


SHORT COMMUNICATION

Non-invasive genomics of respiratory pathogens infecting wild great apes using hybridisation capture

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Abstract

Human respiratory pathogens have repeatedly caused lethal outbreaks in wild great apes across Africa, leading to population declines. Nonetheless, our knowledge of potential genomic changes associated with pathogen introduction and spread at the human-great ape interface remains sparse. Here, we made use of target enrichment coupled with next generation sequencing to non-invasively investigate five outbreaks of human-introduced respiratory disease in wild chimpanzees living in Taï National Park, Ivory Coast. By retrieving 34 complete viral genomes and three distinct constellations of pneumococcal virulence factors, we provide genomic insights into these spillover events and describe a framework for non-invasive genomic surveillance in wildlife.

KEYWORDS

non-invasive health monitoring, pathogen genomics, respiratory disease, wild great apes

1 | MAIN

Spillover of common human respiratory pathogens to wild great apes habituated to human presence for research or tourism has been repeatedly documented.¹⁻³ The development of long-term health monitoring programmes within conservation initiatives,⁴ bringing together behavioural observations and non-invasive sampling (e.g. faeces, urine and performing necropsies on dead wildlife), has

led to the establishment of a framework for studying disease epidemiology in these endangered populations. Viruses of different families have been identified as primary causative agents, with members of the family *Pneumoviridae* being frequently reported. Bacterial co-infections, most often caused by *Streptococcus pneumoniae*,^{5,6} have contributed to disease severity, ultimately leading to mortality.

Emergence of a pathogen in a new host population raises many questions, such as whether the pathogen will spread efficiently,

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whether mutations will arise and be fixed whilst spreading and whether it will remain endemic in the population or not. With the exception of a few recent reports,^{2,3,7} earlier molecular characterisations of the human pathogens causing disease in wild great apes have been limited to a few, partial genes. These investigations were conducted using PCR-based screening approaches aimed at detecting and genotyping common respiratory agents infecting humans.^{6,8} To better understand pathogen introduction, more comprehensive genomic analyses, ideally performed on samples collected at different stages of an outbreak and from different individuals, are required. Unbiased metagenomic/metatranscriptomic analyses may allow to assemble complete genomes,^{2,3} but these are likely to require relatively deep sequencing when applied to samples with a low ratio of target (respiratory pathogen) versus background (host and other microorganisms) RNA/DNA, such as faeces. Implementing a target enrichment step prior to high throughput sequencing (HTS) can help overcome this limitation by focusing sequencing efforts on the RNA/DNA of interest,⁷ thus drastically reducing the sequencing depth required. Here, we made use of hybridisation capture coupled to HTS to re-analyse a set of necropsy and faecal samples collected from wild chimpanzees, aiming at generating complete genomes from human pneumoviruses and characterising virulence proteins of pneumococci that caused outbreaks in the early 2000s.

Between 2004 and 2009 we monitored six outbreaks of respiratory disease in two communities (South and East) of wild, human habituated chimpanzees⁹ in Tai National Park, Ivory Coast^{6,8,10} (Table 1). Faeces were collected from as many individuals as possible and full necropsies were performed on the chimpanzees that succumbed to the infection. Initial PCR diagnostics performed on lung tissue of deceased chimpanzees,¹ and at a later time on faeces of chimpanzees with clinical signs,^{6,8} targeting partial glycoprotein (G) and phosphoprotein (P) viral genes allowed for the identification and genotyping of the causative agents. To start exploring potential genome-wide diversity, we designed an RNA bait set encompassing complete human metapneumovirus (HMPV) and human orthopneumovirus (HRSV) type A and B genomes (synthesised by a service provider, MYBaits[®], supporting information methods). We first validated the technique by performing two rounds of 24-h hybrid

capture on libraries built from 12 lung RNA extracts previously identified as positive for either pneumovirus via PCR (supporting information Table S1). On-target reads represented from 0.1% to 99.08% of all reads and six complete viral genomes (three HMPV, two HRSVB and one HRSVA) could be reconstructed. To investigate whether we could broaden our dataset, we applied the method to 32 pneumovirus-positive faecal samples collected during the different outbreaks and tested in previous studies.^{6,8} Our sample selection aimed at maximising the number of individuals and the time frame tested. On-target reads represented from 69.3% to 99.74% of all reads, and 21 complete viral genomes were reconstructed (28 at 2X coverage), highlighting a good performance also on non-invasive samples (Table 1 and supporting information Table S1). As opposed to necropsy samples, which are collected (at the earliest) at the time of death several days after symptoms' onset, faeces can be collected continuously throughout the different phases of the infection, allowing to retrieve information from animals who survive (here, $n = 27$). This shows how despite not being the gold standard for respiratory viral infection diagnostics, faecal samples are well suited for and extend the reach of genomic studies of such pathogens in wildlife.

In all samples analysed from each outbreak, viral genomes identified in different chimpanzees and over time were identical. By relaxing the consensus calling criteria (from at least 20 reads and 95% agreement to at least two reads and 65% agreement), one or two single nucleotide polymorphisms (SNPs) emerged in all outbreaks studied (supporting information Tables S2 and S3), which is in line with what is generally observed in within-household and nosocomial transmission of pneumoviruses in humans.¹¹ Further to suggesting that these spillovers likely represent single introduction events, these data point towards an efficient replication and spread without the insurgence of adaptive mutations.

Previous analyses based on a partial fragment of the G gene of the HRSVB had suggested that the virus causing the 2005 and 2006 outbreaks in the South group was identical.⁸ Our genome-wide analyses revealed the existence of nine SNPs distributed over five genes that distinguish the two viruses (supporting information Table S2). This highlights the finer resolution allowed for by genome-wide

TABLE 1 Non-invasive samples analysed via in-solution hybridisation capture for viral enrichment

Outbreak period	Chimpanzee group	Virus identified	Lung samples tested	Faecal samples tested	Period covered	Complete genomes 20X (lungs, faeces)	Complete genomes 2X (lungs, faeces)
March 2004	South	HMPV ^a	3	12	08.03.2004 to 02.04.2004	7 (2, 5)	8 (1, 7)
October 2004	South	HMPV	n.a.	1	29.10.2004	0	0
August 2005	South	HRSVB	n.a.	14	16–24.08.2005	14	-
February 2006	South	HRSVB ^a	1	0	10.02.2006	1	-
February 2006	East	HRSVB ^a	2	0	07–09.02.2006	1	1
November 2009	South	HRSVA ^a	6	5	27.11.2009 to 17.12.2009	5 (1, 4)	1 (0, 1)

Note: Shown is a summary of the necropsy and faecal samples selected for viral enrichment and the respective results of genome coverage depth.

Abbreviations: HMPV, human metapneumovirus; HRSVB, human orthopneumovirus.

^aCo-infection with *Streptococcus pneumoniae*.

analyses and confirms that, as observed in humans, an infection with HRSV does not provide protection from re-infection with a highly similar, homologous subtype, even when occurring just 6 months apart. To investigate whether the same was true for HMPV, we compared positive faecal samples from the only individual in which HMPV infection was confirmed in both March and October 2004. Despite retrieving only 37.5% of the viral genome from the October sample, we could assess that this portion was identical to the strain that circulated 7 months earlier. The evidence of re-infection with seasonal human strains stresses even more the threat that these viruses pose to remaining great ape populations and the importance of implementing strict hygiene rules at tourism and research sites.⁴

Whole-genome maximum-likelihood phylogenetic analyses confirmed previous genotyping, with the 2004 HMPV strain falling within the diversity of the B2 lineage (supporting information Figure S1), the HRSVB from 2005 and 2006 within the GB3 genotype (supporting information Figure S2) and the HRSVA from 2009 within the GA2 (supporting information Figure S3). We acknowledge that given the paucity of genomic information available for viral strains circulating in humans in these remote areas, we could not fit these data to seasonal local patterns.

In four out of six outbreaks, several deaths occurred among the chimpanzees and were attributed to co-infections with *S. pneumoniae* (pneumococcus). The availability of necropsy samples allowed for an initial characterisation of pneumococcal strains via Multi Locus Sequence Typing (MLST) and serotyping. These analyses identified three distinct serotypes^{5,6} (ST 2309 in South 2004/2006, ST 2308 in East 2006, ST 8485 in South 2009), one of which was of unequivocal human origin (ST 8485). Bacterial isolation was successful only for the latter,⁶ highlighting the need of alternative tools to characterise more in depth pneumococci threatening wild populations.

To broaden the efficacy of the available polyvalent conjugate vaccines directed against pneumococcal capsular polysaccharides, recent vaccine development has been focused on targeting one or more virulence proteins¹² such as choline binding proteins (e.g. CbpA, PspA, PcpA), serine-rich repeat proteins (Psrp), pneumococcal pili (RrgA, B and C) favouring adherence and surface enzymes like the hyaluronate lyase (HysA), which favours tissue invasion. Presence and genetic variability of these proteins are known to vary substantially across serotypes identified in humans.¹³ To which extent the same occurs in pneumococcal strains circulating in wildlife remains largely unknown. To investigate the diversity of virulence factors identified in the pneumococci infecting the Taï chimpanzee population and to simultaneously design a tool that would allow for the differentiation of *S. pneumoniae* from other commensal streptococci (e.g. *Streptococcus mitis* and *Streptococcus oralis*), we designed a bait set targeting nine (entire or partial) virulence genes thus far only reported in pneumococci (supporting information Table S4). We generated DNA baits by using sheared long-range PCR products to which biotinylated adapters were subsequently attached.

Following hybrid capture on libraries generated from lung samples, 1.01% to 50.19% of the total reads were on target (supporting information Table S5), allowing for the characterisation of the

virulence factors tested in the different strains. In the 2004 and 2006 outbreak samples from the South community, only five (CbpG, CbpA, PspA, HysA and PcpA) of the nine virulence genes tested were detected (supporting information Figure S4). When compared, consensus sequences for these genes were identical, suggesting the same strain was involved in both outbreaks. The same five virulence genes, plus a sixth one (Psrp), were detected in the 2006 East outbreak samples. Sequences, however, differed from those of the 2004/2006 South strain for all genes but HysA, which was identical in its full length (approximately 3 kb). The pneumococcal strain found in South in 2009 had yet another constellation, carrying all nine virulence genes tested. Sequences of the genes shared with the 2004/2006 South and the 2006 East strains differed from each other, in line with the previous (serotype) identification of three distinct strains. The greatest diversity was recorded in the choline binding (CbpA and PspA) and serine-rich repetitive (Psrp) proteins (up to 33%, supporting information Figure S4), which are known to be highly polymorphic.¹⁴ From a functional perspective, the hyaluronidase gene of the 2004 and 2006 strains displayed an early truncation due to a one base pair deletion, suggesting potential functional loss. Inactivity of this enzyme due to indels or mutations in the coding gene has been reported for other virulent streptococci,¹⁵ implying that this gene is not an essential virulence factor.

To test the suitability of the method to faeces, we performed hybridisation capture on three libraries built from faecal extracts of the 2009 outbreak. On-target reads represented 6.27% to 12.38% of all reads, falling within the range of what was observed for the lungs (supporting information Table S5). Similarly, the profile of virulence factors was comparable, with all nine genes being detected and identical sequences. Overall, gene profiling via hybridisation capture added layers of information on pneumococci responsible for mortality in wild chimpanzees, providing a framework for better understanding pathogenesis and tailoring potential emergency interventions (e.g. vaccinations). The power of this tool ultimately lies in the flexibility of simultaneously being able to study multiple aspects that may be relevant to species conservation, for example, adding baits to characterise genes associated with antibiotic resistance.

The results herein reported provide yet another example of the quality and quantity of information that can be obtained from non-invasively monitoring outbreaks of disease in great apes. This should encourage the implementation of similar continuous surveillance programmes at other research sites or at least sampling faeces in the presence of clinical signs. To better understand the links between human and animal health, these should be coupled with similar studies in the local human population. Such comparative data could provide baseline evidence to guide improvements of wildlife conservation measures and local public health at once, for example, targeted vaccination campaigns for staff or local communities living around great ape habitats, following a truly One Health approach.

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CONFLICT OF INTEREST

No potential conflict of interest was reported by the authors.

AUTHOR CONTRIBUTIONS

Livia Victoria Patrono: Conceptualization; formal analysis; investigation; methodology; project administration. **Caroline Röthemeier:** Investigation; methodology. **Leonce Kouadio:** Investigation; resources. **Emmanuel Couacy-Hymann:** Resources; supervision. **Roman M. Wittig:** Funding acquisition; resources. **Sébastien Calvignac-Spencer:** Conceptualization; methodology; supervision; validation. **Fabian H. Leendertz:** Conceptualization; funding acquisition; methodology; resources; supervision.

PEER REVIEW

The peer review history for this article is available at <https://publons.com/publon/10.1111/irv.12984>.

DATA AVAILABILITY STATEMENT

Raw data are available in the European Nucleotide Archive, Project number PRJEB48718. Alignments of full viral genomes and pneumococcal genes are available in Zenodo (doi:10.5281/zenodo.5702462).

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REFERENCES

- Köndgen S, Kühl H, N'Goran PK, et al. Pandemic human viruses cause decline of endangered great apes. *Curr Biol*. 2008;18(4):260-264.
- Negrey JD, Reddy RB, Scully EJ, et al. Simultaneous outbreaks of respiratory disease in wild chimpanzees caused by distinct viruses of human origin. *Emerg Microbes Infect*. 2019;8(1):139-149.
- Scully EJ, Basnet S, Wrangham RW, et al. Lethal respiratory disease associated with human rhinovirus C in wild Chimpanzees, Uganda, 2013. *Emerg Infect Dis*. 2018;24(2):267-274.
- Gilardi K V, Gillespie TR, Leendertz FH, Macfie EJ, Travis DA, Whittier CA, Williamson EA. *Best Practice Guidelines for Health Monitoring and Disease Control in Great Ape Populations*. 2015.
- Chi F, Leider M, Leendertz FH, et al. New Streptococcus pneumoniae clones in deceased wild chimpanzees. *J Bacteriol*. 2007;189(16):6085-6088.
- Köndgen S, Calvignac-Spencer S, Grützmacher K, et al. Evidence for Human Streptococcus pneumoniae in wild and captive chimpanzees: A potential threat to wild populations. *Sci Rep*. 2017;7(1):1-8.
- Patrono LV, Samuni L, Corman VM, et al. Human coronavirus OC43 outbreak in wild chimpanzees, Côte d'Ivoire, 2016 correspondence. *Emerg Microbes Infect*. 2018;7(1):2-5.
- Köndgen S, Schenk S, Pauli G, Boesch C, Leendertz FH. Noninvasive monitoring of respiratory viruses in wild chimpanzees. *Ecohealth*. 2010;7(3):332-341.
- Wittig RM. Taï Chimpanzees. *Encycl Anim Cogn Behav*. 2018;1-7.
- Köndgen S, Kühl H, N'Goran PK, et al. Pandemic Human Viruses Cause Decline of Endangered Great Apes. *Curr Biol*. 2008;18(4):260-264.
- Agoti CN, Phan MVT, Munywoki PK, et al. Genomic analysis of respiratory syncytial virus infections in households and utility in inferring who infects the infant. *Sci Rep*. 2019;9(1):1-14.
- Feldman C, Anderson R. Review: Current and new generation pneumococcal vaccines. *J Infect*. 2014;69(4):309-325.
- Gámez G, Castro A, Gómez-Mejía A, et al. The variome of pneumococcal virulence factors and regulators. *BMC Genomics*. 2018;19(1):1-18.
- Croucher NJ, Campo JJ, Le TQ, et al. Diverse evolutionary patterns of pneumococcal antigens identified by pangenome-wide immunological screening. *Proc Natl Acad Sci U S A*. 2017;114(3):E357-E366.
- Hynes W, Johnson C, Stokes M. A single nucleotide mutation results in loss of enzymatic activity in the hyaluronate lyase gene of Streptococcus pyogenes. *Microb Pathog*. 2009;47(6):308-313.

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of the article at the publisher's website.

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