

Review Article

How to Translate Basic Knowledge into Clinical Application of Biologic Therapy in Spondyloarthritis

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Received 7 November 2012; Accepted 24 May 2013

Academic Editor: Jieruo Gu

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Spondyloarthritis (SpA) is a family of many diseases, and these diseases share some clinical, genetic, and radiologic features. The disease process in the spine at the beginning is spinal inflammation, in which TNF α is the principal cytokine involved. Therefore, the dramatic clinical and pathologic response of anti-TNF α therapy in SpA is based upon the presence of increased TNF α in synovial tissues and sacroiliac joints, which perpetuates chronic inflammation. The increased Toll-like receptors (TLR) 2 and 4 in the serum, peripheral blood mononuclear cells, or synovial tissues of ankylosing spondyloarthritis (AS) or SpA patients suggest that SpA is highly associated with innate immunity. Any drug including anti-TNF α blocker which can downregulate the TCR, infiltrated neutrophils, or CD163+ macrophages in the synovial tissue is the rationale for the management of SpA. Like rheumatoid arthritis, the increased TH22 and TH17 cells either in blood, synovial fluid, or synovial tissues were also demonstrated in SpA. Thus, TH17 and TH22 may be reasonable cellular targets for therapeutic intervention. Drugs (anti-IL6R or anti-IL6) which can reduce the binding of IL6 and IL6R to the cell surface may be beneficial in SpA. Many proteins are implicated in the new bone formation (syndesmophyte) or ankylosis in AS or SpA. The enhanced BMP and Wnt pathway will activate osteoblasts which promote the new bone formation. However, no drug including anti-TNF α can stop or prevent the syndesmophyte in AS. In summary, looking for new targeting therapies for either anti-inflammation (beyond anti-TNF) or anti-bone formation (including anti-TGF β or PDGF) is warranted in the future.

1. Introduction

Spondyloarthritis (SpA) is a family of many diseases that includes ankylosing spondylitis (AS), reactive arthritis (ReA), psoriatic arthritis (PsA), inflammatory bowel disease (IBD), and undifferentiated arthritis (USpA) [1–3]. These diseases share some clinical, genetic, and radiologic features. The most common and important prototype in SpA is AS. Defining preaxial SpA by using the new AS criteria has recently become an important issue in the early diagnosis and management of SpA.

The pathologic process of AS or SpA can be divided into 3 stages: stage 1 is spinal inflammation, in which TNF α is the principle cytokine involved [4]. Stage 2 is erosion, in which cathepsin K or matrix metalloproteinase (MMP) may contribute. Stage 3 is abnormal bone remodeling, which can exhibit new bone formation (syndesmophytes). Bone

morphogenic protein (BMP) and Wnt protein are 2 major proteins that may enhance osteoblast activity with new bone formation.

To reduce inflammation and pain, nonsteroidal anti-inflammatory drugs (NSAIDs) are the first-line remedy in AS or SpA [3, 5–7]. However, only 70%–80% of patients have a response. The synthetic disease modifying antirheumatic drugs (DMARDs), including methotrexate, salazopyrin, and leflunomide, are active therapy for PsA or AS with peripheral arthritis but no efficacy for axial SpA [8–10].

In the past 10 years, TNF α inhibitors have been demonstrated to be very effective in rheumatoid arthritis (RA), AS, and PsA [3, 7, 11–13]. Five anti-TNF α blockers have been approved and are used in the management of inflammatory arthritis. The TNF α blocker is superior in pain relief, joint function, and life quality improvement and in reducing ESR and CRP and decreasing inflammation as seen on MRI,

compared to conventional therapy. Moreover, the rapid onset (usually 2 weeks after injection) and persistence of drug survival are two other major benefits.

From NSAIDs and DMARDs to the recent biologic therapy, what is the mechanism of these drugs that may lead to success or failure in the management of patients with RA or SpA? Why would the drugs be effective in the early stage but then lose efficacy at a later stage? In this review paper, the translation of basic knowledge to the clinical application of biologic therapy may provide some answers.

2. The Role of TNF α in SpA and Why Anti-TNF α Is Very Useful in SpA

Accumulating evidence has shown that TNF α plays a pivotal role in inflammatory arthritis [14, 15]. Many cells from the inflammatory synovium, when activated, can release different cytokines. Among them, TNF α is a potent proinflammatory cytokine exerting pleiotropic effects on various cell types. Our previous study on hip synovitis in AS and knee arthritis in RA and osteoarthritis (OA) demonstrated that TNF α expression was prominent in synovial lining cells in both RA and AS [16]. The MMP3 and CD68+ cells were significantly increased in AS compared to OA. A short-term, open-label and multiple center study in Taiwan demonstrated that etanercept was very effective in Chinese patients with AS [17]. In addition, laboratory investigation also confirmed that etanercept could significantly decrease the serum levels of IL6 and MMP3 [18]. German investigators reported that TNF α was overexpressed in synovial tissues that were obtained from the sacroiliac joint of AS patients through CT-guided needle biopsy [15]. Since 2000, many clinical trials have demonstrated that anti-TNF α therapy could significantly reduce spine and joint inflammations [3, 13, 19, 20]. This clinical response paralleled the pathologic changes [21–23]. Early research with 8 SpA patients found that after infliximab therapy, the reduction of lining cell hyperplasia, vascularity, and mononuclear cells infiltration was very dramatic and that that had a good correlation with the clinical response [23]. A recent study evaluating the effect of adalimumab treatment on synovial tissues suggested that CD3 T cells and MMP-13 might be used as biomarkers that are sensitive to changes after treatment [24]. The abundant synovial expression of RANKL and OPG in SpA was also dramatically decreased after the patients had received an anti-TNF α drug [25].

Therefore, the implication of the efficacy of anti-TNF α therapy in SpA is based on the presence of increased TNF α in synovial tissues and sacroiliac joints, which perpetuates chronic inflammation. The dramatic clinical response and long-term drug survival of TNF α blockers in those active AS patients confirmed that anti-TNF α therapy is the best treatment in AS or other SpA diseases [13, 26].

3. The Role of Innate Immunity in SpA and Future Targeting Therapy

Toll-like receptor (TLR) is a type I transmembrane receptor and can recognize the pathogen-associated molecular

pattern or danger-associated molecular pattern [27–29]. Several TLRs have been identified. Among them, TLR4 most commonly binds with the ligand, lipopolysaccharide (LPS) of Gram-negative bacteria. TLR can protect the host against bacterial infection, but when inappropriately secreted, it can cause chronic inflammation and autoimmunity. The engagement of TLRs results in activation of the NF- κ B pathway, which promotes secretion of proinflammatory cytokines, such as IL6, IL12, TNF, and IFN- γ , that may drive inflammation in AS. TNF α may increase TLR expression.

In one animal study, the enhanced B-cell TLR7 expression permits the specific development of Abs to RNA/protein complexes [30]. Zhao et al. studied TLR2 and TLR4 expression and TH1/TH2-related cytokines in bronchoalveolar lavage fluid (BALF) and peripheral blood and concluded that TLR2 and TLR4 are involved in acute inflammatory response in the lung histology in mice [31]. Study on murine salmonella intestinal infection by Shi et al. suggested in contrast to TLR5 as a “carrier of salmonella,” TLR 11 works as a “blocker of salmonella” to prevent highly invasive salmonella from penetrating into the murine Peyer’s patches [32].

Several studies have shown that TLR2 or TLR4 was significantly increased in the serum, peripheral blood mononuclear cells, or synovial tissues of AS and SpA patients [33, 34]. When those patients were treated with anti-TNF α blockers, significantly reduced TLR2, TLR4, or TLR5 was seen [33]. Innate immunity has been suggested to play a pivotal role in AS. This is based on the evidence of early synovial infiltration of neutrophils, CD163+ macrophages, and CD17+ mast cells, which are innate immune cells [35–38]. Those cells are not only present in synovial tissue, but they are already primed and activated and cause inflammation. The rationale for the management of inflammatory arthritis, particularly at the early stage of SpA [39], includes decreasing chemokine or chemokine receptors in activated innate-related immune cells, increasing the apoptosis of infiltrated macrophages (CD163+), and downregulating TLR expression.

Whether the targeting therapy for TLR can modify the SpA clinical features remains unknown, and this may need further study to elucidate.

4. Are IL17, IL23, and TH17 Cells, and Recently TH22 Cells, Playing an Important Role in AS or SpA, and How Will Future Cellular Targeting Therapy Be?

TH1 cells are important in the pathogenesis of RA. Another subset of CD4+ T cells, CD17 cells, was recently implicated to be associated with the inflammation in RA [40–42]. The proinflammatory cytokine IL17, which is released from TH17 cells, can stimulate many cells, including synovial fibroblasts, macrophages, and synovial lining cells, to produce proinflammatory cytokines (TNF α , IL6, etc.) and RANKL, as well as GM-CSF, which can enhance osteoclast numbers and activity. Several studies have explored the relationship between IL17 and IL23 and SpA [43–48]. In AS or PsA, increased serum IL17 and TH17 cells were demonstrated [43, 46]. Increased IL23R was found in psoriasis skin, and anti-IL23R could

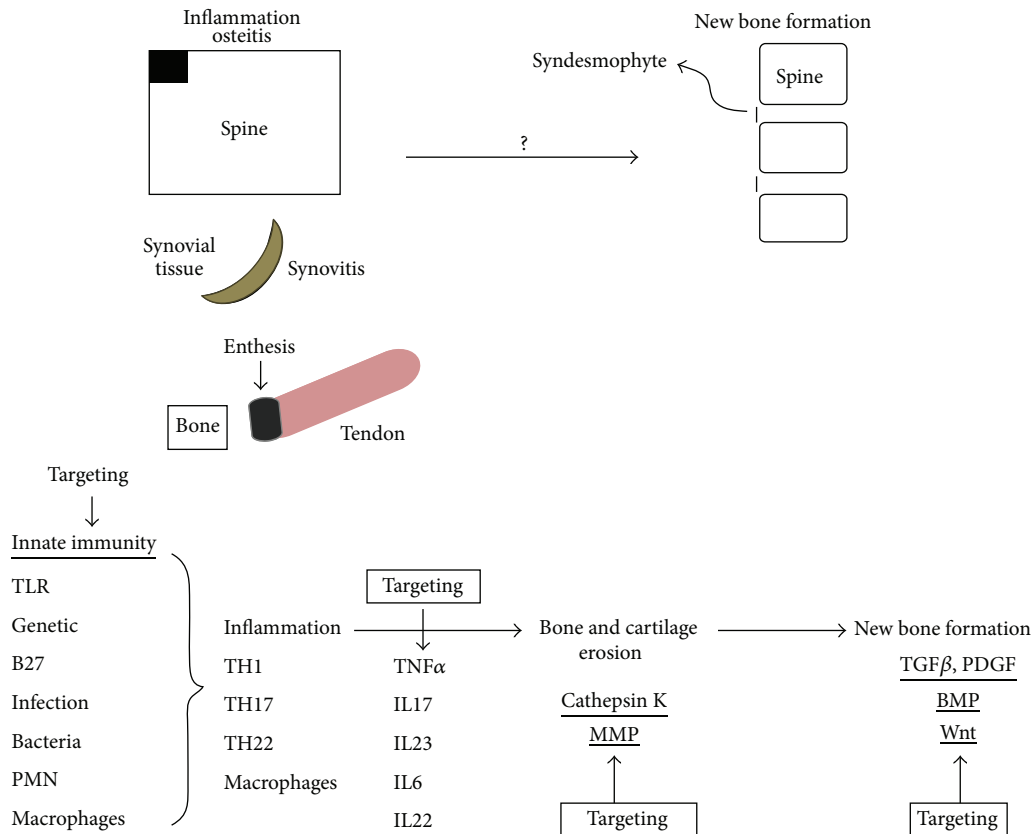


FIGURE 1: SpA: immunopathogenesis and the potential sites for targeting therapy.

improve the psoriasis [49]. Chinese investigators and our group as well have shown that serum IL17 and IL23 levels were significantly increased in AS [44, 47, 50]. Using RT-PCR, they demonstrated that IL23R P19mRNA was significantly increased in the PBMC of AS patients. Therefore, IL17, TH17 cells, and IL23 are important in the pathogenesis of AS. This has been translated into the development of a new targeting therapy, secukinumab, which has shown efficacy in early clinical trial [51].

In addition to TH1 and TH17 cells, the TH22 cells, a new human T-helper subset, were recently defined [52–55]. The naive T cell in the presence of IL6 and TNF α can differentiate toward TH22 cells. Increased IL22 from activated TH22 cells was demonstrated in the blood of RA patients and in the skin of psoriasis patients [52, 55]. The results of an investigation of TH22 in ethnic Chinese with AS showed that compared to OA, the blood TH17 and TH22 cells and serum IL22 were all significantly elevated [55]. In addition, a positive correlation between TH22 cells and TH17 cells or TH17 cells and IL22 was disclosed. Therefore, we suggest that TH22 cells, TH17 cells, and their products, IL17 as well as IL22, are implicated in the pathogenesis of AS. TH22 and TH17 cells may be reasonable cellular targets for therapeutic intervention. However, this requires further elucidation through future clinical trials.

Since TH17 cells are belonging to the upstream inflammatory cells and their products, IL17 can affect the downstream proinflammatory cytokines (TNF α , IL1, IL6, etc.) secretion,

and targeting therapy for IL17 is the rationale to suppress the inflammation in AS or SpA [56].

5. Beyond TNF α , the Role of IL6 in AS and IL6R-Targeting Therapy (Tocilizumab) in AS

A previous study showed that the disease-activity biomarkers in AS were ESR and CRP. A recent review paper indicated that CRP, IL6, and VEGF were inflammation-related markers. Among them, CRP is the most important because CRP levels in serum can predict persistent inflammation, subsequent syndesmophyte formation, and treatment response to anti-TNF α drugs in AS patients [18, 57–59]. CRP is mainly produced by hepatocytes, and 2 cytokines, including IL1 and IL6, are major stimulators. Early studies have shown that serum IL6 was increased in AS. We did a small cohort study treating ethnic Chinese AS patients with etanercept. The serum IL6 and MMP3 were significantly increased in the AS patients compared to the control, and after 12 weeks of treatment with etanercept, a significant decrease in IL6 and MMP3 was demonstrated [18]. Tocilizumab (anti-IL6R) has already been proven as a potential therapy in patients with RA [60]. The anti-IL6R drug can exert its biological effect by reducing the binding of IL6 and IL6R to the cell surface, which is the process of initiating the signal transduction for

cell activation. Anti-IL6R was beneficial in AS patients but only reported in small case studies, and this awaits further large randomized control trial for confirmation [61–63].

6. Abnormal Bone Remodeling in AS and Can We Prevent Aberrant New Bone Formation in AS?

Unlike RA, abnormal bone remodeling can occur in AS [14, 64, 65]. In RA, the increased TNF α can increase DKK1, which is the natural inhibitor of Wnt protein. When the Wnt β -catenin signal increases, it will activate osteoblasts, which cause new bone formation. In RA, the high signal of RANKL and low signal of Wnt (due to the increased DKK1) persistently stimulate osteoclast activity, but have a low probability of stimulating osteoblasts. In AS, the lower DKK1, which enhances Wnt protein, and the high signal of BMP strongly promote osteoblasts and then the development of new bone formation. Recent review paper suggested that profibrotic mediators, including transforming growth factor (TGF β), BMP, and platelet-derived growth factor (PDGF) may induce the myofibroblast phenotype and drive new bone formation [66].

The GESPIC study, with a 2-year followup of AS patients in Germany, demonstrated that with lower DKK1 there is more opportunity to develop syndesmophytes [67]. A similar study from Korea also found that DKK1 was lower in AS patients than in the control [68]. Sclerostin, another molecule like Dkk1, can suppress the Wnt pathway. Appel measured sclerostin in osteocytes and serum in RA, AS, and OA patients and again found that sclerostin was significantly lower in AS patients than in RA and OA patients and healthy controls [69]. The one molecule that can enhance osteoblasts is BMP. Our previous study demonstrated that serum BMP levels were significantly increased in AS patients, particularly in patients with spinal fusion or bamboo spine [58, 70]. However, until now, no targeting therapy for BMP, TGF β , and PDGF is available for the clinical trial to see whether they can suppress the syndesmophyte.

The immunopathogenesis and potential sites for targeting therapy is shown in Figure 1. As of the present, no drugs or biologic therapies are available to prevent or stop the new bone formation in AS. The 3 anti-TNF α blockers have been used in AS, but no one has proved to be effective in suppressing syndesmophytes [71]. Looking for new targeting therapies for those bone-formation proteins may hopefully be able to stop the excessive bone remodeling in AS.

References

- [1] J. Sieper, J. Braun, M. Rudwaleit, A. Boonen, and A. Zink, "Ankylosing spondylitis: an overview," *Annals of the Rheumatic Diseases*, vol. 61, supplement 3, pp. iii8–iii18, 2002.
- [2] M. A. Khan and S. M. van der Linden, "Ankylosing spondylitis and other spondyloarthropathies," *Rheumatic Disease Clinics of North America*, vol. 16, no. 3, pp. 551–579, 1990.
- [3] X. Baraliakos and J. Braun, "Spondyloarthritides," *Best Practice and Research: Clinical Rheumatology*, vol. 25, no. 6, pp. 825–842, 2011.
- [4] L.-S. Tam, J. Gu, and D. Yu, "Pathogenesis of ankylosing spondylitis," *Nature Reviews Rheumatology*, vol. 6, no. 7, pp. 399–405, 2010.
- [5] J. Zochling, D. van der Heijde, R. Burgos-Vargas et al., "ASAS/EULAR recommendations for the management of ankylosing spondylitis," *Annals of the Rheumatic Diseases*, vol. 65, no. 4, pp. 442–452, 2006.
- [6] J. Braun, R. Van Den Berg, X. Baraliakos et al., "2010 update of the ASAS/EULAR recommendations for the management of ankylosing spondylitis," *Annals of the Rheumatic Diseases*, vol. 70, no. 6, pp. 896–904, 2011.
- [7] Z. Ash, C. Gaujoux-Viala, L. Gossec et al., "A systematic literature review of drug therapies for the treatment of psoriatic arthritis: current evidence and meta-analysis informing the EULAR recommendations for the management of psoriatic arthritis," *Annals of the Rheumatic Diseases*, vol. 71, no. 3, pp. 319–326, 2012.
- [8] J. Braun, J. Zochling, X. Baraliakos et al., "Efficacy of sulfasalazine in patients with inflammatory back pain due to undifferentiated spondyloarthritis and early ankylosing spondylitis: a multicentre randomised controlled trial," *Annals of the Rheumatic Diseases*, vol. 65, no. 9, pp. 1147–1153, 2006.
- [9] H. Haibel, M. Rudwaleit, J. Braun, and J. Sieper, "Six months open label trial of leflunomide in active ankylosing spondylitis," *Annals of the Rheumatic Diseases*, vol. 64, no. 1, pp. 124–126, 2005.
- [10] H. Haibel, H. C. Brandt, I. H. Song et al., "No efficacy of subcutaneous methotrexate in active ankylosing spondylitis: a 16-week open-label trial," *Annals of the Rheumatic Diseases*, vol. 66, no. 3, pp. 419–421, 2007.
- [11] M. L. Hetland, I. J. Christensen, U. Tarp et al., "Direct comparison of treatment responses, remission rates, and drug adherence in patients with rheumatoid arthritis treated with adalimumab, etanercept, or infliximab: results from eight years of surveillance of clinical practice in the nationwide Danish DANBIO registry," *Arthritis and Rheumatism*, vol. 62, no. 1, pp. 22–32, 2010.
- [12] P. Emery, F. Breedveld, D. van der Heijde et al., "Two-year clinical and radiographic results with combination etanercept-methotrexate therapy versus monotherapy in early rheumatoid arthritis: a two-year, double-blind, randomized study," *Arthritis and Rheumatism*, vol. 62, no. 3, pp. 674–682, 2010.
- [13] J. Sieper, "Developments in therapies for spondyloarthritis," *Nature Reviews Rheumatology*, vol. 8, no. 5, pp. 280–287, 2012.
- [14] G. Schett, "Osteoimmunology in rheumatic diseases," *Arthritis Research and Therapy*, vol. 11, no. 1, article 210, 2009.
- [15] J. Braun, M. Bollow, L. Neure et al., "Use of immunohistologic and in situ hybridization techniques in the examination of sacroiliac joint biopsy specimens from patients with ankylosing spondylitis," *Arthritis and Rheumatism*, vol. 38, no. 4, pp. 499–505, 1995.
- [16] W. S. Chen, C. H. Chen, K. C. Lin et al., "Immunohistological features of hip synovitis in ankylosing spondylitis with advanced hip involvement," *Scandinavian Journal of Rheumatology*, vol. 38, no. 2, pp. 154–155, 2009.
- [17] C.-T. Chou, C.-Y. Tsai, T.-H. Liang et al., "Better short-term clinical response to etanercept in Chinese than Caucasian patients with active ankylosing spondylitis," *Modern Rheumatology*, vol. 20, no. 6, pp. 580–587, 2010.
- [18] K. H. Hsiao, C. Y. Tsai, T. M. Chang et al., "Significant decrease in serum IL-6 and MMP-3 after treatment in ankylosing spondylitis patients," *Formosan Journal of Rheumatology*, vol. 25, pp. 11–19, 2011.

- [19] C. Henderson and J. C. Davis, "Drug insight: anti-tumor-necrosis-factor therapy for ankylosing spondylitis," *Nature Clinical Practice Rheumatology*, vol. 2, no. 4, pp. 211–218, 2006.
- [20] M. Rudwaleit, X. Baraliakos, J. Listing, J. Brandt, J. Sieper, and J. Braun, "Magnetic resonance imaging of the spine and the sacroiliac joints in ankylosing spondylitis and undifferentiated spondyloarthritis during treatment with etanercept," *Annals of the Rheumatic Diseases*, vol. 64, no. 9, pp. 1305–1310, 2005.
- [21] J. J. Haringman, D. M. Gerlag, A. H. Zwinderman et al., "Synovial tissue macrophages: a sensitive biomarker for response to treatment in patients with rheumatoid arthritis," *Annals of the Rheumatic Diseases*, vol. 64, no. 6, pp. 834–838, 2005.
- [22] E. Izquierdo, J. D. Cañete, R. Celis et al., "Synovial fibroblast hyperplasia in rheumatoid arthritis: clinicopathologic correlations and partial reversal by anti-tumor necrosis factor therapy," *Arthritis and Rheumatism*, vol. 63, no. 9, pp. 2575–2583, 2011.
- [23] D. Baeten, E. Kruithof, F. van den Bosch et al., "Immunomodulatory effects of anti-tumor necrosis factor alpha therapy on synovium in spondylarthropathy: histologic findings in eight patients from an open-label pilot study," *Arthritis and Rheumatism*, vol. 44, no. 1, pp. 186–195, 2001.
- [24] A. W. R. van Kuijk, D. M. Gerlag, K. Vos et al., "A prospective, randomised, placebo-controlled study to identify biomarkers associated with active treatment in psoriatic arthritis: effects of adalimumab treatment on synovial tissue," *Annals of the Rheumatic Diseases*, vol. 68, no. 8, pp. 1303–1309, 2009.
- [25] C.-H. Chen, H.-A. Chen, H.-T. Liao, C.-H. Liu, C.-Y. Tsai, and C.-T. Chou, "Soluble receptor activator of nuclear factor-kappaB ligand (RANKL) and osteoprotegerin in ankylosing spondylitis: OPG is associated with poor physical mobility and reflects systemic inflammation," *Clinical Rheumatology*, vol. 29, no. 10, pp. 1155–1161, 2010.
- [26] N. Busquets-Perez, H. Marzo-Ortega, and P. Emery, "Emerging drugs for axial spondyloarthritis including ankylosing spondylitis," *Expert Opinion on Emerging Drugs*, vol. 18, no. 1, pp. 71–86, 2013.
- [27] F. K. Tan and K. Farheen, "The potential importance of Toll-like receptors in ankylosing spondylitis," *International Journal of Clinical Rheumatology*, vol. 6, no. 6, pp. 649–654, 2011.
- [28] S. Akira, "Mammalian Toll-like receptors," *Current Opinion in Immunology*, vol. 15, no. 1, pp. 5–11, 2003.
- [29] S. Akira and K. Takeda, "Toll-like receptor signalling," *Nature Reviews Immunology*, vol. 4, no. 7, pp. 499–511, 2004.
- [30] S. H. Hwang, H. Lee, M. Yamamoto et al., "B cell TLR7 expression drives anti-RNA autoantibody production and exacerbates disease in systemic lupus erythematosus-prone mice," *The Journal of Immunology*, vol. 189, no. 12, pp. 5786–5796, 2012.
- [31] C. Zhao, J. Liao, W. Chu et al., "Involvement of TLR2 and TLR4 and Th1/Th2 shift in inflammatory responses induced by fine ambient particulate matter in mice," *Inhalation Toxicology*, vol. 24, no. 13, pp. 918–927, 2012.
- [32] Z. Shi, Z. Cai, J. Yu et al., "Toll-like receptor 11 (TLR11) prevents Salmonella penetration into the murine Peyer patches," *The Journal of Biological Chemistry*, vol. 287, no. 52, pp. 43417–43423, 2012.
- [33] L. de Rycke, B. Vandooren, E. Kruithof, F. de Keyser, E. M. Veys, and D. Baeten, "Tumor necrosis factor α blockade treatment down-modulates the increased systemic and local expression of Toll-like receptor 2 and Toll-like receptor 4 in spondylarthropathy," *Arthritis and Rheumatism*, vol. 52, no. 7, pp. 2146–2158, 2005.
- [34] Z.-X. Yang, Y. Liang, Y. Zhu et al., "Increased expression of Toll-like receptor 4 in peripheral blood leucocytes and serum levels of some cytokines in patients with ankylosing spondylitis," *Clinical and Experimental Immunology*, vol. 149, no. 1, pp. 48–55, 2007.
- [35] D. Baeten, P. Demetter, C. A. Cuvelier et al., "Macrophages expressing the scavenger receptor CD163: a link between immune alterations of the gut and synovial inflammation in spondyloarthropathy," *Journal of Pathology*, vol. 196, no. 3, pp. 343–350, 2002.
- [36] E. Kruithof, D. Baeten, L. de Rycke et al., "Synovial histopathology of psoriatic arthritis, both oligo- and polyarticular, resembles spondyloarthropathy more than it does rheumatoid arthritis," *Arthritis Research & Therapy*, vol. 7, no. 3, pp. R569–R580, 2005.
- [37] D. Baeten, E. Kruithof, L. de Rycke et al., "Infiltration of the synovial membrane with macrophage subsets and polymorphonuclear cells reflects global disease activity in spondyloarthropathy," *Arthritis Research & Therapy*, vol. 7, no. 2, pp. R359–R369, 2005.
- [38] T. Noordenbos, N. Yermenko, I. Gofita et al., "Interleukin-17-positive mast cells contribute to synovial inflammation in spondylarthritis," *Arthritis and Rheumatism*, vol. 64, no. 1, pp. 99–109, 2012.
- [39] R. J. Ulevitch, "Therapeutics targeting the innate immune system," *Nature Reviews Immunology*, vol. 4, no. 7, pp. 512–520, 2004.
- [40] J. Leipe, M. Grunke, C. Dechant et al., "Role of Th17 cells in human autoimmune arthritis," *Arthritis and Rheumatism*, vol. 62, no. 10, pp. 2876–2885, 2010.
- [41] E. Lubberts, "Th17 cytokines and arthritis," *Seminars in Immunopathology*, vol. 32, no. 1, pp. 43–53, 2010.
- [42] T. Korn, E. Bettelli, M. Oukka, and V. K. Kuchroo, "IL-17 and Th17 cells," *Annual Review of Immunology*, vol. 27, pp. 485–517, 2009.
- [43] C. Jandus, G. Bioley, J.-P. Rivals, J. Dudler, D. Speiser, and P. Romero, "Increased numbers of circulating polyfunctional Th17 memory cells in patients with seronegative spondylarthritides," *Arthritis and Rheumatism*, vol. 58, no. 8, pp. 2307–2317, 2008.
- [44] W. S. Chen, Y. S. Chang, K. C. Lin et al., "Association of serum interleukin-17 and interleukin-23 levels with disease activity in Chinese patients with ankylosing spondylitis," *Journal of the Chinese Medical Association*, vol. 75, no. 7, pp. 303–308, 2012.
- [45] G. Layh-Schmitt and R. A. Colbert, "The interleukin-23/interleukin-17 axis in spondyloarthritis," *Current Opinion in Rheumatology*, vol. 20, no. 4, pp. 392–397, 2008.
- [46] Y. Mei, F. Pan, J. Gao et al., "Increased serum IL-17 and IL-23 in the patient with ankylosing spondylitis," *Clinical Rheumatology*, vol. 30, no. 2, pp. 269–273, 2011.
- [47] H. Shen, J. C. Goodall, and J. S. Hill Gaston, "Frequency and phenotype of peripheral blood Th17 cells in ankylosing spondylitis and rheumatoid arthritis," *Arthritis and Rheumatism*, vol. 60, no. 6, pp. 1647–1656, 2009.
- [48] R. Singh, A. Aggarwal, and R. Misra, "Th1/Th17 cytokine profiles in patients with reactive arthritis/undifferentiated spondyloarthropathy," *Journal of Rheumatology*, vol. 34, no. 11, pp. 2285–2290, 2007.
- [49] E. Lee, W. L. Trepicchio, J. L. Oestreicher et al., "Increased expression of interleukin 23 p19 and p40 in lesional skin of patients with psoriasis vulgaris," *Journal of Experimental Medicine*, vol. 199, no. 1, pp. 125–130, 2004.

- [50] X. Wang, Z. Lin, Q. Wei, Y. Jiang, and J. Gu, "Expression of IL-23 and IL-17 and effect of IL-23 on IL-17 production in ankylosing spondylitis," *Rheumatology International*, vol. 29, no. 11, pp. 1343–1347, 2009.
- [51] D. Baeten, J. Sieper, P. Emery et al., "The anti-IL-17A monoclonal antibody secukinumab (AIN457) showed good safety and efficacy in the treatment of active ankylosing spondylitis," *Annals of the Rheumatic Diseases*, vol. 70, no. 3, p. 127, 2011.
- [52] H. Ikeuchi, T. Kuroiwa, N. Hiramatsu et al., "Expression of interleukin-22 in rheumatoid arthritis: potential role as a proinflammatory cytokine," *Arthritis and Rheumatism*, vol. 52, no. 4, pp. 1037–1046, 2005.
- [53] S. C. Liang, X.-Y. Tan, D. P. Luxenberg et al., "Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides," *Journal of Experimental Medicine*, vol. 203, no. 10, pp. 2271–2279, 2006.
- [54] Y. Zheng, D. M. Danilenko, P. Valdez et al., "Interleukin-22, a Th17 cytokine, mediates IL-23-induced dermal inflammation and acanthosis," *Nature*, vol. 445, no. 7128, pp. 648–651, 2007.
- [55] L. Zhang, Y.-G. Li, Y.-H. Li et al., "Increased frequencies of Th22 cells as well as Th17 cells in the peripheral blood of patients with ankylosing spondylitis and rheumatoid arthritis," *PLoS One*, vol. 7, no. 4, Article ID e31000, 2012.
- [56] É. Toussiot, "The IL23/Th17 pathway as a therapeutic target in chronic inflammatory diseases," *Inflammation and Allergy—Drug Targets*, vol. 11, no. 2, pp. 159–168, 2012.
- [57] J. Gratacos, A. Collado, X. Filella et al., "Serum cytokines (IL-6, TNF- α , IL- β and IFN- γ) in ankylosing spondylitis: a close correlation between serum IL-6 and disease activity and severity," *British Journal of Rheumatology*, vol. 33, no. 10, pp. 927–931, 1994.
- [58] H.-A. Chen, C.-H. Chen, H.-T. Liao et al., "Factors associated with radiographic spinal involvement and hip involvement in ankylosing spondylitis," *Seminars in Arthritis and Rheumatism*, vol. 40, no. 6, pp. 552–558, 2011.
- [59] S. J. Pedersen, M. L. Hetland, I. J. Sørensen, M. Østergaard, H. J. Nielsen, and J. S. Johansen, "Circulating levels of interleukin-6, vascular endothelial growth factor, YKL-40, matrix metalloproteinase-3, and total aggrecan in spondyloarthritis patients during 3 years of treatment with TNF α inhibitors," *Clinical Rheumatology*, vol. 29, no. 11, pp. 1301–1309, 2010.
- [60] J. A. Schafer, N. K. Kjesbo, and P. P. Gleason, "Formulary review of 2 new biologic agents: tocilizumab for rheumatoid arthritis and ustekinumab for plaque psoriasis," *Journal of Managed Care Pharmacy*, vol. 16, no. 6, pp. 402–416, 2010.
- [61] S. Rajalingham and S. Das, "Antagonizing IL-6 in ankylosing spondylitis: a short review," *Inflammation & Allergy-Drug Targets*, vol. 11, no. 4, pp. 262–265, 2012.
- [62] J.-D. Cohen, R. Ferreira, and C. Jorgensen, "Ankylosing spondylitis refractory to tumor necrosis factor blockade responds to tocilizumab," *Journal of Rheumatology*, vol. 38, no. 7, p. 1527, 2011.
- [63] E. Koumakis, A. Feydy, A. Kahan, and Y. Allanore, "Interleukin 6 blockade in spondyloarthritis," *The Journal of Rheumatology*, vol. 39, no. 5, pp. 1097–1098, 2012.
- [64] G. Schett, "Joint remodelling in inflammatory disease," *Annals of the Rheumatic Diseases*, vol. 66, supplement 3, pp. iii42–iii44, 2007.
- [65] D. Diarra, M. Stolina, K. Polzer et al., "Dickkopf-1 is a master regulator of joint remodeling," *Nature Medicine*, vol. 13, no. 2, pp. 156–163, 2007.
- [66] C. Beyer and J. H. Distler, "Changing paradigms in spondyloarthritis: the myofibroblast signature," *Arthritis & Rheumatism*, vol. 65, no. 1, pp. 24–27, 2013.
- [67] G. R. Heiland, H. Appel, D. Poddubnyy et al., "High level of functional dickkopf-1 predicts protection from syndesmophyte formation in patients with ankylosing spondylitis," *Annals of the Rheumatic Diseases*, vol. 71, no. 4, pp. 572–574, 2012.
- [68] S.-R. Kwon, M.-J. Lim, C.-H. Suh et al., "Dickkopf-1 level is lower in patients with ankylosing spondylitis than in healthy people and is not influenced by anti-tumor necrosis factor therapy," *Rheumatology International*, vol. 32, no. 8, pp. 2523–2527, 2012.
- [69] H. Appel, G. Ruiz-Heiland, J. Listing et al., "Altered skeletal expression of sclerostin and its link to radiographic progression in ankylosing spondylitis," *Arthritis and Rheumatism*, vol. 60, no. 11, pp. 3257–3262, 2009.
- [70] H.-A. Chen, C.-H. Chen, Y.-J. Lin et al., "Association of bone morphogenetic proteins with spinal fusion in ankylosing spondylitis," *Journal of Rheumatology*, vol. 37, no. 10, pp. 2126–2132, 2010.
- [71] D. van der Heijde, R. Landewé, X. Baraliakos et al., "Radiographic findings following two years of infliximab therapy in patients with ankylosing spondylitis," *Arthritis and Rheumatism*, vol. 58, no. 10, pp. 3063–3070, 2008.