Revised: 10 October 2019

REVIEW

Expression, function and clinical application of stanniocalcin-1 in cancer

Fangyu Zhao¹ [Gang Yang¹ | Mengyu Feng¹ | Zhe Cao¹ | Yueze Liu¹ | Jiangdong Qiu¹ | Lei You¹ | Lianfang Zheng² | Taiping Zhang^{1,3} | Yupei Zhao¹

¹Department of General Surgery, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

²Department of Nuclear Medicine, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

³Clinical Immunology Center, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China

Correspondence

Taiping Zhang and Yupei Zhao, Department of General Surgery, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, 1 Shuaifuyuan, Wangfujing Street, Beijing 100730, China. Emails: tpingzhang@yahoo.com (T.Z.); zhao8028@263.net (Y.Z.)

Funding information

This review was supported by grants from the National Natural Science Foundation of China (No. 81772639 and 81802475). Natural Science Foundation of Beijing Municipality (No. 7192157), CAMS Innovation Fund for Medical Sciences (CIFMS) (No. 2016-I2M-1-001), Nonprofit Central Research Institute Fund of Chinese Academy of Medical Sciences (No. 2018PT32014 and 2018PT32002). China Postdoctoral Science Foundation (No. 198831), CAMS Innovation Fund for Medical Students (No. 2017-1002-1-16) and PUMC Youth Fund and the Fundamental Research Funds for the Central Universities (No. 2017320027).

1 | INTRODUCTION

Abstract

The glycoprotein stanniocalcin-1 functions as a regulatory endocrine hormone that maintains the balance of calcium and phosphorus in bony fish and as a paracrine/ autocrine factor involved in many physiological/pathological processes in humans, including carcinogenesis. In this review, we provide an overview of (a) the possible mechanisms through which STC1 affects the malignant properties of cancer, (b) transcriptional and post-transcriptional regulation pathways of STC1 and (c) the potential clinical relevance of STC1 as a cancer biomarker and even a therapeutic target in the future. Exploring the role of STC1 in cancer development may provide a better understanding of the tumorigenesis process in humans and may facilitate finding an effective therapeutic method against cancer.

KEYWORDS

biomarker, cancer development, clinical application, stanniocalcin-1

which is located on the short arm of chromosome 8 (8p11.2-p21).⁵ An additional STC family member was subsequently identified,⁶ and they were renamed as STC1 and STC2. The human *STC1* gene is widely expressed in many tissues, but STC1 is normally undetectable in serum,² suggesting that STC1 may act as a paracrine/autocrine factor rather than an endocrine hormone.⁷ STC1 has appeared to change its

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Journal of Cellular and Molecular Medicine published by Foundation for Cellular and Molecular Medicine and John Wiley & Sons Ltd

Stanniocalcin (STC)-1 is a glycoprotein first discovered in the corpuscles of Stannius in fish^{1,2} and is believed to regulate serum calcium and phosphate homeostasis in the fish body as an endocrine hormone.³ The first mammalian form of STC was discovered in 1996,⁴

Fangyu Zhao and Gang Yang contributed equally to this work.

function during evolution. In mammals, it acts preferentially on phosphate metabolism compared with calcium metabolism. STC1 has been shown to stimulate phosphate reabsorption in the small intestines and proximal tubules of the kidney.⁷ STC1 also participates in many kinds of physiological and pathological processes including pregnancy,^{8,9} lactation,^{7,10} angiogenesis,¹¹ bone and muscle development,^{12,13} organogenesis,¹⁴ cerebral ischaemia,¹⁵⁻¹⁷ ischaemia/reperfusion kidney injury,^{18,19} idiopathic pulmonary fibrosis²⁰ and hypertonic stress.²¹ However, its role in cancer has attracted much attention,⁵ and many studies have shown that STC1 promotes tumour cell viability and proliferation, and facilitates solid tumour invasion and metastasis. STC1 might be modulated by HIF-1 expression under hypoxia condition, thereby initiating expression of several downstream genes. As a result, the Warburg effect will be turned on, reprogramming of tumour metabolism from oxidative to glycolytic metabolism.²² What's more, it has been proved that STC1 is involved in multiple cancer-related signalling pathways, such as NF-kB,²³ ERK1/2²⁴ and JNK signalling pathways.²⁵ Epithelial-mesenchymal transition (EMT) is a process in which epithelial cells acquire mesenchymal features, associated with tumour initiation, invasion, metastasis and resistance to therapy.²⁶ STC1 also participates in EMT process to reshape the tumour microenvironment, promote the initiation of malignant phenotypes.²⁷ Clinical data have also revealed that STC1 may have a negative correlation with prognosis, a higher STC1 expression level in tumour tissues was correlated with shorter DFS (disease-free survival) and OS (overall survival).²⁸⁻³⁰ These evidences suggest that STC1 is a potential diagnostic and prognostic marker as well as therapeutic target of cancer. The secretion of STC1 is intermittent and can be stimulated

by external stimulus, and thus, the secretion of STC1 is more susceptible to external cytokines related to cancer. While the secretion mode of STC2 is constitutive, which implies STC2 participates the body's normal physiological activity.⁵ So we want to explore the function of STC1 thoroughly for it may be regulated by many cancer-related factors. Moreover, the researches about STC1 and cancer are more abundant than STC2. Therefore, we will discuss the possible mechanisms through which STC1 interacts with cancer cells and its clinical relevance in this review.

2 | INTERACTIONS BETWEEN STC1 AND CANCER CELLS

2.1 | STC1 is aberrantly expressed in tumours

An aberrant expression level of STC1 has been found in many kinds of tumours (Table 1). It is usually overexpressed in most human cancer tissues compared with normal tissues, such as ovarian cancer,^{31,32} hepatocellular carcinoma (HCC),^{25,33} non-small cell lung cancer,³⁴ lung adenocarcinoma,³⁵ glioma,^{36,37} thyroid cancer,³⁸ fibrosarcoma,³⁹ oesophageal squamous cell carcinoma,²⁸ laryngeal squamous cell carcinoma (LSCC),⁴⁰ colorectal cancers,⁴¹⁻⁴³ breast carcinoma,^{44,45} leukaemia^{46,47} and gastric cancer.⁴⁸ However, Guo et al and Pan et al observed decreases of STC1 expression in cervical cancer tissues, especial at stages II and III, compared with adjacent normal tissues and stage I cervical cancer tissues.^{49,50} The reason is that NF- κ B pathway is activated in cervical cancer, leading to an

TABLE 1 Expression levels and functions of STC1 in different kinds of cancers

Type of cancer	STC1 expression level	STC1 Effects on tumour	Source
Cervical cancer	↑	Inhibit cell proliferation, migration and invasion	43, 44
Colorectal cancer	\uparrow	Stimulate migration and invasion	35-37
Ovarian cancer	\uparrow	Increase cell proliferation and migration	22
Ovarian cancer	\downarrow	Inhibit cell proliferation, promote apoptosis	46
Breast cancer (TNBC)	\uparrow	Promote metastasis	50, 51
Breast cancer (ER+)	↑	Increase cell proliferation	52
Breast cancer (BRCA1-mutant)	\downarrow	Inhibit cell proliferation, promote apoptosis	47
Laryngeal squamous cell carcinoma	↑	Correlated with advanced clinical stage	34
Leukaemia	1	Promote chemoresistance, marker of MRD (minimal residual disease)	50, 41
Gastric cancer	1	Associated with more lymph metastasis and advanced clinical stage	42
Non-small cell lung cancer	1	Associate with advanced tumour stage and histological subtype	27
Glioma	\uparrow	Associate with high pathological grade	29, 30
Hepatocellular carcinoma	\uparrow	Associate with tumour size (<5 cm in diameter)	24
Hepatocellular carcinoma	\downarrow	Associate with tumour size (>6.3 cm in diameter)	25, 87
Thyroid cancer	\uparrow	Enhance cell proliferation, inhibit cell apoptosis	31
Oesophageal squamous cell carcinoma	\uparrow	Associate with advanced T-stage	33
Lung adenocarcinoma	\uparrow	Increase cell proliferation, inhibit cell apoptosis	28

increased level of phospho-P65 and a corresponding decreased level of non-phosphorylated P65 protein. As Guo et al⁴⁹ reported, NF- κ B p65 protein directly bound to STC1 promoter and activated the expression of STC1 in cervical cancer cells, thus the decreased non-phosphorylated p65 protein level then leads to the reduced expression of STC1, the phenomenon we observed in tumour tissues.

Additionally, the correlation between STC1 expression and breast/ovarian cancer is intricate. Some previous studies have found that STC1 expression down-regulates after loss-of-function of BRCA1, which leads to breast and ovarian cancer.⁵¹⁻⁵³ Because STC1 is critical for normal breast and ovarian physiologies.⁵⁴ and relatively highly expressed in the ovary with expression dramatically increasing during pregnancy and lactation,⁵⁵ the loss of STC expression in tumour cells may be a phenomenon of malignancy because the normal physiological activity has been disrupted. However, this is the opposite view from recent studies showing that STC1 is overexpressed in breast and ovarian cancer tissues compared normal tissues.^{31,56} In this regard, the tumour source and/or variability are the likely causes of this discrepancy. Triple-negative breast cancer (TNBC) tends to have a very high STC1 level compared to normal tissues, 56,57 and ER-positive tumours also possess a relatively higher STC1 expression level than normal,^{58,59} whereas STC1 may be absent in BRCA1-mutant breast tumours.⁵³ Such evidence implies that STC1 may have a tumour-promoting function in most cases, but possesses a tumour-suppressing trait in a few cases.

2.2 | STC1 acts via an autocrine signalling pathway in tumours

It has been reported that STC1 acts in a paracrine manner to regulate corpora lutea functions,⁶⁰ but STC1 might perform its oncogenic function through an autocrine manner. Mccudden et al⁵⁸ showed that STC1 and its receptor were colocalized in 91% of examined human breast cancer biopsies (53/58) by immunocytochemistry and in situ ligand and binding staining, and all cancerous cells were positively stained for both the ligand and receptor. This pattern is analogous with that in the kidney where an autocrine signalling pathway has been identified in the collecting ducts, whose cells express STC and possess high affinity STC receptors.⁶¹ These results suggest STC1 appears to be operating via an autocrine loop in the malignant mammary gland, which differs from its paracrine signalling pathways in tissues such as ovaries.

2.3 | STC1 is regulated by a wide range of factors related to tumours

The *STC1* gene may be the downstream target of several genes and proteins related to cancers (Figure 1), including p53,⁶² BRCA1,⁵³ RET,⁶³ Wnt2,⁶⁴ JMJD3,⁶⁵ PPM1D,⁶⁶ PKC α ,⁶⁷ PPARD,⁶⁸ LINE-1,⁶⁹ IGF-1⁷⁰ and CAIX.⁵⁷

The STC1 gene promoter is repressed by HDAC, a p53-dependent histone deacetylase⁷¹ and a repressor complex formed by transcription

factor Sp1 and Rb inhibits STC1 gene transcription.⁷² It has been reported that one of the key proteins that mediates p53-induced apoptosis, NF-kB, also regulates STC1 gene transcription.²³ Based on these data. Ching et al⁶² showed that, instead of direct involvement of transactivation of STC1 gene expression, p53 actives the NF-kB signalling pathway and inhibits the activity of HDAC then increases the levels of histone H3 acetylation to elevate STC1 gene expression. Another study confirmed that induction of STC1 expression in an apoptotic human nasopharyngeal cancer cell line (CNE2) is mediated by activation of p53.73 Welcsh et al⁵³ described the absence of BRCA1 expression in primary breast tumour cells without expression of STC1 and that BRCA-1induced cells had a much higher expression level of STC1 compared with control cells. They confirmed that BRCA1 is a stimulator of STC1 expression by oligonucleotide array-based expression profiling. RET, an oncogene responsible for multiple endocrine neoplasia (MEN) type 2A and 2B that develops into medullary thyroid carcinoma and pheochromocytoma, has been proven to induce STC1 expression, which is in line with immunohistochemistry results indicating that STC1 is highly expressed in specimens of MEN2B-MTC.⁶³ JMJD3, a kind of histone demethylase that correlates with melanoma growth and metastasis, also up-regulates STC1 expression.⁶⁵ In addition, an increased STC1 expression level after protein phosphatase magnesium-dependent 1 delta (PPM1D) silencing indicated that STC1 was affected by PPM1D, a protein overexpressed in various cancer cell lines, which has been associated with poor prognoses of cancers.⁶⁶ Moreover, STC1 expression is suppressed by PKC α (protein kinase C α), a highly expressed protein in breast cancers. After depleting PKC α expression by siRNA, the STC1 expression level is particularly increased.⁶⁷ Klein et al⁶⁴ demonstrated that STC1 is a target gene of Wnt signalling and can be down-regulated by Wnt2, which acts as an angiogenic growth factor and differentiation factor in vascular development during cancer progression. In addition, VEGF treatment stimulates STC1 gene transcription and increases its mRNA levels.^{74,75} Combined with the aforementioned evidence that STC1 promotes the expression of VEGF, these data suggest positive feedback regulation between STC-1 and VEGF.

Some miRNAs have also been reported to regulate STC1 gene expression post-transcriptionally. Sakata et al reported STC1 as a target of miR-101b by several experiments in which miR-101b bound to three sites in the 3'-UTR of the STC1 gene, thereby blocking the translation of STC1 mRNA, but not inducing mRNA degradation.⁷⁶ Sakata et al also reported that miR-29b, miR-34a, miR-101 and miR-137 have predicted binding sites in STC1 mRNA and down-regulate both the mRNA level and protein concentration.⁷⁷ Similarly, Lu et al⁷⁸ found that STC1 is a potential target gene of miR-21-5p. Furthermore, Groves et al reported that glucocorticoid potently inhibits STC1 gene expression through a post-transcriptional mechanism. Hydrocortisone and dexamethasone down-regulate the STC1 expression level dramatically in several kinds of mouse and human tumour cell lines, and this effect is antagonized by activation of the cAMP signalling pathway. It is possible that cAMP inhibits the activity of glucocorticoid-induced protein(s) that destabilize STC mRNA, thereby preserving STC1 expression.79

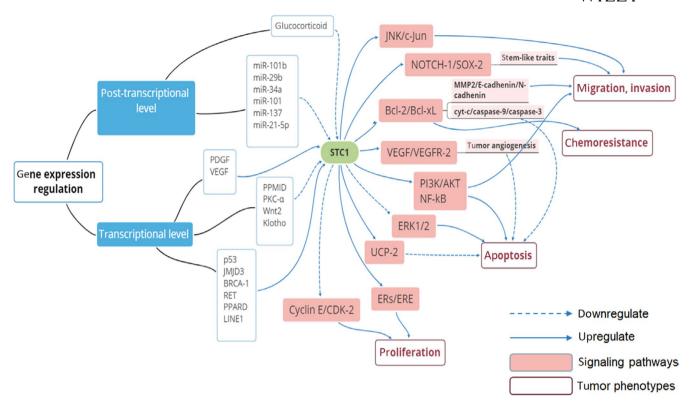


FIGURE 1 Regulation networks of STC1. STC1 is regulated by many proteins, and it can affect several signalling pathways to modulate tumour phenotypes

These factors regulate STC1 expression differently and antagonistically. STC1 expression is regulated by a complex network in which factors that promote tumorigenesis can either induce or suppress STC1 expression, and the same mechanism operates for factors that inhibit tumorigenesis. Such examples indicate that STC1 is an intermediate modulated by many tumour-related factors rather than a determining factor in tumorigenesis, causing it plays distinct functions in different kinds of cancers.

2.4 | Effect of STC1 on apoptosis in cancer

STC1 has been identified as a stimulator of mitochondrial respiration because it stimulates activity of the mitochondrial electron transport chain and calcium transport in a concentration-dependent manner. Therefore, STC1 is likely to be an anti-apoptosis and oncogenic factor because it provides additional energy for tumour cell growth.⁸⁰ However, it is now believed that STC1 performs its cytoprotective function mainly through a hypoxic signalling pathway. It has been observed that STC1 expression is elevated under hypoxic conditions in tumours. For example, Yeung et al confirmed that hypoxia stimulates STC1 gene expression in various human cancer cells including colon carcinomas, nasopharyngeal cancer cell lines (CNE-2, HONE-1 and HK-1), and ovarian cancer cell lines (CaOV3, OVCAR3 and SKOV3).²² This effect is believed to be mediated by hypoxia inducible factor (HIF)-1 α , a key transcriptional factor in the hypoxic response. Hypoxia stimulates the expression of HIFs and thus induces overexpression of stanniocalcin-1.⁸¹ Subsequently, a study by Law et al using a normoxic human nasopharyngeal cancer cell line (CNE2) confirmed that the STC1 gene has an authentic hypoxia response element (HRE) motif located at the upstream region between -2322 and -2335.⁸² This HIF-HRE combination requires recruitment of p300, a transcription coactivator.²² CAIX, a pH-regulating enzyme that plays a key role in maintaining alkaline intracellular pH under hypoxic conditions, is also indispensable for STC1 expression and functions as a mediator during hypoxia-induced STC1 expression. CAIX is up-regulated under hypoxic conditions as a direct transcriptional target of HIF-1 and then elevates STC1 expression.⁵⁷

It has been well demonstrated that STC1 enhances tolerance to hypoxia in tumour cells and thus has an anti-apoptosis effect on tumour cells (Figure 2). In cancer cells, the metabolism changes from aerobic respiration to glycolysis under hypoxia, a much more anaerobic respiratory mode.⁸³ During this process, STC1 up-regulates the expression level of UCP2 (uncoupling protein 2) and decreases the mitochondrial membrane potential.⁸⁴ As a result, the lactate production for anaerobic glycolysis has been increased (also known as the Warburg effect), which promotes the growth of cancers. Oxidative phosphorylation is normally coupled to superoxide generation, such as ROS, which are reactive species that cause oxidative damage to cellular biomolecules and play a major role in pathophysiological processes.⁸⁵ The Warburg effect shifts mitochondrial respiration to a much more glycolytic metabolic profile, reducing ROS generation.⁸⁶ Taken together, STC1 reduces the generation of ROS, diminishes oxidative and ER stresses,⁸⁷

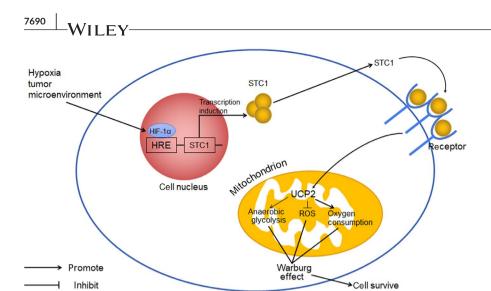


FIGURE 2 STC1 and hypoxia. HIF, hypoxia inducible factor; HRE, hypoxia response element; UCP, uncoupling protein; ROS, reactive oxygen species

and makes tumours resistant to ROS,⁸⁶ thereby facilitating cancer cells to survive and proliferate.⁸⁸ In addition, Wang et al reported that STC1 promotes the expression of Bcl-2, leading to decreased caspase-9/-3-dependent cell death under hypoxia.⁸⁹

It is noteworthy that there are also some studies supporting the notion that STC1 can also be a proapoptotic factor. Despite Nguyen et al reporting a high expression level of STC1 under oxidative stress in accordance with the aforementioned studies, they found that STC1 reduces survival and promote apoptosis by down-regulating a prosurvival signalling pathway, ERK1/2.²⁴ In addition, Pan et al reported that STC1 promotes apoptosis via NF-κB phospho-p65 (Ser536) by PI3K/AKT, $I\kappa B\alpha$ and IKK signalling in cervical cancer cells.⁵⁰ Yeung et al also found a proapoptotic effect of STC1 in hepatocellular carcinoma cell lines Hep3B and MHCC-97L through up-regulating proapoptotic genes interleukin-12 and NOD-like receptor family, pyrin domain-containing 3, thereby slowing down the process of carcinogenesis.⁹⁰ Moreover, trichostatin A [TSA; a histone deacetylase (HDAC) inhibitor], as one of the most common treatment methods to induce apoptosis in human cancer cells, induces overexpression of STC1 through activation of p53 in cancer cells.⁶² In addition, TSAinduced apoptotic processes were found to be significantly reduced by silencing STC1 expression, which supports the notion that STC1 is a proapoptotic factor.²³

The contradictory roles of STC1 in apoptosis may be because STC1 plays different roles on different pathways related to apoptosis. And the particular role of STC1 may be dependent on the extent of cellular stress.⁶²

2.5 | Effect of STC1 on cancer cell proliferation

STC1 is believed to promote the proliferation of several kinds of tumour cells. For example, recently, Dai et al found that STC1 is involved in the interaction of Klotho and tumour progression. Klotho, a newly found anti-ageing gene, reduces proliferation of thyroid cancer cell lines FTC133 and FTC238 and enhances apoptosis. A high level of Klotho was found to be associated with a low level of STC1 in both cell lines. Thus, they speculated that Klotho may inhibit thyroid tumour cell proliferation by inhibiting the expression level of STC1. Furthermore, they verified that recombinant STC1 markedly enhanced cell proliferation. However, the detailed molecular mechanism of Klotho-mediated cell proliferation and apoptosis remains unclear, and Klotho-induced apoptosis was only tested in thyroid cancer cell lines FTC133 and FTC238. Therefore, further validation is required in other types of thyroid cancer cell lines and animal models.⁹¹ Furthermore, in a study aimed to elucidate the function of STC1 in prostate carcinoma, Bai et al found that knock-down of STC1 decreased the proliferation of prostate carcinoma cell lines DU145 and LNCaP2. During this process, the expression levels of cell cycle-related proteins, cyclin E1/CDK2, were elevated. In contrast, overexpression of STC1 in normal prostate cell line RWPE-1 and xenografted tumours promotes cell growth.⁹² Similarly. Ma et al described a consistent result that STC1 promotes cell cycle progression, accelerates G1/S transition through elevating the expression of cyclin D1, Cdk4 and Cdk6, and suppressing the expression of p21.93 Therefore, STC1 promotes cancer cell proliferation via a novel mechanism through which several cyclins and CDKs are recruited to regulate the cell cycle positively.

However, the effect of STC1 on cell proliferation in breast cancer depends on the subtype. In 4T1ch9 and MDA-MB-231HM-luc cells, which are triple-negative (ER-/PR-/HER2-) breast cancer cell lines, STC1 expression has no effect on cell proliferation.⁹⁴ Conversely, in T47D cells, a luminal (ER+/PR+) breast cancer line, reduction of STC1 expression reduces in vitro cell proliferation.⁵¹ This phenomenon can be explained by a proliferation-promoting effect of STC1 on ER + breast cancer cells, which is dependent on the ER, because the STC1 gene is coexpressed with the ER. STC1 and its receptor were found to be present in all oestrogen receptor-positive samples (30/30),⁵⁸ which was in agreement with the finding that STC1 and STC2 proteins and mRNAs are highly correlated with ER levels.95 ERs function as transcription factors when they are bound to oestrogens. Upon hormone binding, ERs bind to DNA with high affinity at specific sites that are termed oestrogen response elements (EREs) to exert the physiological function of oestrogens. In addition to

signalling through EREs, there are alternative, non-classical ER transcriptional response pathways through which the ensuing expression of ER-regulated genes results in cell proliferation.^{96,97}

2.6 | Effect of STC1 on invasion and metastasis of cancer cells

STC1 promotes tumour metastasis via activation of PI3K/Akt and JNK signalling pathways. Li et al showed that the treatment with recombinant human STC1 significantly increases the invasiveness of TNBC cells, which was mediated by phosphorylation of JNK/c-Jun leading to up-regulation of matrix metalloproteinase 9.⁵⁶ Consistent with this result, secretory STC1 enhances the metastatic potential of HCC via the JNK signalling pathway.²⁵ Histone methylation is an important epigenetic mark that regulates gene expression. Aberrant histone methylation patterns caused by deregulated histone demethylases have been associated with carcinogenesis. The histone H3 lysine 27 (H3K27) demethylase JMJD3 up-regulates STC1 expression, activates NF-κB and PI3K pathways and promotes distant metastasis of melanoma, indicating that a high STC1 expression level is related to metastasis.⁶⁵ Analysis of transcriptome profiling identified STC1 as a peroxisome proliferator-activated receptor- δ (PPARD) target gene. Expression of STC1 is up-regulated during the metastasis process induced by overexpression of PPARD in cancer cells, indicating a pro-metastasis effect of STC1 on cancer.⁶⁸ In addition, Li et al pointed out that STC1 interacts with NOTCH1 to activate the SOX2 signalling pathway, augmenting the stem-like traits of GBM cells, and Sakata et al⁷⁷ found STC1 is a promoting factor of metastasis in GBM. Taken together, we speculated STC1 augments the stem-like traits of GBM cells and thus enhancing tumourigenicity and distal metastasis, since stem cell-like properties of cancer cells play a major role in metastasis. Furthermore, expression of STC1 is elevated, which functions as a mediator during the metastasis process induced by platelet-derived growth factor (PDGF), a major functional determinant of cancer-associated fibroblasts (CAFs). Elevated expression of PDGF receptors on stromal CAFs is associated with metastasis and a poor prognosis of colorectal cancer. Taken together, PDGF-stimulated fibroblasts increase the migration and invasion of colorectal cancer cells in an STC1-dependent manner.⁹⁸ Carbonic anhydrase IX (CAIX) is overexpressed in a variety of solid cancers, including breast cancer, which has been implicated in the migration, invasion and stemness of breast cancer cells. The biological effects caused by inhibition of CAIX reduce invasiveness and the self-renewal capacity by blocking STC1 induction, suggesting that STC1 promotes the invasiveness of breast cancer cells.⁵⁷

2.7 | STC1 promotes chemoresistance in tumours

The hypoxic tumour microenvironment plays a prominent role in the induction of chemoresistance.⁹⁹ Because STC1 is closely related to hypoxia in the tumour microenvironment, it may be possible that STC1 contributes to the chemoresistance feature of cancers. Although previous studies have not found a relationship between STC1 expression and chemoresistance,²⁸ recent studies have implied a connection between STC1 and chemoresistance in leukaemia because the STC1 expression level was higher in patients whom chemotherapy was unsuccessful compared with responsive patients.^{46,47} Wang et al also found that, under hypoxic conditions, STC1 promotes chemoresistance in gastric cancer patients, probably via up-regulation of Bcl-2, a well-characterized anti-apoptotic and EMT-related protein.⁸⁹

2.8 | STC1 contributes to the tumour-supporting microenvironment

The tumour microenvironment plays an important role in tumour initiation, progression, metastasis and chemoresistance.¹⁰⁰ A remarkable feature of the tumour microenvironment is the hypoxic condition induced by rapid proliferation and relatively insufficient vascularization of the tumour mass.^{101,102} STC1 promotes tumour neoangiogenesis that in turn changes the tumour microenvironment. The new vascular system enables tumour cells to obtain enough oxygen and nutrients for survival and proliferation and promotes distal metastasis.

STC1 increases both the mRNA and protein levels of eNOS, VEGF and VEGFR2, and stimulates the VEGF signalling pathway, which subsequently enhances tumour angiogenesis. This promotion effect of STC1 on the expression of VEGF depends on the activation of PKCbII and ERK1/2 pathways.^{11,103} It has been elucidated that stromal cells are recruited to constitute tumorigenic microenvironments and can significantly influence cancer phenotypes.¹⁰⁴ Fibroblasts and mesenchymal stem cells are important stromal cells that participate in hypoxia-induced STC1 expression. Fibroblasts can be activated by PDGF, which affect the tumour microenvironment and promote tumourigenicity. STC1 was identified as a fibroblast-secreted protein¹⁰⁵ and functions as a mediator of the PDGF receptor and fibroblasts to promote tumour growth and metastasis of colorectal cancer cells.^{27,98} Multipotent human mesenchymal stroma/stem cells (MSCs) are recruited in the tumour microenvironment during carcinogenesis and are believed to support tumour growth for repairing tissue injuries and inflammatory processes caused by tumours.^{106,107} Several studies have found that the supportive effect of MSCs on tumour cells may be partially due to suppression of apoptosis induced by ROS and enhancing the Warburg effect by secreting STC1.^{86,108}

3 | CLINICAL SIGNIFICANCE OF STC1

3.1 | STC1 serves as a biomarker for cancer diagnosis and early detection

STC1 is highly expressed in various kinds of tumours at both mRNA and protein levels in the blood and tumour tissues of cancer patients

compared with patients with benign diseases and healthy individuals, indicating that it can serve as a potential biomarker for cancer diagnosis.²⁹ For example, 12 cases out of 15 showed higher expression of STC1 mRNA in oesophageal squamous cell carcinoma tissue compared with normal counterparts. Furthermore, immunohistochemistry showed a high level of STC1 protein in 38.9% (89/229) of patients, mainly localized in the cytoplasm of tumour cells.²⁸ Moreover, Arigami et al found that the relative numbers of STC1 mRNA copies were significantly higher in gastric cancer cell lines and in blood specimens from patients with gastric cancer than in blood specimens from healthy volunteers (P = .0001 and P = .003, respectively). They demonstrated that 65 of 93 (69.9%) patients were positive for STC1 by RT-PCR analysis. This result indicated superiority to conventional tumour markers CEA and CA-199 whose positive rates in the same population were 26.9 (25/93) and 24.7% (23/93), respectively. Receiver operating characteristics were used to describe the diagnostic specificity and sensitivity of serum STCs to discriminate patients with gastric cancer from healthy volunteers, and the values were 69.9 and 71.4%, respectively.²⁹ Fang et al also reported supporting experimental results.⁴⁸

3.2 | Relationship between STC1 expression and clinicopathological parameters

Elevated expression of STC1 is significantly associated with the tumour grade, size, invasion and metastasis. For example, higher levels of circulating STC1 mRNA in serum were associated with more advanced tumour stages. The expression level of STC1 was significantly higher in the advanced pathological tumour stages of oesophageal squamous cell carcinoma (P = .019).²⁸ LSCC (P = .083).⁴⁰ gastric cancer (P = .013),²⁹ glioma (P < .001),³⁷ non-small cell lung cancer (P = .018)³⁴ and clear cell renal cell carcinoma (P = .008).⁹³ In addition, the presence of STC1 expression was significantly correlated with the depth of tumour invasion in gastric cancer (P = .032)²⁹ and thyroid-cartilage invasion in laryngeal squamous cell carcinoma (P = .086).⁴⁰ Ma et al designed a study focused on clear cell renal cell carcinoma (ccRCC) and found that the STC1 mRNA level in tumour tissues was positively correlated with the average tumour diameter.93 In line with this result, a high serum STC1 level was correlated with a larger tumour size in hepatocellular cancer (HCC) patients (P = .019, mostly <5 cm).²⁵ However, Yeung et al reported a negative correlation of STC1 expression with the HCC tumour size in large tumours (>6.3 cm in diameter). They found that tumours with higher expression levels of STC1 were significantly smaller than those with lower levels of expression (P = .008). Although STC1 plays a pro-oncogenic role in tumour progression, the counteracting effects STC1 on the pro-inflammatory effects of IL6 and IL8 might slow down the process of carcinogenesis as the tumour grows.^{33,90}

STC1 expression levels were elevated to a greater extent in ccRCC tissues and associated with distant metastasis, but only T1 stage tumours exhibited a statistically significant difference (P = .021) in STC1 expression compared with other tumour stages,

suggesting that STC1 is associated with metastasis of early stage ccRCC.⁹³ STC1 also promotes metastasis of gastric cancer, Fang et al⁴⁸ found out that a high STC1 expression level in GC tissues was associated with more lymph metastasis in GC patients (P = .029). Breast cancer patients with higher STC1 expression levels in tumour tissues were more likely to have lung metastasis than those who had lower levels (P = .02).⁹⁴ Furthermore, the serum level of STC1 mRNA serves as a molecular marker for micrometastases of various human cancers because STC1 mRNA might be useful to detect cancer cells in blood.¹⁰⁹ For example, an interesting study showed that STC1 is a detectable molecular marker for occult breast cancer with metastasis in the blood and bone marrow.^{45,110} Based on these studies, we speculate that overexpression of STC1 is positively correlated with an advanced tumour grade, larger tumour size and deeper tumour invasion with much more distant metastasis.

3.3 | STC1 may be a potential prognostic marker for cancer

The STC1 expression level has a close connection with the prognosis of cancer patients. Many studies have shown that overexpression of STC1 with high levels of mRNA and protein in serum and cancer tissues compared with normal counterparts correlates with a poor prognosis of patients. STC1 protein level in tumour tissues has been shown to be associated with much more lymph and distant metastases, stronger invasiveness of the tumour and poorer 5-year disease-free survival (or 3-year progression-free survival) and the overall survival rate in many kinds of tumours such as ESCC,²⁸ glioma³⁷ and gastric cancer.⁴⁸ Shirakawa et al²⁸ showed that overexpression of STC1 mRNA and protein was significantly correlated with a poor prognosis [overall survival (P < .0006) and disease-free survival (P < .0002)] of ESCC patients who had undergone curative surgery. In basal-type breast cancer patients, high expression of STC-1 protein in tissues indicated poorer relapse-free (P = .044) and overall survival (P = .0054) compared with low expression.⁵⁶ Another study also confirmed a significant association between high STC1 expression and worse OS (P = .025) and RFS (P = .0007) of patients with basal subtype breast cancer. The association was even stronger when a subgroup of basal-type BC cases with TP53 mutations was analysed separately, but no such correlations were found for luminal A and HER2 + subtypes.⁵⁷ Furthermore, STC1 may potentiate late breast cancer recurrence because the expression level of STC1 in primary breast tumours was strongly associated with late versus early recurrence at 6-10 years following diagnosis among ER-positive, tamoxifen-treated patients (OR = 2.70, 95% CI: 1.22, 5.98).¹¹¹ TNBC (triple-negative breast cancer), a subtype of breast cancer, accounts for 15%-20% of breast cancer cases, which is more aggressive and has a poorer prognosis than other subtypes. A study showed that the level of STC1 expression was markedly higher in TNBC cells than in non-TNBC cells.⁵⁶ Therefore, a much higher level of STC1 in breast cancer patients may indicate a more invasive subtype and worse prognosis. These findings may facilitate

development of new methods to diagnose and treat breast cancers using STC1 as a biomarker and treatment target based on the different subtypes of cancer. Nevertheless, the complete function and significance of STC1 in breast cancer is not yet clear and needs further investigation. In addition, STC1 is associated with relapse in leukaemia patients. Patients with AML or ALL, who experienced relapse, had significantly higher STC1 gene expression than those who achieved complete remission (P < .001 and P = .012, respectively).⁴⁶

Therefore, STC1 is probably an adverse prognostic indicator for patients, but this still needs further verification.

4 | CONCLUSION

STC1, a glycoprotein hormone first discovered in the corpuscles of Stannius, the endocrine glands on the kidney of bony fish, is a regulatory factor for calcium/phosphate homeostasis and protects the fish against toxic hypercalcemia. It has lost its regulatory function as an endocrine hormone during evolution in mammals and is believed to function as a paracrine/autocrine factor in several physiological and pathological processes. Recent studies also verified that STC1 promotes carcinogenesis in most kinds of cancer, for it prevents cellular apoptosis, promotes cancer cell viability, proliferation and invasiveness.⁵ However, it plays a cancer-suppressing role in cervical cancer, for it promotes apoptosis, inhibits proliferation and invasion, and enhances the efficacy of chemotherapy.49 Thus, its exact role during malignant processes has attracted attention, and many studies to explore its function have been performed thus far. However, the results are sometimes contradictory and the underlying mechanisms of the interaction of STC1 and cancers remain unclear. Therefore, further and deeper investigations are needed to elucidate the full profile of STC1 in carcinogenesis.

It has been reported that STC1 has a distinct clinical significance and is related to the diagnosis, pathological parameters and prognosis of cancer patients. Thus, it may be a potent diagnostic and prognostic marker for patients, which can detect diseases at a much more early stage and evaluate the outcome of patients more precisely. Furthermore, a specific antagonist of STC1 may be a potential therapeutic drug for cancer. Hence, the discovery and application of such therapeutic drugs may be a future goal.

CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

FZ and GY wrote the draft; MF, ZC, YL and JQ created the figures and table; LY and LZ provided guidance for language expression. TZ and YZ made suggestions for revision. All authors approved the final manuscript.

ORCID Fangyu Zhao D https://orcid.org/0000-0002-4191-7355

REFERENCES

- Chang AC, Dunham MA, Jeffrey KJ, Reddel RR. Molecular cloning and characterization of mouse stanniocalcin cDNA. *Mol Cell Endocrinol.* 1996;124:185-187.
- Yeung BH, Law AY, Wong CK. Evolution and roles of stanniocalcin. Mol Cell Endocrinol. 2012;349:272-280.
- Sundell K, Bjornsson BT, Itoh H, Kawauchi H. Chum salmon (Oncorhynchus keta) stanniocalcin inhibits in vitro intestinal calcium uptake in Atlantic cod (Gadus morhua). J Comp Physiol B. 1992;162:489-495.
- Olsen HS, Cepeda MA, Zhang QQ, Rosen CA, Vozzolo BL. Human stanniocalcin: a possible hormonal regulator of mineral metabolism. Proc Natl Acad Sci USA. 1996;93:1792-1796.
- Chang AC, Jellinek DA, Reddel RR. Mammalian stanniocalcins and cancer. Endocr Relat Cancer. 2003;10:359-373.
- Chang AC, Reddel RR. Identification of a second stanniocalcin cDNA in mouse and human: stanniocalcin 2. *Mol Cell Endocrinol*. 1998;141:95-99.
- Ishibashi K, Imai M. Prospect of a stanniocalcin endocrine/ paracrine system in mammals. Am J Physiol Renal Physiol. 2002;282:F367-375.
- Juhanson P, Rull K, Kikas T, et al. Stanniocalcin-1 hormone in nonpreeclamptic and preeclamptic pregnancy: clinical, life-style, and genetic modulators. J Clin Endocrinol Metab. 2016;101:4799-4807.
- Kikuchi M, Nakano Y, Nambo Y, et al. Production of calcium maintenance factor Stanniocalcin-1 (STC1) by the equine endometrium during the early pregnant period. J Reprod Dev. 2011;57:203-211.
- Takabatake Y, Oxvig C, Nagi C, et al. Lactation opposes pappalysin-1-driven pregnancy-associated breast cancer. EMBO Mol Med. 2016;8:388-406.
- Law AY, Wong CK. Stanniocalcin-1 and -2 promote angiogenic sprouting in HUVECs via VEGF/VEGFR2 and angiopoietin signaling pathways. *Mol Cell Endocrinol.* 2013;374:73-81.
- Jiang WQ, Chang AC, Satoh M, Furuichi Y, Tam PP, Reddel RR. The distribution of stanniocalcin 1 protein in fetal mouse tissues suggests a role in bone and muscle development. J Endocrinol. 2000;165:457-466.
- Johnston J, Ramos-Valdes Y, Stanton L-A, Ladhani S, Beier F, DiMattia GE. Human stanniocalcin-1 or -2 expressed in mice reduces bone size and severely inhibits cranial intramembranous bone growth. *Transgenic Res.* 2010;19:1017-1039.
- 14. Wu SF, Yoshiko Y, De Luca F. Stanniocalcin 1 acts as a paracrine regulator of growth plate chondrogenesis. *J Biol Chem.* 2006;281:5120-5127.
- Durukan Tolvanen A, Westberg JA, Serlachius M, et al. Stanniocalcin 1 is important for poststroke functionality, but dispensable for ischemic tolerance. *Neuroscience*. 2013;229:49-54.
- Holighaus Y, Weihe E, Eiden LE. STC1 induction by PACAP is mediated through cAMP and ERK1/2 but not PKA in cultured cortical neurons. J Mol Neurosci. 2012;46:75-87.
- Zhang K, Lindsberg PJ, Tatlisumak T, Kaste M, Olsen HS, Andersson LC. Stanniocalcin: a molecular guard of neurons during cerebral ischemia. *Proc Natl Acad Sci USA*. 2000;97:3637-3642.
- Huang L, Belousova T, Chen M, DiMattia G, Liu D, Sheikh-Hamad D. Overexpression of stanniocalcin-1 inhibits reactive oxygen species and renal ischemia/reperfusion injury in mice. *Kidney Int.* 2012;82:867-877.
- Pan JS, Huang L, Belousova T, et al. Stanniocalcin-1 inhibits renal ischemia/reperfusion injury via an AMP-activated protein kinase-dependent pathway. J Am Soc Nephrol. 2015;26:364-378.
- Ohkouchi S, Ono M, Kobayashi M, et al. Myriad functions of stanniocalcin-1 (STC1) cover multiple therapeutic targets in the complicated pathogenesis of idiopathic pulmonary fibrosis (IPF). Clin Med Insights Circ Respir Pulm Med. 2015;9:91-96.

WILEY

WILEY

- Sazonova O, James KA, McCudden CR, Segal D, Talebian A, Wagner GF. Stanniocalcin-1 secretion and receptor regulation in kidney cells. *Am J Physiol Renal Physiol*. 2008;294:F788-F794.
- Yeung HY, Lai KP, Chan HY, Mak NK, Wagner GF, Wong CKC. Hypoxia-inducible factor-1-mediated activation of stanniocalcin-1 in human cancer cells. *Endocrinology*. 2005;146:4951-4960.
- Law AY, Lai KP, Lui WC, Wan HT, Wong CK. Histone deacetylase inhibitor-induced cellular apoptosis involves stanniocalcin-1 activation. *Exp Cell Res.* 2008;314:2975-2984.
- Nguyen A, Chang ACM, Reddel RR. Stanniocalcin-1 acts in a negative feedback loop in the prosurvival ERK1/2 signaling pathway during oxidative stress. Oncogene. 2009;28:1982-1992.
- Chan KKS, Leung CO-N, Wong CC-L, et al. Secretory Stanniocalcin 1 promotes metastasis of hepatocellular carcinoma through activation of JNK signaling pathway. *Cancer Lett.* 2017;403:330-338.
- Pastushenko I, Blanpain C. EMT transition states during tumor progression and metastasis. *Trends Cell Biol.* 2019;29:212-226.
- Rajaram M, Li J, Egeblad M, Powers RS. System-wide analysis reveals a complex network of tumor-fibroblast interactions involved in tumorigenicity. *PLoS Genet*. 2013;9:e1003789.
- Shirakawa M, et al. Assessment of stanniocalcin-1 as a prognostic marker in human esophageal squamous cell carcinoma. Oncol Rep. 2012;27:940-946.
- Arigami T, Uenosono Y, Ishigami S, et al. Expression of stanniocalcin 1 as a potential biomarker of gastric cancer. *Oncology-Basel*. 2012;83:158-164.
- 30. Mitsuhiro S, et al. Assessment of stanniocalcin-1 as a prognostic marker in human esophageal squamous cell carcinoma. 2012;27:940-946.
- Liu G, Yang G, Chang B, et al. Stanniocalcin 1 and ovarian tumorigenesis. J Natl Cancer Inst. 2010;102:812-827.
- Zhang S, Mercado-Uribe I, Hanash S, Liu J. iTRAQ-based proteomic analysis of polyploid giant cancer cells and budding progeny cells reveals several distinct pathways for ovarian cancer development. *PLoS ONE*. 2013;8:e80120.
- Leung CC, Wong CK. Effects of STC1 overexpression on tumorigenicity and metabolism of hepatocellular carcinoma. *Oncotarget*. 2018;9:6852-6861.
- Du YZ, Gu XH, Li L, Gao F. The diagnostic value of circulating stanniocalcin-1 mRNA in non-small cell lung cancer. J Surg Oncol. 2011;104:836-840.
- Du YZ, Gu X-H, Cheng S-F, et al. The oncogenetic role of stanniocalcin 1 in lung adenocarcinoma: a promising serum candidate biomarker for tracking lung adenocarcinoma progression. *Tumour Biol.* 2016;37:5633-5644.
- Yoon JH, Kim J, Kim KL, et al. Proteomic analysis of hypoxia-induced U373MG glioma secretome reveals novel hypoxia-dependent migration factors. *Proteomics*. 2014;14:1494-1502.
- Su JY, Guo B, Zhang T, Wang K, Li X, Liang G. Stanniocalcin-1, a new biomarker of glioma progression, is associated with prognosis of patients. *Tumor Biol.* 2015;36:6333-6339.
- Hayase S, Sasaki Y, Matsubara T, et al. Expression of stanniocalcin 1 in thyroid side population cells and thyroid cancer cells. *Thyroid*. 2015;25:425-436.
- Jellinek DA, Chang AC, Larsen MR, Wang X, Robinson PJ, Reddel RR. Stanniocalcin 1 and 2 are secreted as phosphoproteins from human fibrosarcoma cells. *Biochem J*. 2000;350:453-461.
- Zhou H, Li Y-Y, Zhang W-Q, Lin D, Zhang W-M, Dong WD. Expression of stanniocalcin-1 and stanniocalcin-2 in laryngeal squamous cell carcinoma and correlations with clinical and pathological parameters. *PLoS ONE*. 2014;9:e95466.
- Macartney-Coxson DP, Hood KA, Shi H-J, et al. Metastatic susceptibility locus, an 8p hot-spot for tumour progression disrupted in colorectal liver metastases: 13 candidate genes examined at the DNA, mRNA and protein level. *BMC Cancer*. 2008;8:187.

- 42. Rezapour S, Bahrami T, Hashemzadeh S, et al. STC1 and NFkappaB p65 (Rel A) is constitutively activated in colorectal cancer. *Clin Lab.* 2016;62:463-469.
- Tamura S, Oshima T, Yoshihara K, et al. Clinical significance of STC1 gene expression in patients with colorectal cancer. *Anticancer Res.* 2011;31:325-329.
- 44. Joensuu K, Heikkila P, Andersson LC. Tumor dormancy: elevated expression of stanniocalcins in late relapsing breast cancer. *Cancer Lett.* 2008;265:76-83.
- 45. Wascher RA, Huynh KT, Giuliano AE, et al. Stanniocalcin-1: a novel molecular blood and bone marrow marker for human breast cancer. *Clin Cancer Res.* 2003;9:1427-1435.
- Abaza HM, Elmougy MI, El Maraghy HM, Mahmoud HM. Stanniocalcin1 gene expression in patients with acute leukemia: impact on response to therapy and disease outcome. *Int J Lab Hematol.* 2016;38:81-89.
- Tohmiya Y, Koide Y, Fujimaki S, et al. Stanniocalcin-1 as a novel marker to detect minimal residual disease of human leukemia. *Tohoku J Exp Med*. 2004;204:125-133.
- Fang Z, Tian Z, Luo K, Song H, Yi J. Clinical significance of stanniocalcin expression in tissue and serum of gastric cancer patients. *Chin J Cancer Res.* 2014;26:602-610.
- Guo F, Li Y, Wang J, Li Y, Li Y, Li G. Stanniocalcin1 (STC1) inhibits cell proliferation and invasion of cervical cancer cells. *PLoS ONE*. 2013;8:e53989.
- Pan X, Jiang B, Liu J, et al. STC1 promotes cell apoptosis via NFkappaB phospho-P65 Ser536 in cervical cancer cells. *Oncotarget*. 2017;8:46249-46261.
- 51. Daniel AR, Lange CA. Protein kinases mediate ligand-independent derepression of sumoylated progesterone receptors in breast cancer cells. *Proc Natl Acad Sci U S A*. 2009;106:14287-14292.
- Ismail RS, Baldwin RL, Fang J, et al. Differential gene expression between normal and tumor-derived ovarian epithelial cells. *Can Res.* 2000;60:6744-6749.
- Welcsh PL, Lee MK, Gonzalez-Hernandez RM, et al. BRCA1 transcriptionally regulates genes involved in breast tumorigenesis. *Proc Natl Acad Sci USA*. 2002;99:7560-7565.
- Luo CW, Kawamura K, Klein C, Hsueh AJW. Paracrine regulation of ovarian granulosa cell differentiation by stanniocalcin (STC) 1: Mediation through specific STC1 receptors. *Mol Endocrinol.* 2004;18:2085-2096.
- Deol HK, Varghese R, Wagner GF, Dimattia GE. Dynamic regulation of mouse ovarian stanniocalcin expression during gestation and lactation. *Endocrinology*. 2000;141:3412-3421.
- Han J, Jeon M, Shin I, Kim S. Elevated STC-1 augments the invasiveness of triple-negative breast cancer cells through activation of the JNK/c-Jun signaling pathway. Oncol Rep. 2016;36:1764.
- Zandberga E, Zayakin P, Abols A, Pupola D, Trapencieris P, Line A. Depletion of carbonic anhydrase IX abrogates hypoxia-induced overexpression of stanniocalcin-1 in triple negative breast cancer cells. *Cancer Biol Ther.* 2017;18:596-605.
- McCudden CR, Majewski A, Chakrabarti S, Wagner GF. Colocalization of stanniocalcin-1 ligand and receptor in human breast carcinomas. *Mol Cell Endocrinol.* 2004;213:167-172.
- Mccudden CR, Agata M, Subrata C, Wagner GF. Co-localization of stanniocalcin-1 ligand and receptor in human breast carcinomas. *Mol Cell Endocrinol.* 2004;213:167-172.
- Varghese R, Wong CK, Deol H, Wagner GF, DiMattia GE. Comparative analysis of mammalian stanniocalcin genes. Endocrinology. 1998;139:4714-4725.
- McCudden CR, James KA, Hasilo C, Wagner GF. Characterization of mammalian stanniocalcin receptors. Mitochondrial targeting of ligand and receptor for regulation of cellular metabolism. *J Biol Chem.* 2002;277:45249-45258.

- 62. Ching LY, Yeung BH, Wong CK. Synergistic effect of p53 on TSAinduced stanniocalcin 1 expression in human nasopharyngeal carcinoma cells, CNE2. *J Mol Endocrinol*. 2012;48:241-250.
- Watanabe T, Ichihara M, Hashimoto M, et al. Characterization of gene expression induced by RET with MEN2A or MEN2B mutation. Am J Pathol. 2002;161:249-256.
- Klein D, Demory A, Peyre F, et al. Wnt2 acts as an angiogenic growth factor for non-sinusoidal endothelial cells and inhibits expression of stanniocalcin-1. *Angiogenesis*. 2009;12:251-265.
- 65. Park WY, Hong BJ, Lee J, Choi C, Kim MY. H3K27 demethylase JMJD3 employs the NF-kappaB and BMP signaling pathways to modulate the tumor microenvironment and promote melanoma progression and metastasis. *Can Res.* 2016;76:161-170.
- Parssinen J, Alarmo E-L, Khan S, Karhu R, Vihinen M, Kallioniemi A. Identification of differentially expressed genes after PPM1D silencing in breast cancer. *Cancer Lett.* 2008;259:61-70.
- Cornmark L, Lonne GK, Jogi A, Larsson C. Protein kinase Calpha suppresses the expression of STC1 in MDA-MB-231 breast cancer cells. *Tumour Biol.* 2011;32:1023-1030.
- Zuo X, Xu W, Xu M, et al. Metastasis regulation by PPARD expression in cancer cells. JCl Insight. 2017;2:e91419.
- Nguyen THM, Carreira PE, Sanchez-Luque FJ, et al. L1 retrotransposon heterogeneity in ovarian tumor cell evolution. *Cell Rep.* 2018;23:3730-3740.
- Mitsui T, Ishida M, Izawa M, Arita J. Differences between rat strains in the development of PRL-secreting pituitary tumors with long-term estrogen treatment: In vitro insulin-like growth factor-1-induced lactotroph proliferation and gene expression are affected in Wistar-Kyoto rats with low estrogen-susceptibility. *Endocr J.* 2013;60:1251-1259.
- Kaeser MD, Iggo RD. Promoter-specific p53-dependent histone acetylation following DNA damage. Oncogene. 2004;23:4007-4013.
- Law AYS, Yeung BHY, Ching LY, Wong CKC. Sp1 is a transcription repressor to stanniocalcin-1 expression in TSA-treated human colon cancer cells, HT29. J Cell Biochem. 2011;112:2089-2096.
- Lai KP, Law AYS, Yeung HY, Lee LS, Wagner GF, Wong CKC. Induction of stanniocalcin-1 expression in apoptotic human nasopharyngeal cancer cells by p53. *Biochem Biophys Res Commun.* 2007;356:968-975.
- Wary KK, Thakker GD, Humtsoe JO, Yang J. Analysis of VEGFresponsive genes involved in the activation of endothelial cells. *Mol Cancer*. 2003;2:25.
- Kahn J, Mehraban F, Ingle G, et al. Gene expression profiling in an in vitro model of angiogenesis. Am J Pathol. 2000;156:1887-1900.
- 76. De Martino I, Visone R, Fedele M, et al. Regulation of microRNA expression by HMGA1 proteins. *Oncogene*. 2009;28:1432-1442.
- Sakata J, Sasayama T, Tanaka K, et al. MicroRNA regulating stanniocalcin-1 is a metastasis and dissemination promoting factor in glioblastoma. J Neuro-Oncol. 2019;142:241-251.
- Lu M, Song Y, Fu W, et al. MicroRNA and target mRNA selection through invasion and cytotoxicity cell modeling and bioinformatics approaches in esophageal squamous cell carcinoma. *Oncol Rep.* 2017;38:1181-1189.
- Groves TC, Wagner GF, DiMattia GE. cAMP signaling can antagonize potent glucocorticoid post-transcriptional inhibition of stanniocalcin gene expression. J Endocrinol. 2001;171:499-516.
- 80. Ellard JP, et al. The respiratory effects of stanniocalcin-1 (STC-1) on intact mitochondria and cells: STC-1 uncouples oxidative phosphorylation and its actions are modulated by nucleotide triphosphates. *Mol Cell Endocrinol.* 2007;264:90-101.
- 81. Ito Y, Zemans R, Correll K, et al. Stanniocalcin-1 is induced by hypoxia inducible factor in rat alveolar epithelial cells. *Biochem Biophys Res Commun.* 2014;452:1091-1097.

- Law AY, Ching LY, Lai KP, Wong CK. Identification and characterization of the hypoxia-responsive element in human stanniocalcin-1 gene. *Mol Cell Endocrinol*. 2010;314:118-127.
- Hsu PP, Sabatini DM. Cancer cell metabolism: Warburg and beyond. Cell. 2008;134:703-707.
- Wang YL, Huang L, Abdelrahim M, et al. Stanniocalcin-1 suppresses superoxide generation in macrophages through induction of mitochondrial UCP2. *J Leukocyte Biol.* 2009;86:981-988.
- Li R, Jia Z, Trush MA. Defining ROS in biology and medicine. *React Oxyg Species (Apex)*. 2016;1:9-21.
- Ohkouchi S, Block GJ, Katsha AM, et al. Mesenchymal stromal cells protect cancer cells from ROS-induced apoptosis and enhance the Warburg effect by secreting STC1. *Mol Ther*. 2012;20:417-423.
- Ono M, Ohkouchi S, Kanehira M, et al. Mesenchymal stem cells correct inappropriate epithelial-mesenchyme relation in pulmonary fibrosis using stanniocalcin-1. *Mol Ther.* 2015;23:549-560.
- Acharya A, Das I, Chandhok D, Saha T. Redox regulation in cancer: a double-edged sword with therapeutic potential. Oxid Med Cell Longev. 2010;3:23-34.
- Wang Y, Qi Z, Zhou M, et al. Stanniocalcin1 promotes cell proliferation, chemoresistance and metastasis in hypoxic gastric cancer cells via Bcl2. Oncol Rep. 2019;41:1998-2008.
- Yeung BH, Shek FH, Lee NP, Wong CK. Stanniocalcin-1 reduces tumor size in human hepatocellular carcinoma. *PLoS ONE*. 2015;10:e0139977.
- Dai D, Wang QI, Li X, Liu J, Ma X, Xu W. Klotho inhibits human follicular thyroid cancer cell growth and promotes apoptosis through regulation of the expression of stanniocalcin-1. *Oncol Rep.* 2016;35:552-558.
- Bai Y, Xiao Y, Dai Y, et al. Stanniocalcin 1 promotes cell proliferation via cyclin E1/cyclindependent kinase 2 in human prostate carcinoma. Oncol Rep. 2017;37:2465-2471.
- Ma X, Gu L, Li H, et al. Hypoxia-induced overexpression of stanniocalcin-1 is associated with the metastasis of early stage clear cell renal cell carcinoma. *J Transl Med.* 2015;13:56.
- Chang AC, Doherty J, Huschtscha LI, et al. STC1 expression is associated with tumor growth and metastasis in breast cancer. *Clin Exp Metastasis*. 2015;32:15-27.
- 95. Bouras T, Southey MC, Chang AC, et al. Stanniocalcin 2 is an estrogen-responsive gene coexpressed with the estrogen receptor in human breast cancer. *Can Res.* 2002;62:1289-1295.
- 96. Fuqua SA. The role of estrogen receptors in breast cancer metastasis. J Mammary Gland Biol Neoplasia. 2001;6:407-417.
- Yamnik RL, Digilova A, Davis DC, et al. S6 kinase 1 regulates estrogen receptor alpha in control of breast cancer cell proliferation. J Biol Chem. 2009;284:6361-6369.
- Pena C, Cespedes MV, Lindh MB, et al. STC1 expression by cancer-associated fibroblasts drives metastasis of colorectal cancer. *Can Res.* 2013;73:1287-1297.
- Brown JM, Wilson WR. Exploiting tumour hypoxia in cancer treatment. Nat Rev Cancer. 2004;4:437-447.
- 100. Zhou Z, Lu ZR. Molecular imaging of the tumor microenvironment. Adv Drug Deliv Rev. 2017;113:24-48.
- Bertout JA, Patel SA, Simon MC. The impact of O₂ availability on human cancer. Nat Rev Cancer. 2008;8:967-975.
- 102. Patel A, Sant S. Hypoxic tumor microenvironment: opportunities to develop targeted therapies. *Biotechnol Adv.* 2016;34:803-812.
- He LF, Wang T-T, Gao Q-Y, et al. Stanniocalcin-1 promotes tumor angiogenesis through up-regulation of VEGF in gastric cancer cells. J Biomed Sci. 2011;18:39.
- Hanahan D, Coussens LM. Accessories to the crime: functions of cells recruited to the tumor microenvironment. *Cancer Cell*. 2012;21:309-322.

ΊΙΕΥ

- 105. Orr B, Riddick ACP, Stewart GD, et al. Identification of stromally expressed molecules in the prostate by tag-profiling of cancer-associated fibroblasts, normal fibroblasts and fetal prostate. Oncogene. 2012;31:1130-1142.
- 106. Melzer C, Yang Y, Hass R. Interaction of MSC with tumor cells. Cell Commun Signal. 2016;14:20.
- 107. Klopp AH, Gupta A, Spaeth E, Andreeff M, Marini F. Concise review: dissecting a discrepancy in the literature: do mesenchymal stem cells support or suppress tumor growth? Stem Cells. 2011;29:11-19.
- 108. Block GJ, Ohkouchi S, Fung F, et al. Multipotent stromal cells are activated to reduce apoptosis in part by upregulation and secretion of stanniocalcin-1. Stem Cells. 2009;27:670-681.
- 109. Fujiwara Y, Sugita Y, Nakamori S, et al. Assessment of Stanniocalcin-1 mRNA, as a molecular marker for micrometastases of various human cancers. Int J Oncol. 2000;16:799-804.

- 110. Pantel K, Alix-Panabieres C, Riethdorf S. Cancer micrometastases. Nat Rev Clin Oncol. 2009;6:339-351.
- 111. Brantley KD, Kjærsgaard A, Cronin-Fenton D, et al. Stanniocalcin expression as a predictor of late breast cancer recurrence. Cancer Epidem Biomar. 2018;27:653-659.

How to cite this article: Zhao F, Yang G, Feng M, et al. Expression, function and clinical application of stanniocalcin-1 in cancer. J Cell Mol Med. 2020;24:7686-7696. https://doi.org/10.1111/jcmm.15348