadly ncRNAs in the context of iPSCs would certainly benefit future biomedical research aiming to utilize iPSCs in the clinic.

#### 4. lncRNAs in MSCs and the Mesodermal Lineage

Mesenchyme is connective tissue derived from the mesoderm during animal development. MSCs are first identified in the bone marrow (BM) stroma. BM MSCs provide microenvironment for hematopoietic stem cells and can also differentiate into various mesodermal lineages. Later, MSCs are found in many tissues such as placenta, umbilical cord blood, adipose tissue, muscle, corneal stroma, and the dental pulp of deciduous baby teeth. These MSCs with different origins all possess property to differentiate into mesodermal lineages. Thus, MSCs are an important source of multipotent cells with great potential of clinic applications. lncRNAs in MSCs have significant regulatory roles and potentially can be used as biomarkers for disease or therapeutic targets. Wang et al. performed differential expression profiles of lncRNAs and mRNAs of undifferentiated versus chondrodifferentiated human BM MSCs using microarray in 2015 [32]. Some of the identified lncRNAs could be important regulators in chondrogenic differentiation. The same group identified lncRNAs with differential expression and putative functions during the osteogenic differentiation of human BM MSCs [33]. A total of 1,206 differentially expressed lncRNAs were identified, and among them 687 were upregulated and 519 downregulated, more than twofold.

Some lncRNAs are functionally required during adipogenesis [34]. Brown adipose tissue (BAT) and white adipose tissue (WAT) are differentiated from MSCs in adipose tissues. Sun et al. performed RNA sequencing and identified 175 lncRNAs in total from *in vitro* cultured brown and white preadipocytes, *in vitro* differentiated mature brown and white adipocytes, and primary brown and white mature adipocytes directly isolated from mice [34]. These lncRNAs were up- or downregulated during differentiation of both brown and white adipocytes. Activation of some transcriptional factors and cofactors such as peroxisome proliferator-activated receptor  $\gamma$  (PPAR $\gamma$ ) and CCAAT/enhancer-binding protein  $\alpha$  (CEBP $\alpha$ ) is the hallmark of brown adipocyte differentiation. These lncRNAs have promoters with binding of PPAR $\gamma$  and/or CEBP $\alpha$  key transcription factors. RNAi-mediated loss-of-function identified the top 20 lncRNAs that may be involved in the proper differentiation of adipocyte precursors. Later in 2014, substantially more lncRNAs were identified and may be functionally required during proper adipogenesis [35]. Zhao and colleagues identified inducible lncRNAs involved in brown fat development and thermogenesis using transcriptional profiling in adipose tissues and cells during brown adipocyte differentiation [36]. Twenty-one lncRNAs were found to be enriched in BAT, and among them AK038898 significantly impaired brown adipogenesis upon RNAi knocking down. This lncRNA was then renamed brown fat lncRNA 1 (Blnc1). The expression of Blnc1 was highly induced during brown and beige adipogenesis. The induction was correlated with the stimulation of the thermogenic gene program and in consistency with the expression of key thermogenic markers such as Ucp1 in adipose tissues. Blnc1 is

primarily localized in the nucleus and forms a ribonucleoprotein complex with EBF2, which is a member of the EBF family of transcription factors that has been implicated in the control of adipogenesis [37]. Blnc1 itself is also a target of EBF2, thereby forming a feed-forward regulatory loop to regulate adipocyte gene expression. Knowing the distinct roles and functional mechanisms of lncRNAs in adipogenesis have great values in developing new approaches to fight obesity and other related pathological consequences.

lncRNAs also regulate cardiomyocyte differentiation. Zhu et al. identified multiple lncRNAs involved in the development of the lateral mesoderm and in the differentiation of cardiomyocytes with P19 cells, which were isolated from embryo-derived mouse teratocarcinoma and could differentiate into cardiomyocytes [38]. Forty differentially expressed lncRNAs, 28 upregulated and 12 downregulated, were identified using microarrays. In another research, Matkovich and colleagues used genome-wide sequencing and improved bioinformatics to measure dynamic lncRNA regulation during the transition between embryo and adult mouse hearts and identified numerous cardiac-specific lncRNAs [39]. Analyses indicated a broad role of regulated cardiac lncRNAs as modulators of key cardiac transcriptional pathways. Two lncRNAs, Fendrr and Braveheart, were later found to be involved in the development of the lateral mesoderm in the heart and the differentiation of cardiac myocytes, respectively [40, 41]. Fendrr was discovered in a survey of differentially expressed lncRNAs in six different tissues dissected from early somite-stage of mouse embryos by using RNA-seq and ChIP-seq [40]. Knocking out Fendrr by homologous recombination was embryonic lethal. The ChIP data showed that Fendrr interacts directly with the PRC2 and/or TrxG/MLL complexes in mouse embryos, indicating that it acts as a modulator of chromatin signatures defining gene activity in embryonic development. Braveheart also plays roles as an epigenetic factor [41]. This lncRNA is required for the progression of nascent mesoderm towards a cardiac fate by interacting with SUZ12, a component of PRC2 to alter cardiomyocyte differentiation and retain the cardiac phenotype in neonatal cardiomyocyte [41]. Cardiovascular diseases are currently the main cause of morbidity worldwide, and researches on these lncRNAs are critical to achieving a better understanding of heart development and designing new ways for the diagnosis and treatment of cardiac related diseases.

Several important lncRNAs play critical roles in myogenesis [42]. linc-MD1 is a muscle-specific long noncoding RNA expressed during early phases of muscle differentiation. It promotes the switch of muscle differentiation from early to later stages by acting as a miR-133 and miR-135 sponge [43]. What is more interesting is that linc-MD1 is also the primary transcript of miR-133b [44]. The process of linc-MD1 as a pri-microRNA by Drosha is controlled by HuR. The binding of HuR to linc-MD1 suppresses Drosha cleavage of linc-MD1 and leads to accumulation of linc-MD1, which further inhibits the functions of miR-133 as a sponge so that this microRNA could not target the HuR mRNA. Thus, HuR and linc-MD1 form an elegant feed-forward positive

loop [45]. Another lncRNA YAM-1 affects myogenesis via epigenetic transcriptional activation [42]. The transcription factor Yin Yang 1 (YY1) regulates expression multiple genes in myoblasts, and YAM-1 is positively regulated by YY1. YAM-1 expression is downregulated during myogenesis and its expression inhibits myoblast differentiation. YAM-1 regulates the transcription of miR-715 in *cis* and miR-715 is a microRNA that targets Wnt7b, an important signaling modulator of myogenesis [42].

## 5. LncRNAs in Adult Stem Cells

A series of studies focused on lncRNAs in adult stem cells such as hematopoietic stem cells and adult neural stem cells. The milestone discovery of lincRNA HOTAIR in 2007 was carried out with primary human fibroblasts, which are a kind of somatic stem cell [45]. In this and following researches, HOTAIR was found to bind both the polycomb repressive complex 2 (PRC2) and the LSD1/CoREST/REST complex and thus to modulate histone modifications on target genes [45, 46]. The lncRNA ANCR (antidifferentiation noncoding RNA) was also characterized by epidermal progenitors versus differentiating cells [47]. Knocking down ANCR alone led to rapid expression of differentiation genes. ANCR was thought to be essential for the undifferentiated state, although the exact molecular mechanism requires further investigation [47]. On the other hand, an lncRNA called terminal differentiation-induced noncoding RNA (TINCR) promoted epidermal differentiation [48]. TINCR had a very low level in epidermal stem cells, but it was dramatically induced upon differentiation. As a cytoplasmic ncRNA, TINCR could interact with mRNAs of many differentiation genes such as KRT80 and RNA-binding protein STAU1 to mediate mRNA stabilization through binding TINCR box.

In a systematic analysis of adult neural stem cell lineage in the mouse subventricular zone for lncRNA with potential roles in adult neurogenesis, Ramos et al. identified two lncRNAs Six3os and Dlx1as playing crucial roles in the lineage specification of adult stem cells [49]. The same lab later characterized a neural-specific lncRNA named Pnky, which localized in the nucleus of human and mouse neural stem cells [50]. With knockdown of Pnky, neuronal differentiation increased both *in vitro* and *in vivo*. Pnky interacted with the splicing regulator PTBPI and thus regulated the expression and alternative splicing.

Recently, Luo et al. identified 159 unannotated lncRNAs enriched in hematopoietic stem cells with RNA sequencing. Knocking down two of these lncRNAs showed effects on self-renewal and lineage commitment of hematopoietic stem cells [51]. Interestingly, the genomic binding sites of one lncRNA overlapped significantly with binding sites of a key hematopoietic transcription factor E2A, indicating this lncRNA might be a cofactor of the transcription factor.

It is obvious that lncRNAs possess vital roles in a variety of somatic stem cells with diverse mechanisms. Considering the large number of lncRNAs identified, we still have a long

way to understand the full range of roles of these regulators in adult stem cells as well as in the other eukaryotic cells.

## 6. LncRNAs in Cancer Stem Cells

Cancer stem cells (CSCs) play critical roles in tumor initiation, progression, metastasis, chemoresistance, and recurrence [52, 53]. Some studies showed that lncRNAs might be involved in regulating stem cell signaling in CSCs.

A novel lncRNA HIF2PUT (hypoxia-inducible factor-2 $\alpha$  promoter upstream transcript) was identified in a kind of CSCs, the osteosarcoma stem cells [54]. HIF2PUT expression levels were positively correlated with its parent gene *HIF-2 $\alpha$*  in osteosarcoma tissues, indicating a regulatory role of HIF2PUT through HIF-2 $\alpha$ . Knocking down HIF2PUT enhanced the proliferation, migration, and self-renewal of osteosarcoma stem cells while overexpression of HIF2PUT decreased the proliferation, migration, and self-renewal [54]. In liver CSCs, lncTCF7 was identified, and it recruited the SWI/SNF complex and further activated the Wnt signaling to promote self-renewal of liver CSCs and tumor propagation [55].

The lncRNA MALAT-1 (metastasis-associated lung adenocarcinoma transcript 1) was found to be upregulated in CSCs, and high expression levels of this lncRNA positively correlated with the proportion of CSCs in pancreatic cancer cells [10, 56, 57]. MALAT-1 might serve as an oncogenic lncRNA in pancreatic cancer by promoting epithelial-mesenchymal transition and stimulating the expression of self-renewal factors such as Sox2.

An lncRNA H19 known for more than two decades also functions in cancer and cancer stem cells [58, 59]. H19 is an imprinting gene in both mice and human [60, 61]. H19 RNA was identified as a tumor suppressor and was involved in Wilms' tumor long time ago in early 1990s [62, 63]. Despite the fact that H19 is one of the first identified, most abundant and conserved ncRNAs in mammals, its actual physiological functions and functional mechanism have been elusive for a long time. In 2007 and then in 2012, compelling lines of evidence showed that H19 is the primary transcript of a microRNA miR-675 [64, 65]. Overexpression of miR-675 in embryonic and extraembryonic cell lines reduced proliferation. Just like the linc-MD1 (the primary transcript of miR-133b), the processing of H19 into miR-675 is also regulated by HuR. What makes H19 more intriguing is the finding that H19 could be sponge of let-7 and miR-106a (a miR-17-5p family member) [66, 67]. HOTAIR with high expression level was always involved in various cancers as well as CSCs via promoting tumorigenesis, cell proliferation, or tumor metastasis and could be a novel epigenetic molecular target for therapeutics [68, 69]. Linc00617 may be another potential therapeutic target for breast cancer because of functioning as a key regulator of EMT and promotes cancer progression and metastasis via activating the transcription of Sox2 [70].

Knowing the roles of lncRNAs in tumor cells and especially in CSCs would help the development of tumor



TABLE 2: Regulatory roles of lncRNAs in different stem cells.

lncRNA	Stem cell types	Regulations and functions
AK028326	ESCs	Transcriptionally regulated by Oct4 and Nanog
AK141205	ESCs	A coactivator of Oct4
RMST	ESCs	Involved in pluripotency maintenance and interacting with SOX2 and SUZ12
HERVH	ESCs	Function as nuclear lncRNAs with associations with OCT4, coactivators such as p300 and Mediator subunits
SOX2OT	ESCs	Modulate pluripotency through the regulation of SOX2 expression
Linc-RoR	ESCs, iPSCs	Function as microRNA sponge to prevent mRNA of some key transcription factors
Xist	ESCs, iPSCs	An epigenetic regulator of X chromosome inactivation
Blnc1	MSCs	Highly induced during brown and beige adipogenesis and form a feed-forward regulatory loop with EBF2 to regulate adipocyte gene expression
Fendrr	MSCs	Interact directly with the PRC2 and/or TrxG/MLL complexes and act as a modulator of chromatin signatures defining gene activity in embryonic development
Braveheart	MSCs	An epigenetic factor, required for the progression of nascent mesoderm towards a cardiac fate by interacting with SUZ12
Linc-MD1	MSCs	Promote the switch of muscle differentiation from early to later stages by acting as miR-133 and miR-135 sponge
YAM-1	MSCs	Affect myogenesis via epigenetic transcriptional activation
HOTAIR	ASCs, CSCs	Bind both the polycomb repressive complex 2 (PRC2) and the LSD1/CoREST/REST complex and thus modulate histone modifications on target genes
ANCR	ASCs	Essential for the undifferentiated state
TINCR	ASCs	Interact with mRNAs of many differentiation genes such as KRT80 and RNA-binding protein STAU1 to mediate mRNA stabilization through binding TINCR box
Six3os	ASCs	Play crucial roles in the lineage specification of adult stem cells
Dlx1as	ASCs	Play crucial roles in the lineage specification of adult stem cells
Pnky	ASCs	Interact with the splicing regulator PTBPI and regulate the expression and alternative splicing
HIF2PUT	CSCs	A regulatory role through HIF-2 $\alpha$
lncTCF7	CSCs	Recruit the SWI/SNF complex and further activated the Wnt signaling to promote self-renewal of liver CSCs and tumor propagation
MALAT-1	CSCs	Serve as an oncogenic lncRNA in pancreatic cancer by promoting epithelial-mesenchymal transition and stimulating the expression of self-renewal factors such as Sox2
H19	CSCs	Primary transcript of a microRNA miR-675; processing of H19 into miR-675 is also regulated by HuR
Linc00617	CSCs	Function as a key regulator of EMT and promote cancer progression and metastasis via activating the transcription of Sox2

diagnosis and therapeutics [71]. So far, lncRNA research in cancer and CSCs is just emerging, and more efforts are required to push the field forward. Table 2 lists lncRNAs, their regulatory roles, and stem cells where they exert functions.

## 7. Conclusions and Perspectives

lncRNAs and other ncRNAs play substantial roles in diverse stem cell types. Here we have summarized some of the lncRNAs in five classes of stem cells, although we are not trying to summarize all the known lncRNAs in all kinds of stem cells and have to miss a lot of researches about lncRNAs in multiple other somatic and germline stem cells. lncRNAs could participate in the biology of stem cells with diverse mechanisms by serving as transcriptional coactivator

or corepressor, modular scaffold for chromatin modifiers, factors of tuning splicing, primary transcripts of microRNAs, competing endogenous RNAs of microRNAs, and so on (Figure 1). We need to keep in mind that the functions and mechanisms of a vast majority of lncRNAs in stem cells remain unknown. The pervasive transcription of the genome and the growing inventory of ncRNAs with newly identified ncRNAs such as circRNAs and EIciRNAs also make a call for enormous future efforts to investigate ncRNAs in stem cells [72–77]. Nevertheless, the field of ncRNA research in stem cells has emerged and has been making progress towards a systematic understanding of stem cell biology. Researches in this area would eventually bring benefit to the understanding of lncRNAs and Biomedicine.

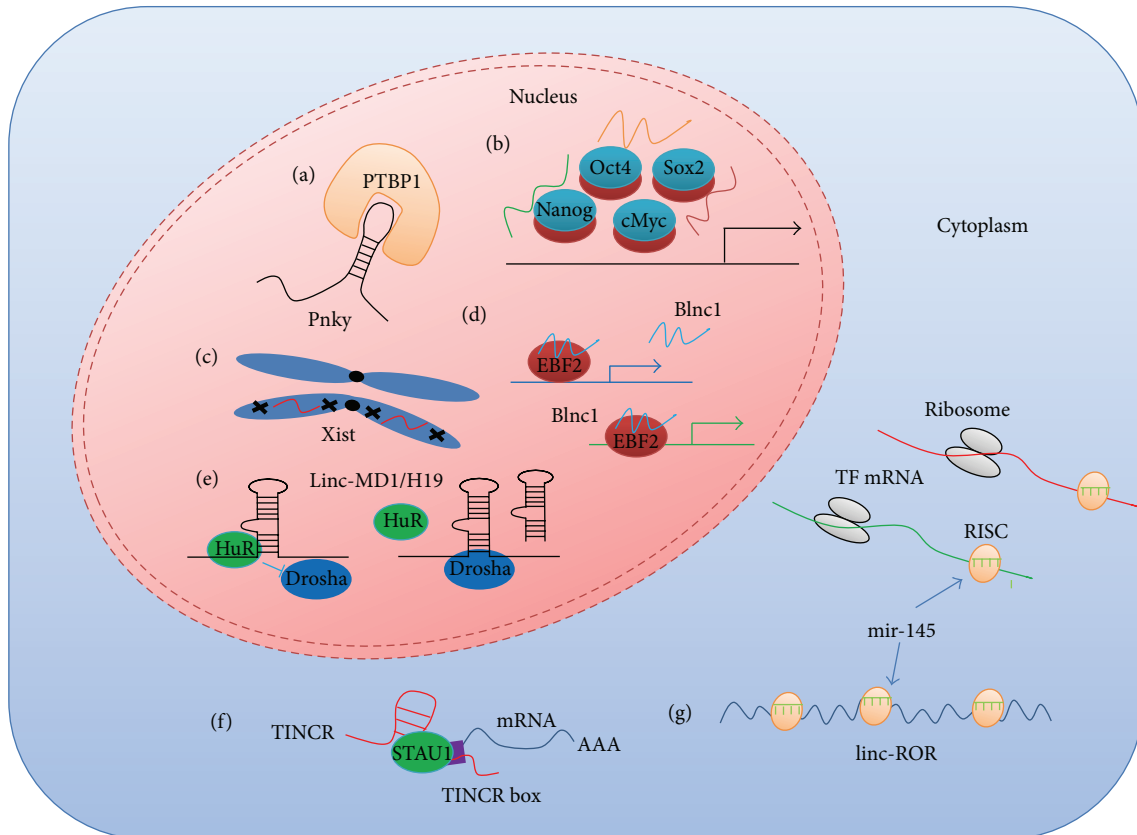


FIGURE 1: Some known functional mechanisms of lncRNAs in stem cells. (a) Pnky localizes to the nucleus and interacts with the mRNA splicing regulator PTBP1 to regulate splicing of key transcripts; (b) some lncRNAs serve as cofactors of key transcription factors (TF) to regulate gene transcription; (c) Xist as an epigenetic regulator of X chromosome inactivation plays roles in female ESCs and iPSCs; (d) Blnc1 is activated by EBF2 and functions as a coactivator of EBF2; (e) linc-MD1 is the primary transcript of miR-133b and H19 is the primary transcript of miR-675, and the Drosha processing of both is regulated by HuR; (f) TINCR interacts with STAUI and multiple mRNAs via an RNA motif called TINCR box; (g) linc-ROR functions as a miR-145 sponge.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

## References

- [1] International Human Genome Sequencing Consortium, "Finishing the euchromatic sequence of the human genome," *Nature*, vol. 431, no. 7011, pp. 931–945, 2004.
- [2] C. P. Ponting, P. L. Oliver, and W. Reik, "Evolution and functions of long noncoding RNAs," *Cell*, vol. 136, no. 4, pp. 629–641, 2009.
- [3] S. Geisler and J. Collier, "RNA in unexpected places: long non-coding RNA functions in diverse cellular contexts," *Nature Reviews Molecular Cell Biology*, vol. 14, no. 11, pp. 699–712, 2013.
- [4] I. Ulitsky and D. P. Bartel, "LincRNAs: genomics, evolution, and mechanisms," *Cell*, vol. 154, no. 1, pp. 26–46, 2013.
- [5] T. Ravasi, H. Suzuki, K. C. Pang et al., "Experimental validation of the regulated expression of large numbers of non-coding RNAs from the mouse genome," *Genome Research*, vol. 16, no. 1, pp. 11–19, 2006.
- [6] J. L. Rinn and H. Y. Chang, "Genome regulation by long noncoding RNAs," *Annual Review of Biochemistry*, vol. 81, pp. 145–166, 2012.
- [7] P. J. Batista and H. Y. Chang, "Long noncoding RNAs: cellular address codes in development and disease," *Cell*, vol. 152, no. 6, pp. 1298–1307, 2013.
- [8] J. T. Lee and M. S. Bartolomei, "X-inactivation, imprinting, and long noncoding RNAs in health and disease," *Cell*, vol. 152, no. 6, pp. 1308–1323, 2013.
- [9] R. A. Gupta, N. Shah, K. C. Wang et al., "Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis," *Nature*, vol. 464, no. 7291, pp. 1071–1076, 2010.
- [10] V. Tripathi, J. D. Ellis, Z. Shen et al., "The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation," *Molecular Cell*, vol. 39, no. 6, pp. 925–938, 2010.
- [11] I. L. Weissman, "Translating stem and progenitor cell biology to the clinic: barriers and opportunities," *Science*, vol. 287, no. 5457, pp. 1442–1446, 2000.
- [12] R. Jaenisch and R. Young, "Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming," *Cell*, vol. 132, no. 4, pp. 567–582, 2008.

- [13] F. H. Gage and S. Temple, "Neural stem cells: generating and regenerating the brain," *Neuron*, vol. 80, no. 3, pp. 588–601, 2013.
- [14] K. N. Ivey, A. Muth, J. Arnold et al., "MicroRNA regulation of cell lineages in mouse and human embryonic stem cells," *Cell Stem Cell*, vol. 2, no. 3, pp. 219–229, 2008.
- [15] K. N. Ivey and D. Srivastava, "MicroRNAs as regulators of differentiation and cell fate decisions," *Cell Stem Cell*, vol. 7, no. 1, pp. 36–41, 2010.
- [16] D. N. Cox, A. Chao, J. Baker, L. Chang, D. Qiao, and H. Lin, "A novel class of evolutionarily conserved genes defined by piwi are essential for stem cell self-renewal," *Genes and Development*, vol. 12, no. 23, pp. 3715–3727, 1998.
- [17] J. Gonzalez, H. Qi, N. Liu, and H. Lin, "Piwi is a key regulator of both somatic and germline stem cells in the *Drosophila* testis," *Cell Reports*, vol. 12, no. 1, pp. 150–161, 2015.
- [18] J. S. Mohamed, P. M. Gaughwin, B. Lim, P. Robson, and L. Lipovich, "Conserved long noncoding RNAs transcriptionally regulated by Oct4 and Nanog modulate pluripotency in mouse embryonic stem cells," *RNA*, vol. 16, no. 2, pp. 324–337, 2010.
- [19] M. Guttman, J. Donaghey, B. W. Carey et al., "LincRNAs act in the circuitry controlling pluripotency and differentiation," *Nature*, vol. 477, no. 7364, pp. 295–300, 2011.
- [20] S.-Y. Ng, R. Johnson, and L. W. Stanton, "Human long non-coding RNAs promote pluripotency and neuronal differentiation by association with chromatin modifiers and transcription factors," *The EMBO Journal*, vol. 31, no. 3, pp. 522–533, 2012.
- [21] S.-Y. Ng and L. W. Stanton, "Long non-coding RNAs in stem cell pluripotency," *Wiley Interdisciplinary Reviews: RNA*, vol. 4, no. 1, pp. 121–128, 2013.
- [22] S.-Y. Ng, G. K. Bogu, B. Soh, and L. W. Stanton, "The long noncoding RNA RMST interacts with SOX2 to regulate neurogenesis," *Molecular Cell*, vol. 51, no. 3, pp. 349–359, 2013.
- [23] X. Lu, F. Sachs, L. Ramsay et al., "The retrovirus HERVH is a long noncoding RNA required for human embryonic stem cell identity," *Nature Structural and Molecular Biology*, vol. 21, no. 4, pp. 423–425, 2014.
- [24] A. Shahryari, M. S. Jazi, N. M. Samaei, and S. J. Mowla, "Long non-coding RNA SOX2OT: expression signature, splicing patterns, and emerging roles in pluripotency and tumorigenesis," *Frontiers in Genetics*, vol. 6, article 196, 2015.
- [25] Y. Wang, Z. Xu, J. Jiang et al., "Endogenous miRNA sponge lincRNA-RoR regulates Oct4, Nanog, and Sox2 in human embryonic stem cell self-renewal," *Developmental Cell*, vol. 25, no. 1, pp. 69–80, 2013.
- [26] S. Loewer, M. N. Cabili, M. Guttman et al., "Large intergenic non-coding RNA-RoR modulates reprogramming of human induced pluripotent stem cells," *Nature Genetics*, vol. 42, no. 12, pp. 1113–1117, 2010.
- [27] D. H. Kim, G. K. Marinov, S. Pepke et al., "Single-cell transcriptome analysis reveals dynamic changes in lincRNA expression during reprogramming," *Cell Stem Cell*, vol. 16, no. 1, pp. 88–101, 2015.
- [28] N. Maherali, R. Sridharan, W. Xie et al., "Directly reprogrammed fibroblasts show global epigenetic remodeling and widespread tissue contribution," *Cell Stem Cell*, vol. 1, no. 1, pp. 55–70, 2007.
- [29] D. Lessing, M. C. Anguera, and J. T. Lee, "X chromosome inactivation and epigenetic responses to cellular reprogramming," *Annual Review of Genomics and Human Genetics*, vol. 14, pp. 85–110, 2013.
- [30] S. F. Briggs, A. A. Dominguez, S. L. Chavez, and R. A. Reijo Pera, "Single-cell XIST expression in human preimplantation embryos and newly reprogrammed female induced pluripotent stem cells," *STEM CELLS*, vol. 33, no. 6, pp. 1771–1781, 2015.
- [31] S. S. Silva, R. K. Rowntree, S. Mekhoubad, and J. T. Lee, "X-chromosome inactivation and epigenetic fluidity in human embryonic stem cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 12, pp. 4820–4825, 2008.
- [32] L. Wang, Z. Li, Z. Li, B. Yu, and Y. Wang, "Long noncoding RNAs expression signatures in chondrogenic differentiation of human bone marrow mesenchymal stem cells," *Biochemical and Biophysical Research Communications*, vol. 456, no. 1, pp. 459–464, 2015.
- [33] L. Wang, Y. Wang, Z. Li, Z. Li, and B. Yu, "Differential expression of long noncoding ribonucleic acids during osteogenic differentiation of human bone marrow mesenchymal stem cells," *International Orthopaedics*, vol. 39, no. 5, pp. 1013–1019, 2015.
- [34] L. Sun, L. A. Goff, C. Trapnell et al., "Long noncoding RNAs regulate adipogenesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 9, pp. 3387–3392, 2013.
- [35] J. Zhang, X. Cui, Y. Shen et al., "Distinct expression profiles of LncRNAs between brown adipose tissue and skeletal muscle," *Biochemical and Biophysical Research Communications*, vol. 443, no. 3, pp. 1028–1034, 2014.
- [36] X.-Y. Zhao, S. Li, G.-X. Wang, Q. Yu, and J. D. Lin, "A Long noncoding RNA transcriptional regulatory circuit drives thermogenic adipocyte differentiation," *Molecular Cell*, vol. 55, no. 3, pp. 372–382, 2014.
- [37] S. Elattar and A. Satyanarayana, "Can brown fat win the battle against white fat?" *Journal of Cellular Physiology*, vol. 230, no. 10, pp. 2311–2317, 2015.
- [38] S. Zhu, X. Hu, S. P. Han et al., "Differential expression profile of long non-coding RNAs during differentiation of cardiomyocytes," *International Journal of Medical Sciences*, vol. 11, no. 5, pp. 500–507, 2014.
- [39] S. J. Matkovich, J. R. Edwards, T. C. Grossenheider, C. de Guzman Strong, and G. W. Dorn II, "Epigenetic coordination of embryonic heart transcription by dynamically regulated long noncoding RNAs," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 111, no. 33, pp. 12264–12269, 2014.
- [40] P. Grote, L. Wittler, D. Hendrix et al., "The tissue-specific lincRNA Fendrr is an essential regulator of heart and body wall development in the mouse," *Developmental Cell*, vol. 24, no. 2, pp. 206–214, 2013.
- [41] C. A. Klattenhoff, J. C. Scheuermann, L. E. Surface et al., "Braveheart, a long noncoding RNA required for cardiovascular lineage commitment," *Cell*, vol. 152, no. 3, pp. 570–583, 2013.
- [42] L. Lu, K. Sun, X. Chen et al., "Genome-wide survey by ChIP-seq reveals YY1 regulation of lincRNAs in skeletal myogenesis," *The EMBO Journal*, vol. 32, no. 19, pp. 2575–2588, 2013.
- [43] M. Cesana, D. Cacchiarelli, I. Legnini et al., "A long noncoding RNA controls muscle differentiation by functioning as a competing endogenous RNA," *Cell*, vol. 147, no. 2, pp. 358–369, 2011.
- [44] I. Legnini, M. Morlando, A. Mangiacavalli, A. Fatica, and I. Bozzoni, "A feedforward regulatory loop between HuR and the long noncoding RNA linc-MD1 controls early phases of myogenesis," *Molecular Cell*, vol. 53, no. 3, pp. 506–514, 2014.

- [45] J. L. Rinn, M. Kertesz, J. K. Wang et al., "Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs," *Cell*, vol. 129, no. 7, pp. 1311–1323, 2007.
- [46] M.-C. Tsai, O. Manor, Y. Wan et al., "Long noncoding RNA as modular scaffold of histone modification complexes," *Science*, vol. 329, no. 5992, pp. 689–693, 2010.
- [47] M. Kretz, D. E. Webster, R. J. Flockhart et al., "Suppression of progenitor differentiation requires the long noncoding RNA ANCR," *Genes and Development*, vol. 26, no. 4, pp. 338–343, 2012.
- [48] M. Kretz, Z. Siprashvili, C. Chu et al., "Control of somatic tissue differentiation by the long non-coding RNA TINCR," *Nature*, vol. 493, no. 7431, pp. 231–235, 2013.
- [49] A. D. Ramos, A. Diaz, A. Nellore et al., "Integration of genome-wide approaches identifies lncRNAs of adult neural stem cells and their progeny in vivo," *Cell Stem Cell*, vol. 12, no. 5, pp. 616–628, 2013.
- [50] A. D. Ramos, R. E. Andersen, S. J. Liu et al., "The long noncoding RNA Pnky regulates neuronal differentiation of embryonic and postnatal neural stem cells," *Cell Stem Cell*, vol. 16, no. 4, pp. 439–447, 2015.
- [51] M. Luo, M. Jeong, D. Sun et al., "Long non-coding RNAs control hematopoietic stem cell function," *Cell Stem Cell*, vol. 16, no. 4, pp. 426–438, 2015.
- [52] F. Islam, V. Gopalan, R. A. Smith, and A. K.-Y. Lam, "Translational potential of cancer stem cells: a review of the detection of cancer stem cells and their roles in cancer recurrence and cancer treatment," *Experimental Cell Research*, vol. 335, no. 1, pp. 135–147, 2015.
- [53] G. A. Dashyan, V. F. Semiglazov, R. V. Donskikh, T. Y. Semiglazova, V. V. Vorotnikov, and V. S. Apollonova, "The role of breast cancer stem cells in metastasis," *Voprosy Onkologii*, vol. 61, no. 2, pp. 169–173, 2015.
- [54] Y. Wang, J. Yao, H. Meng et al., "A novel long non-coding RNA, hypoxia-inducible factor-2 $\alpha$  promoter upstream transcript, functions as an inhibitor of osteosarcoma stem cells in vitro," *Molecular Medicine Reports*, vol. 11, no. 4, pp. 2534–2540, 2015.
- [55] Y. Wang, L. He, Y. Du et al., "The long noncoding RNA lncTCF7 promotes self-renewal of human liver cancer stem cells through activation of Wnt signaling," *Cell Stem Cell*, vol. 16, no. 4, pp. 413–425, 2015.
- [56] F. Jiao, H. Hu, C. Yuan et al., "Elevated expression level of long noncoding RNA MALAT-1 facilitates cell growth, migration and invasion in pancreatic cancer," *Oncology Reports*, vol. 32, no. 6, pp. 2485–2492, 2014.
- [57] F. Jiao, H. Hu, T. Han et al., "Long noncoding RNA MALAT-1 enhances stem cell-like phenotypes in pancreatic cancer cells," *International Journal of Molecular Sciences*, vol. 16, no. 4, pp. 6677–6693, 2015.
- [58] V. Pachnis, A. Belayew, and S. M. Tilghman, "Locus unlinked to  $\alpha$ -fetoprotein under the control of the murine raf and Rif genes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 81, no. 17, pp. 5523–5527, 1984.
- [59] C. I. Brannan, E. C. Dees, R. S. Ingram, and S. M. Tilghman, "The product of the H19 gene may function as an RNA," *Molecular and Cellular Biology*, vol. 10, no. 1, pp. 28–36, 1990.
- [60] M. S. Bartolomei, S. Zemel, and S. M. Tilghman, "Parental imprinting of the mouse H19 gene," *Nature*, vol. 351, no. 6322, pp. 153–155, 1991.
- [61] Y. Zhang and B. Tycko, "Monoallelic expression of the human H19 gene," *Nature Genetics*, vol. 1, no. 1, pp. 40–44, 1992.
- [62] Y. Hao, T. Crenshaw, T. Moulton, E. Newcomb, and B. Tycko, "Tumour-suppressor activity of H19 RNA," *Nature*, vol. 365, no. 6448, pp. 764–767, 1993.
- [63] M. J. C. Steenman, S. Rainier, C. J. Dobry, P. Grundy, I. L. Horon, and A. P. Feinberg, "Loss of imprinting of *IGF2* is linked to reduced expression and abnormal methylation of *H19* in Wilms' tumour," *Nature Genetics*, vol. 7, no. 3, pp. 433–439, 1994.
- [64] X. Cai and B. R. Cullen, "The imprinted H19 noncoding RNA is a primary microRNA precursor," *RNA*, vol. 13, no. 3, pp. 313–316, 2007.
- [65] A. Keniry, D. Oxley, P. Monnier et al., "The H19 lincRNA is a developmental reservoir of miR-675 that suppresses growth and Igflr," *Nature Cell Biology*, vol. 14, no. 7, pp. 659–665, 2012.
- [66] A. N. Kallen, X.-B. Zhou, J. Xu et al., "The imprinted H19 lincRNA antagonizes let-7 microRNAs," *Molecular Cell*, vol. 52, no. 1, pp. 101–112, 2013.
- [67] J. Imig, A. Brunschweiler, A. Brümmer et al., "miR-CLIP capture of a miRNA targetome uncovers a lincRNA H19-miR-106a interaction," *Nature Chemical Biology*, vol. 11, no. 2, pp. 107–114, 2015.
- [68] Z. Yang, L. Zhou, L.-M. Wu et al., "Overexpression of long non-coding RNA HOTAIR predicts tumor recurrence in hepatocellular carcinoma patients following liver transplantation," *Annals of Surgical Oncology*, vol. 18, no. 5, pp. 1243–1250, 2011.
- [69] X. Li, Z. Wu, Q. Mei et al., "Long non-coding RNA HOTAIR, a driver of malignancy, predicts negative prognosis and exhibits oncogenic activity in oesophageal squamous cell carcinoma," *British Journal of Cancer*, vol. 109, no. 8, pp. 2266–2278, 2013.
- [70] H. Li, L. Zhu, L. Xu et al., "Long noncoding RNA linc00617 exhibits oncogenic activity in breast cancer," *Molecular Carcinogenesis*, 2015.
- [71] L. Chen, F. Rashid, A. Shah et al., "The isolation of an RNA aptamer targeting to p53 protein with single amino acid mutation," *Proceedings of the National Academy of Sciences*, vol. 112, no. 32, pp. 10002–10007, 2015.
- [72] J. Salzman, C. Gawad, P. L. Wang, N. Lacayo, and P. O. Brown, "Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types," *PLoS ONE*, vol. 7, no. 2, Article ID e30733, 2012.
- [73] S. Memczak, M. Jens, A. Elefsinioti et al., "Circular RNAs are a large class of animal RNAs with regulatory potency," *Nature*, vol. 495, no. 7441, pp. 333–338, 2013.
- [74] T. B. Hansen, T. I. Jensen, B. H. Clausen et al., "Natural RNA circles function as efficient microRNA sponges," *Nature*, vol. 495, no. 7441, pp. 384–388, 2013.
- [75] Z. Li, C. Huang, C. Bao et al., "Exon-intron circular RNAs regulate transcription in the nucleus," *Nature Structural and Molecular Biology*, vol. 22, no. 3, pp. 256–264, 2015.
- [76] H. Liu, X. Wang, H.-D. Wang et al., "*Escherichia coli* noncoding RNAs can affect gene expression and physiology of *Caenorhabditis elegans*," *Nature Communications*, vol. 3, article 1073, 2012.
- [77] S. Hu, J. Wu, L. Chen, and G. Shan, "Signals from noncoding RNAs: unconventional roles for conventional pol III transcripts," *International Journal of Biochemistry and Cell Biology*, vol. 44, no. 11, pp. 1847–1851, 2012.