REVIEW

∂ OPEN ACCESS

Taylor & Francis

Taylor & Francis Group

Targeted alpha therapy using short-lived alpha-particles and the promise of nanobodies as targeting vehicle

Yana Dekempeneer^a, Marleen Keyaerts^{a,b}, Ahmet Krasniqi^a, Janik Puttemans^a, Serge Muyldermans ^b, Tony Lahoutte^{a,b}, Matthias D'huyvetter^{a*} and Nick Devoogdt ^b

^aVrije Universiteit Brussel, In Vivo Cellular and Molecular Imaging, Brussels, Belgium; ^bNuclear Medicine Department, UZ Brussel, Brussels, Belgium; ^cVrije Universiteit Brussel, Laboratory of Cellular and Molecular Immunology, Brussels, Belgium

ABSTRACT

Introduction: The combination of a targeted biomolecule that specifically defines the target and a radionuclide that delivers a cytotoxic payload offers a specific way to destroy cancer cells. Targeted radionuclide therapy (TRNT) aims to deliver cytotoxic radiation to cancer cells and causes minimal toxicity to surrounding healthy tissues. Recent advances using α -particle radiation emphasizes their potential to generate radiation in a highly localized and toxic manner because of their high level of ionization and short range in tissue.

Areas covered: We review the importance of targeted alpha therapy (TAT) and focus on nanobodies as potential beneficial vehicles. In recent years, nanobodies have been evaluated intensively as unique antigen-specific vehicles for molecular imaging and TRNT.

Expert opinion: We expect that the efficient targeting capacity and fast clearance of nanobodies offer a high potential for TAT. More particularly, we argue that the nanobodies' pharmacokinetic properties match perfectly with the interesting decay properties of the short-lived α -particle emitting radionuclides Astatine-211 and Bismuth-213 and offer an interesting treatment option particularly for micrometastatic cancer and residual disease.

ARTICLE HISTORY

Received 15 January 2016 Accepted 29 April 2016 Published online 19 May 2016

KEYWORDS

Cancer; targeting vehicles; targeted alpha therapy; radionuclide labeling; nanobody; bismuth-213; astatine-211

1. Introduction

1.1. Targeted radionuclide therapy

The evolution of modern medicine during the second half of the twentieth century has improved the clinical outcome of patients with numerous forms of cancer. Today, the treatment of cancer generally consists of surgery, systemic chemotherapy, radiation therapy (including external beam radiation), immunotherapy, antihormone therapy, targeted radionuclide therapy (TRNT). The choice depends upon the location and grade of the tumor and the stage of the disease, as well as the general state of the patient. Presently, tumor reduction by chemotherapy is increasingly being used in combination with surgery in multiple cancer types. Chemotherapy interacts with vital processes of the cell cycle or cell metabolism, thereby stopping or reversing cancer growth. Chemotherapy does not distinguish cancer cells from certain healthy cells, making it a less specific treatment option. External beam radiation is not suited for disseminated disease and immunotherapy often has to deal with specific resistance issues.[1]

The main objective of TRNT is the ability to selectively deliver cytotoxic radiation to cancer cells that causes minimal toxicity to surrounding healthy tissues, using optimized vehicles that deliver a nuclear payload into the tumor cells. TRNT is a growing and favorable treatment option for cancer. Currently, two principal categories can be distinguished. First, there are agents that accumulate naturally in tumor tissue. Examples are lodine-131 (¹³¹I) for the treatment of differentiated thyroid cancer [2] and Strontium-89 (89Sr) and Radium-223 (²²³Ra) for the treatment of bone metastases.[3,4] ¹³¹I and ⁸⁹Sr are both β^- -particle-emitting radionuclides, while ²²³Ra is an α-particle-emitting radionuclide. The second category includes agents that target tumor-associated antigens that are aberrantly present in malignant tissue. Examples are Yttrium-90 (⁹⁰Y)- and Lutetium-177 (¹⁷⁷Lu)-octreotide as radiolabeled peptides to treat somatostatin-overexpressing neuroblastoma.[5-7] In addition, monoclonal antibodies (mAbs) are also used as vehicles to target tumor-associated antigens and hereby providing a specific internal radiotherapy.[8] The only regulatory-approved radiolabeled mAb is ⁹⁰Y-ibritumomab to treat non-Hodgkin lymphoma.[9,10]

Thus, a radiopharmaceutical usually consists of two parts: a targeting biomolecule that specifically determines the localization of the radiopharmaceutical and a radionuclide that delivers the mechanism of action through its decay. Today, radiopharmaceuticals are used as either diagnostics for non-invasive imaging through the detection of γ -rays using positron emission tomography (PET) or single-photon emission computerized tomography (SPECT), and/or as therapeutics to deliver radiation to the targeted tumor cells. When

CONTACT Yana Dekempeneer vana.dekempeneer@vub.ac.be Research Cluster Imaging and Physical Sciences (BEFY), In vivo Cellular and Molecular Imaging Lab (ICMI), Vrije Universiteit Brussel (VUB), Campus Jette, Laarbeeklaan 103, B-1090 Jette, Belgium

*These authors contributed equally to this work.

 $\ensuremath{\mathbb{C}}$ 2016 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article highlights

- Due to the short range in tissue and high linear energy transfer of aparticles, targeted alpha therapy (TAT) is ideal for micrometastatic or residual disease
- Nanobodies are the smallest antibody-derived antigen-binding fragments and have superior characteristics compared to classical mAbs and their derived fragments for *in vivo* cell targeting
- Nanobodies are being evaluated intensively as both diagnostic tracers for nuclear imaging and vehicles for TRNT
- The combination of the short-lived α-particle emitters ²¹¹At and ²¹³Bi and nanobodies offer new possibilities for their application in TAT, which will most likely be demonstrated by ongoing and planned research work.

The box summarizes key points contained in the article.

radiopharmaceuticals are employed both for diagnosis and therapy, they are referred to as 'theranostic agents.' This combined diagnostic-therapeutic procedure uses a diagnostic test to determine whether a patient may benefit from a specific therapeutic drug, allowing personal, structural, and functional characterization of a tumor during therapy. Moreover, the therapy response could be measured throughout the therapy.

In general, there are three types of radiation that can be used for TRNT: β^- -particles, Auger electrons, and α -particles. Each radionuclide is characterized by its own decay properties, tissue range, half-life, and chemistry, proposing the opportunity to adapt the features of the radionuclide to a particular type of cancer and in the long run to the needs of an individual patient.[11] Until now, TRNT has been mainly explored using β^- -particle-emitting radionuclides. β^- -Particles have a low linear energy transfer (LET) (0.2 keV/µm), producing repairable DNA damage including single- or double-stranded DNA breaks, base chemical modifications, and protein crosslinks. In case of low-LET radiation, like for example β^- -particles, the damage caused by direct ionization of the target might only be sublethal, if dosed insufficiently high. Indirect effects caused by reactive oxygen species (ROS) also contribute to the eventual damage. β^{-} -Particles have a relatively long range in tissue (1–10 mm), causing cytotoxic damage in surrounding nontargeted cells, referred to as 'crossfire effect.' This might be useful for the treatment of heterogeneous, bulky tumors, but it has the disadvantage of damaging surrounding normal tissue. Most progress with β^- -particle radiation has been made in hematological malignancies, while the progress in epithelial-derived tumors has been slow. One of the shortcomings of low-LET β^- -particle-emitters is that much more of the radioactivity need to reach the tumor tissue to effectively kill it, compared to high-LET α -particles. A single α -particle is sufficient to destroy the cell nucleus, as cell death due to the a-radiation is largely independent of oxygenation or active cell proliferation. β^{-} -Particles on the other hand need much more hits at the level of the cell nucleus as they produce sparse ionization and individual DNA lesions, mostly repairable. This disadvantage is one of the reasons for the average success of agents labeled with β^- -particle-emitting radionuclides in clinical trials. Theoretically, Auger-electron emitters present multiple advantageous characteristics, making it an attractive candidate for TRNT. Auger emitters have a short effect range (subcellular, order of nanometers), a LET of 4-26 keV/µm, and

are able to produce a high level of cytotoxicity due to Auger electron cascades. These cascades, by which electrons, carrying a characteristic kinetic energy, are ejected from atoms in response to a downward transition by another electron in the atom. In contrast to α -radiation, Auger radiation is of low toxicity when decaying outside the cell nucleus, such as in the cytoplasm or outside of cells, and will therefore cause little damage to nontargeted cells. Some studies have shown that Auger electrons can be effective when targeted only to the cell membrane.[12] However, it is generally considered that the radioisotope needs to be delivered close to the cell nucleus in order to be effective, which makes internalization into the cell crucial.[13]

1.2. General considerations of targeted alpha therapy

The selection of the appropriate radionuclide depends on its decay properties, namely the physical half-life and emission characteristics. For the management of bulky and heterogeneous tumors, treatment with β^{-} -particle-emitting radionuclides might be the preferred approach. However, for the eradication of small-volume tumors and small clusters of cancer cells, agents that emit high-energy a-particles would be more beneficial due to their highly specific toxic load to the targeted tumor cells and their short range in tissue. Thus, the main strength of targeted alpha therapy (TAT) is the potential to deliver radiation in a highly localized and toxic manner, because of their high level of ionization produced and short range in tissue.[14] An α -particle consists of a ⁴He nucleus; therefore, it is much heavier than other subatomic particles emitted from decaying radionuclides and nuclear reactions. The main characteristics of currently available α -particle-emitting radionuclides are summarized in Table 1.[12] With a charge of +2, α -particles are effective ionization agents with a high LET (50–230 keV/µm) at a short range of 50–100 µm in tissue. They induce clusters of DNA damage such as doublestranded DNA breaks and base chemical modifications that evoke a large number of cellular responses and pathways that include apoptosis, autophagy, necrosis, and cell-cycle arrest. This type of damage is difficult to repair by the cell. Moreover, the damage is independent from the generation of indirect ROS, leaving their effectiveness potentially unabated by tumor hypoxia.[12] These characteristics make α-emitters effective in eradicating small clusters or isolated cancerous cells with little exposure to surrounding healthy tissue. Thus, TAT is of high interest for the treatment of micrometastatic and minimal residual disease after surgery. Moreover, the concept of TAT has moved from bench to bedside, with increasing clinical experience in, for example, ovarian cancer, metastatic prostate cancer, gliomas, and acute myeloid leukemia (Table 2). A median survival of 8.9 months could be achieved after intravenous administration of the a-immunoconjugate, Bismuth-213 (²¹³Bi)-cyclic diethylenetriaminepentaacetic acid anhydride (cDTPA)-9.2.27, in patients with metastatic melanoma in a phase I trial.[15] Using TAT to treat metastatic melanoma, a-particles reach the endothelial cell nuclei, causing cell death and leading to capillary closure and interruption of nutritional support to the tumor. If enough capillaries are closed down, the tumor might regress and could even disappear. Thus, this

Table 1. Main characteristics of the currently available α -particle-emitting radionuclides.

Maximum						
	Daughter	Physical	energy	Occurrence	Associated	
lsotope	isotopes*	half-life	(keV)	(%)	emissions	
²¹¹ At	_	7.2 h	5.867	α (41.8%)	α, γ, LEE	
	²¹¹ Po	516 ms	7.450	α (100%)		
²²⁵ Ac	_	10 days	5.830	α (100%)	α, γ, Auger, $β^-$	
	²²¹ Fr	4.9 min	6.341	α (100%)		
	²¹⁷ At	32.3 ms	7.069	α (99.98%)/β-		
				(0.01%)		
	²¹³ Bi	45.6 min	6.051	α (2.2%)/		
				β [–] (97.8%)		
	²¹³ Po	4.2 µs	8.377	α (100%)		
²¹³ Bi	-	45.6 min	6.051	α (2.2%)/β-	α, γ, Auger, β ⁻	
				(97.8%)		
	²¹³ Po	4.2 µs	8.377	α (100%)		
²¹² Bi	-	61 min	5.870	α (36%)/β-	α, γ, Auger, β ⁻	
				(64%)		
	²¹² Po	298 ns	8.785	α (100%)		
²²⁷ Th	_	18.72 days	6.038	α (100%)	α, γ, Auger, β ⁻	
	²²³ Ra	11.4 days	5.871	α (100%)		
	²¹⁹ Rn	4 s	6.819	α (100%)		
	²¹⁵ Po	1.8 ms	7.386	α (100%)		
	²¹¹ Bi	2.14 min	6.623	α (99.7%)/β-		
24.2				(0.3%)		
²¹² Pb	_	10.64 h		β [–] (100%)	β-	
	²¹² Bi	61 min	5.870	α (36%)/β-	α, γ, Auger, β ⁻	
				(64%)		
	²¹² Po	0.3 µs	8.785	α (100%)		
²²³ Ra	-	11.4 days	5.871	α (100%)	α, γ, Auger, β ⁻	
	²¹⁹ Rn	4 s	6.819	α (100%)		
	²¹⁵ Po	1.8 ms	7.386	α (100%)		
	²¹¹ Bi	2.14 min	6.623	α (99.7%)/β-		
				(0.3%)		

*Generated α -particle emitter after decay of the conjugated parent. LEE: Low-energy electron emission; NS: yield not significant.

subtype of TAT targets specifically the vasculature and has been referred to as 'tumor anti-vascular a-therapy (TAVAT).'[16] TAT has been compared to β^- -particle-emitting radionuclides in several clinical trials, highlighting their promising therapeutic potential. For example, investigators compared ¹³¹I-labeled bisphosphonates with their Astatine-211 (²¹¹At)-labeled counterparts for pain relief in patients with bone metastasis.[17] In addition, Henriksen et al. explored the bone-seeking properties of ²²³Ra and compared it with those of the β^- -particle-emitting radionuclide ⁸⁹Sr.[3] The conclusion of both studies was that a particle radiation showed a lower toxic effect to the healthy bone marrow compared to β^{-} -particle emitters, which is attributed to the reduced crossfire effect. This and other studies indicated that the strength and short distance of high-LET a-particles make them more suitable than low-LET β^- -particles in particular circumstances. Despite its positive features, the translation of TAT into the clinic has been slow, mainly due to the limited radionuclide availability and the short physical half-life and daughter aparticles of some of the available α -emitters. Furthermore, several other issues concerning a-particle emitters should be addressed as well, which are discussed in the following paragraphs.

1.2.1. Radiolysis

Radiolysis is the dissociation of molecules by nuclear radiation. The magnitude of energy deposits by volume of α -particle emitters is two times greater than that of β^- -emitters such as

 90 Y or 131 I. Because of this, the potential impact of radiolysis effects when using α -particles is noticeably higher. Hence, the radiolabeling of certain vectors with an α -particle emitter using high levels of radioactivity while maintaining appropriate biological properties may be challenging.[51] Studies by Zalutsky et al. indeed emphasize the potential importance of radiolysis-mediated effects on the chemistry of α -particle emitting radiopharmaceuticals and the need to evaluate their labeling chemistry and stability at high doses required for clinical use.[63,64]

1.2.2. The radiation-induced biological bystander effect

The radiation-induced biological bystander effect (RIBBE) is a process whereby nontargeted healthy cells are damaged, not as a result of directly being hit by radiation, but via the radiation-induced death or stress of neighboring cells. As aparticle-emitting radionuclides have a range in tissue that is equivalent to only a few cell diameters, the physical crossfire effect will be limited. To date, the majority of studies of RIBBE have been performed in vitro using single-cell or multicellular systems ex vivo or in artificial three-dimensional human tissue systems. Boyd et al. demonstrated that cell death in adjacent cells after treatment with a-particle-emitting radionuclides might be enhanced via RIBBE.[65] Furthermore, evidence on the in vivo effectiveness of RIBBE has been limited, but new findings indicate that they may affect tumor development in susceptible mouse models. For example, Mancuso et al. demonstrated that DNA double strand breaks and apoptotic cell death could be induced by bystander responses in mouse cerebellum after X-ray exposure of the remainder of the body. [66] Mice were whole-body exposed or irradiated with individual cylindrical lead shields providing protection of heads. Whole-body-irradiated animals developed cerebellar tumors. A high percentage of mice (62%) died of aggressive disease by 23 weeks, with median survival of 14 weeks. Significantly, they also observed a remarkably increased medulloblastoma rate (39%) in lead shielded-irradiated mice, indicating that bystander effects are factual in vivo events with carcinogenic potential. However, the underlying mechanisms are incompletely characterized and it remains unclear how processes involving oxidative metabolism and stress-inducible proteins lead to (oxidative) DNA damage in bystander cells.[67]

1.2.3. Distribution of recoil daughters in the body

Another important aspect that should be taken into account is the unstable bond of daughter isotopes upon α -decay due to the different chemical properties of the daughters. This could result in an immediate loss of the daughter atom from the chelating chemistry.[68] In addition, the recoil energy of the recoiling daughters is more than 1000 times higher than the binding energy of any chemical compound, which will lead to the rupture of the chemical bonds of the daughter atom with the targeting vehicle, as well as to the ionization of the surrounding medium. The released daughter isotopes that are often themselves α -emitters might cause substantial harm since they will no longer be bound to the targeting vehicle. Therefore, it is of utmost importance to study the fate of both mother and daughter isotopes. For instance, the biodistribution of the bone-targeting radiopharmaceutical

Table 2.	Vehicles	used in	targeted	α-particle	therapy in	n preclinical	and clinic	al settings.
----------	----------	---------	----------	------------	------------	---------------	------------	--------------

		1 II	A	Reference	Reference
Radionuclide	IAI agent	Indication	Antigen	(preclinical data)	(clinical phase)
²²⁵ Ac	Anti-CD33 IgG (HuM195)	Leukemia	CD33	[18]	l [19,20]
²²⁵ Ac	Anti-HER2 IgG (trastuzumab)	Ovarian cancer	HER2	[21]	-
²²⁷ Th	Anti-HER2 IgG (trastuzumab)	Breast and ovarian cancer	HER2	[22,23]	
²²⁷ Th	Anti-CD20 IgG (rituximab)	Non-Hodgkin lymphoma	CD20	[24,25]	
²¹³ Bi	Anti-CD33 IgG (HuM195)	Leukemia	CD33	[26,27]	I and I/II [28,29]
²¹³ Bi	Anti-CD20 lgG (rituximab)	Non-Hodgkin lymphoma	CD20	[30,31]	I [32]
²¹³ Bi	Plasminogen activator inhibitor	Breast cancer, pancreatic cancer	Urokinase plasminogen	[33–35]	
	type 2		activator receptor		
²¹³ Bi	Anti-MUC1 IgG (C595 IgG)	Ovarian cancer, pancreatic cancer	MUC1	[36,37]	
²¹³ Bi	Substance P	Glioblastoma	Neurokinin type-1 receptor		0/I [38,39]
²¹³ Bi	Anti-NG2 lgG (9.2.27 lgG)	Melanoma	NG2 proteoglycan	[40,41]	l [15,42,43]
²¹³ Bi	Anti-CD138 lgG	Multiple myeloma	CD138	[44]	
²¹³ Bi	Anti-PSMA lgG (J591 lgG)	Prostate cancer	PSMA	[45]	
²¹³ Bi	C6.5K-A scFv, C6.5K-A diabody	Breast and ovarian carcinomas	HER2	[46]	
²¹² Pb/ ²¹² Bi	Anti-HER2 IgG (TCMC-	Ovarian cancer	HER2	[47,48]	[48–50]
	trastuzumab)				
²¹¹ At	Chimeric 81C6 lgG	Glioblastoma	Tenascin-C	[51,52]	II [53]
²¹¹ At	MX35 F(ab') ₂	Ovarian cancer	NaPi2b	[54]	I [55]
²¹¹ At	Anti-FRA IgG (Mov18)	Ovarian cancer	Folate receptor alpha	[56]	
²¹¹ At	Anti-EGFRvIII IgG	Glioblastoma	EGFRvIII	[57]	
²¹¹ At	Anti-HER2 C6.5 diabody	Breast cancer	HER2	[58]	
²¹¹ At	Z _{HER2:342} and (Z _{HER2:4}) ₂ affibody	Breast and ovarian carcinomas	HER2	[59]	
	molecules				
²²³ Ra	²²³ Ra-chloride	Skeletal breast and prostate cancer	Hydroxyapatite	[60]	I–III [61,62]
		metastases			

NG2: Neural/glial antigen 2; PSMA: prostate-specific membrane antigen; EGFRvIII: epidermal growth factor receptor variant III.

²²³Ra, which naturally targets the hydroxyapatite matrix in the bone, has been studied extensively in vivo.[3,69] Although the daughter isotopes are not intrinsically bone-seeking, the rapid cascade of a-particle-emitting daughters will deliver high doses to bone metastases. However, their short half-life appears to prevent them from causing major damage to healthy tissue. An in vivo study demonstrated that less than 2% of the daughters migrate away from the bone surface within 6 h after administration of ²²³Ra, and after 3 days, this number has dropped down to less than 1%.[3] Another example is the decay of actinium-225 (²²⁵Ac) with the formation of potentially disadvantageous radiotoxic daughter products such as ²¹³Bi. It is critically important to reduce the redistribution of the daughter isotopes to nontarget tissues and to diminish systemic radiotoxic events. Therefore, the ²²⁵Ac 'nanogenerator' approach was designed in which the delivery system is engineered to be internalized into the targeted tumor cell.[70] McDevitt and colleagues demonstrated the ability to safely and efficiently use ²²⁵Ac as a potent tumorselective generator in both established solid carcinomas and disseminated cancers.[71] Although these results were very promising, additional development of this modality is warranted to optimize the stability of the nanogenerator to maximize the retention of the tumor while avoiding uptake in healthy organs.

1.2.4. Dosimetry

Radiation dosimetry is the measurement of the absorbed dose delivered by the ionizing radiation and provides a basis for understanding the effects and efficacy of different radiationbased treatments. One of the major impediments of TRNT is the heterogeneous distribution of the radiopharmaceutical in normal and tumor tissues. In the case of α -particle radiation, their short path length and high LET need to be taken into account, posing an enormous challenge on the methods needed for relevant dosimetry.[72] For high-LET irradiation, the effect of a single incident in the nucleus of the cell is so abundant that the variations in absorbed dose (specific energy) to the nucleus can be very large and therefore might be a misleading index of the biologic effect. The clinical quantification of the absorbed doses with the y-camera is only able to give an estimate about the uptake of the radiopharmaceutical in whole organs and in macroscopic tumors, while quantification of absorbed doses in smaller compartments in organs or microscopic tumors is barely feasible. Thus, smallscale dosimetry or microdosimetry, which takes into account the stochastic nature of energy deposited in small targets, would generate improved dosimetric calculations for a-particle radiation. Due to the limited clinical experience with aparticles to date, unknown maximum tolerable doses in humans are the major issue in TAT. In mice, absorbed doses of a-particle radiation can be calculated in tissues at a macroscopic level (organs and substructures) using Monte Carlo techniques based on fundamental physical principles.[73,74] In addition to that, Bäck and colleagues developed the acamera, which is a quantitative imaging technique developed to detect a-particles in tissues ex vivo at suborgan level, to get a better view on the biodistribution of internal a-radiation on a cellular level.[75] The high-resolution (35 μ m or less) α camera was able to measure the activity distribution on a cellular level by virtue of the short path length of α -particles, making it a promising tool in the evaluation of future TAT.

2. The current developments

2.1. A milestone for TAT: radium-223

Radium (Ra) and polonium (Po) were first described by Marie and Pierre Curie in 1898 while investigating the radioactive

properties of a complex ore, which had radioactive emissions in excess. ²²³Ra and ⁸⁹Sr are bone-targeting radiopharmaceuticals with hydroxyapatite ($Ca_5[PO_4]_3OH$) as target, which is an essential component of the inorganic bone matrix. Ra, barium (Ba), Sr, and calcium (Ca) are all chemicals in the alkaline earth metal family on the periodic table and each will localize in the areas of osteoblastic metastases. ²²³Ra is currently the most commonly used radioisotope for medical therapeutics, showing an increased survival in patients with metastatic castration-resistant prostate cancer [61] and has a half-life of 11.4 days (Table 1). ²²³Ra is the first α -emitter approved by the US Food and Drug Administration.[76] In addition, ²²³Ra is the first α -particle-based therapy that results in pain relief and extends survival in patients with progressive castration-resistant prostate cancer and bone metastasis in the absence of visceral metastasis. Thus, ²²³Ra is naturally incorporated in areas of increased bone turnover in bone metastases.[77] More than 90% of patients with metastatic resistant prostate cancer have radiologic evidence of bone metastases. ²²³Ra dichloride has been evaluated in two phase I trials and three double-blind phase II trials. The phase III ALSYMPCA (Alpharadin in the Treatment of Patients With Symptomatic Bone Metastases in Castration-Resistant Prostate Cancer) trial showed an improved overall survival of 3 months and pain relief in patients with osseous metastasis.[61] The success of ²²³Ra as a therapeutic further stimulates TAT-based preclinical and clinical research. In a way, ²²³Ra could be considered as a game changer in nuclear medicine, as it might facilitate the future use of additional high-LET particle emitters.

2.2. Other promising a-particle-emitting radionuclides

Besides ²²³Ra, many other α -particle emitters have suitable characteristics for therapeutic applications (Table 2). ²¹¹At, ²¹³Bi, lead-212 (²¹²Pb)/bismuth-212 (²¹²Bi), and ²²⁵Ac are the most frequently used α -particle-emitting radionuclides in clinical molecular targeting applications to date.[78]

2.2.1. Actinium-225

 ^{225}Ac is a parent $\alpha\text{-particle}$ emitter in a decay cascade that produces three net a-particle isotopes, ²²¹Fr (half-life 4.8 min), ²¹⁷At (half-life 32.3 ms), and ²¹³Bi (half-life 45.6 min), making it a very effective and potent option for TAT (Table 1). ²²⁵Ac has a half-life of 10 days and can be produced by natural decay of ²³³U in Oak Ridge National Laboratory, USA [79] or by acceleratorbased methods in Karlsruhe.[80] However, the latter production of ²²⁵Ac also results in the production of ²²⁷Ac which decays with a half-life of 21.772 years. The biggest disadvantage concerning ²²⁵Ac is its cost, which might reach to \$1200/mCi. In addition, the recoiled daughters of ²²⁵Ac can do significant damage to healthy tissue when not retained at the tumor site. Encapsulation in a nano-carrier, fast uptake of the α -particle-emitting radionuclides in tumor cells, and local administration are some approaches to minimize toxic effects caused by a-particle-emitting daughters. [68] On the other hand, the relatively long half-life of ²²⁵Ac allows a centralized production and shipment of the irradiated targets to further users so that any investigator is able to exploit the power of this α -particle. Furthermore, ²²⁵Ac decays to ²¹³Bi, of which the latter also results in a 440 keV y-ray emission that can be useful for imaging of the therapeutic biodistribution. It should be remarked that it is uncertain whether the measured radioactive decay represents intact radiopharmaceutical or released daughter radioisotopes. Moreover, ²²⁵Ac can be conjugated to peptides or antibodies, using an optimized radiochemistry with standard widely available macrocyclic bifunctional chelators. [81,82] In vivo experiments showed that the ²²⁵Ac complex 1,4,7,10-tetraazacyclododecane-*N*,*N*',*N*'',*N*'''-tetra-acetic with acid (DOTA) was more stable than the ²²⁵Ac complex with 4,7,10,13,16-hexaazacyclohexadecane- N,N',N'',N''',N'''',N''''-hexaacetic acid.[70] The biodistribution aspects of ²²⁵Ac-labeled mAbs and other carriers, together with their pharmacokinetic properties, radiobiology, and dosimetry, have been reviewed by Miederer et al.[70] A successful phase I trial has demonstrated that a humanized anti-CD33 mAb HuM195 conjugated to ²²⁵Ac (Actimab-A) is safe to use at doses $\leq 0.1 \text{ MBq/kg}$ [19] (Table 2).

2.2.2. Bismuth-213

²¹³Bi is most often produced through an ²²⁵Ac-generator. The principal drawbacks of using ²¹³Bi are its very short physical half-life of 46 min and limitations regarding availability and cost as for ²²⁵Ac. Pippin and colleagues were the first to label ²¹³Bi with mAbs.[83] Moreover, McDevitt and colleagues labeled ²¹³Bi via the bifunctional metal cDTPA complex with a humanized mAb (HuM195) directed against CD33, a glycoprotein expressed on the majority of myeloid leukemia cells.[26] In subsequent studies, the stability of this radiopharmaceutical has been improved to achieve a clinically applicable ²¹³Bi-CHX-A-DTPA-HuM195.[84] A phase I clinical study on 18 patients with acute myelogenous leukemia (AML) or chronic myelomonocytic leukemia showed no significant extramedullary toxicity, although myelosuppression was seen in all patients.[28] The phase I/II trials showed that sequential administration of cytarabine and ²¹³Bi-CHX-A-DTPA-HuM195 was reported to be tolerable and produced remissions in some patients with AML, although myelosuppression was again a common adverse effect.[29] The responses in this high-risk population persisted up to 12 months. In addition, patients with non-Hodgkin lymphoma, malignant melanoma, and glioblastoma have been enrolled in clinical trials with other ²¹³Bi-labeled compounds, showing its relevant potential for TAT (Table 2).

2.2.3. Astatine-211

²¹¹At is an α-particle-emitting radionuclide with a physical half-life of 7.2 h and its decay does not result in the production of any relevant daughter isotopes. The first branch decays to ²¹¹Po (half-life 526 ms), after which it decays through α-particle radiation to stable ²⁰⁷Pb. In the second branch, ²¹¹At α-decays to ²⁰⁷Bi, which then results in stable ²⁰⁷Pb after emission of X-rays. Theoretically, this offers significant advantages for TAT regarding minimal toxicity and quantitative α-particle emission. However, additional clinical research is needed in order to confirm this as a real advantage. The chemical features of ²¹¹At are similar to those of iodine, its nearest halogen neighbor, but ²¹¹At contrarily also tends to behave as a metalloid. Moreover, the exact behavior of ²¹¹At is far from understood due to the limited knowledge of the chemistry of elemental ²¹¹At and the lack of any stable equivalent, which excludes the use of conventional analytical techniques for its characterization.[85] Reasonable yields (0.8-2.5 GBg) of ²¹¹At are obtained via the bombardment of natural bismuth targets with α-particles through the $^{209}\text{Bi}(\alpha,~2n)$ ^{211}At nuclear reaction in a cyclotron.[86] The 7.2-h half-life of ²¹¹At is well suited for a multistep synthetic procedure. Consequently, a wide variety of tumor-associated antigens that are aberrantly expressed on the cancer cell surface have been targeted by ²¹¹At-labeled radiopharmaceuticals.[87,88] To date, ²¹¹At has been investigated bound to antibodies, thymidine analogs, [89] biotin analogs, [90] colloids, [91] melanin precursors, [92] substrate carriers,[93] and bisphosphonate complexes. [94] Only two clinical studies have been reported so far with ²¹¹At-labeled molecules.[53,55] The first clinical study for the treatment of recurrent brain tumor provides a proof-of-concept for regional targeted radiotherapy with ²¹¹At-labeled mAbs.[53] This clinical study demonstrated that the regional administration of ²¹¹At-ch81C6 was feasible, safe, and resulted in a possible therapeutic benefit for patients with malignant brain tumors. In the second reported clinical study of ²¹¹At using the MX35 F(ab')₂, the compound was delivered successfully through intraperitoneal administration without observed toxicity.[55] These two clinical trials showed no subjective toxicity related to the immunoconjugate and the overall outcomes were highly encouraging. However, there are no clinical data on the toxicity of ²¹¹At-labeled immunoconjugates after intravenous administration. Further clinical evaluation of ²¹¹At-labeled compounds in metastatic tumors or residual disease is warranted.

3. Vehicles for TAT

The attractive feature of TRNT is its adaptable nature. The radionuclide and the targeting vehicle should in principle be matched to each other in the context of the route of administration, disease stage, target accessibility, and site of action. The selection of both the optimal tumor-associated antigen and the targeting vehicle is a crucial step in the development of a new probe for TRNT. The ideal antigen should be overexpressed on cancer cells, while the expression levels on normal, healthy cells should be extremely low.[95] Examples of biomarkers that are targeted in TAT studies are epidermal growth factor receptor variant III, human epidermal growth factor receptor 2 (HER2), folate receptor alpha, tenascin-C, CD20, CD33, and prostate-specific membrane antigen (Table 2). The vehicle molecules should be optimized to provide a high degree of selectivity and specificity toward the target site or 'biomarker.' Below, a section of important vehicles are discussed.

mAbs are Y-shaped proteins that contain two identical fragment antigen-binding (Fab) fragments and a fragment crystallizable (Fc) region (Figure 1). They are produced by plasma cells (mature, activated B cells) and are recruited by the immune system to identify and destroy foreign objects. Moreover, they have the capacity to bind any potential antigen epitope with high affinity, including tumor-associated biomarkers. Today, a variety of preclinical and clinical investigations were conducted using mAbs labeled with a-particleemitting radionuclides (Table 2). The melanoma trials (Table 2) using ²¹³Bi-cDTPA-9.2.27 show that solid tumors can be regressed by TAVAT. Moreover, these clinical results demonstrated that TAVAT for melanoma patients were locally efficacious and nontoxic up to 1.4 mCi. In the ²¹³Bi-HuM195 phase I study described above, the authors provided a proof-of-concept for the use of α -particle immunotherapy to treat myeloid leukemia. Although ²¹³Bi-HuM195 was well tolerated and 14 (78%) of 18 patients had reductions in the percentage of bone marrow blasts, myelosuppression was seen in all treated patients.[28] Similarly, myelosuppression and liver function abnormalities were observed in a phase I/II trial investigating antileukemic effects of ²¹³Bi-HuM195 after partial cytoreductive chemotherapy.[29] These toxicities could be explained by the suboptimal pharmacokinetic properties of mAbs as vehicles for TAT. The high molecular weight of mAbs (150 kDa) and the presence of an Fc-region result in a long serum halflife (several days or weeks) and in interactions with Fc-receptors in myeloid and hepatic sinusoidal cells, resulting in higher bone marrow toxicity and accumulation in the liver. Improvement in antibody engineering has led to the development of antibody fragments that are smaller and devoid of Fc, such as 25-kDa single-chain Fv (scFv), Fab (50 kDa), F(ab'), (110 kDa), diabodies (55 kDa), and minibodies (80 kDa) without compromising their affinity and specificity (Figure 1).[96]



Figure 1. Schematic representation of antibodies and their derived antigen-binding fragments. a. Conventional mAb and the derived Fab, scFv, Fv domains V_L or V_{H} , Fab'₂, minibody and diabody. b. Camelid heavy-chain-only antibody and its V_HH (also known as nanobody).

Smaller engineered mAb derivatives are more rapidly delivered to the tumor and mediate more effective tumor penetration. Because of their smaller size and lack of Fc, they are more rapidly cleared from the circulation, which is indirectly proportional to the level of kidney retention. Therefore, their administration results in fast tumor uptake with high tumor-tobackground ratios. One study reported the successful conjugation of ²¹³Bi to anti-HER2 C6.5 scFv and diabody molecules. However, a lack of tumor-specific therapeutic effect was shown, probably resulting from instability of the scFv and diabody molecules in vivo.[46] Here, it was concluded that the physical half-life of 45.6 min of ²¹³Bi was too short to allow the systemically administered diabody to specifically localize in an established solid tumor. In a subsequent study, ²¹¹At was coupled to the stable *N*-succinimidyl-*N*-(4- $[^{211}At]$ astatophenethyl) succinamate and subsequently conjugated to the C6.5 diabody (Table 2).[58] Here, the somewhat longer physical half-life of ²¹¹At matches more closely to the rapid tumor targeting and rather fast systemic clearance of the C6.5 diabody. In the ²¹¹At-MX35 F(ab')₂ phase I trial, therapeutic doses were reached for the treatment of ovarian cancer.[55] However, 50% of the initial activity concentration of this radionuclide remained in the peritoneal fluids 24 h after injection, indicating a higher toxicity risk related to this immunoconjugate.

Besides antibodies and antibody derivatives, ligands (e.g. folate), synthetic protein scaffolds (e.g. affibodies), and substrate analogs (e.g. peptides) can also be used as targeting agents in order to specifically deliver the toxic radionuclide. [97-99] Affibody molecules are small single domain proteins with a molecular weight of 6.5 kDa that are derived from one of the immunoglobulin binding domains of staphylococcal protein A.[100] Previous research demonstrated that affibody molecules can bind to their targets within minutes after administration. The binding kinetics of affibodies are similar to that of nanobodies, but faster than the larger sized mAb and its derived fragments. With regard to TAT, affibody molecules directed against the membrane protein HER2 (Z_{HER2:342} and the bivalent version [Z_{HER2:4}]₂) were radiolabeled with ²¹¹At using the precursor *N*-succimidyl-*para*(trimethylstannyl) benzoate. Based on preliminary results, the authors concluded that the labeling chemistry needs to be improved before this strategy can be translated to clinical studies.[59]

So far, significant improvements have been made in the development and application of optimized vehicles for TAT. While these preliminary results are promising, there is still considerable room for improvement, mainly in the development of new coupling chemistries and elucidation and optimization of the *in vivo* biodistribution.

4. Nanobodies: potential vehicles to specifically deliver toxic α-radiation

Recently, there has been a growing interest in the use of nanobodies as vehicles for TRNT. Nanobodies are the smallest, antigen-binding fragments from unique heavy-chain-only antibodies naturally occurring in *Camelidae* (Figure 1). [101] Several applications of nanobodies as *in vivo* diagnostic tracers have been and are currently being developed.

[102] Nanobodies have many favorable characteristics as targeted tracers, including high stability in harsh conditions, such as elevated temperatures and extreme pHs offering the potential to use a broader range of radiochemistry methods. Other favorable characteristics include high affinity and specificity for their cognate antigen and facile production (Figure 2(a,b)). As such, nanobodies have been developed as efficient radiotracers directed against a variety of membrane-bound biomarkers [103] in various animal models of cancer,[104-107] inflammation,[108] and cardiovascular diseases [102] using SPECT/PET. Because of their exceptional targeting specificity that is unaffected by labeling with various radionuclides, nanobodies have become valuable vehicles for both nuclear imaging and TRNT.[105-107] Furthermore, nanobodies possess various advantages over mAbs. First, the molecular weight of nanobodies kDa) is one-tenth of that of conventional Abs (15 (150 kDa), making it possible to recognize and bind hidden isotopes. Second, nanobodies have a low immunogenicity because of their rapid blood clearance and high sequence identity to human variable domains of the heavy chain. Furthermore, previous studies by our group demonstrated that nanobodies efficiently penetrate tumor tissues and bind tumor antigens rapidly and specifically in vivo.



Figure 2. Nanobodies possess numerous advantageous characteristics, including their high antigen specificity (a) and high tumor targeting potential (b). a. ^{99m}Tc-labeled-nanobody targeting the complement receptor of the lg superfamily, CRIg, expressed on Kupffer cells in the liver. 3D-rendered SPECT/micro-CT images of naive wild-type (A.1.) and CRIg^{-/-} mice (A.2.) 1 h after intravenous injection of ^{99m}Tc-labeled-nanobody. Representative images for 3 mice per group are shown. Figures adapted with permission from.[115] b. Dosimetry calculation of untagged ¹⁷⁷Lu-DTPA-anti-HER2 nanobody coinfused with 150 mg/kg Gelofusin, in HER2^{pos} tumor xenografted mice. Radiolabeling of nanobodies is characterized by significant retention of radioactivity at the kidneys, due to the charged-based aspecific tubular reuptake after glomerular filtration. Figure adapted with permission from.[109].

Meanwhile, there is very little nonspecific binding to other tissues, which, along with the rapid blood clearance, results in high tumor-to-background ratios as early as 1 h after injection.[109] Therefore, the nanobody technology could provide an adequate solution to the off-target toxicity problem caused by long blood circulation, as is observed during mAb-based TRNT. A first-in-human PET study with a GMP-grade HER2-targeting nanobody-based tracer for breast cancer has recently been completed at our university hospital [110] and new clinical trials with nanobodies targeting HER2 and tumor-associated macrophages are planned for 2016. The first clinical study confirmed the fast clearance of nanobodies in patients, with only 10% of the injected activity remaining in the blood at 1 h p.i. (Figure 3(a)). In addition, high tumor-to-background ratios could be observed in 17 out of 19 primary tumors, with mean standard uptake values ranging between 0.7 and 11.8 (Figure 3(b)). Furthermore, the utility of nanobodies as vehicles for TRNT has been investigated in preclinical models using the β^{-} -particle-emitting radionuclide ¹⁷⁷Lu. The most relevant in vivo study demonstrated that ¹⁷⁷Lu-labeled anti-HER2 nanobody efficiently targeted HER2^{pos} s.c. xenografts in a 5-day follow-up study, while radioactivity levels in normal organs were low (Figure 2(b)).[109] Weekly i.v. administrations of ¹⁷⁷Lu-labeled anti-HER2 nanobody in mice with small HER2^{pos} tumors completely prevented tumor growth, while tumors grew exponentially in untreated mice or in mice receiving a control, nontargeting nanobody. In addition, TRNT using a ¹⁷⁷Lu-labeled anti-5T2 multiple myeloma nanobody led to an inhibition of disease progression in treated mice compared to control animals. [111] These proof-of-concept TRNT studies show that nanobodies display a more beneficial toxicity profile than mAbs and can deliver a specific lethal radiation dose to a developing tumor. The low molecular weight of nanobodies,

below the kidney cut-off for glomerular filtration, and the subsequent charged-based nonspecific tubular reuptake result in significant accumulation and retention of radioactivity in the kidneys. To avoid potential kidney-related toxicities, strategies were tested to reduce renal retention. Both the removal of nonessential positively charged amino acids in the nanobody sequence and co-infusion with positively charged amino acids or the plasma expander Gelofusin were able to lower kidney retention significantly. [107,109] Another approach to reduce the kidney retention is to use optimized radiolabeling procedures. For instance, Zalutsky and colleagues labeled an anti-HER2 nanobody with iodine-131 (^{131}I) , using the prosthetic group N-succinimidyl-4-guanidinomethyl-3-iodobenzoate (SGMIB). SGMIB is a prosthetic group used for antibody and small-protein radioiodination and possesses improved properties as a group that stabilizes ¹³¹I and maximizes the retention in tumor cells.[112,113] Remarkably, ¹³¹I-SGMIB-anti-HER2nanobody was not retained in the kidneys, while tumor targeting was maintained. In addition, Zalutsky and coworkers recently labeled an anti-HER2 nanobody with ²¹¹At, using this similar residualizing agent, referred to as N-succinimidyl-3-[²¹¹At]astato-4-guanidinomethylbenzoate

(SAGMB).[114] Paired-label biodistribution studies directly compared the *in vivo* behavior of ²¹¹At-SAGMB-nanobody to that of its ¹³¹I analog SGMIB-nanobody in athymic mice, showing excellent preservation of HER2 binding after ²¹¹At labeling in combination with high internalization and optimal tumor uptake. Further investigation of this ²¹¹At-SAGMB-nanobody compound is warranted.

5. Conclusion

TAT is an emerging and promising treatment modality that has the ability to specifically kill isolated cancer cells or cell



Figure 3. Diagnostic tumor imaging using 68 Ga-HER2-nanobody in patients with HER2 pos -breast cancer. **a**. Time-activity curve of total blood activity, expressed in % of injected activity (%IA) (n=20). **b**. Fusion PET/CT images of the uptake of 68 Ga-HER2-nanobody in breast carcinoma lesions. (B.1.) Patient with the highest tracer uptake (SUVmean 11.8) in a primary breast carcinoma. (B.2.) Patient with moderate tracer uptake in the left breast, which is easily discernable from background (SUVmean 4.9). (B.3.) Patient with invaded lymph nodes in the mediastinum and left hilar region. Lesions are indicated by red arrows. Figures are adapted with permission from.[110].

clusters and might only cause little damage to healthy nontarget tumor cells. The combination of preclinical and clinical data affirms the potential of TAT. However, more research is needed to identify the ideal combination of targeting vehicle and α -particle radionuclide, its specific way of linking both, and all this optimized toward specific target expression, disease stage, target accessibility, and site of action.

6. Expert opinion: nanobodies coupled to α-particleemitting radionuclides in cancer therapy

There is an unmet need to treat minimal residual disease and micrometastatic spread of tumor cells, as the current cancer treatment options like chemotherapy, surgery, and external beam radiotherapy are less effective once the tumor has metastasized. Targeted a-particle therapy or TAT allows, due to the high LET of the associated radioactivity, precise delivery of a highly toxic radiation to target cells with reduced harm to normal untargeted cells in the vicinity. This strategy might be ideal for the treatment of small malignant cell populations that are located in the proximity of essential normal tissue structures and could be used in addition to other existing treatment modalities. Increased production and evaluation of α -particle emitters has improved their availability, enhancing the development for new TATs. Currently, TAT has mainly been explored using mAbs. However, the high molecular weight of mAbs and the presence of the Fc-region result in a long serum half-life and interactions with cells containing Fcreceptors. Consequently, the systemic administration of radiolabeled mAbs results in a prolonged presence of radioactivity in blood and highly perfused organs, and unwanted radiation exposure of nontargeted cells. Unsurprisingly, myelotoxicity has been shown to be a limiting factor in several preclinical and clinical studies. Moreover, the dose delivered to carcinomas is often inadequate, owing to the limited penetration of mAb-based vehicles. Hence, we claim that mAbs are not the ideal vehicles to couple with an α -particle emitter. Ab engineering is an interesting approach to overcome some of the limitations of mAbs. Nanobodies in particular have emerged as excellent Ab fragments, as they exhibit high affinity and specificity, fast diffusion and clearance kinetics in vivo, high tumor-to-normal-tissue ratios, and a high stability. Moreover, nanobodies have already proven their value in both diagnostic and therapeutic applications. We believe that nanobodies, with their improved properties compared to full-size mAbs and larger Ab-fragments, could be ideal vehicles for TAT.

A key element in the design of radiopharmaceuticals is attuning the properties of the therapeutic radionuclide with those of the tumor-targeting vehicle. The main goal here is to optimize the vehicle in such a way that it fits the characteristics of the α -particle-emitting radionuclide, resulting in optimal tumor targeting and minimal exposure of normal organs. Due to their half-life in the range of minutes to hours, ²¹³Bi and ²¹¹At could be ideal radioactive partners for fast and specific targeting nanobodies. However, both radioisotopes have both their advantages and disadvantages. Currently, the most important limitation of ²¹¹At is the limited availability of accelerators that are able to generate the 28 MeV α -particle

beam required to produce useful levels of ²¹¹At.[86] Therefore, production and supply of sufficient amounts of ²¹¹At is still challenging, although over the past few years some progress has been made in the recruitment of new cyclotrons for commercial ²¹¹At production. Today, about 30 cyclotrons in the world have the beam characteristics (28 MeV) capable for the production of ²¹¹At. Furthermore, Lindegren and colleagues developed a fully automated procedure that enables automatic, reproducible, rapid, high-yield production of clinically relevant amounts of ²¹¹At and ²¹¹At-labeled radiopharmaceuticals.[116] To date, only two clinical trials have been reported using ²¹¹At-labeled molecules (Table 2). In the first clinical trial, the median survival for patients with glioblastoma multiforme, anaplastic astrocytoma, and oligodendroglioma was 54, 52, and 116 weeks after ²¹¹At-labeled chimeric antitenascin 81C6 therapy.[53] In the second phase I study, ovarian cancer patients were injected with ²¹¹At-MX35 F(ab')₂. Intraperitoneal administration of this immunoconjugate showed that it was possible to achieve therapeutic absorbed doses (15.6 \pm 1.0 mGy/[MBq/L]) in the peritoneal peritoneum, where the microscopic tumor clusters are situated, without significant toxicity.[55]

Targeting vehicles can be astatinated via a variety of prosthetic groups.[85] However, many prosthetic groups fail to deliver relevant amounts of astatinated end product, as well as proper in vivo stability. In addition, automatable chemistries with high radiochemical yields are yet to be developed. Therefore, many ²¹¹At-labeled compounds labeled have been abandoned in the past. To this, a more in-depth understanding of the chemistry of ²¹¹At is required to provide future, useful astatinated radiopharmaceuticals. The production of ²¹³Bi is more straightforward, through the actinium-225/bismuth-213 generator system. However, the use of ²¹³Bi has been limited by the availability of ²²⁵Ac. In numerous clinical studies, ²¹³Bi ($t_{1/2}$ = 46 min) has shown to be effective to treat patients with malignant melanoma, metastatic breast cancer, prostate cancer, pancreatic cancer, and other metastatic diseases. The labeling of targeting vehicles with ²¹³Bi is generally performed using straightforward chelating agents such as DTPA and DOTA. In addition, ²¹³Bi decays via a branched pathway by α and β emissions to stable ²⁰⁹Bi, leading to low toxicity due to the minimal recoil energy the daughter experience upon α -decay. However, the short half-life of ²¹³Bi might eventually limit its clinical applicability, as relevant therapeutic doses of ²¹³Bi need to be available on a regular basis. Based on these characteristics and on the corresponding features of nanobodies, we claim that nanobodies are ideal for radiolabeling with short-lived radionuclides such as ²¹¹At and ²¹³Bi.

 α -Particle recoiling daughter isotopes pose serious problems during TAT as they can do significant harm to healthy tissue when they are not retained at the tumor site. Different approaches to limit the distribution of recoiling daughter isotopes have been found such as encapsulation in a nano-carrier and fast internalization of the α -particle inside the tumor cells. In general, monovalent nanobodies only show limited degree of internalization inside tumor cells after binding. However, it has been shown that internalization can be stimulated by the development of multivalent nanobody constructs, which

would augment the amount of α -particles trapped inside the tumor cell.[117]

In order to become valuable, some aspects concerning nanobody-based TAT need to be considered.[1] In general, nanobody targeting is characterized by only moderate absolute uptake in tumor tissue (compared to longer circulating mAbs) and fast blood clearance. To this, it will be important to assess the maximum dose that can be delivered to target tissues. The fast clearance and very specific way of targeting of nanobodies allows repeated injections. In the past, we have shown that therapeutic doses can be reached through fractiousing ¹⁷⁷Lu-labeled nanobodies. nated administration [109,111] We therefore believe that therapeutic TAT doses will be achieved by means of repeated administration. The high LET of a-particles will have their beneficial effect on tumor tissues, but can in parallel cause toxicity in tissues with elevated uptake or retention. It is known that nanobodies can interact with the negatively charged lumen of kidney tubuli during filtration from blood. It is therefore of upmost importance to assess the effect of nanobody-TAT at the level of the kidneys. Based on previous published work, there are several countermeasures that can be taken to reduce renal retention of radiolabeled nanobodies. Kidney retention can be reduced significantly by removal of the nanobodies' aminoacid tag or through co-infusion with the plasma expander Gelofusin and positively charged amino acids.[107,109] In addition, it has been shown that the linker between radioisotope and targeting vehicle can have a dramatic influence on the degree of kidney retention. Recently, anti-HER2 nanobodies were radiolabeled with ¹³¹I using the prosthetic group SGMIB. It was shown that while the retention in tumor cells was maintained, a complete absence of kidney retention was observed.[112] Interestingly, this exact prosthetic group can be used for astatination of nanobodies. The short path length of a-particles causes a heterogenous distribution in both tumor and tissues, which can lead to very localized toxicity (suborgan or subtissue level). Novel methods that allow microand small-scale dosimetry will be essential to realistically estimate dosimetry of TAT-based radiopharmaceuticals. Emerging strategies include the recent development of the a-camera that allows ex vivo imaging of a-particle deposits at a cellular level

In conclusion, the superior characteristics of α -particle emitters ²¹³Bi and ²¹¹At as toxic payload and nanobodies as targeting vehicles offer exciting possibilities in TAT. We therefore expect that the pairing of short-lived α -particle emitters and fast and specific targeting nanobodies will show their potential in the future.

Declaration of interests

Y Dekempeneer is supported by a personal grant from Kom op tegen Kanker, and the authors have received funding from the IWT and Research Foundation Flanders (FWO). T Lahoutte, M D'huyvetter and N Devoogdt are co-founders of Camel-IDS. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties.

ORCID

Serge Muyldermans () http://orcid.org/0000-0002-3678-3575 Nick Devoogdt () http://orcid.org/0000-0001-9220-4833

References

Papers of special note have been highlighted as:

- of interest
- of considerable interest
 - Restifo NP, Smyth MJ, Snyder A. Acquired resistance to immunotherapy and future challenges. Nat Rev Cancer. 2016 ;16 (2):121–126. Nature Publishing Group.
 - Middendorp M, Grünwald F. Update on recent developments in the therapy of differentiated thyroid cancer. Semin Nucl Med. 2010 ;40 (2):145–152. Elsevier Inc.
 - 3. Henriksen G, Fisher DR, Roeske JC, et al. Targeting of osseous sites with alpha-emitting 223Ra: comparison with the beta-emitter 89Sr in mice. J Nucl Med. 2003;44(2):252–259.
 - 4. Van Dodewaard-de Jong J, Oprea-Lager D, Hooft L, et al. Radiopharmaceuticals for palliation of bone pain in patients with castration-resistant prostate cancer metastatic to bone: a systematic review. Eur Urol. 2015. European Association of Urology.
 - Kwekkeboom DJ, Mueller-Brand J, Paganelli G, et al. Overview of results of peptide receptor radionuclide therapy with 3 radiolabeled somatostatin analogs. J Nucl Med. 2005;46(1):625–65.
 - Valkema R, Pauwels S, Kvols L, et al. Survival and response after peptide receptor radionuclide therapy with [90Y-DOTA0,Tyr3] octreotide in patients with advanced gastroenteropancreatic neuroendocrine tumors. Semin Nucl Med. 2006;36(2):147–156.
 - De Jong M, Valkema R, Jamar F, et al. Somatostatin receptortargeted radionuclide therapy of tumors: preclinical and clinical findings. Semin Nucl Med. 2002;32(2):133–140.
 - 8. Koppe MJ, Postema EJ, Aarts F, et al. Antibody-guided radiation therapy of cancer. Cancer Metastasis Rev. 2005;24(4):539–567.
 - Ghobrial I, Witzig T. Radioimmunotherapy: a new treatment modality for B-cell non-Hodgkin's Lymphoma. Oncology (Williston Park). 2004;18(3):623–630.
 - Chamarthy MR, Williams SC, Moadel RM. Radioimmunotherapy of non-Hodgkin's lymphoma: from the "magic bullets" to "radioactive magic bullets. Yale J Biol Med. 2011;84(4):391–407.
 - Volkert WA, Goeckleler WF, Ehrhardt GJ, et al. Therapeutic radionuclides: production and decay property considerations. J Nucl Med. 1991;32(1):174–185.
 - Pouget JP, Navarro-Teulon I, Bardiès M, et al. Clinical radioimmunotherapy-the role of radiobiology. Nat Rev Clin Oncol. 2011;8 (12):720–734.
 - Buchegger F, Perillo-Adamer F, Dupertuis YM, et al. Auger radiation targeted into DNA: a therapy perspective. Eur J Nucl Med Mol Imaging. 2006;33(11):1352–1363.
 - Mulford DA, Scheinberg DA, Jurcic JG. The promise of targeted αparticle therapy. J Nucl Med. 2005;46:199–205.
 - Allen B, Singla A, Rizvi S, et al. Analysis of patient survival in a Phase
 I trial of systemic targeted α-therapy for metastatic melanoma. Immunotherapy. 2011;3(9):1041–1050.
 - Allen BJ, Raja C, Rizvi S, et al. Tumour anti-vascular alpha therapy: a mechanism for the regression of solid tumours in metastatic cancer. Phys Med Biol. 2007;52(13):L15–9.
 - 17. Larsen RH, Murud KM, Akabani G, et al. 211At-and 1311-labeled bisphosphonates with high in vivo stability and bone accumulation. J Nucl Med. 1999;40:1197–1204.
 - Miederer M, McDevitt MR, Sgouros G, et al. Pharmacokinetics, dosimetry, and toxicity of the targetable atomic generator, 225Ac-HuM195, in nonhuman primates. J Nucl Med. 2004;45(1):129–137.
 - Jurcic JG, Rosenblat TL, McDevitt MR. Phase I trial of the targeted alpha-particle nano-generator actinium-225 (225Ac)-lintuzumab (anti-CD33; HuM195) in acute myeloid leukemia (AML). J Clin Oncol. 2011;29:Abstract 6516.
 - 20. Jurcic J, Ravandi F, Pagel JM, et al. Phase I trial of the targeted alpha-particle nano-generator actinium-225 (225Ac)-lintuzumab

(anti-CD33) in combination with low-dose cytarabine (LDAC) for older patients with untreated acute myeloid leukemia (AML). Blood. 2013;122(21):1460–1460.

- 21. Borchardt PE, Yuan RR, Miederer M, et al. Targeted actinium-225 in vivo generators for therapy of ovarian cancer. Cancer Res. 2003;63 (16):5084–5090.
- 22. Heyerdahl H, Abbas N, Brevik EM, et al. Fractionated therapy of HER2-expressing breast and ovarian cancer xenografts in mice with targeted alpha emitting 227Th-DOTA-p-benzyl-trastuzumab. PLoS One. 2012;7:8.
- 23. Heyerdahl H, Abbas N, Sponheim K, et al. Targeted alpha therapy with ovarian cancer in nude mice 227Th-trastuzumab of intraperitoneal. Curr Radiopharm. 2013;6(2):106–116.
- 24. Melhus KB, Larsen RH, Stokke T, et al. Evaluation of the binding of radiolabeled rituximab to CD20-positive lymphoma cells: an in vitro feasibility study concerning low-dose-rate radioimmunotherapy with the α -emitter 227 Th. Cancer Biother Radiopharm. 2007;22 (4):469–479.
- Dahle J, Jonasdottir TJ, Heyerdahl H, et al. Assessment of long-term radiotoxicity after treatment with the low-dose-rate alpha-particleemitting radioimmunoconjugate 227Th- rituximab. Eur J Nucl Med Mol Imaging. 2010;37(1):93–102.
- McDevitt M, Nikula T, Finn R. Bismuth labeled antibodies for therapy of leukemias, lymphomas, and carcinomas: preclinical studies. Tumor Target. 1996;2(Abstract):182.
- Schwartz J, Jaggi JS, O'Donoghue JA, et al. Renal uptake of bismuth-213 and its contribution to kidney radiation dose following administration of actinium-225-labeled antibody. Phys Med Biol. 2011;56(3):721–733.
- Jurcic JG, Larson SM, Sgouros G, et al. Targeted alpha particle immunotherapy for myeloid leukemia. Blood. 2002;100(4):1233– 1239.
- The first proof-of-concept for systemic-targeted alpha immunotherapy in humans using a monoclonal antibody as targeting vehicle.
- Rosenblat TL, McDevitt MR, Mulford DA, et al. Sequential cytarabine and α-particle immunotherapy with bismuth-213–lintuzumab (HuM195) for acute myeloid leukemia. Clin Cancer Res. 2010;16 (21):5303–5311.
- 30. Heeger S, Moldenhauer G, Egerer G, et al. Conventional and pretargeted radioimmunotherapy using bismuth-213 to target and treat non-Hodgkin lymphomas expressing CD20: a preclinical model toward optimal consolidation therapy to eradicate minimal residual disease. Blood. 2010;116(20):4231–4239.
- Roscher M, Hormann I, Leib O, et al. Targeted alpha-therapy using [Bi-213] anti-CD20 as novel treatment option for radio-and chemoresistant non-Hodgkin lymphoma cells. Oncotarget. 2013;4 (2):218–230.
- Heeger S, Moldenhauer G, Egerer G. Alpha-radioimmunotherapy of B-lineage non-Hodgkin's lymphoma using 213Bi-labelled anti-CD19-and anti-CD20-CHX-A"-DTPA conjugates. Abstr Pap Am Chem Soc. 2003;225:U261.
- Song EY, Abbas Rizvi SM, Qu CF, et al. Pharmacokinetics and toxicity of213Bi-labeled PAI2 in preclinical targeted alpha therapy for cancer. Cancer Biol Ther. 2007;6(6):898–904.
- Allen BJ, Tian Z, Rizvi SMA, et al. Preclinical studies of targeted alpha therapy for breast cancer using 213Bi-labelled-plasminogen activator inhibitor type 2. Br J Cancer. 2003;88(6):944–950.
- Qu CF, Song EY, Li Y, et al. Pre-clinical study of 213Bi labeled PAI2 for the control of micrometastatic pancreatic cancer. Clin Exp Metastasis. 2005;22(7):575–586.
- Song EY, Qu CF, Rizvi SMA, et al. Bismuth-213 radioimmunotherapy with C595 anti-MUC1 monoclonal antibody in an ovarian cancer ascites model. Cancer Biol Ther. 2008;7(1):76–80.
- Qu CF, Songl YJ, Rizvi SMA, et al. In vivo and in vitro inhibition of pancreatic cancer growth by targeted alpha therapy using 213Bi-CHX.A"-C595. Cancer Biol Ther. 2005;4(8):848–853.
- Cordier D, Forrer F, Bruchertseifer F, et al. Targeted alpha-radionuclide therapy of functionally critically located gliomas with

213Bi-DOTA-[Thi8, Met (O2) 11]-substance P: a pilot trial. Eur J Nucl Med Mol Imaging. 2010;37(7):1335–1344.

- Morgenstern A, Krolicki L, Kunikowska J, et al. Targeted alpha therapy of glioblastoma multiforme: first clinical experience with 213Bi-substance P. J Nucl Med. 2014;55(supplement 1):390–390.
- Rizvi SMA, Allen BJ, Tian Z, et al. In vitro and preclinical studies of targeted alpha therapy (TAT) for colorectal cancer. Color Dis. 2001;3 (5):345–353.
- Li Y, Wang J, Abbas Rizvi SM, et al. In vitro targeting of NG2 antigen by 213Bi-9.2.27 α-immunoconjugate induces cytotoxicity in human uveal melanoma cells. Investig Ophthalmol Vis Sci. 2005;46 (12):4365–4371.
- 42. Raja C, Graham P, Abbas Rizvi S, et al. Interim analysis of toxicity and response in phase 1 trial of systemic targeted alpha therapy for metastatic melanoma. Cancer Biol Ther. 2007;6(6):846–852.
- Allen BJ, Raja C, Rizvi S, et al. Intralesional targeted alpha therapy for metastatic melanoma. Cancer Biol Ther. 2005;4(12):1318–1324.
- 44. Fichou N. Single-dose anti-CD138 radioimmunotherapy: bismuth-213 is more efficient than lutetium-177 for treatment of multiple myeloma in a preclinical model. Front Med. 2015;2(76):1–11.
- 45. Li Y, Tian Z, Rizvi SMA, et al. In vitro and preclinical targeted alpha therapy of human prostate cancer with Bi-213 labeled J591 antibody against the prostate specific membrane antigen. Prostate Cancer Prostatic Dis. 2002;5(1):36–46.
- 46. Adams GP, Shaller CC, Chappell LL, et al. Delivery of the α -emitting radioisotope bismuth-213 to solid tumors via single-chain Fv and diabody molecules. Nucl Med Biol. 2000;27(4):339–346.
- 47. Tan Z, Chen P, Schneider N, et al. Significant systemic therapeutic effects of high-LET immunoradiation by 212Pb-trastuzumab against prostatic tumors of androgen-independent human prostate cancer in mice. Int J Oncol. 2012;40(6):1881–1888.
- 48. Yong K, Brechbiel M. Application of 212Pb for Targeted α-particle Therapy (TAT): pre-clinical and mechanistic understanding through to clinical translation. AIMS Med Sci. 2015;2(3):228–245.
- 49. Meredith R, Torgue J, Shen S, et al. Dose escalation and dosimetry of first-in-human α radioimmunotherapy with 212Pb-TCMC-trastuzumab. J Nucl Med. 2014;55(10):1636–1642.
- Meredith RF, Torgue J, Azure MT, et al. Pharmacokinetics and imaging of 212Pb-TCMC-trastuzumab after intraperitoneal administration in ovarian cancer patients. Cancer Biother Radiopharm. 2013;29(1):12–18.
- 51. Zalutsky MR, Garg PK, Friedman HS, et al. Labeling monoclonal antibodies and F (ab ') 2 fragments with the a-particle-emitting nuclide astatine-211: Preservation of immunoreactivity and in vivo localizing capacity. Proc Natl Acad Sci. 1989;86(18): 7149–7153.
- 52. Zalutsky MR, Stabin MG, Larsen RH, et al. Tissue distribution and radiation dosimetry of astatine-211-labeled chimeric 81C6, an α-particle-emitting immunoconjugate. Nucl Med Biol. 1997;24 (3):255–261.
- 53. Zalutsky MR, Reardon DA, Akabani G, et al. Clinical experience with α-particle–emitting 211At: treatment of recurrent brain tumor patients with 211At-labeled chimeric antitenascin monoclonal antibody 81C6. J Nucl Med. 2007;49(1):30–38.
- 54. Elgqvist J, Andersson H, Bäck T, et al. Fractionated radioimmunotherapy of intraperitoneally growing ovarian cancer in nude mice with 211 At-MX35 F (ab') 2: therapeutic efficacy and myelotoxicity. Nucl Med Biol. 2006;33(8):1065–1072.
- 55. Andersson H, Cederkrantz E, Bäck T, et al. Intraperitoneal α -particle radioimmunotherapy of ovarian cancer patients: pharmacokinetics and dosimetry of 211At-MX35 F (ab') 2—a phase I study. J Nucl Med. 2009;50(7):1153–1160.
- 56. Andersson H, Lindegren S, Bäck T, et al. The curative and palliative potential of the monoclonal antibody MOv18 labelled with 211At in nude mice with intraperitoneally growing ovarian cancer xenografts-a long-term study. Acta Oncol (Madr). 2000;39(6):741–745.
- Reist CJ, Foulon CF, Alston K, et al. Astatine-211 labeling of internalizing anti-EGFRvIII monoclonal antibody using N-succinimidyl 5-[211 At] astato-3-pyridinecarboxylate. Nucl Med Biol. 1999;26 (4):405–411.

- 58. Robinson MK, Shaller C, Garmestani K, et al. Effective treatment of established human breast tumor xenografts in immunodeficient mice with a single dose of the α-emitting radioisotope astatine-211 conjugated to anti-HER2/neu diabodies. Clin Cancer Res. 2008;14(3):875–882.
- 59. Steffen A-C, Almqvist Y, Chyan M-K, et al. Biodistribution of 211At labeled HER-2 binding affibody molecules in mice. Oncol Rep. 2007;17(5):1141–1147.
- Henriksen G, Breistol K, Bruland OS, et al. Significant antitumor effect from bone-seeking, alpha-particle-emitting (223)Ra demonstrated in an experimental skeletal metastases model. Cancer Res. 2002;62(11):3120–3125.
- Parker C, Nilsson S, Heinrich D, et al. Alpha emitter radium-223 and survival in metastatic prostate cancer. N Engl J Med. 2013;369 (3):213–223.
- 62. Nilsson S, Larsen RH, Fosså SD, et al. First clinical experience with alpha-emitting radium-223 in the treatment of skeletal metastases. Clin Cancer Res. 2005;11(12):4451–4459.
- Pozzi OR, Zalutsky MR. Radiopharmaceutical chemistry of targeted radiotherapeutics, part 2 : radiolytic effects of 211At alpha-particles influence N-succinimidyl 3-211At-astatobenzoate synthesis. J Nucl Med. 2005;46(8):1393–1400.
- Pozzi OR, Zalutsky MR. Radiopharmaceutical chemistry of targeted radiotherapeutics, Part 3: alpha-particle-induced radiolytic effects on the chemical behavior of 211At. J Nucl Med. 2007;48(7):1190– 1196.
- Boyd M, Sorensen A, McCluskey AG, et al. Radiation quality-dependent bystander effects elicited by targeted radionuclides. J Pharm Pharmacol. 2008;60(8):951–958.
- Mancuso M, Pasquali E, Leonardi S, et al. Oncogenic bystander radiation effects in patched heterozygous mouse cerebellum. Proc Natl Acad Sci. 2008;105(34):12445–12450.
- 67. Havaki S, Kotsinas A, Chronopoulos E, et al. The role of oxidative DNA damage in radiation induced bystander effect. Cancer Lett. 2015;356(1):43–51. Elsevier Ireland Ltd.
- De Kruijff R, Wolterbeek H, Denkova A. A critical review of alpha radionuclide therapy—how to deal with recoiling daughters? Pharmaceuticals. 2015;8(2):321–336.
- Lewis B, Chalhoub E, Chalouhy C, et al. Radium-223 in bone-metastatic prostate cancer: current data and future prospects. Oncology (Williston Park). 2015;29(7):483–488.
- Manuscript describes the successful alpha emitter ²²³Ra that has been approved by the US Food and Drug Administration (FDA).
- Miederer M, Scheinberg DA, McDevitt MR. Realizing the potential of the actinium-225 radionuclide generator in targeted alpha particle therapy applications. Adv Drug Deliv Rev. 2008;60(12):1371– 1382.
- McDevitt MR, Ma D, Lai LT, et al. Tumor therapy with targeted atomic nanogenerators. Science. 2001;294:1537–1541.
- Sgouros G, Roeske JC, Mcdevitt MR, et al. MIRD pamphlet no. 22 radiobiology and dosimetry of alpha- particle emitters for targeted radionuclide therapy. J Nucl Med. 2010;51(22):311–328.
- Huang C-Y, Oborn BM, Guatelli S, et al. Monte Carlo calculation of the maximum therapeutic gain of tumor antivascular alpha therapy. Med Phys. 2012;39(3):1282–1288.
- Huang CY, Guatelli S, Oborn BM, et al. Microdosimetry for targeted alpha therapy of cancer. Comput Math Methods Med. 2012;2012.
- Bäck T, Jacobsson L. The alpha-camera: a quantitative digital autoradiography technique using a charge-coupled device for ex vivo high-resolution bioimaging of alpha-particles. J Nucl Med. 2010;51 (10):1616–1623.
- FDA Approves Xofigo for Advanced Prostate Cancer [Internet]. 2013. Available from: http://www.fda.gov/NewsEvents.
- Bruland ÒS, Nilsson S, Fisher DR, et al. High-linear energy transfer irradiation targeted to skeletal metastases by alpha-emitter 223Ra: adjuvant or alternative to conventional modalities? Clin Cancer Res. 2006;12(21):6250–6257.

- Wadas TJ, Pandya DN, Solingapuram Sai KK, et al. Molecular targeted α-particle therapy for oncologic applications. Am J Roentgenol. 2014;203:253–260.
- 79. McDevitt MR, Sgouros G, Finn RD, et al. Radioimmunotherapy with alpha-emitting nuclides. Eur J Nucl Med. 1998;25(9):1341–1351.
- Apostolidis C, Molinet R, McGinley J, et al. Cyclotron production of Ac-225 for targeted alpha therapy. Appl Radiat Isot. 2005;62 (3):383–387.
- McDevitt MR, Ma D, Simon J, et al. Design and synthesis of 225Ac radioimmunopharmaceuticals. Appl Radiat Isot. 2002;57:841–847.
- 82. Maguire WF, McDevitt MR, Smith-Jones PM, et al. Efficient 1-step radiolabeling of monoclonal antibodies to high specific activity with 225Ac for α-particle radioimmunotherapy of cancer. J Nucl Med. 2014;55(9):1492–1498.
- Pippin CG, Gansow OA, Brechbiel MW, et al. Recovery of Bi-213 from an Ac-225 cow: application to the radiolabeling of antibodies with Bi-213. Chemists' Views of Imaging Centers. Springer US. 1995;315–322.
- McDevitt MR, Finn RD, Ma D, et al. Preparation of (alpha)-emitting (213) Bi-labeled antibody constructs for clinical use. J Nucl Med. 1999;40(10):1722.
- Buérard F, Gestin J-F, Brechbiel MW. Production of (211At)-astatinated radiopharmaceuticals and applications in targeted α-particle therapy. Cancer Biother Radiopharm. 2013;28(1):1–20.
- Zalutsky MR, Pruszynski M. Astatine-211: production and availability. Curr Radiopharm. 2011;4(3):177–185.
- Paper describes the potential advantages of ²¹¹At for targeted α-particle therapy.
- Zalutsky MR, Vaidyanathan G. Astatine-211-labeled radiotherapeutics an emerging approach to targeted alpha-particle radiotherapy. Curr Pharm Des. 2000;6(14):1433–1455.
- Zalutsky MR, Zhao X, Alston KL, et al. High-level production of αparticle–emitting 211At and preparation of 211At-labeled antibodies for clinical use. J Nucl Med. 2001;42(10):1508–1516.
- Vaidyanathan G, Larsen RH, Zalutsky MR. 5-[211At] Astato-2'-deoxyuridine, an α-particle-emitting endoradiotherapeutic agent undergoing DNA incorporation. Cancer Res. 1996;56(6):1204–1209.
- Foulon CF, Alston KL, Zalutsky MR. Astatine-211-labeled biotin conjugates resistant to biotinidase for use in pretargeted radioimmunotherapy. Nucl Med Biol. 1998;25(2):81–88.
- Bloomer WD, McLaughlin WH, Neirinckx RD, et al. Astatine-211– tellurium radiocolloid cures experimental malignant ascites. Science. 1981;212(4492):340–341.
- Link EM, Carpenter RN. 211At-methylene blue for targeted radiotherapy of human melanoma xenografts: treatment of micrometastases. Cancer Res. 1990;50(10):2963–2968.
- Vaidyanathan G, Zalutsky MR. No-carrier-added meta-[123 l] iodobenzylguanidine: synthesis and preliminary evaluation. Nucl Med Biol. 1995;22(1):61–64.
- 94. Murud KM, Larsen RH, Bruland OS, et al. Influence of pretreatment with 3-amino-1-hydroxypropylidene-1, 1-bisphosphonate (APB) on organ uptake of 211 At and 125 I-labeled amidobisphosphonates in mice. Nucl Med Biol. 1999;26(7):791–794.
- Larson SM, Carrasquillo JA, Cheung N-KV, et al. Radioimmunotherapy of human tumours. Nat Rev Cancer. 2015;15(6):347–360. Nature Publishing Group.
- 96. Holliger P, Hudson PJ. Engineered antibody fragments and the rise of single domains. Nat Biotechnol. 2005;23(9):1126–1136.
- 97. Müller C, Schibli R. Prospects in folate receptor-targeted radionuclide therapy. Front Oncol. 2013;3:249.
- Vergote I, Leamon CP. Vintafolide: a novel targeted therapy for the treatment of folate receptor expressing tumors. Ther Adv Med Oncol. 2015;7(4):206–218.
- 99. Dong C, Liu Z, Wang F. Peptide-based radiopharmaceuticals for targeted tumor therapy. Curr Med Chem. 2014;21(1):139–152.
- 100. Löfblom J, Feldwisch J, Tolmachev V, et al. Affibody molecules: engineered proteins for therapeutic, diagnostic and biotechnological applications. FEBS Lett. 2010;584(12):2670–2680. Federation of European Biochemical Societies.

- 101. Hamers-Casterman C, Atarhouch T, Muyldermans S, et al. Naturally occurring antibodies devoid of light chains. Nature. 1993;363 (6428):446–448.
- 102. De Vos J, Devoogdt N, Lahoutte T, et al. Camelid single-domain antibody-fragment engineering for (pre)clinical in vivo molecular imaging applications: adjusting the bullet to its target. Expert Opin Biol Ther. 2013;13(8):1149–1160.
- 103. D'Huyvetter M, Xavier C, Caveliers V, et al. Radiolabeled nanobodies as theranostic tools in targeted radionuclide therapy of cancer. Expert Opin Drug Deliv. 2014;131(12):1–16. Informa UK, Ltd.
- 104. Huang L, Gainkam LOT, Caveliers V, et al. SPECT imaging with 99mTc-labeled EGFR-specific nanobody for in vivo monitoring of EGFR expression. Mol Imaging Biol. 2008;10(3):167–175.
- 105. Vaneycken I, Govaert J, Vincke C, et al. In vitro analysis and in vivo tumor targeting of a humanized, grafted nanobody in mice using pinhole SPECT/micro-CT. J Nucl Med. 2010;51(7):1099–1106.
- 106. Vaneycken I, Devoogdt N, Van Gassen N, et al. Preclinical screening of anti-HER2 nanobodies for molecular imaging of breast cancer. Faseb J. 2011;25(7):2433–2446.
- 107. Tchouate Gainkam LO, Caveliers V, Devoogdt N, et al. Localization, mechanism and reduction of renal retention of technetium-99m labeled epidermal growth factor receptor-specific nanobody in mice. Contrast Media Mol Imaging. 2011;6(2):85–92.
- 108. Schoonooghe S, Laoui D, Van Ginderachter JA, et al. Novel applications of nanobodies for in vivo bio-imaging of inflamed tissues in inflammatory diseases and cancer. Immunobiology. 2012;217 (12):1266–1272. Elsevier GmbH.
- D'Huyvetter M, Vincke C, Xavier C, et al. Targeted radionuclide therapy with A 177Lu-labeled anti-HER2 nanobody. Theranostics. 2014;4(7):708–720.
 - First paper that describes the successful nanobody-based targeted radionuclide therapy of HER2^{pos} cancer.

- 110. Keyaerts M, Xavier C, Heemskerk J, et al. Phase I study of 68Ga-HER2-nanobody for PET/CT assessment of HER2 expression in breast carcinoma. J Nucl Med. 2015;57(1):27–33.
- •• The first clinical trial with ⁶⁸Ga-labeled anti-HER2 nanobody in HER2^{pos} breast carcinoma (EudraCT number 2012-001135-31).
- 111. Lemaire M, D'Huyvetter M, Lahoutte T, et al. Imaging and radioimmunotherapy of multiple myeloma with anti-idiotypic nanobodies. Leukemia. 2014;28(2):444–447.
- 112. Pruszynski M, Koumarianou E, Vaidyanathan G, et al. Improved tumor targeting of anti-HER2 nanobody through N-succinimidyl 4-guanidinomethyl-3-iodobenzoate radiolabeling. J Nucl Med. 2014;55(4):650–656.
- •• Successful radioiodination of an anti-HER2 nanobody with SGMIB as the residualizing agent.
- 113. Choi J, Vaidyanathan G, Koumarianou E, et al. N-succinimidyl guanidinomethyl iodobenzoate protein radiohalogenation agents: influence of isomeric substitution on radiolabeling and target cell residualization. Nucl Med Biol. 2014;41 (10):802–812.
- 114. Choi J, Vaidyanathan G, Koumarianou E, et al. A 211At-labeled nanobody for α-particle therapy of HER2-expressing cancers. J Nucl Med. 2015;56(Supplement 3):171.
- 115. Zheng F, Put S, Bouwens L, et al. Molecular imaging with macrophage CRIg-targeting nanobodies for early and preclinical diagnosis in a mouse model of rheumatoid arthritis. J Nucl Med. 2014;55 (5):824–829.
- Aneheim E, Albertsson P, Bäck T, et al. Automated astatination of biomolecules – a stepping stone towards multicenter clinical trials. Sci Rep. 2015;5:12025. Nature Publishing Group
- 117. Oliveira S, Schiffelers RM, Van Der Veeken J, et al. Downregulation of EGFR by a novel multivalent nanobody-liposome platform. J Control Release. 2010;145(2):165–175. Elsevier B.V.