## **EDITORIAL**

## Cartilage breakdown in microgravity—a problem for longterm spaceflight?

npj Regenerative Medicine (2017)2:10: doi:10.1038/s41536-017-0016-1

From the early days of spaceflight when humans first escaped the gravitational pull of Earth, it became clear that the unloading environment of microgravity (10<sup>-6</sup>G) had an impact on the human skeleton. The musculoskeletal system is acutely sensitive to changes in the biomechanical environment and prolonged exposure to microgravity causes bones to demineralize, and skeletal muscles to lose mass and strength. 1-5 The lost tissue appears to be restored following return to normal gravity, although long-term studies have identified incomplete recovery of bone mineral density and architecture and permanent damage to bone remains a concern.<sup>1</sup>

The musculoskeletal system is more than just bone and skeletal muscle and the effects of microgravity on other biomechanically sensitive elements are largely unknown. Of particular interest are articulating synovial joints, which are precisely engineered to accommodate large tensile and compressive biomechanical forces and, simultaneously, facilitate smooth mechanical action. Synovial joints are composed of articular cartilage integrated with subchondral bone, meniscal fibro-cartilage, tendon and several ligaments. These tissues are bathed in synovial fluid and enclosed in a fibrous capsule. Damage to any of these functionally interconnected elements lead to joint instability and compensatory connective tissue changes that can result in joint degradation. For the affected individual, these changes are often accompanied by progressively worsening pain, loss of mobility, and eventually, osteoarthritis (OA).6, 7 It is generally accepted that once the collagen II network in articular cartilage is broken down following proteoglycan degradation, restoration of authentic cartilage is no longer feasible. This is probably due to the lack of resident blood vessels, lymphatic vessels, and nerves, and access to mesenchymal or circulating stem cells. From a clinical perspective, no effective clinical options exist for the amelioration of late-stage OA and options for patients remain limited to symptomatic treatment until becoming candidates for total joint replacement. In recent years, treatments that attempt to repair or restore the cartilage lesion have started to be developed to slow or stop the progression towards OA but, in general, these are unsatisfactory with variable and unpredictable results.<sup>8</sup> Given the poor regenerative capacity of cartilage, any microgravitational degradation would compromise flight crew mobility with the potential to negatively impact mission activities. Importantly, this damage may compromise the long-term joint health of flight personnel.

Is there evidence that biomechanical unloading impacts the joint? Data from terrestrial hind-limb unloading (HLU) animal experiments and human bed-rest studies demonstrate that reduced mechanical forces associated with joint unloading and immobilization leads to the proteoglycan loss in articular cartilage.9-12 Together, the joint trauma and unloading studies support the notion that a normal range of mechanical forces are critical for maintaining healthy joints. Interestingly, consistent with this hypothesis is the finding that active exercise prevented cartilage degradation in rats subjected to HLU<sup>13</sup> suggesting that exercise may be useful as a countermeasure to minimize joint pathology in microgravity. With the goal of minimizing long-term musculoskeletal problems, pre-flight and post-flight physiotherapy and conditioning regimes to complement mid-flight exercise are being considered. 14

If biomechanical unloading on Earth leads to cartilage breakdown, is there evidence that exposure to microgravity causes joint degradation? An early microgravity study reported smaller chondrogenic pellets, less proteoglycan synthesis and reduced dynamic stiffness of three-dimensional engineered cartilage constructs grown for 4 months on the Russian MIR craft. <sup>15</sup> More recently, experiments using simulated microgravity, 16, 17 and on parabolic flights, where repeated but short duration periods of microgravity are possible, have been conducted.<sup>18, 19</sup> In studies on cultured human chondrocytes, parabolic flight resulted in upregulation of cytoskeletal network genes and proteins suggesting even with short duration microgravity, cells respond by reorganizing the cytoskeleton.<sup>18</sup> Cytoskeletal reorganization were also reported in simulated microgravity experiments<sup>16, 17</sup> as was reduction in extracellular matrix production and mineralization.<sup>20, 21</sup> Our studies indicate that extended exposure to microgravity results in articular cartilage proteoglycan loss in mice (m/s in preparation). What about in humans? Cartilage degradation products would be present in synovial fluid, urine and blood of flight personnel. While elevated levels of bone resorption markers and calcium metabolism in body fluids of flight personnel have been reported,<sup>22</sup> there are no studies that focused on cartilage breakdown products released by damaged joints, such as carboxy-terminal telopeptides of Type II Collagen. This deficiency can be addressed by sampling the stored fluids of astronauts for cartilage biomarkers.

If cartilage loss in microgravity does occur, as seems likely based on unloading studies on Earth and our mouse findings, several fascinating questions arise.

- Degradation mechanisms—trauma vs. HLU vs. microgravity: Does the microgravity-induced cartilage breakdown occur via the same mechanisms as in the HLU model? If so, then HLU can be used to simulate the effects of microgravity. However, the possibility of a compounding effect of microgravity in addition to the unloading cannot be discounted. Similarly, are there common mechanistic steps between unloading and trauma that lead to cartilage destruction? The existence of a common response would suggest that findings from microgravity studies could inform mechanisms of OA with the potential to reveal new therapies.
- Biochemical threshold for tissue restoration: Are there specific biochemical or molecular thresholds that need to be breached for tissue restoration to fail? Our studies suggest that microgravity is sufficient for reversible proteoglycan loss in mouse femoral cartilage. In these studies, the lack of surface fibrillation and damage suggests that the collagen network is intact and that the point-of-no-return has not yet been reached. Microgravity studies on joint tissues could be used to determine the relationship between cellular and

Received: 22 November 2016 Revised: 13 February 2017 Accepted: 6 March 2017 Published online: 11 April 2017



- **n**pj
- molecular events and the potential for restorative tissue repair.
- 3. Gravitational threshold for tissue restoration: What is the minimal amount of gravity required to maintain healthy cartilage? If microgravity is detrimental to cartilage, would gravity at the surface of the moon (0.16 G), or Mars (0.38 G), be sufficient to maintain tissue regeneration pathways and prevent uncontrolled cartilage destruction? Efforts are underway to explore the use of fractional gravity generated by centrifugal forces to counteract the detrimental effects of microgravity. The application of partial gravity on long duration spaceflight may be the best countermeasure for a host of microgravity-induced pathologies.
- 4. Radiation and tissue regeneration: Does the increased radiation exposure of spaceflight affect tissue healing? The Earth's magnetic field shields craft in low earth orbit (LEO) from significant radiation. However, beyond LEO and protection from radiation, the effects of radiation on biological systems may be significant. A recent report suggesting that radiation worsens cartilage loss caused by HLU suggests that radiation has a compounding effect on cartilage destruction.<sup>10</sup>

If the goal of long duration spaceflight is to be achieved then more research is needed into the effects of microgravity on cells, tissue explants, and whole animals on orbiting platform both in LEO, and beyond. Monitoring of joint health on the ISS can be achieved by fluid biomarker analysis and the expansion of existing imaging technologies<sup>23</sup> to include cartilage. Understanding mechanisms of joint tissue damage in microgravity and the limitations governing consequent tissue repair on return to 1 G will provide insights into repair and regeneration processes for OA here on Earth.

## **COMPETING INTERESTS**

The author declares that he has no competing interests.

Jamie Fitzgerald<sup>1</sup>

Bone and Joint Center, Department of Orthopedic Surgery, Henry
Ford Hospital System, Integrative Biosciences building, 6135

Woodward Ave, Detroit, MI 48202, USA

Correspondence: Jamie Fitzgerald (jfitzge2@hfhs.org)

## **REFERENCES**

- Sibonga, J. D. et al. Recovery of spaceflight-induced bone loss: bone mineral density after long-duration missions as fitted with an exponential function. *Bone* 41, 973–978 (2007).
- Tesch, P. A., Berg, H. E., Bring, D., Evans, H. J. & LeBlanc, A. D. Effects of 17-day spaceflight on knee extensor muscle function and size. *Eur. J. Appl. Physiol.* 93, 463–468 (2005).
- Tanaka, K., Nishimura, N. & Kawai, Y. Adaptation to microgravity, deconditioning, and countermeasures. J. Physiol. Sci. 67, 271–281 (2017).
- Smith, S. M. et al. Fifty years of human space travel: implications for bone and calcium research. *Annu. Rev. Nutr.* 34, 377–400 (2014).
- Ohira, T., Kawano, F., Ohira, T., Goto, K. & Ohira, Y. Responses of skeletal muscles to gravitational unloading and/or reloading. J. Physiol. Sci. 65, 293–310 (2015).

- Buckwalter, J. A. & Martin, J. A. Osteoarthritis. Adv. Drug Deliv. Rev. 58, 150–167 (2006).
- 7. Kurz, B. et al. Pathomechanisms of cartilage destruction by mechanical injury. *Ann. Anat.* **187**, 473–485 (2005).
- Correa, D. & Lietman, S. A. Articular cartilage repair: current needs, methods and research directions. Semin. Cell Dev. Biol. doi:10.1016/j.semcdb.2016.07.013 (2016).
- Videman, T. Connective tissue and immobilization. Key factors in musculoskeletal degeneration? Clin. Orthop. Relat. Res. 221, 26–32 (1987).
- Willey, J. S. et al. Spaceflight-relevant challenges of radiation and/or reduced weight bearing cause arthritic responses in knee articular cartilage. *Radiat. Res.* 186, 333–344 (2016).
- Souza, R. B. et al. Effects of unloading on knee articular cartilage T1rho and T2 magnetic resonance imaging relaxation times: a case series. J. Orthop. Sports Phys. Ther. 42, 511–520 (2012).
- Ganse, B. et al. Muscular forces affect the glycosaminoglycan content of joint cartilage: unloading in human volunteers with the HEPHAISTOS lower leg orthosis. Acta Orthop. 86, 388–392 (2015).
- Luan, H. Q. et al. Use of micro-computed tomography to evaluate the effects of exercise on preventing the degeneration of articular cartilage in tail-suspended rats. Life Sci. Space Res. (Amst). 6, 15–20, doi:10.1016/j.lssr.2015.06.001 (2015).
- Lambrecht, G. et al. The role of physiotherapy in the European Space Agency strategy for preparation and reconditioning of astronauts before and after long duration space flight. *Musculoskelet. Sci. Pract.* 27, S15–S22 (2017).
- Freed, L. E., Langer, R., Martin, I., Pellis, N. R. & Vunjak-Novakovic, G. Tissue engineering of cartilage in space. *Proc. Natl. Acad. Sci. USA* 94, 13885–13890 (1997).
- Aleshcheva, G. et al. Changes in morphology, gene expression and protein content in chondrocytes cultured on a random positioning machine. PLoS One 8, e79057, doi:10.1371/journal.pone.0079057 (2013).
- Ulbrich, C. et al. Characterization of human chondrocytes exposed to simulated microgravity. Cell. Physiol. Biochem. 25, 551–560 (2010).
- Aleshcheva, G. et al. Moderate alterations of the cytoskeleton in human chondrocytes after short-term microgravity produced by parabolic flight maneuvers could be prevented by up-regulation of BMP-2 and SOX-9. FASEB J. 29, 2303–2314 (2015).
- Wehland, M. et al. Differential gene expression of human chondrocytes cultured under short-term altered gravity conditions during parabolic flight maneuvers. *Cell Commun. Signal.* 13, 18 (2015).
- Zhang, X. et al. The effects of simulated microgravity on cultured chicken embryonic chondrocytes. Adv. Space Res. 32, 1577–1583 (2003).
- Klement, B. J. & Spooner, B. S. Mineralization and growth of cultured embryonic skeletal tissue in microgravity. Bone 24, 349–359 (1999).
- Smith, S. M. et al. Bone markers, calcium metabolism, and calcium kinetics during extended-duration space flight on the mir space station. J. Bone Miner. Res. 20, 208–218 (2005).
- Orwoll, E. S. et al. Skeletal health in long-duration astronauts: nature, assessment, and management recommendations from the NASA Bone Summit. J. Bone Miner. Res. 28, 1243–1255, doi:10.1002/jbmr.1948 (2013).



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing,

adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <a href="https://creativecommons.org/licenses/by/4-0/">https://creativecommons.org/licenses/by/4-0/</a>.

© The Author(s) 2017