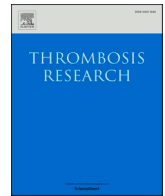




Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Letter to the Editors-in-Chief

Invited commentary to: ADAMTS13 deficiency is associated with abnormal distribution of von Willebrand factor multimers in patients with COVID-19 by Tiffany Pascreau et al. Letter to the Editors-in-Chief, Thrombosis Research

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) infection is the cause of a world-wide pandemic that has caused more than 2.5 million deaths worldwide as of mid February 2021 since its onset in December 2019. Severely affected patients suffer from pneumonia, necessitating invasive ventilation and several groups showed a high incidence and prevalence of venous and arterial thromboembolic events of large vessels and also microvascular thrombosis of the pulmonary circulation [1–3]. Hemostatic laboratory parameters at admission of COVID-19 patients, especially strongly elevated D-dimer levels, were predictive of mortality [4]. Tang et al. described that 15 of the 21 non-survivors among 183 consecutive patients with COVID-19 pneumonia fulfilled the ISTH criteria for overt disseminated intravascular coagulation (DIC) [5], whereas DIC was present in only 1 of 162 survivors [6]. Diagnosis of DIC in these patients was based on often strongly increased D-dimers, mildly/moderately prolonged prothrombin times and often mild thrombocytopenia [6]. In contrast to typical DIC associated with bacterial sepsis, obstetrical complications and other inflammatory conditions, fibrinogen levels in COVID-19 are mostly highly elevated and rarely fall below the critical limit of 100 mg/dL and thrombocytopenia is often mild or lacking [7,8]. Several authors suggested that the hemostaseologic alterations in COVID-19 together with autopsy and histopathologic findings reflected a form of thrombotic microangiopathy (TMA) rather than DIC [9–12]. Direct endothelial invasion by SARS-CoV-2 resulting in endothelial cell apoptosis [13], severe alveolar damage with occlusion of the pulmonary microcirculation [10,14–16] by fibrinous microthrombi [14–16], distinctive features of abnormal new vessel growth with intussusceptive angiogenesis [15], and deposition of complement activation products hinting at activation of the alternative and lectin based complement pathways [16] were reported in autopsy studies. Besides predominant pulmonary involvement, microvascular injury to the skin was observed in histopathologic studies of biopsies of COVID-19-associated skin rashes [16].

Compatible with severe endothelial damage, several authors noted marked elevations of Von Willebrand factor activity (VWF:act), VWF antigen (VWF:Ag) and factor VIII clotting activity (FVIII:C) in COVID-19 patients [7,8,11,17–20] and some found low normal or mildly decreased ADAMTS13 (A Disintegrin And Metalloprotease with Thrombospondin type 1 motif, number 13) activity [8,9,11,18–20]. The VWF:Ag/ADAMTS13 activity ratio was strongly elevated in these studies [8,11,18,20].

In this issue of Thrombosis Research, Pascreau et al. present a single tertiary care center study on 70 patients with COVID-19 pneumonia of variable severity ($n = 4$, treated as outpatients; $n = 44$, needing hospitalization on non-intensive care units; and $n = 22$, needing treatment on intensive care units) [21]. These authors are to be congratulated for

having performed a systematic study on the ADAMTS13-VWF axis in a large number of COVID-19 patients. VWF:act and VWF:Ag levels were significantly increased and the VWF:act/VWF:Ag ratio was slightly lower in both non-ICU and ICU patients with COVID-19 as compared to healthy controls. ADAMTS13 antigen values, on the other hand, were significantly lower in hospitalized COVID-19 patients than in controls, whereby the ICU patients tended to have lower ADAMTS13 antigen than the non-ICU patients (no significant difference). The ADAMTS13:Ag in the 70 patients correlated negatively with the VWF:Ag and also with C-reactive protein. In a subgroup of 10 patients, Pascreau et al. analyzed the VWF multimeric distribution using the commercial Hydrigel 5 von Willebrand factor multimer kit (Sebia) and found a relative decrease of high molecular weight VWF multimers and a relative increase of intermediate and low molecular VWF multimers in 8/10 patients [21]. This imbalance of a markedly increased VWF and moderately decreased ADAMTS13 levels with an elevated VWF/ADAMTS13 ratio is suggested to represent a «consumption» of ADAMTS13 by the massively increased VWF and proposed to contribute to the pulmonary microthrombi formation [21]. The authors note that in contrast to the disease thrombotic thrombocytopenic purpura (TTP), defined by a very severe autoantibody-mediated or congenital deficiency of the ADAMTS13 [22,23], the hallmarks of severe thrombocytopenia and microangiopathic hemolytic anemia are lacking in almost all COVID-19 patients [7,8,20]. Also, the thrombi of the pulmonary microcirculation have mostly been reported as predominantly «fibrinous» [14–16], even though Fox et al. suggested the presence of platelets and VWF in the microthrombi of pulmonary alveolar capillaries by immunostaining [24].

Pascreau et al. [21] as well as other investigators [9,11,18,20] propose that the VWF-ADAMTS13 dysbalance may be causally related to the prothrombotic tendency and the (predominantly pulmonary) microvascular thrombosis. Based on this hypothesis, it is proposed that supplementation of ADAMTS13 [21] or the use of caplacizumab blocking the VWF A1 domain interaction with platelet glycoprotein Ib-alpha might be useful therapeutically in severe COVID-19 [11]. Caplacizumab has been found efficacious as an adjunctive therapy to plasma exchange, fresh frozen plasma replacement and immunosuppression in patients suffering from acute autoimmune TTP with severe acquired ADAMTS13 deficiency, resulting in a faster normalization of the severely decreased platelet count as compared to placebo [25,26]. Nevertheless, in Covid-19 patients there is mostly no severe thrombocytopenia and no microangiopathic hemolytic anemia [8,20] which are the diagnostic hallmarks of classic TMAs [22].

We offer the following pathophysiologic hypothesis: the severe endothelial damage caused by direct SARS-CoV-2 invasion leads to

<https://doi.org/10.1016/j.thromres.2021.02.030>

Received 19 February 2021; Accepted 22 February 2021

Available online 3 March 2021

0049-3848/© 2021 Elsevier Ltd. All rights reserved.

profound and prolonged release of VWF from endothelial storage sites, the Weibel-Palade-bodies. The ADAMTS13 is partially trapped to the endothelial surface where it cleaves the nascent unusually large VWF on the endothelial surface [27] resulting in a mild to moderate decrease of circulating ADAMTS13 similar to the situation in severe sepsis or septic shock [28] or, as a short-lived phenomenon, after excessive VWF release induced by endotoxin or desmopressin application in healthy volunteers [29]. This is compatible with the highly significant negative correlation of the ADAMTS13 antigen with increasing VWF antigen as shown by Pascreau et al. in 70 COVID-19 patients [21] and the mildly decreased ADAMTS13 could represent a mere epiphenomenon. The widespread thrombi in the pulmonary microcirculation consist mainly of fibrin [14–16] similar to the predominantly renal microthrombi in hemolytic uremic syndrome [30]. The strongly elevated FVIII:C levels in severe COVID-19 [7,19] may enhance thrombin and fibrin generation. The fact that COVID-19-associated coagulopathy with its predominantly pulmonary thrombotic microangiopathy is not accompanied by severe thrombocytopenia and intravascular red blood cell fragmentation with schistocytes [8,20], in contrast to the classic TMAs such as TTP or HUS [22], may hypothetically be due to the much lower blood pressure gradient in the pulmonary as compared to the peripheral arterial circulation [8]. Certainly, this hypothesis needs further confirmation.

Declaration of competing interest

B.L. is Chairman of the Data Safety Monitoring Committee of the BAXALTA 281102 and the SHIRE SHP655-201 studies (now both run by TAKEDA), investigating recombinant ADAMTS13 therapy in hereditary and acquired TTP, respectively. He is on the Advisory Board of Sanofi for caplacizumab, and received travel and accommodation support for participating at scientific meetings and/or lecture fees from Ablynx, Alexion, Bayer, Roche, Sanofi and Siemens.

References

- [1] S. Cui, S. Chen, X. Li, S. Liu, F. Wang, Prevalence of venous thromboembolism in patients with severe novel coronavirus pneumonia, *J. Thromb. Haemost.* 18 (2020) 1421–1424.
- [2] F.A. Klok, M.J.H.A. Kruip, N.J.M. Van Der Meer, M.S. Arbous, D. Gommers, K. M. Kant, et al., Confirmation of the high cumulative incidence of thrombotic complications in critically ill ICU patients with COVID-19: an updated analysis, *Thromb. Res.* 191 (2020) 148–150.
- [3] C. Lodigiani, G. Iapichino, L. Carenzo, M. Cecconi, P. Ferrazzi, T. Sebastian, et al., Venous and arterial thromboembolic complications in COVID-19 patients admitted to an academic hospital in Milan, Italy, *Thromb. Res.* 191 (2020) 9–14.
- [4] L. Zhang, X. Yan, Q. Fan, H. Liu, X. Liu, Z. Liu, et al., D-dimer levels on admission to predict in-hospital mortality in patients with Covid-19, *J. Thromb. Haemost.* 18 (2020) 1324–1329.
- [5] F.B. Taylor, C.H. Toh, W.K. Hoots, H. Wada, M. Levi, Towards definition, clinical and laboratory criteria, and a scoring system for disseminated intravascular coagulation, *Thromb. Haemost.* 86 (2001) 1327–1330.
- [6] N. Tang, D. Li, X. Wang, Z. Sun, Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia, *J. Thromb. Haemost.* 18 (2020) 844–847.
- [7] J. Helms, C. Tacquard, F. Severac, I. Leonard-Lorant, M. Ohana, X. Delabranche, et al., High risk of thrombosis in patients with severe SARS-CoV-2 infection: a multicenter prospective cohort study, *Intensive Care Med.* 46 (2020) 1089–1098.
- [8] T. Falter, H. Rossmann, P. Menge, J. Goetje, S. Groenwoldt, A. Weinmann, et al., No evidence for classic thrombotic microangiopathy in COVID-19, *J. Clin. Med.* 10 (2021) 671.
- [9] N. Martinelli, M. Montagnana, F. Pizzolo, S. Friso, G.L. Salvagno, G.L. Forni, et al., A relative ADAMTS13 deficiency supports the presence of a secondary microangiopathy in COVID-19, *Thromb. Res.* 193 (2020) 170–172.
- [10] F.S. Beigee, M.P. Toutkaboni, N. Khalili, S.A. Nadji, A. Dorudinia, M. Rezaei, et al., Diffuse alveolar damage and thrombotic microangiopathy are the main histopathological findings in lung tissue biopsy samples of COVID-19 patients, *Pathol. Res. Pract.* 216 (2020) 153228.
- [11] B.M. Henry, S.W. Benoit, M.H. Santos de Oliveira, G. Lippi, E.J. Favaloro, J. L. Benoit, ADAMTS13 activity to von Willebrand factor antigen ratio predicts acute kidney injury in patients with COVID-19: evidence of SARS-CoV-2 induced secondary thrombotic microangiopathy, *Int. J. Lab. Hematol.* (2020) 1–8.
- [12] C. Diorio, K.O. Mc Nerney, M. Lambert, P. Paessler, E.M. Anderson, S. E. Henrickson, et al., Evidence of thrombotic microangiopathy in children with SARS-CoV-2 across the spectrum of clinical presentations, *Blood Adv.* 4 (2020) 6051–6063.
- [13] Z. Varga, A.J. Flammer, P. Steiger, M. Haberecker, R. Andermatt, A.S. Zinkernagel, et al., Endothelial cell infection and endotheliitis in COVID-19, *Lancet* 395 (2020) 1417–1418.
- [14] M. Dolnikoff, A.N. Duarte-Neto, R.A. de Almeida Monteiro, L.F. Ferraz da Silva, E. P. de Oliveira, P.H. Nascimento Saldiva, et al., Pathological evidence of pulmonary thrombotic phenomena in severe COVID-19, *J. Thromb. Haemost.* 18 (2020) 1517–1519.
- [15] M. Ackermann, S.E. Verleden, M. Kuehnel, A. Haverich, T. Welte, F. Laenger, et al., Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in Covid-19, *N. Engl. J. Med.* 383 (2020) 120–128.
- [16] C. Magro, J.J. Mulvey, D. Berlin, G. Nuovo, S. Salvatore, J. Harp, et al., Complement associated microvascular injury and thrombosis in the pathogenesis of severe COVID-19 infection: a report of five cases, *Transl. Res.* 220 (2020) 1–13.
- [17] R. Escher, N. Breakey, B. Lämmle, Severe COVID-19 infection associated with endothelial activation, *Thromb. Res.* 190 (2020) 62.
- [18] A. Huisman, R. Beun, M. Sikma, J. Westerink, N. Kusadasi, Involvement of ADAMTS13 and von Willebrand factor in thromboembolic events in patients infected with SARS-CoV-2, *Int. J. Lab. Hematol.* 42 (2020) e211–e212.
- [19] R. Escher, N. Breakey, B. Lämmle, ADAMTS13 activity, von Willebrand factor, factor VIII and D-dimers in COVID-19 inpatients, *Thromb. Res.* 192 (2020) 174–175.
- [20] I. Mancini, L. Baronciani, A. Artoni, P. Colpani, M. Biganzoli, G. Cozzi, et al., The ADAMTS13-Von Willebrand factor axis in COVID-19 patients, *J. Thromb. Haemost.* (2020), <https://doi.org/10.1111/jth.15191>.
- [21] T. Pascreau, S. Zia-Chahabi, B. Zuber, C. Tcherakian, E. Farfour, M. Vasse. ADAMTS13 deficiency is associated with abnormal distribution of von Willebrand factor multimers in patients with COVID-19, *Thromb. Res.*
- [22] M. Scully, S. Cataland, P. Coppo, J. de la Rubia, K.D. Friedman, J.A. Kremer Hovinga, et al., Consensus on the standardization of terminology in thrombotic thrombocytopenic purpura and related thrombotic microangiopathies, *J. Thromb. Haemost.* 15 (2017) 312–322.
- [23] X.L. Zheng, S.K. Vesely, S.R. Cataland, P. Coppo, B. Geldziler, A. Iorio, et al., ISTH guidelines for the diagnosis of thrombotic thrombocytopenic purpura, *J. Thromb. Haemost.* 18 (2020) 2486–2495.
- [24] S.E. Fox, A. Akmatbekov, J.L. Harbert, G. Li, J. Quincy Brown, R.S. Vander Heide, Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans, *Lancet Respir. Med.* 8 (2020) 681–686.
- [25] M. Scully, S.R. Cataland, F. Peyvandi, P. Coppo, P. Knöbl, J.A. Kremer Hovinga, et al., Caplacizumab treatment for acquired thrombotic thrombocytopenic purpura, *N. Engl. J. Med.* 380 (2019) 335–346.
- [26] P. Coppo, M. Bubenheim, E. Azoulay, L. Galicier, S. Malot, N. Bigé, et al., A regimen with caplacizumab, immunosuppression, and plasma exchange prevents unfavorable outcomes in immune-mediated TTP, *Blood* 137 (2021) 733–742.
- [27] J.F. Dong, J.L. Moake, L. Nolasco, A. Bernardo, W. Arceneaux, C.N. Shrimpton, et al., ADAMTS-13 rapidly cleaves newly secreted ultralarge von Willebrand factor multimers on the endothelial surface under flowing conditions, *Blood* 100 (2002) 4033–4039.
- [28] J.A. Kremer Hovinga, S. Zeerleder, P. Kessler, T. Romani de Wit, J.A. van Mourik, C.E. Hack, et al., ADAMTS-13, von Willebrand factor and related parameters in severe sepsis and septic shock, *J. Thromb. Haemost.* 5 (2007) 2284–2290.
- [29] R.A. Reiter, K. Varadi, P.L. Turecek, B. Jilma, P. Knöbl, Changes in ADAMTS13 (von Willebrand factor-cleaving protease) activity after induced release of von Willebrand factor during acute systemic inflammation, *Thromb. Haemost.* 93 (2005) 554–558.
- [30] H.M. Tsai, W.L. Chandler, R. Sarode, R. Hoffman, S. Jelacic, R.L. Habeeb, et al., Von Willebrand factor and Von Willebrand factor-cleaving metalloprotease activity in Escherichia coli O157:H7-associated hemolytic uremic syndrome, *Pediatr. Res.* 49 (2001) 653–659.

Bernhard Lämmle^{a,b,c,*}, Heidi Rossmann^{a,d}

^a Center for Thrombosis and Hemostasis, University Medical Center of the Johannes Gutenberg University Mainz, 55131 Mainz, Germany

^b Department of Hematology and Central Hematology Laboratory, Inselspital, Bern University Hospital, University of Bern, CH 3010 Bern, Switzerland

^c Haemostasis Research Unit, University College London, London WC1E 6BT, UK

^d Institute of Clinical Chemistry and Laboratory Medicine, University Medical Center of the Johannes Gutenberg University Mainz, 55131 Mainz, Germany

* Corresponding author at: Schützenweg 3, CH 3065, Bolligen, Switzerland.

E-mail address: bernhard.laemmle@uni-mainz.de (B. Lämmle).