

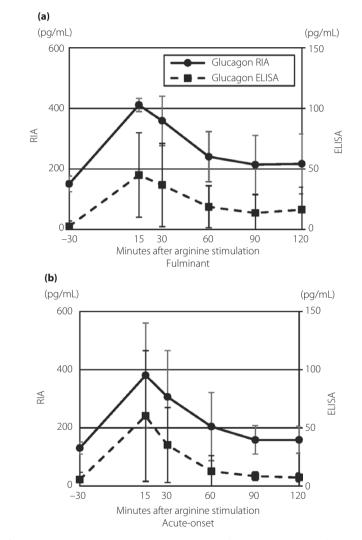
LETTER TO THE EDITOR

Response to "Preserved" glucagon secretion in fulminant type 1 diabetes

We thank Murase-Mishiba et al. for their comments<sup>1</sup> on our case report<sup>2</sup>. The Authors reported that glucagon responses to mixed meal test were not impaired in patients with fulminant type 1 diabetes whose diabetes duration was >4 years. We evaluated glucagon responses to arginine stimulation in five patients with fulminant type 1 diabetes, and those in five age- and diabetes duration-matched patients with acute-onset type 1 diabetes who were enrolled in our study<sup>3</sup> (their age and diabetes duration were 54.8  $\pm$ 20.9 vs 55.6  $\pm$  18.1 years and 3.0  $\pm$  6.3 vs  $3.2 \pm 6.9$  years, respectively; data were presented as mean  $\pm$  standard deviation). As shown in Figure 1, the curves of glucagon levels were similar between the two groups. Another 44-year-old woman underwent an arginine stimulation test at the onset of fulminant type 1 diabetes and 11 months later, and her glucagon response did not decrease (her glucagon levels at pre-loading, at peak and the area under the curve of the glucagon were; 139 vs 204 pg/mL, 406 vs 425 pg/mL and  $3.7 \times 10^4$  vs  $4.6 \times 10^4$  pg/mL/min at the onset of diabetes and 11 months later, respectively). On the contrary, another study reported that immunohistochemically stained glucagon-positive cell areas in the pancreas of patients with fulminant type 1 diabetes were significantly lower than those with autoimmune type 1 diabetes for short duration of  $2.8 \pm 1.9$  months after the onset of diabetes<sup>4</sup>. The change in pancreatic  $\alpha$ -cell

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**Figure 1** | Glucagon response to arginine stimulation in five patients with (a) fulminant type 1 diabetes, and (b) five age- and diabetes duration-matched patients with acute-onset type 1 diabetes. Solid lines with circles show glucagon levels measured by radioimmunoassay (RIA; Sceti Medical Labo, Tokyo, Japan; the intra- and interassay coefficients of variation were <20 and <15%, respectively). Dashed lines with squares show glucagon levels measured by enzyme-linked immunosorbent assay (ELISA; Mercodia AB, Sweden; the intra- and interassay coefficients of variation were 7.3–9.4% and 7.5–8.5%, respectively). Patients were enrolled in our study<sup>3</sup>. Variables are presented as mean ± standard deviation.

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Regarding the methods to measure the glucagon level, Murase-Mishiba et al. used conventional radioimmunoassav (RIA). In our case report, we measured the glucagon levels by not only RIA, but also with quantitative sandwich enzyme-linked immunosorbent assays (ELISA), and the curves of the glucagon levels were similar. We also measured the glucagon levels of patients with fulminant and acute-onset type 1 diabetes by ELISA, as shown in Figure 1. Their glucagon levels measured by RIA and ELISA were similar. However, a recent study showed that the trend of glucagon response to a mixed meal test measured by RIA was different from that measured by ELISA and novel liquid chromatography-high resolution mass spectroscopy, which can measure the glucagon levels more specifically<sup>5</sup>. Further prospective, long-term longitudinal studies with large numbers of patients are warranted to gain a complete understanding of the pancreatic  $\alpha$ -cell function in fulminant type 1 diabetes.

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## DISCLOSURE

The authors declare no conflict of interest.

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