## ORIGINAL ARTICLE



# Evaluation of the Safety, Tolerability, and Pharmacokinetics of Gammaplex<sup>®</sup> 10% Versus Gammaplex<sup>®</sup> 5% in Subjects with Primary Immunodeficiency

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#### Abstract

*Purpose* This phase 3, multicenter, open-label, randomized, two-period, crossover bioequivalence trial evaluated the safety, tolerability, and pharmacokinetics of intravenous immunoglobulins (IVIGs) Gammaplex 5% and Gammaplex 10% in 33 adults and 15 children with primary immunodeficiency diseases (PIDs).

*Methods* Eligible adults received five Gammaplex 5% infusions followed by five Gammaplex 10% infusions, or vice versa, stratified by a 21- or 28-day dosing regimen. Pediatric subjects received five Gammaplex 10% infusions only.

*Results* The primary objective, to demonstrate the bioequivalence of Gammaplex 10% and Gammaplex 5% at the 28-day dosing interval, was met based on the Gammaplex 10%/ Gammaplex 5% ratio of area under the concentration versus time curve (AUC<sub>0-28</sub>) values. Throughout the study, total

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immunoglobulin G trough levels were well maintained, with total values generally  $\geq 600 \text{ mg/dL}$  (minimum level for study inclusion). At the dosing schedules and infusion rates used in this study, safety and tolerability were comparable and acceptable in adult and pediatric PID subjects treated with Gammaplex 10% and 5%.

*Conclusions* In this study, the first direct comparison of 5% IVIG and 10% IVIG products in PID subjects, the pharmacokinetic analysis demonstrated bioequivalence of Gammaplex 10% and Gammaplex 5% at the 28-day dosing interval. The Gammaplex 10% formulation was safe and well tolerated in pediatric and adult PID subjects. Based on the results from this bridging study in PID subjects, Gammaplex 10% could be expected to have a therapeutic effect similar to the licensed Gammaplex 5%, which has demonstrated efficacy and tolerability in patients with PID and idiopathic thrombocytopenic purpura.

**Keywords** Primary immunodeficiency disease (PID)  $\cdot$ intravenous immunoglobulin (IVIG)  $\cdot$  Gammaplex<sup>®</sup> 5%  $\cdot$  Gammaplex<sup>®</sup> 10%

# Introduction

Primary immunodeficiency diseases (PID) comprise a diverse range of disorders, most of which are characterized by defects in antibody production and increased susceptibility to infection [1]. Replacement therapy with immunoglobulin G (IgG) purified from human plasma has been the standard of care since the early 1950s. By the 1960s, IgG products were administered intramuscularly, but efficacy was limited by the relatively small quantities that could be administered by that route due to discomfort and painful local side effects [2, 3]. In the 1980s, intravenous immunoglobulin (IVIG) became available in the USA and became the most common treatment for PID patients with impaired humoral immunity [4–6]. Currently, IVIG is available at concentrations of 3%, 5%, 6%, 9%, 10%, and 12%, with more products available at 10% than at any other concentration [7, 8].

Gammaplex<sup>®</sup> 5% is a highly purified human IgG liquid product intended for intravenous (IV) administration (Bio Products Laboratory Ltd., Elstree, UK) and approved for use in the USA and the UK for PID and immune thrombocytopenic purpura (ITP) [9, 10]; Gammaplex 5% is also licensed in Brazil, Brunei, Israel, Lebanon, and Malta. Previous studies of Gammaplex 5% demonstrated efficacy in preventing serious acute bacterial infections (SABIs) and demonstrated tolerability both in patients with PID [11, 12] and those with ITP [13]. Like Gammaplex 5% (Gammaplex [5%] 5 g in 100 mL), Gammaplex 10% (Gammaplex [10%] 10 g in 100 mL) is a ready-prepared solution for IV administration containing the active ingredient human normal IgG. Both products are manufactured using the same process, but Gammaplex 10% is more concentrated, with an IgG concentration of 100 g/L, and is stabilized with glycine, whereas Gammaplex 5% is stabilized with sorbitol. This study evaluated the bioequivalence of the currently licensed formulation (Gammaplex 5%) and the new formulation (Gammaplex 10%), as well as pharmacokinetics (PK), safety, and tolerability of Gammaplex 10%, in adult and pediatric subjects with PID. This article is the first report of a clinical study of Gammaplex 10% and of the first direct comparison between a 5% and a 10% IVIG product in PID patients.

## Methods

## **Study Design**

This bridging study was a phase 3, multicenter, open-label, randomized, two-period, crossover bioequivalence trial to evaluate the PK, safety, and tolerability of Gammaplex 10% in PID (ClinicalTrials.gov, NCT01963143; EudraCT, 2013-002290-21) versus the PK, safety, and tolerability of Gammaplex 5% [12]. Approximately 1 to 3 days before the first planned infusion (visit 1), eligible adult subjects were randomized (1:1) to the following treatment sequences and stratified by dosing regimen (21 or 28 days): either five infusions of Gammaplex 5% followed by five infusions of Gammaplex 10% (sequence 1), or five infusions of Gammaplex 10% followed by five infusions of Gammaplex 5% (sequence 2). Pediatric subjects received five infusions of Gammaplex 10% only. The enrollment of the pediatric cohort was delayed until acceptable safety and tolerability had been demonstrated in  $\geq$ 5 adult subjects. Samples for PK profiling were collected at specified times for the final infusion of each

treatment (visits 5 and 10 for adult subjects; visit 5 for pediatric subjects).

Gammaplex 5% and Gammaplex 10% were dosed at 300-800 mg/kg per infusion every 21 or 28 days. The selected dose was the same as the dose of IVIG administered during the 3 months prior to study entry. Subjects were to be adequately hydrated prior to infusion, but a pre-infusion hydration step was not required for either product. To provide an initial equivalent protein dose, the initial infusion rates were 0.01 and 0.005 mL/kg/min for Gammaplex 5% and Gammaplex 10%, respectively, for the first 15 min. Subjects who tolerated the first infusion with minimal adverse reactions could begin subsequent infusions at 0.02 mL/kg/min (Gammaplex 5%) and 0.01 mL/kg/min (Gammaplex 10%) (Table 1). For each infusion, the rate was increased incrementally at 15-min intervals, reaching a maximum infusion rate for both Gammaplex 5% and Gammaplex 10% of 0.08 mL/kg/min, depending on the subjects' ability to tolerate slower infusions. Additional infusions were permitted to ensure that subjects received five infusions of Gammaplex 10% or Gammaplex 5% at the same dose and were on stable treatment prior to the PK sampling. Infusion visits were scheduled every 21 or 28 days, depending on the subject's cycle of infusions during prior IVIG treatment. Individual infusions could be administered  $\pm 3$  days of the planned schedule. Subjects returned to the clinic 6 to 7 days after the first infusion of either treatment to assess safety (after visit 1 for pediatric subjects; after visits 1 and 6 for adult subjects). Subjects returned for an end-of-study visit 28 days after the final study infusion. The total study duration

 Table 1
 Infusion rates for Gammaplex 10% and Gammaplex 5%

	Infusion rate	Elapsed time	
	(mL/kg/min)	(mg/kg/h)	(min)
Gammaplex 5%			
Starting infusion	0.01 <sup>a</sup>	30	0-15
Incremental rate increases	0.02	60	16–30
	0.04	120	31-45
	0.06	180	46-60
Maximum rate	0.08	240	61 until end of infusion
Gammaplex 10%			
Starting infusion rate	$0.005^{a}$	30	0-15
Incremental rate increases	0.01	60	16–30
	0.02	120	31-45
	0.04	240	46-60
	0.06	360	61–75
Maximum rate	0.08	480	76 until end of infusion

<sup>a</sup> Subjects who tolerated the first infusion with minimal adverse reactions could begin subsequent infusions at 0.02 mL/kg/min (Gammaplex 5%) or 0.01 mL/kg/min (Gammaplex 10%)

(screening to end-of-study visit) was 38–48 weeks for any given adult subject and 23–28 weeks for any given pediatric subject, depending on the subject's frequency of infusion.

## **Compliance with Ethical Standards**

The protocol for this study was approved by an institutional review board or independent ethics committee for each of 16 study centers in the USA (13 centers), the UK (2 centers), and Hungary (1 center). This study was conducted in accordance with the ICH Harmonised Tripartite Guidelines for Good Clinical Practice [14], and each study participant (or their parent/guardian, if applicable) provided written informed consent to participate in the study. Assents were obtained according to local regulations.

## **Subject Selection**

Study candidates met the following eligibility criteria: aged 16–55 years inclusive (adult cohort) or aged 2–15 years inclusive and body weight  $\geq$ 10 kg (pediatric cohort); diagnosis of PID with hypogammaglobulinemia; currently receiving a licensed (or investigational stage 3 or 3b) IVIG at a dose that had not changed by ±50% of the mean dose for  $\geq$ 3 months before study entry and was 300–800 mg/kg per infusion, with an infusion interval of every 21 or 28 days; and trough IgG level of  $\geq$ 6 g/L (600 mg/dL), with  $\geq$ 1 documented trough level available from the 3 months before screening.

Excluded subjects were those with a history of anaphylactic reaction to blood or blood-derived products; abnormal liver and/or renal function; selective immunoglobulin A (IgA) deficiency, history of reaction to products containing IgA, or history of antibodies to IgA; cellular or innate impaired immunity; evidence of an active infection at the time of enrollment; currently receiving or had received any investigational agent other than an IVIG product within the prior 3 months; pregnant or nursing; positive HIV-1 and HIV-2, hepatitis C virus, or hepatitis B surface antigen testing; history of deep vein thrombosis or thrombotic complications of IVIG therapy; or intolerance to any component of Gammaplex. The use of local anesthetics, antipyretics, antihistamines, analgesics, and antiemetics prior to infusion was allowed, but routine longterm use of corticosteroids (apart from oral and parenteral steroids if average daily dose was <0.15 mg of prednisone equivalent/kg/day) was not permitted.

## Outcomes

As this was a bridging study to evaluate the bioequivalence and compare the safety and tolerability of the two formulations of Gammaplex, no formal efficacy analyses were performed. The primary analysis was the bioequivalence assessment of Gammaplex 10% and Gammaplex 5% in adult subjects at their final infusion (PK infusion) on a 28-day treatment schedule. The primary outcome was area under the concentration versus time curve from time=0 to time=28 days (AUC<sub>0-28</sub>; calculated by the linear trapezoidal rule). Secondary outcomes included a bioequivalence assessment of Gammaplex 10% and Gammaplex 5% using area under the concentration versus time curve from time=0 to time=21 days (AUC<sub>0-21</sub>; calculated by the linear trapezoidal rule) determined in adult subjects on a 21-day treatment schedule at the final infusion in each treatment period (PK infusion).

Additional PK parameters measured during and after the PK infusion included maximum observed concentration ( $C_{\text{max}}$ ), time at which  $C_{\text{max}}$  was apparent ( $t_{\text{max}}$ ), systemic clearance (CL, calculated as dose/AUC<sub>0- $\tau$ </sub> [where  $\tau$ is the dosing interval]), apparent volume of distribution at steady state ( $V_{ss}$ , calculated as CL·MRT; MRT, the mean residence time at steady state, was calculated as AUMC<sub>0- $\tau$ </sub> +  $\tau$ ·AUC<sub> $\tau-\infty$ </sub>/AUC<sub>0- $\tau$ </sub> - T/2 [where AUMC is the area under the first moment curve, T is the infusion duration, and  $\tau$  is the dosing interval]), apparent terminal half-life  $(t_{1/2})$ , and trough IgG levels. Trough levels of total IgG were measured before every infusion and at end of study, and trough levels by IgG subclass and of IgG antibodies to specific antigens (Haemophilus influenzae type b and Streptococcus pneumoniae) were measured before the first infusion of each Gammaplex formulation and at end of study. Baseline IgG levels for each formulation were defined as those prior to the first study dose of Gammaplex. At the end of each treatment period, IgG levels were assessed within 10 min before the end of the infusion and at the following times after the end of the infusion: 60 min; 3, 6, 24, and 48 h; and 4, 7, 14, and 21 days (and at 28 days for subjects on the 28-day regimen).

Safety assessments included the number and percentage of adverse events (AEs; defined as those events with onset between the first infusion date and 28 days after the last infusion); number and percentage of product-related AEs (defined as AEs in which the relationship to the study drug was recorded as "possible," "probable," or "very likely/certain," or in which information regarding the relationship to the study drug was unavailable); number and percentage of adverse reactions (ARs; defined as AEs [irrespective of causality] that occurred during the infusion or within 72 h after completion of the infusion, any product-related AEs, or any AEs with unknown causality); vital signs; clinical laboratory tests (including tests for hemolysis); transmission of viruses; and physical examination.

Safety variables were determined for Gammaplex 10% in all adult and pediatric subjects and for Gammaplex 5% in all adult subjects. AEs were coded using the Medical Dictionary for Regulatory Activities (version 17.0).

## **Statistical Analysis**

PK analyses (AUC<sub>0-72</sub>  $C_{\text{max}}$ ,  $t_{\text{max}}$ , CL,  $V_{\text{ss}}$ ,  $t_{1/2}$ , and trough IgG levels for PK infusions) were performed on the PK population, defined as all subjects who received regular doses of Gammaplex (i.e., assumed to be at steady state) and from whom an appropriate PK profile was obtained over the 21or 28-day period. Gammaplex PK was based on absolute (unadjusted) and baseline-adjusted (i.e., adjusted for predose concentrations) values and were calculated using a noncompartmental method. All trough IgG levels were analyzed unadjusted. Actual sample times, rather than nominal times, were used to calculate PK parameters. The AUC<sub> $0-\tau$ </sub> on which bioequivalence was based was log-transformed prior to analysis. Adult subjects on the 28-day regimens were analyzed using an analysis of variance model, including fixed effect terms for sequence, subject within sequence, period, and formulation (Gammaplex 10% or Gammaplex 5%). The 90% confidence interval (CI) for the difference between formulations was obtained and back transformed to obtain a CI for the ratio between formulations on the original scale. To show bioequivalence of the two formulations, this backtransformed 90% CI needed to lie between 0.8 and 1.25. Safety parameters (including AEs and adverse reactions) and trough levels of total IgG, IgG subclasses, and IgG antibodies to specific antigens were summarized using the intent-to-treat population, which was defined as all subjects who received ≥1 infusion of Gammaplex 10% or Gammaplex 5%.

# Results

#### **Subject Disposition**

A total of 48 subjects were enrolled in this study; 33 adults were randomized to sequence 1 or sequence 2 over 21-day (n = 14) or 28-day (n = 19) infusion schedules, and 15 nonrandomized pediatric subjects were enrolled to receive Gammaplex 10% over 21-day (n = 7) or 28-day (n = 8)

infusion schedules (Fig. 1). Thirty-two of 33 adult subjects completed the study; one adult withdrew during the first infusion due to inconvenience of study visits. Of the 32 adult subjects who completed the study, 30 were eligible for the PK analysis (n = 14 and n = 15 for the 21- and 28-day infusion schedules, respectively). Fourteen of 15 pediatric subjects completed the study; one child was withdrawn from the study after the fourth infusion at the investigator's discretion due to consistently low IgG trough levels despite a dose increase. Of the 14 pediatric subjects who completed the study, 13 were eligible for the PK analysis (n = 7 and n = 6 for the 21- and 28-day infusion schedules, respectively).

## **Baseline Characteristics**

Upon study entry, the median age of the adult subjects was 42.0 years (range, 17-55 years); the median age of the pediatric subjects was 8.0 years (range, 3-15 years) (Table 2). Most enrolled subjects had a diagnosis of common variable immunodeficiency (30 of 33 adults [91%] and 8 of 15 pediatric subjects [53%]). The remaining subjects had diagnoses of X-linked or autosomal forms of agammaglobulinemia (3 of 33 adults [9%] and 5 of 15 pediatric subjects [33%]) and hypogammaglobulinemia (2 of 15 pediatric subjects [13%]). Both subjects with hypogammaglobulinemia were aged  $\geq 8$  years and were therefore too old to have transient hypogammaglobulinemia of infancy. In accordance with study eligibility criteria, all subjects had received IGIV therapy prior to enrollment. One adult had also been treated with a subcutaneous immunoglobulin product prior to receiving IGIV treatment and subsequent study enrollment.

#### **Primary Outcome**

For adults treated with Gammaplex 10%, the geometric mean  $AUC_{0-\tau}$  for the 28-day regimen was 34,900 mg·day/dL for absolute (unadjusted) values and 7830 mg·day/dL for baseline-adjusted (adjusted for pre-dose concentration) values (Supplemental Table S1). For adults treated with Gammaplex





<sup>c</sup> Pediatric subjects received five infusions of Gammaplex 10% only. <sup>d</sup> One subject withdrew consent. <sup>e</sup> One subject was withdrawn at investigator discretion

**Table 2**Subject demographicand baseline characteristics

Demographic characteristic	Adults $(n = 33)$	Pediatrics $(n = 15)$	All subjects $(N = 48)$	
Age (years)				
Mean (SD)	39.5 (11.99)	9.6 (4.15)	30.1 (17.29)	
Median (range)	42.0 (17–55)	8.5 (3–15)	30.5 (3-55)	
Age group, $n$ (%)				
2–5 years	0	2 (13.3)	2 (4.2)	
6–11 years	0	7 (46.7)	7 (14.6)	
12-15 years	0	6 (40.0)	6 (12.5)	
16-55 years	33 (100)	0	33 (68.8)	
Female, $n$ (%)	21 (63.6)	7 (46.7)	28 (58.3)	
White, <i>n</i> (%)	33 (100)	15 (100)	48 (100)	
Diagnosis, n (%)				
Common variable immunodeficiency	30 (90.9)	8 (53.3)	38 (79.2)	
X-linked and autosomal forms of agammaglobulinemia	3 (9.1)	5 (33.3)	8 (16.7)	
Hypogammaglobulinemia	0	2 (13.3)	2 (4.2)	
Baseline chest X-ray/CT scan, n (%)				
Normal	28 (84.8)	15 (100)	43 (89.6)	
Abnormal <sup>a</sup>	5 (15.2)	0	5 (10.4)	
Weight at screening (kg)				
Mean (SD)	78.81 (20.279)	38.32 (16.028)	66.15 (26.761)	
Median (range)	75.50 (51.6–140.0)	34.70 (14.8-65.4)	65.65 (14.8–140.0)	

CT computed tomography, CVID common variable immunodeficiency, SD standard deviation

<sup>a</sup> Observed abnormal chest X-ray/CT scans included a possible emphysematous change or hyperplasia involving the right upper lobe (judged to be not clinically significant), a chronic left lower lobe bronchiectasis, an elevation of the left hemidiaphragm, a subject with radiographic findings suggestive of pulmonary fibrosis (subject had a history of CVID-related lung disease), and a subject with minimal parenchymal scarring with parenchymal configuration that suggested chronic air trapping (no evidence of acute disease was noted)

5%, the geometric mean AUC<sub>0- $\tau$ </sub> for the 28-day regimen was 34,800 mg·day/dL for absolute values and 7230 mg·day/dL for baseline-adjusted values. Mean absolute serum IgG concentrations at steady state following repeated IV doses of Gammaplex 10% and Gammaplex 5% (28-day regimen) were almost superimposable (Fig. 2a, b). Based on the ratios (Gammaplex 10%/Gammaplex 5%) of least squares (LS) geometric means for AUC<sub>0- $\tau$ </sub> for the 28-day regimen (ratio for absolute IgG concentrations 1.01 [90% CI 0.98–1.03]; ratio for baseline-adjusted IgG concentrations 1.07 [90% CI 0.93–1.23]), both 90% CIs were within the prescribed bioequivalence range of 0.8–1.25. Thus, based on these results, the two products were bioequivalent.

## **Secondary Outcomes**

#### Adult Subjects

For the 21-day dosing interval, geometric means of the AUC<sub>0- $\tau$ </sub> of IgG in adults following repeated IV infusions of Gammaplex 10% (30,000 mg·day/dL) and Gammaplex 5% (30,400 mg·day/dL) were bioequivalent for absolute serum IgG concentrations (ratio of LS geometric means 0.99 [90%

CI 0.95–1.02]). Results for baseline-adjusted IgG concentrations were similar (AUC<sub>0- $\tau$ </sub><sup>-</sup> Gammaplex 10%, 6980 mg·day/ dL; Gammaplex 5%, 6380 mg·day/dL); however, the upper 90% confidence limit for the ratio of geometric mean (1.10 [90% CI 0.96–1.26]) was just outside of the upper prescribed bound of 1.25. Thus, the secondary objective, to demonstrate bioequivalence between the 21-day dosing regimens of Gammaplex 10% and 5%, was met using absolute serum IgG concentrations but just missed the defined bioequivalence bounds for baseline-adjusted IgG concentrations.

Similar to that observed for the 28-day regimen, the mean absolute serum concentrations of IgG among adults at steady state for the 21-day regimen following repeated IV doses of Gammaplex 10% and Gammaplex 5% were almost superimposable (Fig. 2c, d). For each dosing regimen, mean absolute trough IgG concentrations at the end of dosing following repeated ( $\geq$ 5) IV infusions of Gammaplex 10% (28-day regimen, 918 mg/dL; 21-day regimen, 1103 mg/dL) and Gammaplex 5% (28-day regimen, 945 mg/dL; 21-day regimen, 1120 mg/dL) were also equivalent based on ratio of geometric mean data (28-day regimen, 0.98 [90% CI 0.94–1.02]; 21-day regimen, 0.95 [90% CI 0.92–0.99]). Adults treated with Gammaplex 10% had mean total IgG trough

Fig. 2 Mean absolute serum concentrations of immunoglobulin G (IgG) following intravenous infusion of Gammaplex 10% to pediatric subjects, Gammaplex 10% to adults, and Gammaplex 5% to adults: 28-day regimen linear (**a**) and semi-logarithmic (**b**) scales; 21-day regimen linear (**c**) and semi-logarithmic (**d**) scales. Predose samples were taken up 30 min prior to infusion; samples at time zero were taken within 10 min prior to the end of infusion



Time after infusion (h)

levels of 879 mg/dL (28-day regimen) and 1074 mg/dL (21day regimen) at baseline, with mean changes from baseline to after the last Gammaplex 10% infusion of 58 and 29 mg/dL, respectively (Fig. 3). Adults treated with Gammaplex 5% had mean total IgG trough levels of 891 mg/dL (28-day regimen) and 1074 mg/dL (21-day regimen) at baseline, with mean changes from baseline to after the last Gammaplex 5% infusion of 71 and 46 mg/dL, respectively.

Fig. 3 Mean trough (preinfusion) serum concentrations of immunoglobulin G (IgG) following intravenous infusion of Gammaplex 10% in adults (**a**), Gammaplex 5% in adults (**b**), and Gammaplex 10% in pediatric (**c**) subjects. Threshold at 600 mg/dL (minimum trough serum IgG level for inclusion in study) is indicated by the *gray line. SD* standard deviation



Throughout the study, total IgG trough levels in adult subjects were well maintained, with total IgG trough levels consistently  $\geq 600 \text{ mg/dL}$  (the minimum trough serum IgG level for inclusion in the study) (Fig. 3a, b). Trough levels of IgG antibodies to specific antigens (*H. influenzae* type b and *S. pneumoniae* serotypes) were comparable after treatment with Gammaplex 10% or Gammaplex 5%, and no clinically relevant changes from baseline were observed (Supplemental Table S2).

Similar results were observed for trough levels of IgG subclasses 1, 2, and 3, whereas trough levels of IgG subclass 4 were slightly lower after the last Gammaplex infusion (mean [standard deviation, SD]: Gammaplex 10%, 8.3 [5.8] mg/dL; Gammaplex 5%, 8.6 [7.8] mg/dL) compared with baseline (mean [SD]: Gammaplex 10%, 13.0 [7.7] mg/dL; Gammaplex 5%, 13.0 [7.6] mg/dL) (Supplemental Table S3). All other PK parameters were similar between Gammaplex 10% and Gammaplex 5% (Supplemental Table S1).

#### Pediatric Subjects

For the 28-day dosing interval, the geometric mean AUC<sub>0- $\tau$ </sub> for pediatric subjects treated with Gammaplex 10% was 32,600 mg·day/dL for absolute values and 7280 mg·day/dL for baseline-adjusted values (Supplemental Table S1). For the 21-day dosing interval, the geometric mean AUC<sub>0- $\tau$ </sub> was 27,100 mg·day/dL for absolute values and 6000 mg·day/dL for baseline-adjusted values.

Mean absolute serum concentrations of IgG among pediatric subjects at steady state following repeated IV doses of Gammaplex 10% were similar to those of adults receiving Gammaplex 10% and Gammaplex 5% for the 28-day (Fig. 2a, b) and 21-day (Fig. 2c, d) dosing regimens.

Each of the 14 pediatric subjects who completed the study maintained total IgG trough levels of  $\geq 600 \text{ mg/dL}$  throughout the study, and total IgG trough levels were well maintained throughout (Fig. 3c). For the pediatric subject who did not complete the study and was withdrawn by the investigator,

the total IgG trough level prior to initiating Gammaplex 10% was 507 mg/dL and thereafter ranged from 343 to 470 mg/dL, despite dose increases (infusion 1, 488 mg/kg; infusion 2, 494 mg/kg; infusion 3, 651 mg/kg; infusion 4, 659 mg/kg). Twelve days after infusion 4, and without having received any other IVIG, the total IgG trough level in this subject was 846 mg/dL.

Among pediatric subjects, mean trough levels of total IgG at baseline and after the last infusion were 875 and 908 mg/dL, respectively (Fig. 3c). No clinically relevant changes from baseline were observed in trough levels of IgG subclasses (Supplemental Table S3) or IgG antibodies to *H. influenzae* and *S. pneumoniae* antigens (Supplemental Table S2) in pediatric subjects after treatment with Gammaplex 10%. Additional PK parameters of IgG following treatment with Gammaplex 10% in pediatric subjects are shown in Supplemental Table S1.

#### Safety

Overall, the mean (SD) total duration of exposure to Gammaplex (10% and/or 5%) was 252.2 (56.79) days among adults and 138.5 (30.26) days among pediatric subjects, with median (range) durations of 281.0 (1-316) and 139.0 (97-196) days, respectively. With the crossover study design, the mean (SD) durations of exposure among adults were similar for Gammaplex 10% (132.6 [22.58] days) and Gammaplex 5% (124.6 [27.59] days), with median (range) durations of exposure of 141.0 (97-181) and 140.0 (1-150) days, respectively. For adults, the mean (SD) total dose of Gammaplex 10% was 202.21 (77.732) g, with a mean (SD) dose per infusion of 494.9 (135.95) mg/kg; the mean (SD) total dose of Gammaplex 5% was 190.01 (67.094) g, with a mean (SD) dose per infusion of 485.6 (145.68) mg/kg. Among pediatric subjects, the mean (SD) total dose of Gammaplex 10% was 111.91 (52.988) g, with a mean (SD) dose per infusion of 535.1 (121.25) mg/kg. Among adults, the mean (SD) durations of infusion were 111.4 (24.98) min for Gammaplex 10% and 168.7 (47.02) min for Gammaplex 5% (Fig. 4); among pediatric subjects, the mean (SD) duration of infusion was 139.3 (42.89) min for Gammaplex 10%. Similar proportions of adults on each

Fig. 4 Mean infusion duration and mean dose per infusion for Gammaplex 5% in adults and Gammaplex 10% in adult and pediatric subjects. *SD* standard deviation product received pre-infusion medications (Gammaplex 5%: n = 3, 9.1%; Gammaplex 10%: n = 3, 9.4%), including cetirizine, diphenhydramine, and paracetamol; six pediatric subjects (40%) received pre-infusion medications, including diphenhydramine, ibuprofen, loratadine, methylprednisolone, paracetamol, and saline. Although hydration was not required prior to infusion of Gammaplex 5% or Gammaplex 10%, three pediatric patients received IV saline during the study; no adults received saline prior to any infusion.

AEs were reported more frequently by subjects receiving Gammaplex 10% (44 of 47 subjects [93.6%]) than those receiving Gammaplex 5% (23 of 33 subjects [69.7%]). In contrast, product-related AEs occurred in similar proportions of subjects receiving Gammaplex 10% (16 of 47 subjects [34.0%]) and Gammaplex 5% (12 of 33 subjects [36.4%]) (Supplemental Table S4). Among adults, product-related AEs that occurred in more than one subject were headache (12.5% [Gammaplex 10%] and 18.2% [Gammaplex 5%]), migraine (6.3% and 6.1%, respectively), pyrexia (6.3% and 0%, respectively), fatigue (3.1% and 6.1%, respectively), and nausea (3.1% and 6.1%, respectively). Among pediatric subjects, the only AE that occurred in more than one subject was headache (20.0%). ARs (i.e., AEs that occurred during the infusion or within 72 h after completion of the infusion were product-related or had unknown causality) are summarized in Table 3.

No serious product-related AEs occurred with either formulation, no subject withdrew due to an AE, and no thromboembolic events occurred during the study. No notable differences in adult subjects were observed between the two formulations of Gammaplex in the results of clinical laboratory, vital signs, physical examination, or body weight assessments. The Gammaplex 10% formulation was also well tolerated in pediatric subjects with PID.

## Discussion



This study is the first direct comparison of 5% IVIG and 10% IVIG products in subjects with PID. In this study, the results of the PK analysis met the primary objective of demonstrating the

**Table 3** Adverse reactions occurring in  $\geq$ 5% of subjects receiving Gammaplex 5% (*n* = 33) or Gammaplex 10% (*n* = 47) (ITT population)

Preferred term, n (%)	Gammaple	Gammaplex 5% Adult		Gammaplex 10%					
	Adult			Adult		Pediatric		All	
	Subjects $(n = 33)$	Infusions $(n = 163)$	Subjects $(n = 32)$	Infusions $(n = 166)$	Subjects $(n = 15)$	Infusions $(n = 82)$	Subjects $(n = 47)$	Infusions $(n = 248)$	
Headache	6 (18.2)	15 (9.2)	4 (12.5)	12 (7.2)	4 (26.7)	5 (6.1)	8 (17.0)	17 (6.9)	
Rash/rash generalized	0	0	4 (12.5)	4 (2.4)	0	0	4 (8.5)	4 (1.6)	
Rhinitis/rhinorrhea	1 (3.0)	4 (2.5)	2 (6.3)	6 (3.6)	1 (6.7)	1 (1.2)	3 (6.4)	7 (2.8)	
Migraine	2 (6.1)	3 (1.8)	2 (6.3)	2 (1.2)	1 (6.7)	1 (1.2)	3 (6.4)	3 (1.2)	
Pyrexia	0	0	2 (6.3)	2 (1.2)	1 (6.7)	1 (1.2)	3 (6.4)	3 (1.2)	
Fatigue	3 (9.1)	7 (4.3)	1 (3.1)	5 (3.0)	0	0	1 (2.1)	5 (2.0)	
Lower respiratory tract infection/bronchitis/fibrinous bronchitis	2 (6.1)	3 (1.8)	1 (3.1)	1 (0.6)	0	0	1 (2.1)	1 (0.4)	
Sinusitis/chronic sinusitis	2 (6.1)	2 (1.2)	1 (3.1)	1 (0.6)	0	0	1 (2.1)	1 (0.4)	
Nausea	2 (6.1)	1 (0.6)	1 (3.1)	1 (0.6)	0	0	1 (2.1)	1 (0.4)	

Adverse reactions were adverse events (irrespective of causality) that occurred during the infusion or within 72 h after completion of the infusion, any product-related adverse events, or any adverse events with unknown causality

ITT intent to treat

bioequivalence of Gammaplex 10% IVIG and Gammaplex 5% IVIG at the 28-day dosing interval. The secondary objectives of demonstrating the bioequivalence of Gammaplex 10% and Gammaplex 5% were also met, with the exception of adjusted AUC<sub>0- $\tau$ </sub> over the 21-day dosing interval, which had an upper confidence limit (1.26) that was just outside the upper prescribed bound of 1.25. In both primary and secondary PK analyses, the 90% CIs for baseline-adjusted values were wider than those for the respective analyses of the absolute IgG measurements. Subtracting baseline values from subsequent values magnifies experimental error and may explain the observed upper confidence limit of 1.26 for the secondary endpoint.

Trough levels of IgG, IgG subclasses 1 to 3, and antibodies to specific antigens were well maintained with Gammaplex 5% and Gammaplex 10% throughout the study. The reduction in trough levels of IgG subclass 4 observed across all treatments and age groups was expected, since IgG4 is reduced in proportion to other IgG subclasses during the Gammaplex manufacturing process. The role of  $IgG_4$  has not been fully elucidated, and the reductions in levels seen during this study were not clinically significant. In one pediatric subject, consistently low trough IgG levels were observed despite a dose increase; the cause of this was unclear but may have been related to a change in metabolism of IVIG prior to initiating Gammaplex 10% treatment. Since terminating this study, this subject has continued to exhibit inconsistent but increasing IgG catabolism despite product changes and a dosing increase (Bio Products Laboratory, data on file). Among the adult subjects, the mean infusion duration was 34% shorter for Gammaplex 10% than for Gammaplex 5%. Infusion durations were not reduced by half, as might be expected with equivalent doses administered via a 10% versus a 5% product, because the starting infusion rate per study protocol for Gammaplex 10% was half that of Gammaplex 5% (Table 1), to allow for an equivalent initial protein dose.

To minimize infusion-related side effects [15–17], previous studies of IVIG products have increased infusion rates incrementally at intervals of 30 or 60 min [18, 19]. As infusion time is an important factor for patients, this study was prospectively designed to increase infusion rates incrementally at 15-min intervals, if tolerated, to shorten overall infusion times; as reported above, this approach resulted in mean infusion times of 1.9 h for Gammaplex 10% in adults, 2.8 h for Gammaplex 5% in adults, and 2.3 h for Gammaplex 10% in children. Using the dosing schedules and infusion rates employed in this study (including the use of a 15-min titration schedule), Gammaplex 10% and Gammaplex 5% were both safe and well tolerated in adult subjects with PID. No notable differences were observed between the safety profiles of the two formulations.

The Gammaplex 10% formulation was also safe and well tolerated in pediatric subjects with PID. Based on the bioequivalence and PK analysis from this bridging study in PID subjects, Gammaplex 10% can be expected to have a therapeutic effect similar to the licensed Gammaplex 5%, which has demonstrated tolerability and efficacy in adult and pediatric patients with PID and ITP [11–13].

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Authorship Contributions Acquisition, analysis, and interpretation of data: RLW, IRM, MRS, SJ, MN, JNM

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Approval of the manuscript for submission/publication: RLW, IRM, MRS, SJ, MN, JNM

#### **Compliance with Ethical Standards**

**Conflict of Interest** RLW has served as a consultant for ADMA Biologics, Bio Products Laboratories, Grifols, Kedrion, Korean Green Cross, ProMetic, Shire, and Therapure; as an investigator for ADMA Biologics, Bio Products Laboratories, CSL Behring, Kedrion, Shire, and ProMetic; and as a speaker for Baxter/Shire and CSL Behring.

IRM has served as a clinical trial investigator for Bio Products Laboratory.

MRS has served as a consultant and speaker for Baxter Healthcare, Green Cross, and CSL Behring; and as an investigator for ADMA Biologics, Green Cross, ProMetic, Bio Products Laboratories, and Kedrion.

SJ reports consulting, speaker, travel, advisory board, and research support from CSL Behring, Baxter, Bio Products Laboratory, Biotest, SOBI, Shire, Viropharma, and Octapharma.

MN is an employee of Bio Products Laboratory.

JNM has served as a clinical trial investigator for Bio Products Laboratory, a consultant for Baxter and ProMetic, and an iDMC member for MacroCure and Octapharma.

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