## Corrigendum

## The structure of the TsaB/TsaD/TsaE complex reveals an unexpected mechanism for the bacterial t<sup>6</sup>A tRNA-modification

Sophia Missoury<sup>1</sup>, Stéphane Plancqueel<sup>1</sup>, Ines Li de la Sierra-Gallay<sup>1</sup>, Wenhua Zhang<sup>1</sup>, Dominique Liger<sup>1</sup>, Dominique Durand<sup>1</sup>, Raoudha Dammak<sup>1</sup>, Bruno Collinet<sup>1,2,\*</sup> and Herman van Tilbeurgh<sup>1,\*</sup>

<sup>1</sup>Institute for Integrative Biology of the Cell (I2BC), CEA, CNRS UMR 9198, Univ. Paris-Sud, Université Paris-Saclay, 91198 Gif sur Yvette Cedex, France and <sup>2</sup>Institut de Minéralogie, de Physique des Matériaux et de Cosmochimie, UMR7590 CNRS/Sorbonne-Université, UPMC, Paris, France

## Nucleic Acids Research, 2018, 46(11): 5850–5860, https://doi.org/10.1093/nar/gky323

Subsequent to publication of this study, we obtained crystals that yielded slightly better diffraction data for the *Tm*TsaBDE complex (Table 1). These data resulted in improved electron density maps and allowed us to identify and correct a few errors in the previously deposited structure (originally deposited as PDB 6FPE). The latter (which is still available for download and comparison, at https://www.rcsb.org/structure/removed/6FPE) has been superceded by a new set of revised coordinates (PDB 6S84).

The most significant differences between the two coordinate sets consist of:

- 1. The C-terminus of one of the TsaB copies in the asymmetric unit: amino acids 194–205 that were missing in the 6FPE structure could be constructed into the density.
- 2. Modeling of a nucleotide bound at the active site of TsaD. We initially cautiously interpreted the residual electron density by glycerol and PEG moieties present in the crystal freezing liquor. The new maps clearly showed that a nucleotide was bound at this location (Figure 1). We could easily fit the density by AMPCPP, present as a ligand in the crystallization solution.

The new structure does not alter the primary conclusions of our manuscript:

- The C-terminal part of the active site of *Tm*TsaD remains well-structured and is still capable of binding a nucleotide in the context of the ternary *Tm*TsaBDE complex.
- The AMPCPP occupies exactly the same position as the carboxy-AMP compound present in the recent structure of *Tm*TsaBDE, reported by Swairjo *et al.* (1). We further confirm that in our structure the N-terminal part of the *Tm*TsaD active site remains partially disordered and that neither Zn nor Fe ions are bound. This contrasts with the structure reported by Swairjo *et al.*, which has an ordered metal binding site occupied by Zn. This latter structure was obtained in presence of ATP, and we suspect that the nature of the bound nucleotide (AMPCPP versus ATP) might play a role in the metal binding. The two structures of the TsaBDE complex (6N9A and 6S84) represent probably different snapshots along the catalytic pathway.

<sup>&</sup>lt;sup>\*</sup>To whom correspondence should be addressed. Herman van Tilbeurgh. Email: herman.van-tilbeurgh@u-psud.fr Correspondence may also be addressed to Bruno Collinet. Email: Bruno.collinet@i2bc.paris-saclay.fr Present address: Wenhua Zhang, School of Life Sciences, Lanzhou University, 730000 Lanzhou, China.

<sup>©</sup> The Author(s) 2019. Published by Oxford University Press on behalf of Nucleic Acids Research.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License

<sup>(</sup>http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com



Figure 1.  $2F_o$ - $F_c$  difference map (blue) of the AMPCPP surrounding at the active site of TmTsaD.  $F_o$ - $F_c$  map : positive and negative densities are represented as green and red grids respectively. The metal coordinating histidines 109 and 113 are also shown. They are partially disordered and not in a configuration compatible with metal binding.

Table 1.	New	(vs.	original)	) data	collection	and	refinement	statis	tics
		<b>`</b>							

	PDB 6S84	PDB 6FPE
Wavelength (Å)	0.978570	0.9801
Resolution range (Å)	46.14-2.90 (3.00-2.90)	48.44-3.14 (3.33-3.14)
Space group	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	P 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Unit cell a,b,c (Å)	a = 85.16, b = 108.21, c = 176.65	a = 84.31, b = 113.94, c = 177.62
Total reflections	324005	138146
Unique reflections	36849	30297
Completeness (%)	99.1 (94.8)	99.1 (95.2)
Mean I/sigma(I)	9.1 (0.87)	7.1 (1.03)
R-meas	0.23	0.19
CC1/2	99.7 (42.1)	99.2 (43.9)
R-work	0.209	0.23
R-free	0.282	0.29
Number of non-hydrogen atoms	10678	10418
RMSD bonds (Å)	0.003	0.011
RMSD angles (°)	1.011	1.258
Ramachandran favored (%)	94.77	95.3
Average B-factor (Å <sup>2</sup> )	89.66	80.67

Statistics for the highest-resolution shell are shown in parentheses.

## REFERENCE

 Luthra, A., Paranagama, N., Swinehart, W., Bayooz, S., Phan, P., Quach, V., Schiffer, J.M., Stec, B., Iwata-Reuyl, D. and Swairjo, M.A. (2019) Conformational communication mediates the reset step in t<sup>6</sup>A biosynthesis. *Nucleic Acids Res.*, 47, 6551–6567.