

RESEARCH ARTICLE

Tangeretin Alleviates Cisplatin-Induced Acute Hepatic Injury in Rats: Targeting MAPKs and Apoptosis

Hany A. Omar^{1,2}, Wafaa R. Mohamed¹, Hany H. Arab³, El-Shaimaa A. Arafa^{1,4*}

1 Department of Pharmacology and Toxicology, Faculty of Pharmacy, Beni-Suef University, Beni-Suef 62514, Egypt, **2** Sharjah Institute for Medical Research, College of Pharmacy, University of Sharjah, Sharjah 27272, United Arab of Emirates, **3** Department of Biochemistry, Faculty of Pharmacy, Cairo University, Cairo 11562, Egypt, **4** Department of Pharmacology and Toxicology, Faculty of Pharmacy, Taif University, Taif 21974, Saudi Arabia

* elshimaa.arafa@pharm.bsu.edu.eg; shaimaarafa@yahoo.com



OPEN ACCESS

Citation: Omar HA, Mohamed WR, Arab HH, Arafa E-SA (2016) Tangeretin Alleviates Cisplatin-Induced Acute Hepatic Injury in Rats: Targeting MAPKs and Apoptosis. *PLoS ONE* 11(3): e0151649. doi:10.1371/journal.pone.0151649

Editor: Krishnendu Acharya, University of Calcutta, INDIA

Received: July 28, 2015

Accepted: March 2, 2016

Published: March 31, 2016

Copyright: © 2016 Omar et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper.

Funding: The current work was financially supported by grant number 1-434-2830 from the Deanship of Scientific Research, Taif University, Saudi Arabia.

Competing Interests: The authors have declared that no competing interests exist.

Abbreviations: ALT, alanine aminotransferase; AST, aspartate transaminase; GPx, glutathione peroxidase; GSH, glutathione; H&E, hematoxylin and eosin; IL-10, interleukin-10; MDA, malondialdehyde; NO, nitric oxide; NRF-2, nuclear factor erythroid 2-

Abstract

Despite its broad applications, cisplatin affords considerable nephro- and hepatotoxicity through triggering inflammatory and oxidative stress cascades. The aim of the current investigation was to study the possible protective effects of tangeretin on cisplatin-induced hepatotoxicity. The impact of tangeretin on cisplatin-evoked hepatic dysfunction and histopathologic changes along with oxidative stress, inflammatory and apoptotic biomarkers were investigated compared to silymarin. Tangeretin pre-treatment significantly improved liver function tests (ALT and AST), inhibited cisplatin-induced lipid profile aberrations (total cholesterol and triglycerides) and diminished histopathologic structural damage in liver tissues. Tangeretin also attenuated cisplatin-induced hepatic inflammatory events as indicated by suppression of tumor necrosis factor- α (TNF- α) and enhancement of interleukin-10 (IL-10). Meanwhile, it lowered malondialdehyde (MDA), nitric oxide (NO) and nuclear factor erythroid 2-related factor 2 (NRF-2) levels with restoration of glutathione (GSH), and glutathione peroxidase (GPx). Regarding mitogen-activated protein kinase (MAPK) pathway, tangeretin attenuated cisplatin-induced increase in phospho-p38, phospho-c-Jun N-terminal kinase (p-JNK) and phospho-extracellular signal-regulated kinase (p-ERK1/2) in liver tissues. In addition, tangeretin downregulated Bax expression with augmentation of Bcl-2 promoting liver cell survival. Our results highlight the protective effects of tangeretin against cisplatin-induced acute hepatic injury via the concerted modulation of inflammation, oxidative stress, MAPKs and apoptotic pathways.

Introduction

Cisplatin is one of the most widely used anticancer agents in the management of different malignancies. While 70–80% of patients respond to platinum treatment, such an initial effect is not robust, and results from a 5-year patient survival study revealed that the response is

related factor 2; ROS, reactive oxygen species; TNF- α , tumor necrosis factor- α .

only 15–20% due to the development of resistance [1]. The relapse of the disease and the emergence of resistance in initially responsive tumors occur within 18–24 months [2, 3]. The dose scale necessary to overcome even a small increase in cellular resistance can lead to severe cytotoxicity in normal cells, such as nephrotoxicity, hepatotoxicity and spermiotoxicity which radically limits the clinical usefulness of cisplatin-based therapy [4–6]. Cisplatin induces nephrotoxicity through multiple mechanisms, including hypoxia, the generation of free radicals, inflammation, and apoptosis with an increase in the pro-apoptotic protein Bax and a decrease in the anti-apoptotic protein Bcl-2 [7]. While, the mechanisms of cisplatin-induced hepatotoxicity are not fully understood [8], the link between oxidative stress and cisplatin toxicity was suggested in many experimental models [9, 10]. Several reports have implicated free radicals and reactive oxygen species (ROS) in cisplatin toxicity associated with an increase in lipid peroxidation (LPO), decreased levels of protein bound sulfhydryl groups and glutathione [11].

There is an increasing interest in the use of phytochemicals for evaluating their synergistic efficacy in combination with chemotherapeutic agents [12]. This is supported by the vast epidemiological data indicating the protective effect of vegetables and fruit intake rich in naturally occurring compounds against various diseases including cancer [13]. Flavonoids are dietary compounds that are widespread in fruits and vegetables. They have demonstrated a good potential as anticancer agents via their antiproliferative activity against human tumor cell lines [14], therefore, they were employed in cancer combination therapies for greater efficacy and safety [15].

Tangeretin, a citrus flavonoid concentrated in the peel of citrus fruits, has exhibited significant anti-inflammatory and antioxidant activities [16, 17]. Tangeretin oral bioavailability and safety have been reported previously [18, 19]. Notably, it has inhibited cancer cell proliferation in human cancer cell lines derived from squamous cell carcinoma, gliosarcoma, leukemia, melanoma, colorectal cancer, gastric carcinoma, lung carcinoma, breast carcinoma and oral cancer cells [20–22]. In a previous study from our laboratory, we have reported that pretreatment of cisplatin-resistant human ovarian cancer cells with tangeretin synergistically enhanced the growth inhibitory effects induced by low dose of cisplatin [23]. The combination markedly triggered apoptosis and arrested the cell cycle at G₂-M. Additionally, the phosphoinositide3-kinase (PI3K)/protein kinase B (Akt) survival pathway was effectively downregulated. The fact that tangeretin enhances the cytotoxic actions of cisplatin encouraged us to investigate whether tangeretin, as a bioactive flavonoid, can protect against cisplatin-evoked hepatotoxicity, a serious complication of cisplatin that may limit its therapeutic utility. To this end, the hepatoprotective effects of tangeretin were assessed by measuring its ability to antagonize cisplatin-induced inflammation, oxidative stress and apoptotic cell death in rat liver. The effect of tangeretin was compared to silymarin, a naturally occurring flavonoid used as a reference hepatoprotective agent. Several studies have reported its hepatoprotective effects in acute and chronic hepatic diseases [24–26]. Silymarin has been reported to protect against CCl₄- [27–29], acetaminophen- [30, 31], D-galactosamine- [32] and cisplatin- [33] induced liver injuries.

Material and Methods

Ethics Statement

The experimental protocol of the current study was approved by the Ethics Committee of the Faculty of Pharmacy, Beni-Suef University (Reference Number: 2014/A-32). The Guidelines for the Care and Use of Laboratory Animals declared by the US National Institute of Health were followed in all the experimental procedures.

Animals

Adult male Wistar rats weighing 160–200 g were obtained from the National Institute for Research, Cairo, Egypt. Rats were kept under controlled conditions of temperature ($22\pm 1^\circ\text{C}$), humidity ($60\pm 10\%$) and normal photoperiod (12–12 h light-dark cycles) with free access to a standard commercial pellet diet and water.

Materials

Cisplatin was purchased from Sigma-Aldrich, MD, USA, whereas tangeretin was purchased from Shaanxi Huike Botanical Development Co. (Xi'an, China). All other chemical reagents used in the study were of analytical grade (AR). Antibodies against Bax, Bcl-2 and β -actin were purchased from Santa Cruz Biotechnology (Santa Cruz, CA) while Erk1/2, p-Erk1/2, JNK, p-JNK, p38 and p-p38 antibodies were purchased from cell Signaling Technology (Beverly, MA).

Experimental design and treatment protocol

The animals were divided into 7 groups, each with eight rats, according to their experimental treatment as follows: (1) Normal (control) group: rats received only oral vehicle (2% Tween 80) p.o for 7 consecutive days and a single i.p injection of isotonic saline on the 2nd day of the experiment; (2) Silymarin group: rats received silymarin (100 mg/kg/day p.o) for 7 days and a single i.p injection of isotonic saline on the 2nd day of the experiment; (3) Tangeretin group: received tangeretin (100 mg/kg/day p.o) for 7 days and a single i.p injection of isotonic saline on the 2nd day of the experiment; (4) Cisplatin group: received oral vehicle for 7 days and a single dose of cisplatin (7.5 mg/kg i.p) on the 2nd day of the experiment; (5) Cisplatin-Silymarin group: rats received silymarin (100 mg/kg/day p.o) for 7 days and a single dose of cisplatin (7.5 mg/kg i.p) on the 2nd day of the experiment, 1 hour after the dose of silymarin; (6) Cisplatin-Tangeretin 50 group: rats received tangeretin (50 mg/kg/day p.o) for 7 days and a single dose of cisplatin (7.5 mg/kg i.p) on the 2nd day of the experiment day, 1 hour after the dose of tangeretin; (7) Cisplatin-Tangeretin 100 group: rats received tangeretin (100 mg/kg/day p.o) for 7 days and a single dose of cisplatin (7.5 mg/kg i.p) on the 2nd day of the experiment day, 1 hour after the dose of tangeretin.

The selected doses of cisplatin and tangeretin were chosen after performing preliminary experiments based on previous dose-response studies that have been reported to cause hepatotoxicity [33, 34] and marked antitumor effects in rats [17, 35].

Tissue collection and preparation

Animals were sacrificed under ether anesthesia on the last day of the experiment. For the different biochemical measurements, blood samples were collected and allowed to stand for 30 min at 37°C , and then centrifuged at $1000 \times g$ for 15 min at 4°C to separate serum and were stored at -70°C . The liver tissues were quickly harvested and one part of the liver tissue was instantly fixed in 10% phosphate buffered formaldehyde for histological and immunohistochemical studies. For the biochemical determinations, another part of liver was homogenized in lysis buffer containing protease and phosphatase inhibitor cocktails (Sigma-Aldrich, St. Louis, MD, USA).

Serum Biochemical Tests

The collected serum was used for the colorimetric estimation of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) as mentioned before [36] by measuring the amount of pyruvate or oxaloacetate produced by forming 2,4-dinitrophenylhydrazone. The

produced color was measured spectrophotometrically at 546 nm. Concentrations of triglycerides (TGs) and serum total cholesterol (TC) were measured by enzymatic colorimetric methods using commercial kits (Spinreact, Gerona, Spain).

Oxidative Stress Biomarkers. Liver homogenates were used to determine nitric oxide (NO) essentially as mentioned before, using Nitric Oxide ($\text{NO}_2^-/\text{NO}_3^-$) assay Kit (Assay Designs, Ann Arbor, MI, USA) [37]. Griess reagent was used for the quantitative colorimetric determination of NO levels as total nitrate/nitrite. To reduce nitrate to nitrite, vanadium trichloride and then Griess reagent was added and incubated at 37°C. The absorbance was measured after allowing the mixture to cool at 540 nm. Results were expressed as $\mu\text{mol/g}$ tissue.

Glutathione (GSH) levels in liver tissues were, measured as mentioned before [38]. Briefly, after the precipitation of liver proteins by 10% trichloroacetic acid, 10 mM DTNB (5,5'-dithio-bis 2-nitrobenzoic acid) solution was added to develop the color that was measured at 412 nm. Results were expressed as $\mu\text{mol/g}$ tissue.

Enzymatic and Transcriptional Antioxidant Status

The homogenates of liver tissues were utilized in the determination of glutathione peroxidase (GPx) activity using the corresponding assay kit (Sigma-Aldrich, St. Louis, MD, USA) according to the manufacturer's guidelines. The decrease in the absorbance of NADPH was determined at 340 nm. The amount of enzyme which oxidizes one μmol of NADPH per min at 25°C is defined as one unit of enzyme.

Lipid peroxides in liver tissues were expressed as malondialdehyde (MDA) and determined as mentioned before [39]. The reaction proceeded in trichloroacetic acid, the precipitate was removed and the absorbance was measured at 535 nm. Results were presented as nmol/g tissue. The nuclear factor erythroid 2-related factor (NRF-2) is a transcription factor that controls the redox homeostatic gene network. Hepatic tissue NRF-2 was estimated using Total NRF-2 Cell-Based Colorimetric ELISA Kit (Immuno Way Biotechnology, Newark, DE, USA) as directed by the manufacturer.

Histopathologic Examination

Samples of tissues were fixed in 10% neutral formalin for 24 h, and paraffin blocks were then processed for light microscopy examination. Slices of 4–5 μm were obtained from the prepared blocks and stained with hematoxylin-eosin (H&E). The preparations obtained were visualized using a Nikon microscopy at a magnification of 400 \times .

Immunohistochemistry of Bax and Bcl-2 expression

Antigen retrieval and immunohistochemistry was performed essentially as mentioned before [40]. In brief, paraffin-embedded tissue samples were rehydrated then blocked by 5% bovine serum albumin (BSA) in Tris buffered saline. The samples were then incubated overnight at 4°C with primary antibodies against Bax or Bcl-2 (Santa Cruz Biotechnology Inc, CA, USA). The slides were then washed and incubated with secondary antibodies. The sections were then washed and visualized using 3,3'-diaminobenzidine tetrahydrochloride (DAB Horseradish Peroxidase Substrate Kit, Vector Laboratories Inc, Burlingame, CA, USA). Counter staining with hematoxylin was used and the slides were observed under a light microscope (Leica Microsystems, Germany) by an experienced observer blinded to the identity of the sample.

Western blotting

Liver tissue homogenates were prepared for Western blotting as mentioned before [41]. Briefly, protein concentration in the tissue lysate was determined using DC protein assay kit (Bio Rad). Protein samples were then separated by SDS-PAGE (30 µg per lane) and transferred to a nitrocellulose membrane. The membrane were blocked using 5% (w/v) non-fat dry milk in Tris buffered saline-tween 20 (0.025 M Tris; 0.15 M NaCl; 0.05% Tween 20; pH 7.4), incubated overnight at 4°C with primary antibodies, rinsed, and then incubated with horseradish peroxidase-conjugated secondary antibodies (Santa Cruz Biotechnology, Santa Cruz, CA) for 1 h at room temperature before the detection using Super Signal West Pico chemiluminescent substrate (Pierce, Rockford, IL). β-actin was used as a loading control. Band density in intermediately exposed films was quantitated using ImageJ image processing (ImageJ, National Institutes of Health, USA).

Caspase 3/7 assay

Caspase-3/7 activities in liver tissue homogenate were measured using a Caspase-Glo assay kit (Promega, Madison, WI, USA) following the manufacturer’s instructions.

Statistical analysis

The data were expressed as mean ± SEM. Statistical analysis was done using one-way analysis of variance (ANOVA), followed by Tukey-Kramer post hoc multiple comparisons among treatment means. The analysis was done using SPSS program, version 17. Differences were considered significant at $p < 0.05$.

Results

Tangeretin pre-treatment improves liver function and inhibits cisplatin-induced aberrations in lipids profile

To assess the severity of cisplatin-induced liver injury, liver function tests were performed. Results revealed that the used dose of cisplatin triggered severe liver injury as indicated by elevated serum ALT and AST enzyme activities. Meanwhile, increased serum total cholesterol and triglyceride in cisplatin-treated group was observed showing an extensive damage to the liver tissues (Table 1).

Table 1. Tangeretin pre-treatment improved liver function and inhibited cisplatin-induced aberrations in lipids profile.

	ALT (U/l)	AST (U/l)	Triglyceride (mg/dl)	Total Cholesterol (mg/dl)
Normal	24.5 ± 1.7	112.3 ± 13.6	53.3 ± 3.5	72.3 ± 4.9
Silymarin (100 mg/kg)	25.5 ± 3.1	112.5 ± 12.5	57.5 ± 5.2	69.5 ± 8.2
Tangeretin (100 mg/kg)	27.5 ± 2.4	114.3 ± 17.2	62.2 ± 8.4	71.6 ± 4.2
Cisplatin	77.3 ± 7.0*	290.0 ± 38.3*	175.2 ± 19.7*	138.0 ± 12.8*
Cisplatin + Silymarin (100 mg/kg)	38.8 ± 4.6 [#]	136.2 ± 12.7 [#]	111.0 ± 12.5 [#]	89.6 ± 10.5 [#]
Cisplatin + Tangeretin (50 mg/kg)	68.0 ± 5.1	252.8 ± 36.1	159.7 ± 20.2	122.4 ± 5.4
Cisplatin + Tangeretin (100 mg/kg)	35.8 ± 4.2 [#]	132.2 ± 12.3 [#]	84.8 ± 13.1 [#]	82.5 ± 6.3 [#]

Values are mean ± SD of the mean (n = 4–5 independent values). Statistical analysis was carried out by using one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparisons test

* Significantly different from normal control at $p < 0.05$.

[#] Significantly different from cisplatin-treated (7.5 mg/kg) animals at $p < 0.05$.

doi:10.1371/journal.pone.0151649.t001

On the other hand, pretreatment with tangeretin for 1 week significantly alleviated liver damage as evidenced by a dose-dependent restoration of the normal liver functions and lipid profile. At the same time, the reference hepatoprotective, silymarin afforded remarkable hepatic protection against cisplatin-induced liver injury. These data suggest the hepatoprotective activity of tangeretin against the development of cisplatin-induced liver tissue damage which were similar to that obtained by silymarin (Table 1).

Tangeretin attenuates liver histopathological aberrations

We next explored the ability of tangeretin to protect against the histopathologic changes accompanying cisplatin-induced liver injury. Liver sections of control, silymarin and tangeretin groups showed the normal architecture of the liver tissues with minor Kupffer cells activation (Fig 1). On the other hand, the administration of cisplatin caused a severe liver injury (Table 2), as reflected by the dilatation and congestion of central vein and hepatic sinusoids with Kupffer cell activation and focal hepatic necrosis. This was accompanied with diffuse inflammatory cell infiltration. Pretreatment with tangeretin decreased the high pathologic changes, suggesting the attenuation of liver damage with the preservation of the liver wall architecture. These effects were closely analogous to those afforded by silymarin (Fig 1).

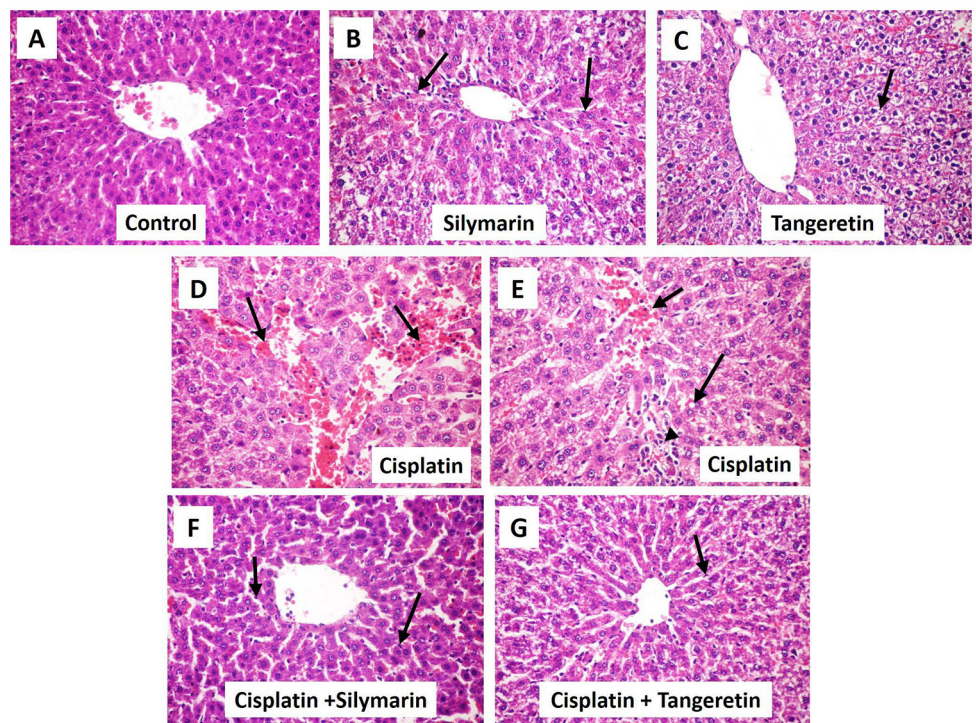


Fig 1. Tangeretin alleviates cisplatin-induced hepatic histopathologic injury in rats. Representative photomicrographs of sections from liver tissues stained by hematoxylin and eosin ($\times 400$ magnification). (A) Control rats receiving saline vehicle showed normal histological structure of the hepatic lobule. (B) Rats which received silymarin (100 mg/kg p.o.) showed normal hepatic histology with few Kupffer cell activation (arrow). (C) Rats which received tangeretin (100 mg/kg p.o.) showed slight hydropic degeneration of hepatocytes (arrow). (D) Cisplatin-treated group showed dilatation and congestion of central vein and hepatic sinusoids (arrow). (E) Liver of rat from cisplatin-treated group demonstrated congestion of central vein (short arrow), cytoplasmic vacuolization of hepatocytes (long arrow) and focal hepatic necrosis associated with inflammatory cell infiltration (arrow head). (F, G) Silymarin and tangeretin pre-treatment revealed attenuated morphological modifications with resolving Kupffer cell activation (arrow).

doi:10.1371/journal.pone.0151649.g001

Table 2. Liver microscopic damage.

Histopathological alterations	Control	Silymarin	Tangeretin	Cisplatin	Cisplatin+Silymarin	Cisplatin +Tangeretin
Kupffer cells activation	-	+	-	++	+	+
Congestion of central vein and hepatic sinusoids	-	-	-	+++	+	-
Cytoplasmic vacuolation of hepatocytes	-	-	-	++	-	+
Focal hepatic necrosis associated with inflammatory cell infiltration	-	-	-	++	-	-
Hydropic degeneration of hepatocytes	-	-	+	-	-	-

+++ Extensive

++ Moderate

+ Mild

-Nil

doi:10.1371/journal.pone.0151649.t002

Tangeretin attenuates cisplatin-induced inflammatory response and apoptosis in hepatic tissues

Cisplatin administration caused an inflammatory response as demonstrated by a 3.1 fold increase of hepatic TNF- α and a decline in IL-10 (32%) levels (Fig 2). Pretreatment with tangeretin at 100 mg/kg dose decreased the levels of TNF- α by 46% and slightly increased the IL-10. These results were similar to silymarin. In addition, the immunohistochemical detection of Bax and Bcl-2 revealed an extensive expression of Bax (Fig 3A–3E) and decreased Bcl-2 (Fig 3G–3L) in the hepatic tissues of rats treated with cisplatin; events which were markedly mitigated by administration of tangeretin. These results were confirmed by Western blot analysis of Bax and Bcl-2 (Fig 4A and 4B) and caspase 3/7 activation (Fig 4C) which indicated the ability of tangeretin to attenuate cisplatin-induced apoptosis in hepatic tissues. Analogous to silymarin, tangeretin counteracted these changes in favor of cell survival. Together, these

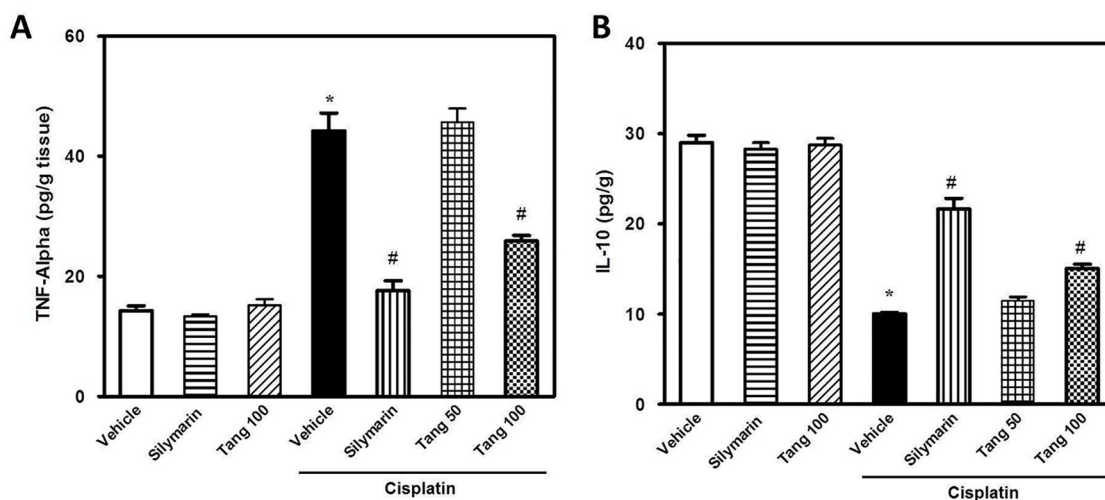


Fig 2. Tangeretin mitigates cisplatin-induced inflammatory response in liver tissues. The effect of tangeretin pretreatment on cisplatin-induced inflammatory response in liver tissues as indicated by the modulation of TNF- α (A) and IL-10 (B). Columns, mean; bars, \pm SEM (n = 8 independent values). Statistical analysis was carried out by using one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparisons test. *Significant difference from normal control (vehicle) group at p < 0.05, #Significant difference from cisplatin group at p < 0.05. Tang 50; tangeretin (50 mg/kg), Tang 100; tangeretin (100 mg/kg).

doi:10.1371/journal.pone.0151649.g002

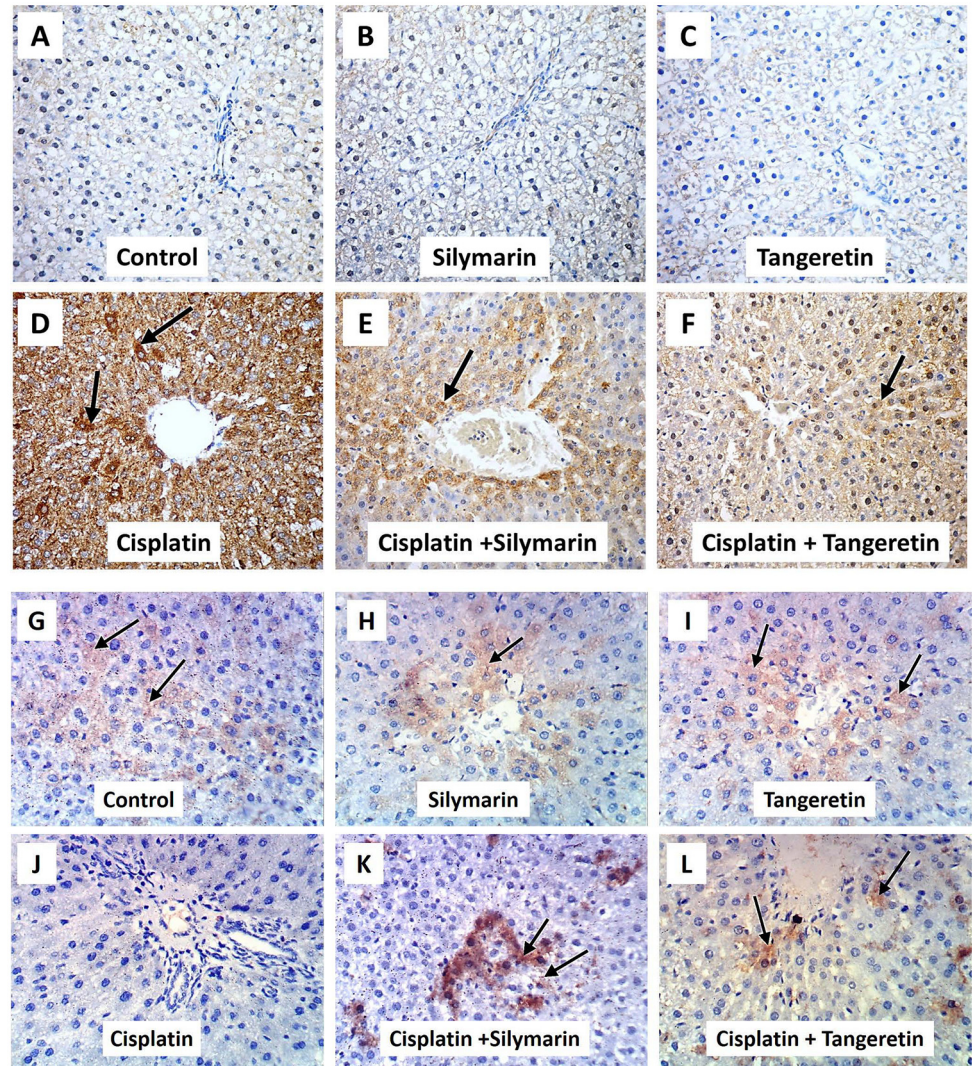


Fig 3. Tangeretin modulates cisplatin-induced protein expression of Bax and Bcl-2 in rat liver. Representative images for the immunohistochemical detection of Bax (A-F) and Bcl-2 (G-L) expression in liver tissues (arrows) (magnification: $\times 400$).

doi:10.1371/journal.pone.0151649.g003

observations suggested the modulation of inflammatory cytokines and suppression of apoptosis as crucial events in tangeretin protection against cisplatin-induced hepatic insult.

Tangeretin limits oxidative stress and boosts hepatic antioxidant defense

Cisplatin administration elicited hepatic oxidative stress as demonstrated by elevation of NO levels (195%) and decline of GSH content (29%) (Fig 5). In addition, cisplatin caused a marked decrease of hepatic antioxidant defenses as indicated by the decrease in GPx activity (29%) together with elevation of MDA (210%) and NRF-2 (375%) as compared to the control group (Fig 6). Pretreatment with tangeretin significantly protected against the oxidative stress as indicated by lowering of MDA, NO and NRF-2 in addition to the restoration of GSH and GPx, events which were analogous to the actions of silymarin. Together, these data suggest that the

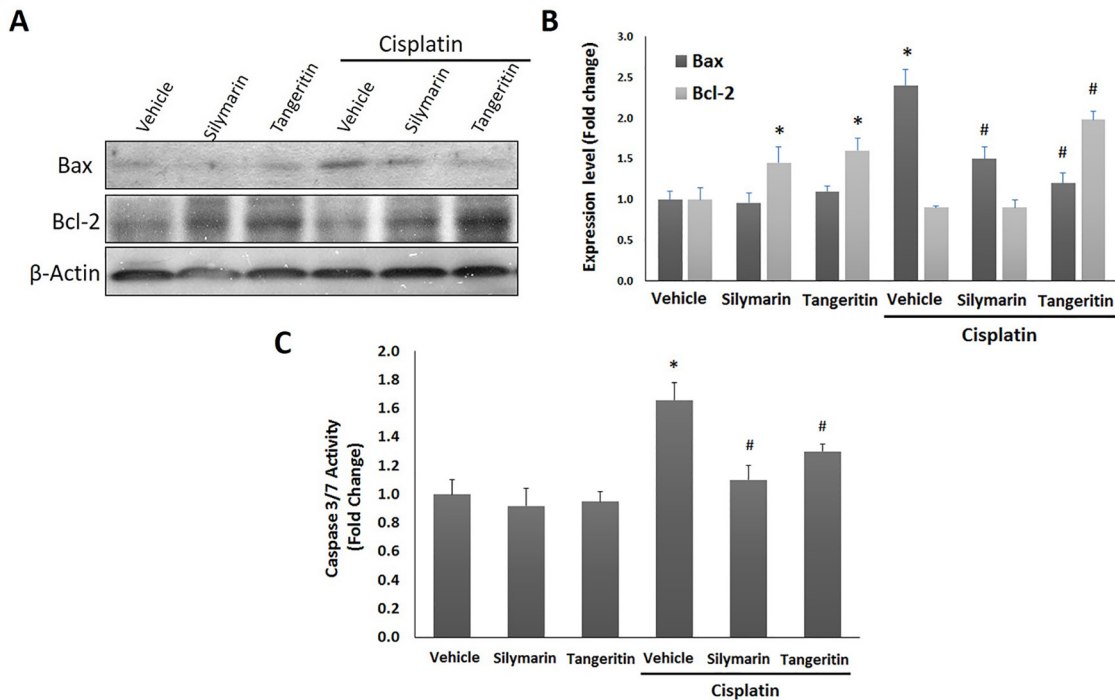


Fig 4. Tangeretin counteracts cisplatin-induced apoptotic changes of Bax and Bcl-2 in protein expression in rat liver. (A) Western blot analysis showed the expression levels of Bax and Bcl-2 in liver tissues after the indicated treatments. (B) Relative expression levels of Bax and Bcl-2. The amount of immunoblotted proteins was quantitated by densitometry and normalized to that of β -actin. Columns, mean; bars, \pm SD (n = 3 independent experiments). (C) Relative caspase 3/7 activity in liver tissues after the indicated treatments. Columns, mean; bars, \pm SD (n = 4 independent values). Statistical analysis was carried out by using one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparisons test. Silymarin (100 mg/kg); Tangeretin (100 mg/kg); Cisplatin (7.5 mg/kg). *Significant difference from normal control (vehicle) group at $p < 0.05$, #Significant difference from cisplatin group at $p < 0.05$.

doi:10.1371/journal.pone.0151649.g004

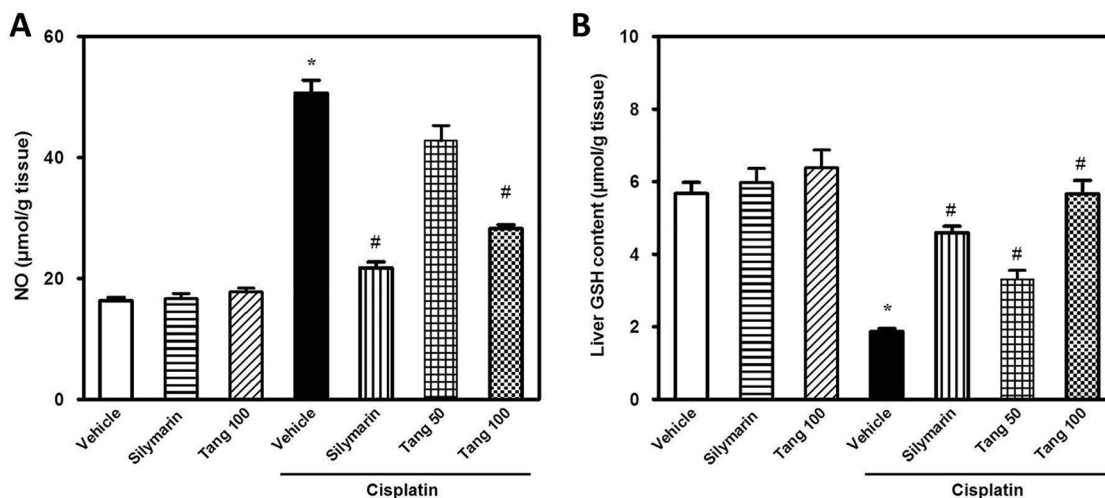


Fig 5. Tangeretin limits cisplatin-induced oxidative stress in liver tissues. The effect of tangeretin pretreatment on cisplatin-induced oxidative stress in liver tissues as indicated by the modulation of hepatic NO (A) and GSH content (B). Columns, mean; bars, \pm SEM (n = 8 independent values). Statistical analysis was carried out by using one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparisons test. *Significant difference from normal control (vehicle) group at $p < 0.05$, #Significant difference from cisplatin group at $p < 0.05$. Tang 50; tangeretin (50 mg/kg), Tang 100; tangeretin (100 mg/kg).

doi:10.1371/journal.pone.0151649.g005

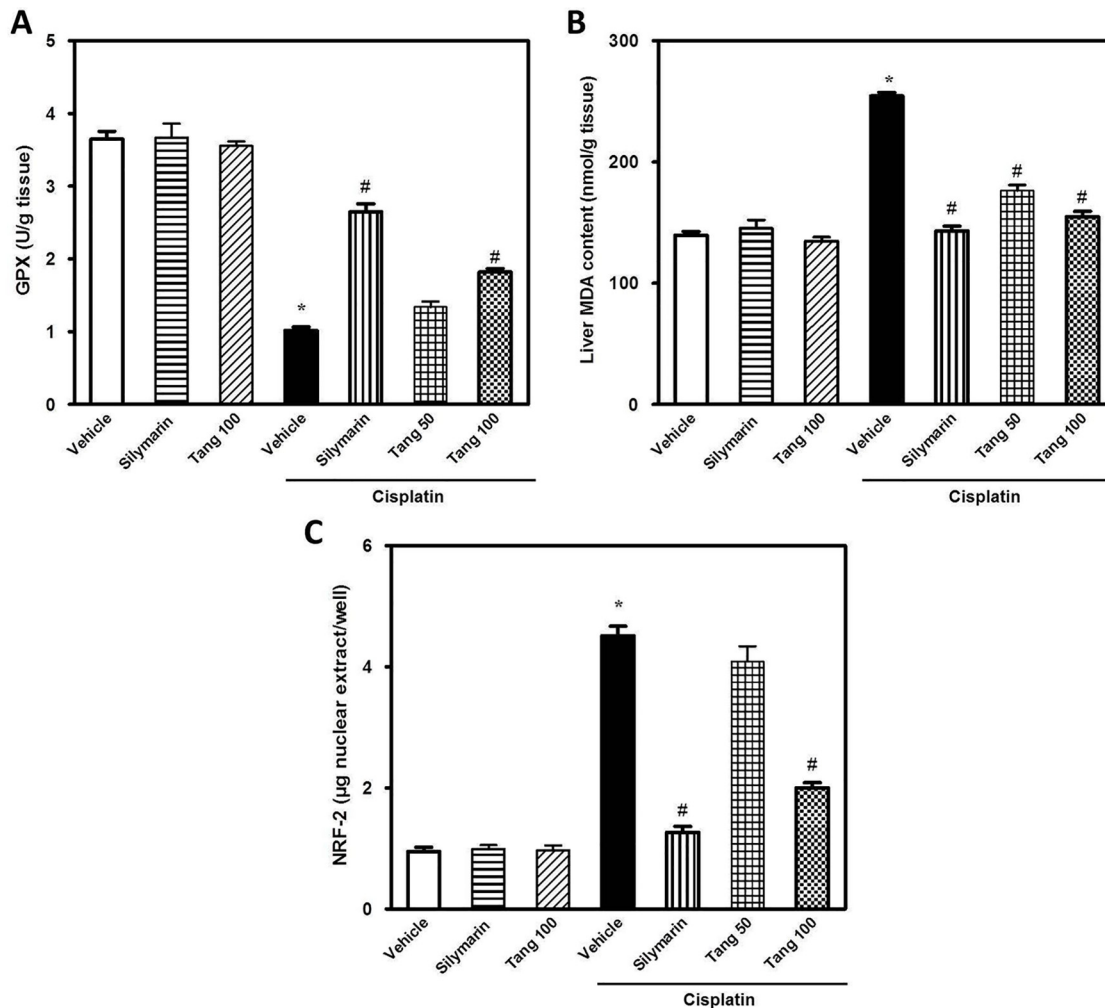


Fig 6. Tangeretin boosts hepatic antioxidant defense in cisplatin-induced oxidative stress in liver tissues. The effect of tangeretin pretreatment on cisplatin-induced oxidative stress in liver tissues as indicated by the modulation of GPx (A), MDA (B) and NRF-2 (C) content. Columns, mean; bars, \pm SEM (n = 8 independent values). Statistical analysis was carried out by using one way analysis of variance (ANOVA) followed by Tukey-Kramer multiple comparisons test. *Significant difference from normal control (vehicle) group at $p < 0.05$, #Significant difference from cisplatin group at $p < 0.05$. Tang 50; tangeretin (50 mg/kg), Tang 100; tangeretin (100 mg/kg).

doi:10.1371/journal.pone.0151649.g006

ability of tangeretin to counteract cisplatin-induced oxidative stress and enhance the antioxidant defenses plays a role in the defense against cisplatin-induced hepatic injury.

Tangeretin downregulates MAPK pathway in cisplatin-induced acute hepatic injury

The MAPK pathway has been reported to be activated in cisplatin-induced hepatic cytotoxicity [42]. Western blotting analysis revealed that administration of cisplatin activated MAPK signal transduction as evidenced by increased phosphorylation of p38 MAPK, JNK and ErK1/2 without affecting the corresponding total protein levels compared to the control group (Fig 7). Pretreatment with tangeretin attenuated the phosphorylation of the three hepatic MAPKs; effects which were comparable to those afforded by silymarin. Together, these data suggest that tangeretin downregulation of MAPK pathway plays a role in alleviating cisplatin-induced hepatic insult.

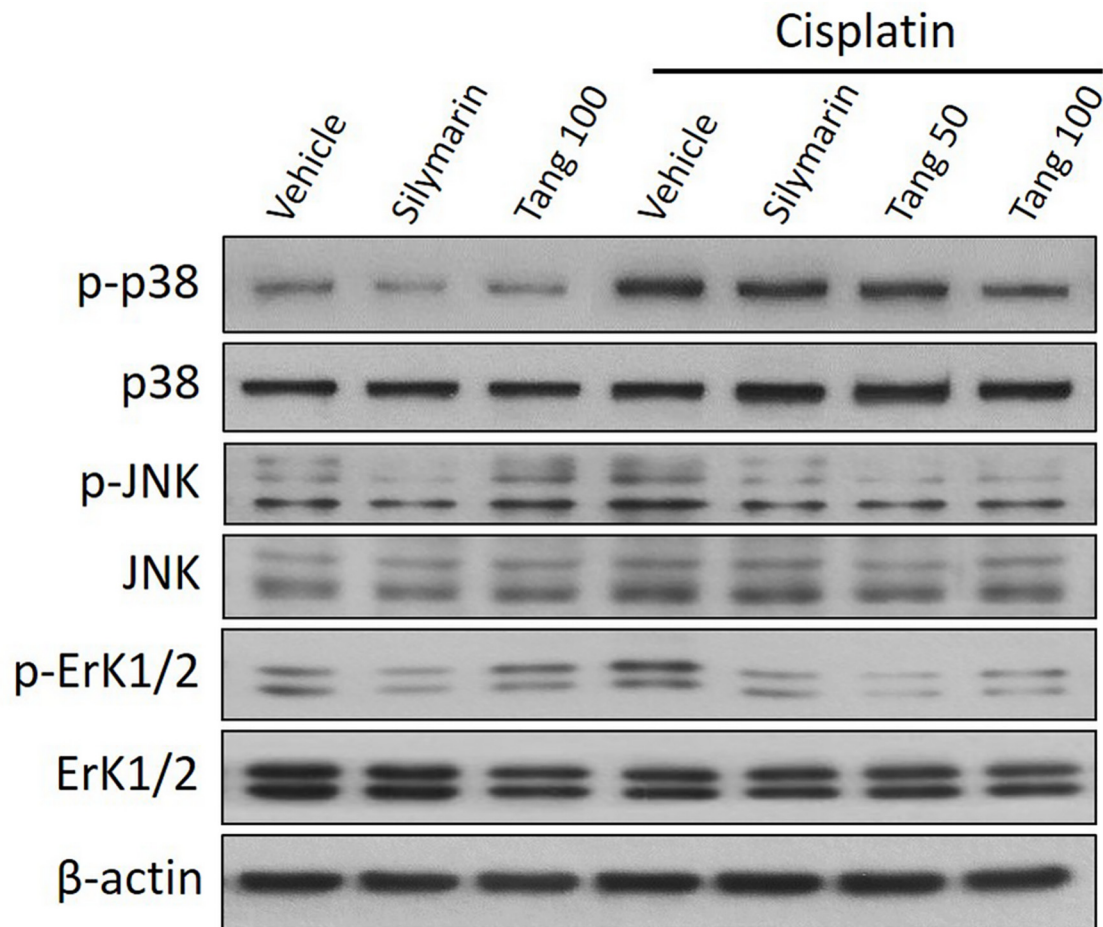


Fig 7. Tangeretin downregulates MAPK pathway in cisplatin-induced acute hepatic injury. Western blot analysis demonstrates that cisplatin increased the phosphorylation of p38 MAPK (upper panel), JNK (middle panel) and ErK1/2 (lower panel) without affecting the corresponding total protein levels, compared to the control group. Tangeretin and Silymarin pretreatments mitigated the phosphorylation of the three MAPKs. Tang 50; tangeretin (50 mg/kg), Tang 100; tangeretin (100 mg/kg).

doi:10.1371/journal.pone.0151649.g007

Discussion

The present study sheds light on the possible protective actions of tangeretin, a natural flavone found in citrus peels, against cisplatin-induced hepatic injury in rats. In a previous study, we have addressed the ability of tangeretin to synergize the anticancer activity of cisplatin in ovarian cancer cell lines study through targeting cancer cell survival pathways such as phosphoinositide 3-kinase/Akt signaling [23]. The focus of the current study was whether tangeretin can alleviate cisplatin-induced hepatotoxicity, one of the serious complications of cisplatin that may limit its therapeutic utility and to move a step towards the translational application of tangeretin in cancer combinatorial protocols. Notwithstanding its valuable applications as an anti-cancer agent, cisplatin has significant nephrotoxic and hepatotoxic side effects. Cisplatin inflicts hepatic injury through the activation of inflammatory and oxidative stress pathways along with associated apoptosis and anomalies in liver structure and function [9, 10]. These complications besides the chemoresistance are the most important limiting factors for the clinical application of cisplatin in the cancer treatment [10, 43].

TNF- α and IL-10 play a crucial role in mediating the interplay between inflammatory, oxidative stress and apoptotic pathways [44–46]. Tangeretin afforded significant protection against cisplatin-induced hepatic injury in rats mainly through the suppression of TNF- α and upregulation of IL-10 and indirectly via combating ROS production. In addition, tangeretin protected against cisplatin effect through its anti-apoptotic effects which were similar to those afforded by the reference hepatoprotective, silymarin. These actions signify the potential use of tangeretin in attenuating cisplatin-induced liver lesions.

Cisplatin induces a massive inflammatory response in liver tissues which was observed by the elevation of the proinflammatory, TNF- α and the decline of the anti-inflammatory, IL-10 [47, 48]. TNF- α intensifies the hepatic tissue inflammation via the chemotaxis of immune cells and activation other cytokines [49]. On the other hand, the decline in IL-10 exacerbates hepatic lesion because of its ability to downregulate antigen presentation and the pro-inflammatory cytokines release [50, 51].

Tangeretin significantly inhibited TNF- α and restored IL-10 levels, which can be assumed as a chief mechanism for its hepatoprotective role in cisplatin-induced hepatic injury. The findings were parallel to the histopathologic results that showed the ability of tangeretin to attenuate the inflammatory cell infiltration and hepatic necrosis. The anti-inflammatory actions of tangeretin are mediated through the modulation of TNF- α and other inflammatory mediators [52].

Cisplatin administration triggered lipid peroxide formation and depleted the hepatic GSH and GPx. Oxidative stress was reported to be involved in cisplatin-induced acute hepatic injury [9, 10]. The depletion of GSH and GPx, which play major roles in the cellular defense against oxidative stress and cellular damage, renders hepatic tissues more susceptible to oxidative stress [53, 54]. Tangeretin administration alleviated oxidative stress and improved the antioxidant status in rats which was in agreement with previous studies [55, 56]. The observed antioxidant actions of tangeretin play a role in the protection against cisplatin injury.

Cisplatin induced apoptotic cell death in hepatic tissues by the modulation of Bax and Bcl-2 expression levels [57]. The current data revealed that tangeretin inhibited the pro-apoptotic Bax and increased the anti-apoptotic Bcl-2, indicating diminished hepatic apoptosis. The attenuation of hepatic tissue apoptosis can be related to the observed suppression of oxidative stress and TNF- α which enhance liver tissue apoptosis [58].

Cisplatin-induced hepatotoxicity has been reported to be associated with MAPK signal transduction pathway which plays a major role in mediating cellular inflammatory response and apoptosis [59, 60]. Our results revealed the ability of cisplatin to induce the phosphorylation of p38, JNK and ERK1/2 in liver tissues which are in accordance with previous reports [59]. These changes were abrogated by pretreatment with tangeretin which underlie the modulation of MAPK signaling pathway as a putative mechanism for the protection against cisplatin injury. This action could be secondary to its antioxidant and anti-inflammatory activities. The reactive oxygen species generated by cisplatin treatment can activate various downstream proteins that mediate apoptosis and necrosis, in particular, MAPK family proteins [61]. The MAPK family comprises of three major serine/Threonine kinase proteins such as ERK, JNK and p38 which are associated with cell growth and differentiation, and are extensively linked to inflammation, apoptosis and cell death [62]. Cisplatin treatment shifts the balance between pro- and anti-apoptotic signals towards proapoptotic cascade [63]. It affords upregulation of Bax, a proapoptotic effector and diminishes Bcl-2, an anti-apoptotic protein. It also induces translocation of Bax from cytosol to mitochondria releasing cytochrome c to cytosol [64]. Cytochrome c further activates caspase 8, 9 and ultimately 3, thereby triggering apoptotic cell death [65]. Previous reports have regarded caspase activation as a crucial cellular mechanisms for induction of apoptosis in renal tubular cells in cisplatin-induced acute renal injury [66].

Many *in vitro* and *in vivo* studies have demonstrated the central role of p38, JNK and ERK1/2 in cisplatin-induced oxidative stress and apoptosis [65]. It's noteworthy that due to the mutual interplay between MAPKs and ROS with consequent deleterious effects on liver tissues, the current study cannot specifically confirm the activation of MAPK being a cause or a result for the hepatoprotection [67, 68].

In the same context, evidence has indicated that cisplatin treatment is associated with reactive oxygen species (ROS) generation which activates p38 MAPK resulting in apoptotic cell death in rat kidney [69]. The cross-talk between p38 MAPK and caspase signaling cascade was also reported in cisplatin-treated carcinoma and rat renal tissues [69]. It has been suggested that p38 and JNK favor apoptotic cell death [70, 71]. It is also reported that JNK is associated with TNF- α -induced apoptosis [72]. The interplay between p38 MAPK and TNF- α has been described in cisplatin-evoked renal injury [69] where p38 MAPK acts as an upstream signal

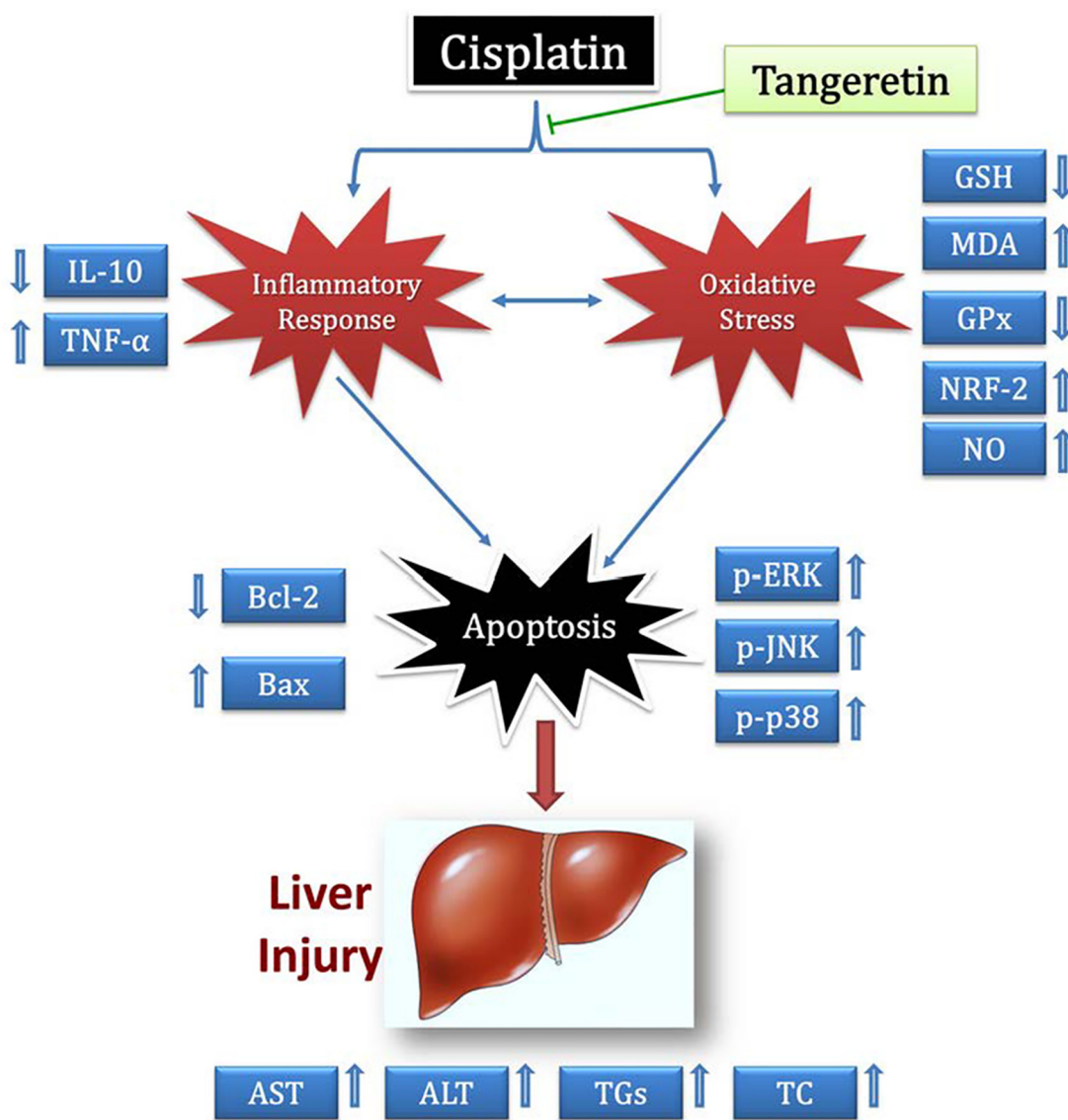


Fig 8. Diagram depicting the proposed protective mechanisms of tangeretin against cisplatin-induced hepatic injury.

doi:10.1371/journal.pone.0151649.g008

which activates the transcription factor NF- κ B with the consequent generation of TNF- α [73]. Abrogation of ROS production/ p38 MAPK activation by the antioxidants such as N-acetyl cysteine has considerably protected the renal tissues from cisplatin insult [69].

In conclusion, the present work underscores the possible protective effects of tangeretin against cisplatin-induced acute hepatic injury in rats (Fig 8). These favorable actions confirm tangeretin benefits as an effective and safe approach for the management of one of cisplatin drawbacks. Further studies addressing the impact of tangeretin after repeated administration of cisplatin and at different time points are warranted, in order to delineate the exact underlying molecular mechanisms of tangeretin in cisplatin-evoked hepatotoxicity and the implication of other cell signaling networks.

Acknowledgments

The current work was financially supported by grant number 1-434-2830 from the Deanship of Scientific Research, Taif University, Saudi Arabia. The authors are thankful to Dr. Kawkab A. Ahmed, Professor of pathology, Faculty of Veterinary Medicine, Cairo University, Egypt for her kind help in the histopathology and immunohistochemistry.

Author Contributions

Conceived and designed the experiments: HAO WRM HHA EAA. Performed the experiments: HAO WRM HHA EAA. Analyzed the data: HAO WRM HHA EAA. Contributed reagents/materials/analysis tools: HAO WRM HHA EAA. Wrote the paper: HAO WRM HHA EAA.

References

1. Siddik ZH. Cisplatin: mode of cytotoxic action and molecular basis of resistance. *Oncogene*. 2003; 22(47):7265–79. Epub 2003/10/25. doi: [10.1038/sj.onc.1206933](https://doi.org/10.1038/sj.onc.1206933) PMID: [14576837](https://pubmed.ncbi.nlm.nih.gov/14576837/).
2. Cui W, Yazlovitskaya EM, Mayo MS, Pelling JC, Persons DL. Cisplatin-induced response of c-jun N-terminal kinase 1 and extracellular signal-regulated protein kinases 1 and 2 in a series of cisplatin-resistant ovarian carcinoma cell lines. *Molecular carcinogenesis*. 2000; 29(4):219–28. Epub 2001/02/15. PMID: [11170260](https://pubmed.ncbi.nlm.nih.gov/11170260/).
3. Shi R, Huang Q, Zhu X, Ong YB, Zhao B, Lu J, et al. Luteolin sensitizes the anticancer effect of cisplatin via c-Jun NH2-terminal kinase-mediated p53 phosphorylation and stabilization. *Molecular cancer therapeutics*. 2007; 6(4):1338–47. Epub 2007/04/14. doi: [10.1158/1535-7163.MCT-06-0638](https://doi.org/10.1158/1535-7163.MCT-06-0638) PMID: [17431112](https://pubmed.ncbi.nlm.nih.gov/17431112/).
4. Antunes LM, Darin JD, Bianchi MD. Protective effects of vitamin c against cisplatin-induced nephrotoxicity and lipid peroxidation in adult rats: a dose-dependent study. *Pharmacological research: the official journal of the Italian Pharmacological Society*. 2000; 41(4):405–11. Epub 2000/03/08. doi: [10.1006/phrs.1999.0600](https://doi.org/10.1006/phrs.1999.0600) PMID: [10704263](https://pubmed.ncbi.nlm.nih.gov/10704263/).
5. Zicca A, Cafaggi S, Mariggio MA, Vannozzi MO, Ottone M, Bocchini V, et al. Reduction of cisplatin hepatotoxicity by procainamide hydrochloride in rats. *European journal of pharmacology*. 2002; 442(3):265–72. Epub 2002/06/18. PMID: [12065080](https://pubmed.ncbi.nlm.nih.gov/12065080/).
6. Atessahin A, Karahan I, Turk G, Gur S, Yilmaz S, Ceribasi AO. Protective role of lycopene on cisplatin-induced changes in sperm characteristics, testicular damage and oxidative stress in rats. *Reprod Toxicol*. 2006; 21(1):42–7. Epub 2005/06/28. doi: [10.1016/j.reprotox.2005.05.003](https://doi.org/10.1016/j.reprotox.2005.05.003) PMID: [15979841](https://pubmed.ncbi.nlm.nih.gov/15979841/).
7. Tsuruya K, Ninomiya T, Tokumoto M, Hirakawa M, Masutani K, Taniguchi M, et al. Direct involvement of the receptor-mediated apoptotic pathways in cisplatin-induced renal tubular cell death. *Kidney international*. 2003; 63(1):72–82. Epub 2002/12/11. doi: [10.1046/j.1523-1755.2003.00709.x](https://doi.org/10.1046/j.1523-1755.2003.00709.x) PMID: [12472770](https://pubmed.ncbi.nlm.nih.gov/12472770/).
8. Hong KO, Hwang JK, Park KK, Kim SH. Phosphorylation of c-Jun N-terminal Kinases (JNKs) is involved in the preventive effect of xanthorrhizol on cisplatin-induced hepatotoxicity. *Archives of toxicology*. 2005; 79(4):231–6. Epub 2004/11/13. doi: [10.1007/s00204-004-0623-7](https://doi.org/10.1007/s00204-004-0623-7) PMID: [15536542](https://pubmed.ncbi.nlm.nih.gov/15536542/).
9. Bentli R, Parlakpınar H, Polat A, Samdancı E, Sarihan ME, Sagir M. Molsidomine prevents cisplatin-induced hepatotoxicity. *Archives of medical research*. 2013; 44(7):521–8. Epub 2013/10/15. doi: [10.1016/j.arcmed.2013.09.013](https://doi.org/10.1016/j.arcmed.2013.09.013) PMID: [24120390](https://pubmed.ncbi.nlm.nih.gov/24120390/).

10. Lu Y, Cederbaum AI. Cisplatin-induced hepatotoxicity is enhanced by elevated expression of cytochrome P450 2E1. *Toxicological sciences: an official journal of the Society of Toxicology*. 2006; 89(2):515–23. Epub 2005/10/28. doi: [10.1093/toxsci/kfj031](https://doi.org/10.1093/toxsci/kfj031) PMID: [16251482](https://pubmed.ncbi.nlm.nih.gov/16251482/).
11. Pratibha R, Sameer R, Rataboli PV, Bhiwgade DA, Dhume CY. Enzymatic studies of cisplatin induced oxidative stress in hepatic tissue of rats. *European journal of pharmacology*. 2006; 532(3):290–3. Epub 2006/02/07. doi: [10.1016/j.ejphar.2006.01.007](https://doi.org/10.1016/j.ejphar.2006.01.007) PMID: [16458885](https://pubmed.ncbi.nlm.nih.gov/16458885/).
12. Bava SV, Puliappadamba VT, Deepti A, Nair A, Karunagar D, Anto RJ. Sensitization of taxol-induced apoptosis by curcumin involves down-regulation of nuclear factor-kappaB and the serine/threonine kinase Akt and is independent of tubulin polymerization. *The Journal of biological chemistry*. 2005; 280(8):6301–8. Epub 2004/12/14. doi: [10.1074/jbc.M410647200](https://doi.org/10.1074/jbc.M410647200) PMID: [15590651](https://pubmed.ncbi.nlm.nih.gov/15590651/).
13. Singh RP, Agarwal R. Natural flavonoids targeting deregulated cell cycle progression in cancer cells. *Current drug targets*. 2006; 7(3):345–54. Epub 2006/03/07. PMID: [16515531](https://pubmed.ncbi.nlm.nih.gov/16515531/).
14. Chen KH, Weng MS, Lin JK. Tangeretin suppresses IL-1beta-induced cyclooxygenase (COX)-2 expression through inhibition of p38 MAPK, JNK, and AKT activation in human lung carcinoma cells. *Biochemical pharmacology*. 2007; 73(2):215–27. Epub 2006/10/28. doi: [10.1016/j.bcp.2006.09.018](https://doi.org/10.1016/j.bcp.2006.09.018) PMID: [17067555](https://pubmed.ncbi.nlm.nih.gov/17067555/).
15. Sak K. Cytotoxicity of dietary flavonoids on different human cancer types. *Pharmacognosy reviews*. 2014; 8(16):122–46. Epub 2014/08/16. doi: [10.4103/0973-7847.134247](https://doi.org/10.4103/0973-7847.134247) PMID: [25125885](https://pubmed.ncbi.nlm.nih.gov/25125885/); PubMed Central PMCID: [PMC4127821](https://pubmed.ncbi.nlm.nih.gov/PMC4127821/).
16. Sundaram R, Shanthi P, Sachdanandam P. Tangeretin, a polymethoxylated flavone, modulates lipid homeostasis and decreases oxidative stress by inhibiting NF-κB activation and proinflammatory cytokines in cardiac tissue of streptozotocin-induced diabetic rats. *Journal of Functional Foods*. 2015; 16(0):315–33. doi: [10.1016/j.jff.2015.03.024](https://doi.org/10.1016/j.jff.2015.03.024)
17. Periyasamy K, Baskaran K, Ilakkia A, Vanitha K, Selvaraj S, Sakthisekaran D. Antitumor efficacy of tangeretin by targeting the oxidative stress mediated on 7,12-dimethylbenz(a)anthracene-induced proliferative breast cancer in Sprague-Dawley rats. *Cancer Chemother Pharmacol*. 2015; 75(2):263–72. Epub 2014/11/29. doi: [10.1007/s00280-014-2629-z](https://doi.org/10.1007/s00280-014-2629-z) PMID: [25431347](https://pubmed.ncbi.nlm.nih.gov/25431347/).
18. Kurowska EM, Manthey JA. Hypolipidemic effects and absorption of citrus polymethoxylated flavones in hamsters with diet-induced hypercholesterolemia. *Journal of agricultural and food chemistry*. 2004; 52(10):2879–86. Epub 2004/05/13. doi: [10.1021/jf035354z](https://doi.org/10.1021/jf035354z) PMID: [15137829](https://pubmed.ncbi.nlm.nih.gov/15137829/).
19. Vanhoecke BW, Delporte F, Van Braeckel E, Heyerick A, Depypere HT, Nuytinck M, et al. A safety study of oral tangeretin and xanthohumol administration to laboratory mice. *In Vivo*. 2005; 19(1):103–7. Epub 2005/03/31. PMID: [15796161](https://pubmed.ncbi.nlm.nih.gov/15796161/).
20. Hirano T, Abe K, Gotoh M, Oka K. Citrus flavone tangeretin inhibits leukaemic HL-60 cell growth partially through induction of apoptosis with less cytotoxicity on normal lymphocytes. *British journal of cancer*. 1995; 72(6):1380–8. Epub 1995/12/01. PMID: [8519648](https://pubmed.ncbi.nlm.nih.gov/8519648/); PubMed Central PMCID: [PMC2034105](https://pubmed.ncbi.nlm.nih.gov/PMC2034105/).
21. Pan MH, Chen WJ, Lin-Shiau SY, Ho CT, Lin JK. Tangeretin induces cell-cycle G1 arrest through inhibiting cyclin-dependent kinases 2 and 4 activities as well as elevating Cdk inhibitors p21 and p27 in human colorectal carcinoma cells. *Carcinogenesis*. 2002; 23(10):1677–84. Epub 2002/10/12. PMID: [12376477](https://pubmed.ncbi.nlm.nih.gov/12376477/).
22. Walle T, Walle UK. Novel methoxylated flavone inhibitors of cytochrome P450 1B1 in SCC-9 human oral cancer cells. *The Journal of pharmacy and pharmacology*. 2007; 59(6):857–62. Epub 2007/07/20. doi: [10.1211/jpp.59.6.0012](https://doi.org/10.1211/jpp.59.6.0012) PMID: [17637178](https://pubmed.ncbi.nlm.nih.gov/17637178/).
23. Arafa el SA, Zhu Q, Barakat BM, Wani G, Zhao Q, El-Mahdy MA, et al. Tangeretin sensitizes cisplatin-resistant human ovarian cancer cells through downregulation of phosphoinositide 3-kinase/Akt signaling pathway. *Cancer Res*. 2009; 69(23):8910–7. Epub 2009/11/12. doi: [10.1158/0008-5472.CAN-09-1543](https://doi.org/10.1158/0008-5472.CAN-09-1543) PMID: [19903849](https://pubmed.ncbi.nlm.nih.gov/19903849/); PubMed Central PMCID: [PMC3319094](https://pubmed.ncbi.nlm.nih.gov/PMC3319094/).
24. Flora K, Hahn M, Rosen H, Benner K. Milk thistle (*Silybum marianum*) for the therapy of liver disease. *The American journal of gastroenterology*. 1998; 93(2):139–43. Epub 1998/02/19. doi: [10.1111/j.1572-0241.1998.00139.x](https://doi.org/10.1111/j.1572-0241.1998.00139.x) PMID: [9468229](https://pubmed.ncbi.nlm.nih.gov/9468229/).
25. Naveau S. [Acute alcoholic hepatitis: treatments]. *Presse Med*. 2001; 30(20):1024–30. Epub 2001/07/04. PMID: [11433695](https://pubmed.ncbi.nlm.nih.gov/11433695/).
26. Laekeman G, De Coster S, De Meyer K. [St. Mary's Thistle: an overview]. *Journal de pharmacie de Belgique*. 2003; 58(1):28–31. Epub 2003/05/02. PMID: [12722542](https://pubmed.ncbi.nlm.nih.gov/12722542/).
27. Syed SN, Rizvi W, Kumar A, Khan AA, Moin S, Ahsan A. In vitro antioxidant and in vivo hepatoprotective activity of leave extract of *Raphanus sativus* in rats using CCL4 model. *African journal of traditional, complementary, and alternative medicines: AJTCAM / African Networks on Ethnomedicines*. 2014; 11(3):102–6. Epub 2014/11/06. PMID: [25371570](https://pubmed.ncbi.nlm.nih.gov/25371570/); PubMed Central PMCID: [PMC4202426](https://pubmed.ncbi.nlm.nih.gov/PMC4202426/).

28. Clichici S, Olteanu D, Nagy AL, Oros A, Filip A, Mircea PA. Silymarin inhibits the progression of fibrosis in the early stages of liver injury in CCl(4)-treated rats. *Journal of medicinal food*. 2015; 18(3):290–8. Epub 2014/08/19. doi: [10.1089/jmf.2013.0179](https://doi.org/10.1089/jmf.2013.0179) PMID: [25133972](https://pubmed.ncbi.nlm.nih.gov/25133972/).
29. Singh D, Arya PV, Sharma A, Dobhal MP, Gupta RS. Modulatory potential of alpha-amyryn against hepatic oxidative stress through antioxidant status in Wistar albino rats. *Journal of ethnopharmacology*. 2015; 161:186–93. Epub 2014/12/30. doi: [10.1016/j.jep.2014.12.025](https://doi.org/10.1016/j.jep.2014.12.025) PMID: [25542388](https://pubmed.ncbi.nlm.nih.gov/25542388/).
30. Gopi KS, Reddy AG, Jyothi K, Kumar BA. Acetaminophen-induced Hepato- and Nephrotoxicity and Amelioration by Silymarin and Terminalia chebula in Rats. *Toxicology international*. 2010; 17(2):64–6. Epub 2010/12/21. doi: [10.4103/0971-6580.72672](https://doi.org/10.4103/0971-6580.72672) PMID: [21170247](https://pubmed.ncbi.nlm.nih.gov/21170247/); PubMed Central PMCID: PMC2997457.
31. Iroanya OO, Adebisin OA, Okpuzor J. Evaluation of the Hepato and Nephron-Protective Effect of a Polyherbal Mixture using Wistar Albino Rats. *Journal of clinical and diagnostic research: JCDR*. 2014; 8(6):HC15–21. Epub 2014/08/15. doi: [10.7860/JCDR/2014/5875.4501](https://doi.org/10.7860/JCDR/2014/5875.4501) PMID: [25121002](https://pubmed.ncbi.nlm.nih.gov/25121002/); PubMed Central PMCID: PMC4129255.
32. Pushpavalli G, Veeramani C, Pugalendi KV. Influence of chrysin on hepatic marker enzymes and lipid profile against D-galactosamine-induced hepatotoxicity rats. *Food and chemical toxicology: an international journal published for the British Industrial Biological Research Association*. 2010; 48(6):1654–9. Epub 2010/04/07. doi: [10.1016/j.fct.2010.03.040](https://doi.org/10.1016/j.fct.2010.03.040) PMID: [20362027](https://pubmed.ncbi.nlm.nih.gov/20362027/).
33. Mansour HH, Hafez HF, Fahmy NM. Silymarin modulates Cisplatin-induced oxidative stress and hepatotoxicity in rats. *Journal of biochemistry and molecular biology*. 2006; 39(6):656–61. Epub 2006/11/30. PMID: [17129399](https://pubmed.ncbi.nlm.nih.gov/17129399/).
34. Zicca A, Cafaggi S, Mariggio MA, Vannozzi MO, Ottone M, Bocchini V, et al. Reduction of cisplatin hepatotoxicity by procainamide hydrochloride in rats. *Eur J Pharmacol*. 2002; 442(3):265–72. Epub 2002/06/18. PMID: [12065080](https://pubmed.ncbi.nlm.nih.gov/12065080/).
35. Arivazhagan L, Sorimuthu Pillai S. Tangeretin, a citrus pentamethoxyflavone, exerts cytostatic effect via p53/p21 up-regulation and suppresses metastasis in 7,12-dimethylbenz(alpha)anthracene-induced rat mammary carcinoma. *The Journal of nutritional biochemistry*. 2014; 25(11):1140–53. Epub 2014/08/26. doi: [10.1016/j.jnutbio.2014.06.007](https://doi.org/10.1016/j.jnutbio.2014.06.007) PMID: [25151216](https://pubmed.ncbi.nlm.nih.gov/25151216/).
36. Reitman S, Frankel S. A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. *Am J Clin Pathol*. 1957; 28(1):56–63. Epub 1957/07/01. PMID: [13458125](https://pubmed.ncbi.nlm.nih.gov/13458125/).
37. Miranda KM, Espey MG, Wink DA. A rapid, simple spectrophotometric method for simultaneous detection of nitrate and nitrite. *Nitric oxide: biology and chemistry / official journal of the Nitric Oxide Society*. 2001; 5(1):62–71. Epub 2001/02/17. doi: [10.1006/niox.2000.0319](https://doi.org/10.1006/niox.2000.0319) PMID: [11178938](https://pubmed.ncbi.nlm.nih.gov/11178938/).
38. Ellman GL. Tissue sulfhydryl groups. *Arch Biochem Biophys*. 1959; 82(1):70–7. Epub 1959/05/01. PMID: [13650640](https://pubmed.ncbi.nlm.nih.gov/13650640/).
39. Buege JA, Aust SD. Microsomal lipid peroxidation. *Methods in enzymology*. 1978; 52:302–10. Epub 1978/01/01. PMID: [672633](https://pubmed.ncbi.nlm.nih.gov/672633/).
40. Arab HH, Salama SA, Omar HA, Arafa el SA, Maghrabi IA. Diosmin Protects against Ethanol-Induced Gastric Injury in Rats: Novel Anti-Ulcer Actions. *PloS one*. 2015; 10(3):e0122417. Epub 2015/03/31. doi: [10.1371/journal.pone.0122417](https://doi.org/10.1371/journal.pone.0122417) PMID: [25821971](https://pubmed.ncbi.nlm.nih.gov/25821971/); PubMed Central PMCID: PMC4378914.
41. Omar HA, Sargeant AM, Weng JR, Wang D, Kulp SK, Patel T, et al. Targeting of the Akt-nuclear factor-kappa B signaling network by [1-(4-chloro-3-nitrobenzenesulfonyl)-1H-indol-3-yl]-methanol (OSU-A9), a novel indole-3-carbinol derivative, in a mouse model of hepatocellular carcinoma. *Molecular pharmacology*. 2009; 76(5):957–68. doi: [10.1124/mol.109.058180](https://doi.org/10.1124/mol.109.058180) PMID: [19706731](https://pubmed.ncbi.nlm.nih.gov/19706731/); PubMed Central PMCID: PMC2774988.
42. Sun Y, Yang J, Wang LZ, Sun LR, Dong Q. Crocin attenuates cisplatin-induced liver injury in the mice. *Hum Exp Toxicol*. 2014; 33(8):855–62. Epub 2013/11/28. doi: [10.1177/0960327113511475](https://doi.org/10.1177/0960327113511475) PMID: [24275644](https://pubmed.ncbi.nlm.nih.gov/24275644/).
43. Iseri S, Ercan F, Gedik N, Yuksel M, Alican I. Simvastatin attenuates cisplatin-induced kidney and liver damage in rats. *Toxicology*. 2007; 230(2–3):256–64. Epub 2007/01/02. doi: [10.1016/j.tox.2006.11.073](https://doi.org/10.1016/j.tox.2006.11.073) PMID: [17196726](https://pubmed.ncbi.nlm.nih.gov/17196726/).
44. Tadagavadi RK, Reeves WB. Endogenous IL-10 attenuates cisplatin nephrotoxicity: role of dendritic cells. *J Immunol*. 2010; 185(8):4904–11. Epub 2010/09/17. doi: [10.4049/jimmunol.1000383](https://doi.org/10.4049/jimmunol.1000383) PMID: [20844196](https://pubmed.ncbi.nlm.nih.gov/20844196/); PubMed Central PMCID: PMC3169908.
45. Chou WY, Lu CN, Lee TH, Wu CL, Hung KS, Concejero AM, et al. Electroporative interleukin-10 gene transfer ameliorates carbon tetrachloride-induced murine liver fibrosis by MMP and TIMP modulation. *Acta pharmacologica Sinica*. 2006; 27(4):469–76. Epub 2006/03/17. doi: [10.1111/j.1745-7254.2006.00304.x](https://doi.org/10.1111/j.1745-7254.2006.00304.x) PMID: [16539848](https://pubmed.ncbi.nlm.nih.gov/16539848/).

46. Shaw J, Chen B, Huang WH, Lee AR, Media J, Valeriote FA. The small-molecule TNF-alpha modulator, UTL-5g, reduces side effects induced by cisplatin and enhances the therapeutic effect of cisplatin in vivo. *J Exp Ther Oncol*. 2011; 9(2):129–37. Epub 2011/06/28. PMID: [21699020](#).
47. Rehman MU, Ali N, Rashid S, Jain T, Nafees S, Tahir M, et al. Alleviation of hepatic injury by chrysin in cisplatin administered rats: probable role of oxidative and inflammatory markers. *Pharmacological reports: PR*. 2014; 66(6):1050–9. Epub 2014/12/03. doi: [10.1016/j.pharep.2014.06.004](#) PMID: [25443734](#).
48. Tadagavadi RK, Reeves WB. Endogenous IL-10 attenuates cisplatin nephrotoxicity: role of dendritic cells. *J Immunol*. 2010; 185(8):4904–11. Epub 2010/09/17. doi: [10.4049/jimmunol.1000383](#) PMID: [20844196](#); PubMed Central PMCID: PMC3169908.
49. Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harbor perspectives in biology*. 2009; 1(6):a001651. Epub 2010/05/12. doi: [10.1101/cshperspect.a001651](#) PMID: [20457564](#); PubMed Central PMCID: PMC2882124.
50. Koppelman B, Neefjes JJ, de Vries JE, de Waal Malefyt R. Interleukin-10 down-regulates MHC class II alphabeta peptide complexes at the plasma membrane of monocytes by affecting arrival and recycling. *Immunity*. 1997; 7(6):861–71. Epub 1998/01/16. PMID: [9430231](#).
51. Knolle PA, Uhrig A, Hegenbarth S, Loser E, Schmitt E, Gerken G, et al. IL-10 down-regulates T cell activation by antigen-presenting liver sinusoidal endothelial cells through decreased antigen uptake via the mannose receptor and lowered surface expression of accessory molecules. *Clinical and experimental immunology*. 1998; 114(3):427–33. Epub 1998/12/09. PMID: [9844054](#); PubMed Central PMCID: PMC1905120.
52. Shu Z, Yang B, Zhao H, Xu B, Jiao W, Wang Q, et al. Tangeretin exerts anti-neuroinflammatory effects via NF-kappaB modulation in lipopolysaccharide-stimulated microglial cells. *International immunopharmacology*. 2014; 19(2):275–82. Epub 2014/01/28. doi: [10.1016/j.intimp.2014.01.011](#) PMID: [24462494](#).
53. Louvet A, Mathurin P. Alcoholic liver disease: mechanisms of injury and targeted treatment. *Nature reviews Gastroenterology & hepatology*. 2015; 12(4):231–42. Epub 2015/03/18. doi: [10.1038/nrgastro.2015.35](#) PMID: [25782093](#).
54. Ramos AS, Correia AT, Antunes SC, Goncalves F, Nunes B. Effect of acetaminophen exposure in *Oncorhynchus mykiss* gills and liver: detoxification mechanisms, oxidative defence system and peroxidative damage. *Environmental toxicology and pharmacology*. 2014; 37(3):1221–8. Epub 2014/05/13. doi: [10.1016/j.etap.2014.04.005](#) PMID: [24816177](#).
55. Lakshmi A, Subramanian SP. Tangeretin ameliorates oxidative stress in the renal tissues of rats with experimental breast cancer induced by 7,12-dimethylbenz[a]anthracene. *Toxicol Lett*. 2014; 229(2):333–48. Epub 2014/07/06. doi: [10.1016/j.toxlet.2014.06.845](#) PMID: [24995432](#).
56. Lakshmi A, Subramanian S. Chemotherapeutic effect of tangeretin, a polymethoxylated flavone studied in 7, 12-dimethylbenz(a)anthracene induced mammary carcinoma in experimental rats. *Biochimie*. 2014; 99:96–109. Epub 2013/12/05. doi: [10.1016/j.biochi.2013.11.017](#) PMID: [24299963](#).
57. Martins NM, Santos NA, Curti C, Bianchi ML, Santos AC. Cisplatin induces mitochondrial oxidative stress with resultant energetic metabolism impairment, membrane rigidification and apoptosis in rat liver. *Journal of applied toxicology: JAT*. 2008; 28(3):337–44. Epub 2007/07/03. doi: [10.1002/jat.1284](#) PMID: [17604343](#).
58. Wang K. Molecular mechanisms of hepatic apoptosis. *Cell death & disease*. 2014; 5:e996. Epub 2014/01/18. doi: [10.1038/cddis.2013.499](#) PMID: [24434519](#); PubMed Central PMCID: PMC4040708.
59. Sohn JH, Han KL, Kim JH, Rukayadi Y, Hwang JK. Protective Effects of macelignan on cisplatin-induced hepatotoxicity is associated with JNK activation. *Biological & pharmaceutical bulletin*. 2008; 31(2):273–7. Epub 2008/02/02. PMID: [18239286](#).
60. Zarubin T, Han J. Activation and signaling of the p38 MAP kinase pathway. *Cell research*. 2005; 15(1):11–8. Epub 2005/02/03. doi: [10.1038/sj.cr.7290257](#) PMID: [15686620](#).
61. Malik S, Suchal K, Gamad N, Dinda AK, Arya DS, Bhatia J. Telmisartan ameliorates cisplatin-induced nephrotoxicity by inhibiting MAPK mediated inflammation and apoptosis. *European journal of pharmacology*. 2015; 748:54–60. doi: [10.1016/j.ejphar.2014.12.008](#) PMID: [25510231](#).
62. Wada T, Penninger JM. Mitogen-activated protein kinases in apoptosis regulation. *Oncogene*. 2004; 23(16):2838–49. doi: [10.1038/sj.onc.1207556](#) PMID: [15077147](#).
63. Indran IR, Tufo G, Pervaiz S, Brenner C. Recent advances in apoptosis, mitochondria and drug resistance in cancer cells. *Biochimica et biophysica acta*. 2011; 1807(6):735–45. doi: [10.1016/j.bbabbio.2011.03.010](#) PMID: [21453675](#).
64. Ramesh G, Reeves WB. p38 MAP kinase inhibition ameliorates cisplatin nephrotoxicity in mice. *American journal of physiology Renal physiology*. 2005; 289(1):F166–74. doi: [10.1152/ajprenal.00401.2004](#) PMID: [15701814](#).

65. Francescato HDC, Costa RS, Júnior FB, Coimbra TM. Effect of JNK inhibition on cisplatin-induced renal damage. *Nephrology Dialysis Transplantation*. 2007; 22(8):2138–48. doi: [10.1093/ndt/gfm144](https://doi.org/10.1093/ndt/gfm144)
66. Helmy MM, Helmy MW, Abd Allah DM, Abo Zaid AM, Mohy El-Din MM. Selective ETA receptor blockade protects against cisplatin-induced acute renal failure in male rats. *European journal of pharmacology*. 2014; 730:133–9. doi: [10.1016/j.ejphar.2014.03.002](https://doi.org/10.1016/j.ejphar.2014.03.002) PMID: [24632086](https://pubmed.ncbi.nlm.nih.gov/24632086/)
67. Bubici C, Papa S, Dean K, Franzoso G. Mutual cross-talk between reactive oxygen species and nuclear factor-kappa B: molecular basis and biological significance. *Oncogene*. 2006; 25(51):6731–48. Epub 2006/10/31. doi: [10.1038/sj.onc.1209936](https://doi.org/10.1038/sj.onc.1209936) PMID: [17072325](https://pubmed.ncbi.nlm.nih.gov/17072325/).
68. Nakagawa H, Maeda S. Molecular mechanisms of liver injury and hepatocarcinogenesis: focusing on the role of stress-activated MAPK. *Pathology research international*. 2012; 2012:172894. Epub 2012/06/06. doi: [10.1155/2012/172894](https://doi.org/10.1155/2012/172894) PMID: [22666632](https://pubmed.ncbi.nlm.nih.gov/22666632/); PubMed Central PMCID: [PMC3361329](https://pubmed.ncbi.nlm.nih.gov/PMC3361329/).
69. Luo J, Tsuji T, Yasuda H, Sun Y, Fujigaki Y, Hishida A. The molecular mechanisms of the attenuation of cisplatin-induced acute renal failure by N-acetylcysteine in rats. *Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association—European Renal Association*. 2008; 23(7):2198–205. doi: [10.1093/ndt/gfn090](https://doi.org/10.1093/ndt/gfn090) PMID: [18385389](https://pubmed.ncbi.nlm.nih.gov/18385389/).
70. Chang L, Karin M. Mammalian MAP kinase signalling cascades. *Nature*. 2001; 410(6824):37–40. doi: [10.1038/35065000](https://doi.org/10.1038/35065000) PMID: [11242034](https://pubmed.ncbi.nlm.nih.gov/11242034/).
71. Parlakpınar H, Sahna E, Ozer MK, Ozugurlu F, Vardi N, Acet A. Physiological and pharmacological concentrations of melatonin protect against cisplatin-induced acute renal injury. *Journal of pineal research*. 2002; 33(3):161–6. PMID: [12220331](https://pubmed.ncbi.nlm.nih.gov/12220331/).
72. Ding WX, Yin XM. Dissection of the multiple mechanisms of TNF-alpha-induced apoptosis in liver injury. *Journal of cellular and molecular medicine*. 2004; 8(4):445–54. PMID: [15601573](https://pubmed.ncbi.nlm.nih.gov/15601573/).
73. Hayakawa K, Meng Y, Hiramatsu N, Kasai A, Yao J, Kitamura M. Spontaneous activation of the NF-kappaB signaling pathway in isolated normal glomeruli. *American journal of physiology Renal physiology*. 2006; 291(6):F1169–76. doi: [10.1152/ajprenal.00513.2005](https://doi.org/10.1152/ajprenal.00513.2005) PMID: [16705144](https://pubmed.ncbi.nlm.nih.gov/16705144/).