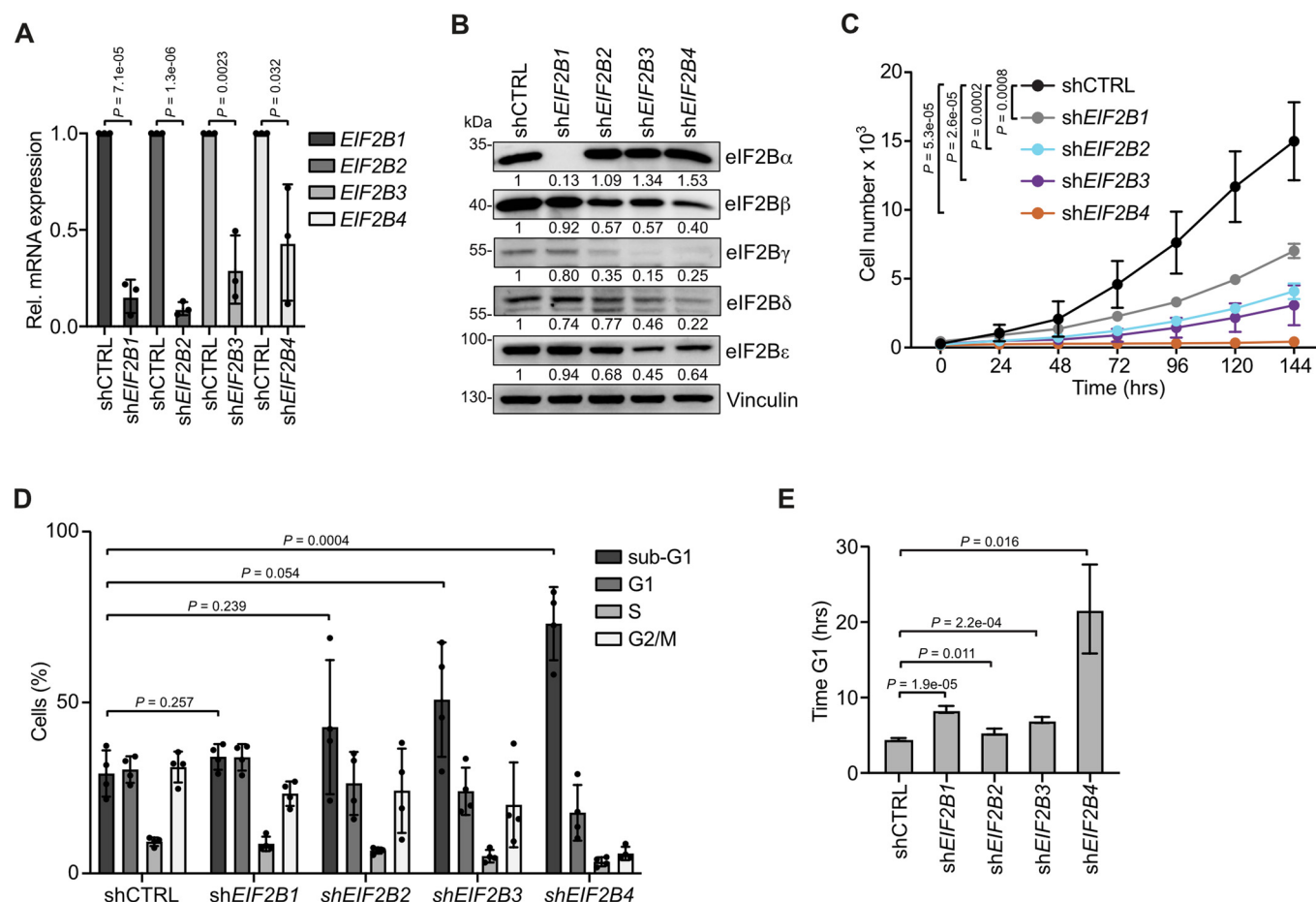
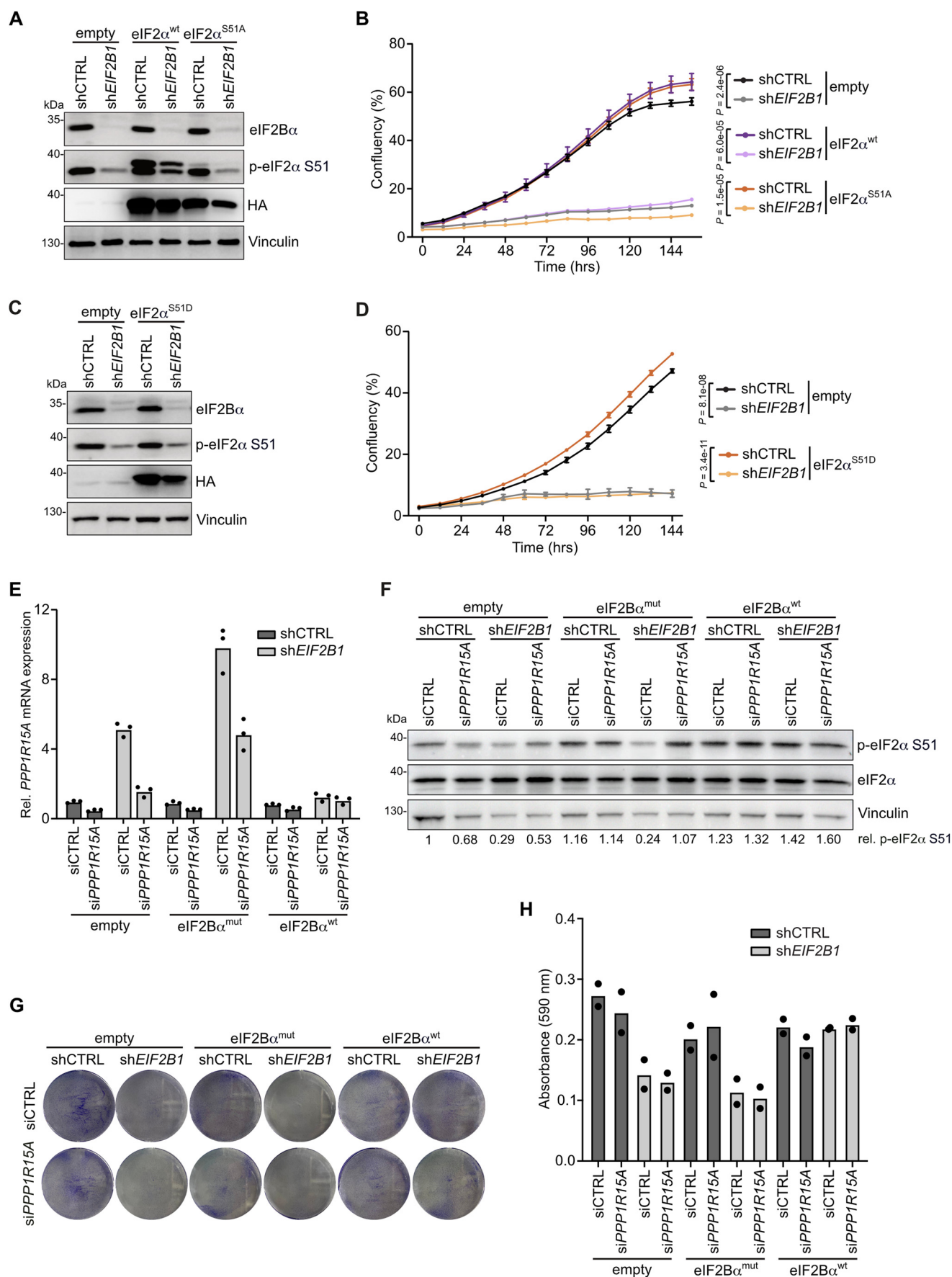


Expanded View Figures

**Figure EV1. DLD1 cells show reduced viability upon eIF2B subunit depletion.**

(A) mRNA expression of indicated genes in SW480 cells transduced with shCTRL or shRNAs against *EIF2B1-4*. Data show mean \pm s.d. ($n = 3$ biological replicates); Student's t test. (B) Western blot of indicated proteins in DLD1 cells transduced with shCTRL or shRNAs against *EIF2B1-4*, representative of three biological replicates with similar results. Levels of the respective eIF2B subunits, relative to vinculin, are given below each corresponding panel. (C) Growth curve of DLD1 cells transduced as described in (B), measured with Operetta screening microscope. Data show mean \pm s.d. ($n = 6$ biological replicates); Student's t test. (D) PI cell cycle FACS analysis of DLD1 cells transduced as described in (B). Data show mean \pm s.d. ($n = 4$ biological replicates); Student's t test. (E) Length of G1 cell cycle phase of DLD1 cells transduced as described in (B), calculated with data acquired from growth curve in (C) and PI cell cycle FACS in (D). Data show mean \pm s.e.m.; Student's t test. Source data are available online for this figure.



◀ **Figure EV2. Restoration of eIF2 α phosphorylation does not rescue the viability defect upon eIF2B α modulation.**

(A) Western blot of indicated proteins in shCTRL- or shEIF2B1-transduced SW480 cells stably overexpressing eIF2 α WT (eIF2 α^{wt}), eIF2 α S51A mutant (eIF2 α^{S51A}) construct, or without any overexpression (empty). The western blot is representative of three biological replicates with similar results. (B) Growth curve of SW480 cells transduced as described in (A), measured with Incucyte[®] live-cell imaging system. Data show mean \pm s.d. ($n = 4$ biological replicates); Student's t test. (C) Western blot of indicated proteins in shCTRL- or shEIF2B1-transduced SW480 cells stably overexpressing eIF2 α S51D mutant (eIF2 α^{S51D}) construct, or without any overexpression (empty). The western blot is representative of three biological replicates with similar results. (D) Growth curve of SW480 cells transduced as described in (C), measured with Incucyte[®] live-cell imaging system. Data show mean \pm s.d. ($n = 4$ biological replicates); Student's t test. (E) mRNA expression of PPP1R15A in shCTRL- or shEIF2B1-transduced SW480 cells stably overexpressing eIF2B α mutant (eIF2B α^{mut}), eIF2B α WT (eIF2B α^{wt}) construct, or without any overexpression (empty), transfected with siCTRL or siPPP1R15A for 72 hr. Data show mean of technical triplicates of one representative experiment ($n = 2$ biological replicates). (F) Western blot of indicated proteins in SW480 cells transduced and transfected as described in (E). The western blot is representative of two biological replicates with similar results. Levels of p-eIF2 α S51, relative to total eIF2 α , are given below. (G) Crystal violet staining of SW480 cells transduced and transfected as described in (E). Staining was done 7 days after seeding. Pictures are representative of two biological replicates with similar results. (H) Quantification of crystal violet staining described in (G). Data show mean ($n = 2$ biological replicates). Source data are available online for this figure.

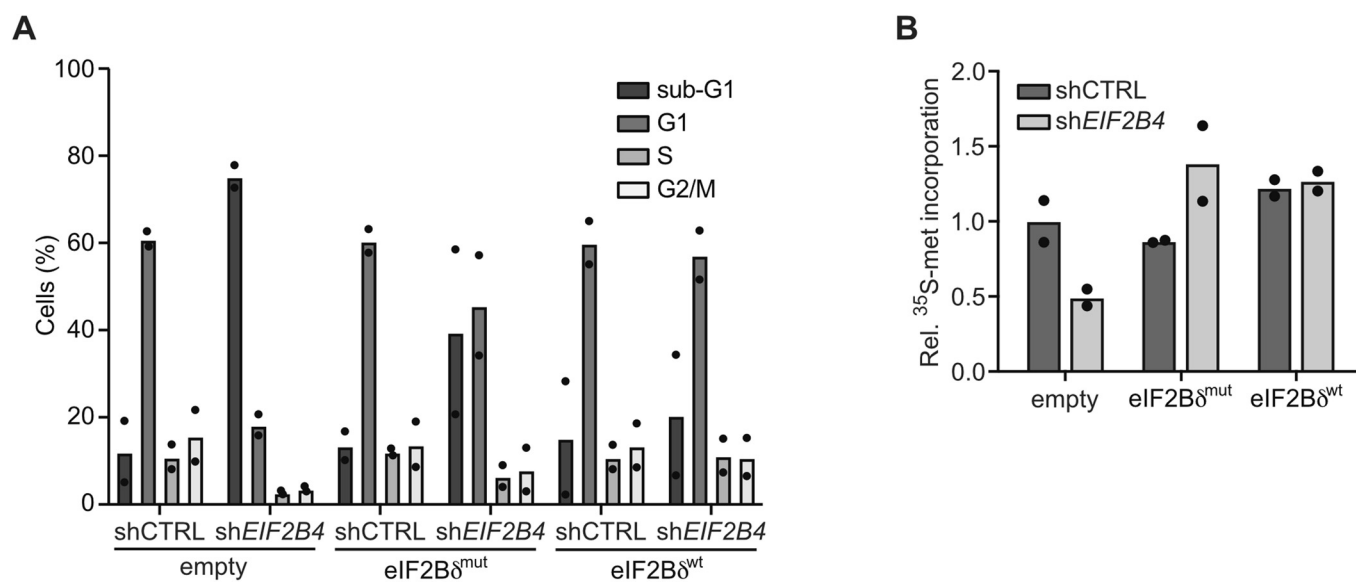


Figure EV3. eIF2B δ^{mut} expression rescues protein synthesis rates but not the viability defect upon eIF2B δ depletion.

(A) PI cell cycle FACS analysis of shCTRL- or shEIF2B4-transduced SW480 cells stably overexpressing eIF2B δ mutant (eIF2B δ^{mut}), eIF2B δ WT (eIF2B δ^{wt}) construct, or without any overexpression (empty). Data show mean ($n = 2$ biological replicates). (B) Relative ^{35}S -methionine incorporation of SW480 cells transduced as described in (A). Data show mean ($n = 2$ biological replicates). Source data are available online for this figure.

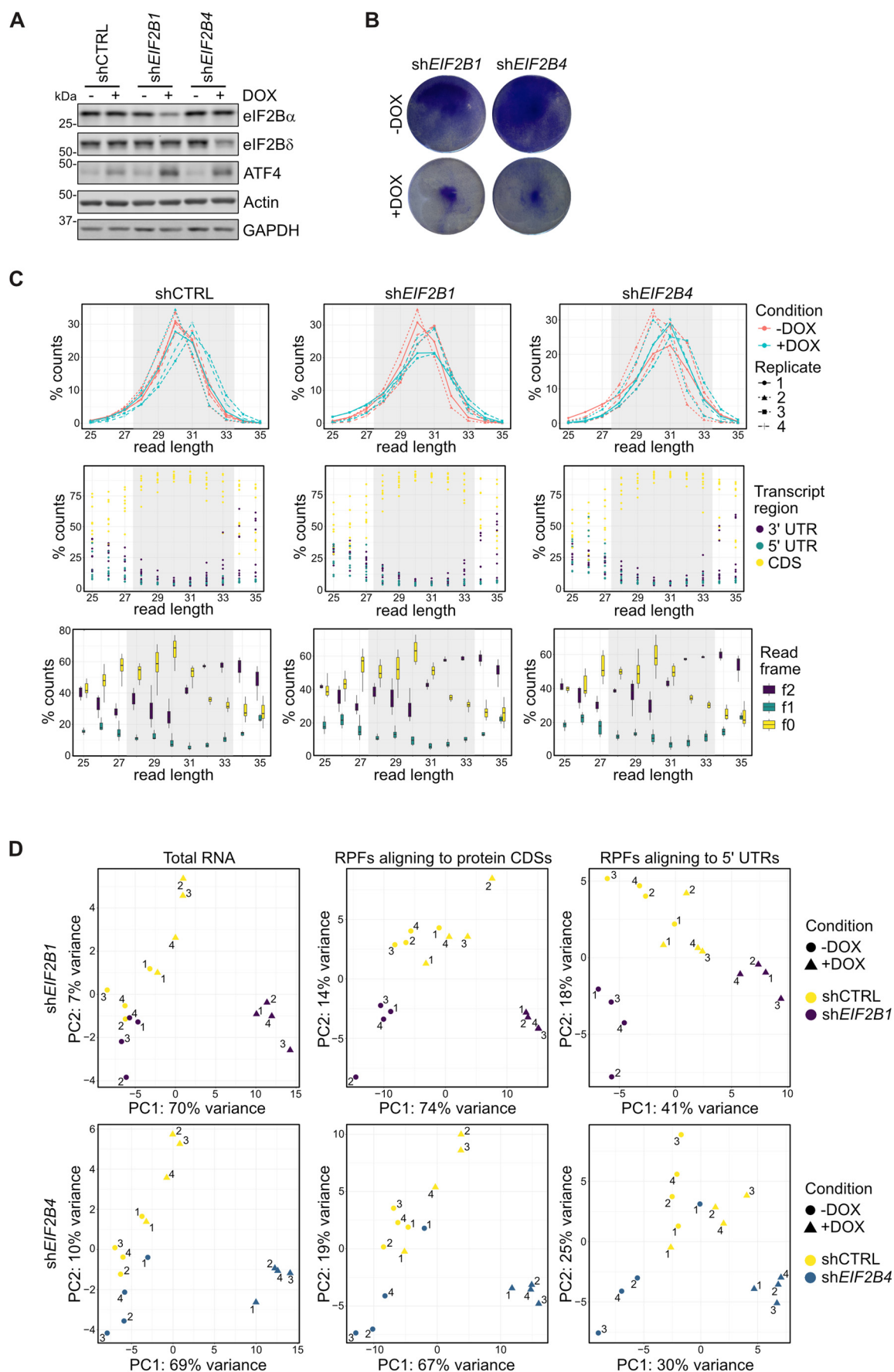




Figure EV4. Ribo-seq quality controls (QC).

(A) Western blot of indicated proteins in SW480 cells transduced with doxycycline-inducible shCTRL, shEIF2B1 or shEIF2B4 (5 days of doxycycline treatment (+ DOX), DMSO as control (-DOX)), representative of four biological replicates with similar results. GAPDH is probed on the same membrane as eIF2B δ , and is used as its loading control; Actin is probed on the same membrane as ATF4 and eIF2B α , and is used as loading control for these two targets. (B) Crystal violet staining of SW480 cells transduced with doxycycline-inducible shEIF2B1 or shEIF2B4 after 7 days of DOX treatment (+ DOX, EtOH as control (-DOX)). The experiment was performed once. (C) Ribo-seq QC plots; the gray shaded area in all graphs indicates reads of length 28–33 nt, representing the extracted reads for downstream analyses. Columns show results from cells transduced with respective doxycycline-inducible shRNA. The first row shows distribution of ribosome-protected fragments (RPFs) reads for -DOX and +DOX samples, with four replicates plotted independently. The second row shows the transcript region where the RPF reads align to; each dot represents an individual sample. The third row shows the read frame respective to the known codon position within the RPFs. Box plot summary statistics: boxes' lower and upper hinges represent first and third quantiles, lines represent the median, whiskers extend until last data point to a maximum of 1.5* inter-quantile range. (D) PCA plots for total cytoplasmic RNA sequencing (first column), RPFs aligning to protein-coding sequences (second column), or RPFs aligning to the 5' UTRs (third column). Data from each shRNA against either of the two subunits is plotted with the data for the shCTRL (yellow in all plots). Variance is displayed on each axis; size of axes is not variance-scaled. Source data are available online for this figure.

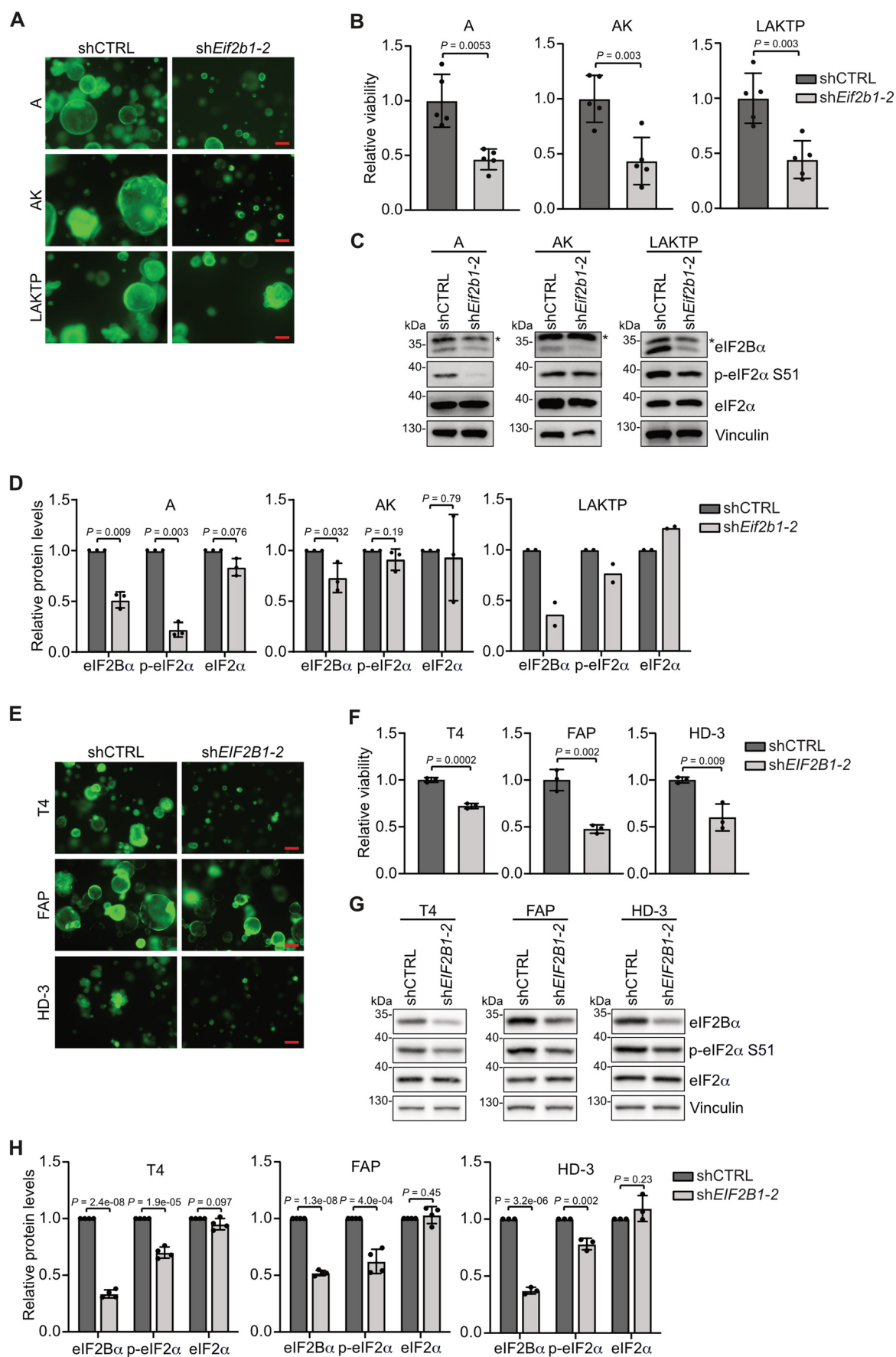




Figure EV5. Depletion of eIF2B α by a second independent shRNA reduces viability of murine and human tumor organoids.

(A) Pictures of murine A, AK and LAKTP intestinal organoids transduced with doxycycline-inducible shCTRL or sh*Elf2b1-2* (7 days of doxycycline treatment), representative of five biological replicates with similar results. Green signal (GFP) indicates shRNA induction. Scale bar = 200 μ m. (B) Relative viability of A, AK and LAKTP organoids transduced and treated as described in (A). Data show mean \pm s.d. ($n = 5$ biological replicates); Student's t test. (C) Western blot of indicated proteins in A, AK and LAKTP organoids transduced as described in (A), representative of two or three biological replicates with similar results (96 h of doxycycline treatment); *unspecific bands. (D) Quantification of eIF2B α , p-eIF2 α S51 and total eIF2 α levels, normalized to vinculin, of western blots described in (C). Data show mean \pm s.d. ($n = 2$ or 3 biological replicates); Student's t test. (E) Pictures of human T4, FAP, HD-3 PDOs transduced with doxycycline-inducible shCTRL or sh*EIF2B1-2* (7 days of doxycycline treatment), representative of three biological replicates with similar results. Green signal (GFP) indicates shRNA induction. Scale bar = 200 μ m. (F) Relative viability of T4, FAP, HD-3 PDOs transduced and treated as described in (E). Data show mean \pm s.d. ($n = 3$ biological replicates); Student's t test. (G) Western blot of indicated proteins in T4, FAP, HD-3 PDOs transduced as described in (E), representative of three or four biological replicates with similar results (96 h of doxycycline treatment). (H) Quantification of eIF2B α , p-eIF2 α S51 and total eIF2 α levels, normalized to vinculin, of western blots described in (G). Data show mean \pm s.d. ($n = 3$ or 4 biological replicates); Student's t test. Source data are available online for this figure.