

ORIGINAL RESEARCH

Pretreatment lymphopenia is an easily detectable predictive and prognostic marker in patients with metastatic esophagus squamous cell carcinoma receiving first-line chemotherapy

Furong Kou, Zhihao Lu, Jian Li, Xiaotian Zhang, Ming Lu, Jun Zhou, Xicheng Wang, Jifang Gong, Jing Gao, Jie Li, Yan Li & Lin Shen

Key laboratory of Carcinogenesis and Translational Research (Ministry of Education), Department of GI Oncology, Peking University School of Oncology, Beijing Cancer Hospital & Institute, Beijing 100142, China

Keywords

Chemotherapy, efficacy, esophageal squamous cell carcinoma, lymphopenia, prognosis, toxicity

Correspondence

Lin Shen, Research Building 602, 52 Fucheng Road, Haidian District, Beijing, China 100142. Tel: 86-10-88196561; Fax: 86-10-88196561; E-mail: lin100@medmail.com.cn

Funding Information

No funding information provided.

Received: 3 September 2015;
Revised: 22 November 2015;
Accepted: 16 December 2015

Cancer Medicine 2016; 5(5):778–786

doi: 10.1002/cam4.638

Furong Kou and Zhihao Lu contributed equally to this work

Abstract

To explore the influence of pretreatment lymphopenia on the toxicity and efficacy of first-line chemotherapy in patients with metastatic esophagus squamous cell carcinoma (ESCC). In total, 215 patients were included in this retrospective study. Correlations between pretreatment lymphopenia (lymphocyte count $<1 \times 10^9/L$) and the occurrence of toxicity and the efficacy of first-line palliative chemotherapy were investigated. Pretreatment lymphopenia was found in 19.1% of the patients. The overall response rate (ORR) was 35.5% (65 of 183 patients). Patients with pretreatment lymphopenia had a lower ORR to chemotherapy compared with those without lymphopenia (22.2% vs. 38.8%, respectively; $P = 0.045$). Furthermore, the patients with pretreatment lymphopenia have higher grade 3–4 hematological toxicity than that of patients without pretreatment lymphopenia (19 of 41 patients, 46.3% vs. 54 of 174 patients, 31.0%; $P = 0.048$). Pretreatment lymphopenia was not correlated with grade 3–4 nonhematological toxicity. Multivariate analysis showed that pretreatment lymphopenia is an independent prognostic factor. Patients with pretreatment lymphopenia had a significantly shorter overall survival time than those without lymphopenia (8.2 months vs. 12.7 months; $P = 0.020$). This study shows that pretreatment lymphopenia is a good prognostic factor as well as a predictive factor for tumor response and chemotherapy-related hematological toxicity in metastatic ESCC.

Introduction

Esophageal carcinoma is one of the most common cancers in the world, with an incidence that ranks eighth among all of the malignant cancers [1]. In contrast with the predominant adenocarcinoma in western countries, ~95% of esophageal carcinoma in China can be classified as esophagus squamous cell carcinoma (ESCC). Most patients were diagnosed at the advanced stage, yet, treatment is mainly palliative and based on systemic chemotherapy.

The combination of cisplatin and 5-fluorouracil was considered the standard regimen, with an efficacy of 33–35% [2, 3]. Unfortunately, new cytotoxic drugs (paclitaxel,

docetaxel, irinotecan, etc.) [4–6] and targeted drugs (cetuximab [7], gefitinib [8], etc.) did not make a breakthrough with regard to efficacy and improving survival of patients with ESCC. Moreover, chemotherapy-related toxicities, especially hematological toxicities such as leukopenia, neutropenia, and febrile neutropenia were common. The rate of grade 3–4 hematological toxicity was 14–34.1% when treated with cisplatin, 5-fluorouracil, docetaxel, or paclitaxel [2, 3, 5, 6]. In one study, febrile neutropenia was also found in 6.1% patients [5].

It is necessary to screen patients who are most likely to benefit from chemotherapy with high efficacy and low toxicity, which may lead to a good prognosis. Previously,

too much attention was focused on the tumor itself. Cancer is a systemic disease, and the immune system of the host is thought to play a central role in cancer suppression [9]. Lymphocytes are crucial components of the immune system that may affect tumor growth and the survival of patients. Recently, pretreatment lymphopenia has been shown to be a poor prognostic factor for various cancers [10–15]. Furthermore, lymphopenia has been shown to be a powerful predictor of chemotherapy-induced toxicity, as well as of the efficacy of chemotherapy in colorectal cancer, breast cancer, lung cancer, etc. [10, 16–18]. The overall response rate (ORR) in patients with pretreatment lymphopenia was significantly lower than in patients with normal lymphocyte counts [10, 16–18]. However, no studies focused on metastatic ESCC patients receiving first-line chemotherapy.

Therefore, the purpose of this retrospective study was to examine the impact of pretreatment lymphocyte counts on survival, tumor response, and treatment-related toxicity in metastatic ESCC patients receiving first-line chemotherapy.

Patients and Methods

Patients

This study is a retrospective analysis of a cohort of patients with esophageal cancer at the Peking University Cancer Hospital Gastrointestinal Medical Oncology Department between January 2005 and January 2013. Detailed clinical data for patients were recorded in a regularly updated electronic database. Eligibility criteria included the following: (1) all pathologically confirmed ESCC with metastatic diseases (the 7th edition of the American Joint Committee on Cancer Staging); (2) patients receiving first-line chemotherapy; (3) life expectancy ≥ 3 months; and (4) pretreatment and follow-up laboratory values measured at our institution are available in the electronic medical record. Patients were excluded if they fulfilled the following criteria: (1) they experienced recurrence within 6 months after adjuvant chemotherapy or chemoradiation therapy; or (2) they had pathologically confirmed esophageal adenocarcinoma, small cell carcinoma, lymphoma, or adenosquamous cell carcinoma. The study was approved by the Research Ethics Committee of Peking University Cancer Hospital.

Data collection

Demographic, treatment, and laboratory-based characteristics were obtained from the electronic medical records of each patient. Patient-specific variables included age, sex, Karnofsky Performance Status (KPS), height, weight,

and weight loss before chemotherapy. Tumor-specific variables included primary tumor location, histologic grade, and sites of metastasis. Treatment parameters consisted of regimens of first-line chemotherapy, second-line therapy, radiation, and whether or not patients received surgery. Pretreatment laboratory values, including complete blood counts (white blood cell, neutrophil, lymphocyte, platelet values), albumin, and tumor makers (CEA, CYFRA2-11 and SCC), were recorded before first-line chemotherapy was administered. The dichotomization of these variables was based on the upper (white blood cells, neutrophils, platelet, and tumor markers) and lower (albumin and lymphocytes) ranges of the normal measurements for these markers. Neutrophil–lymphocyte ratio (NLR) and platelet–lymphocyte ratio (PLR) were also calculated and defined as variables for analysis. We used the medians of NLR and PLR as cutoff points for dichotomization.

Assessment of toxicity, response and survival

Hematological toxicity and nonhematological toxicity were recorded according to the National Cancer Institute Common Toxicity Criteria Version 3.0 (NCI-CTC.V3.0) based on direct questioning, physical examination, and laboratory tests. All patients who had received at least one course of chemotherapy were evaluated for toxicity. Tumor assessment was performed every 6 weeks or earlier in cases of clinical suspicion of progression using computed tomography scanning. The objective response to treatment was classified using the Response Evaluation Criteria in Solid Tumors (RECIST 1.0). For this analysis, patients with complete or partial response were classified as responders, and those with stable or progressive disease were classified as nonresponders. ORR was the percentage of responders of the total patients. Overall survival (OS) was calculated from the first day of chemotherapy to death from any cause or last follow-up, at which time data were censored. Survival data were available for all patients.

Statistical analysis

Different potential predictive variables for OS were considered to be dichotomous. Accordingly, pretreatment lymphocyte count was considered to be $<1 \times 10^9/L$ (defined as lymphopenia) or $\geq 1 \times 10^9/L$ (Fig. 1). The chi-squared test was used to compare proportions between groups. Univariate survival analysis was performed using the Kaplan–Meier method with the log-rank test. Multivariate survival analysis was performed using a Cox regression model including those factors that were significant

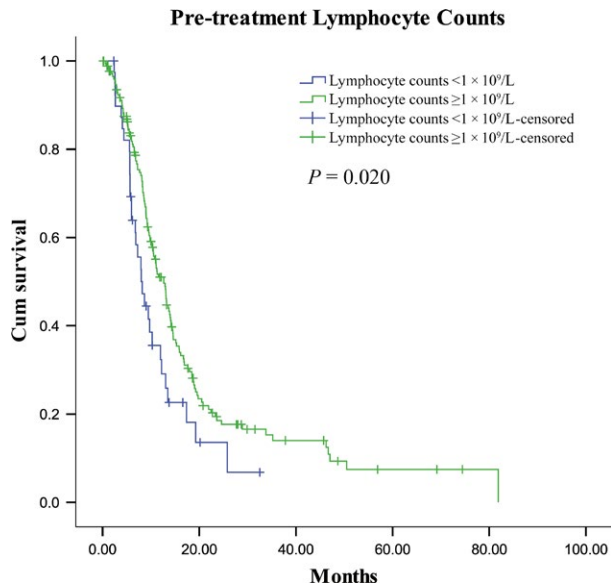


Figure 1. Kaplan–Meier curves of overall survival (OS) according to pretreatment lymphocyte counts. Patients with pretreatment lymphocyte counts $<1 \times 10^9/L$ had shorter OS (median 12.7 months, $n = 41$) than patients with lymphocyte counts $\geq 1 \times 10^9/L$ (median 8.2 months, $n = 174$; $P = 0.020$).

($P < 0.1$) on univariate analysis. SPSS (version 17.0, Inc., Chicago, IL) statistical software was used for the statistical analysis. All P -values were two sided, and $P < 0.05$ was considered statistically significant.

Results

Patient characteristics

From January 2005 to January 2013, 215 patients were eligible for this retrospective analysis. The last follow-up time was April 1, 2015. Demographic, treatment and pretreatment laboratory characteristics of these patients are summarized in Table 1.

The median age was 58 years with a range of 42–82. The majority of patients were male (85.6%). All patients presented with metastases, while 61 (28.4%) patients were afflicted with liver, 74 (34.4%) with lung, and 20 (9.3%) with bone metastasis. Other metastatic sites, such as brain, adrenal glands, kidney, and pleura, were rare. All patients received first-line chemotherapy, with 74.4% of the patients receiving a paclitaxel-based regimen. Other drugs in non-paclitaxel-based regimens included fluorouracil, irinotecan, and cisplatin etc. A small proportion of patients (24.2%) used nimotuzumab-targeted therapy during chemotherapy. Ten (4.7%) patients received radical esophagus radiation, while palliative radiation was applied in 125 (58.1%) patients. The targets of palliative radiotherapy included the primary

Table 1. Patient characteristics ($N = 215$).

| Clinicopathological characteristics | N (%) |
|--|--|
| Median age (years) [range] | 58 [42–82] |
| Gender: male vs. female | 184 (85.6) vs. 31 (14.4) |
| KPS: >80 vs. ≤ 80 | 151 (70.2) vs. 64 (29.8) |
| Weight loss: $\leq 5\%$ vs. $>5\%$ | 132 (61.4) vs. 83 (38.6) |
| Grade: well or moderately vs. poorly or undifferentiated | 112 (52.1) vs. 103 (47.9) |
| Primary tumor location: cervical or upper vs. middle or lower | 36 (16.7) vs. 179 (83.3) |
| Liver metastasis | 61 (28.4) |
| Lung metastasis | 74 (34.4) |
| Bone metastasis | 20 (9.3) |
| Distant lymph node metastasis | 171 (79.5) |
| Number of metastatic sites: <3 vs. ≥ 3 | 175 (81.4) vs. 40 (18.6) |
| First-line chemotherapy: PTX-based regimen vs. non-PTX-based regimens ¹ | 160 (74.4) vs. 55 (25.6) |
| Therapy after first-line chemotherapy | 137 (63.7) |
| Second-line chemotherapy | 47 (21.9) |
| PTX-based chemotherapy | 18 (8.3) |
| Irinotecan-based chemotherapy | 24 (11.2) |
| Other chemotherapy regimens ² | 5 (2.3) |
| Palliative radiotherapy | 120 (55.8) |
| Other treatments ² | 5 (2.3) |
| Surgery history | 76 (35.3) |
| Radiation: radical vs. palliative ² | 10 (4.7) vs. 125 (58.1) |
| White blood cell count: $>10 \times 10^9/L$ [range] | 19 (8.8) [2.91–18 $\times 10^9/L$] |
| Neutrophil count: $>8 \times 10^9/L$ [range] | 14 (6.5) [0.84–14.73 $\times 10^9/L$] |
| Lymphocyte count: $<1 \times 10^9/L$ [range] | 41 (19.1) [0.3–5.14 $\times 10^9/L$] |
| Platelet count: $>300 \times 10^9/L$ [range] | 35 (16.3) [81–615 $\times 10^9/L$] |
| NLR: median [range] | 3.0 [0.47–12.79] |
| PLR: median [range] | 153.0 [19.64–723.33] |
| Albumin: ≤ 40 g/L [range] | 42 (19.5) [18.1–51.5 g/L] |
| CEA: >5 ng/mL [range] | 45 (20.9) [0.2–994.2 ng/mL] |
| CYFRA2-11: >3.3 ng/mL [range] | 102 (47.4) [0.8–237.3 ng/mL] |
| SCC: >1.5 ng/mL [range] | 91 (42.3) [0–96 ng/mL] |

KPS, Karnofsky Performance Status; PTX, Paclitaxel; NLR, neutrophil-lymphocyte ratio; PLR, platelet-lymphocyte ratio.

¹Non-PTX-based regimens: fluorouracil-based, irinotecan-based, cisplatin-based, gemcitabine-based, capecitabine-based, S-1-based and etoposide-based.

²Other chemotherapy regimens: fluorouracil-based, cisplatin-based, gemcitabine-based, etoposide-based, oxaliplatin-based and S-1-based. Other treatments: third-line chemotherapy, transarterial chemoembolization, radiofrequency ablation. Palliative radiation fields: brain, bone, lung, liver, lymph node, and anastomotic stoma.

tumor or the sites of metastasis (brain, bone, lung, liver, lymph node, etc.). After first-line chemotherapy, 137 (63.7%) patients received subsequent therapy, including second-line chemotherapy (47 patients), palliative radiotherapy (120 patients), and other treatments (5 patients). Most patients had normal pretreatment complete blood count values, while 19.1% of the patients had pretreatment lymphopenia. The median values of NLR and PLR were 3.0 and 153.0.

Univariate and multivariate analyses of prognostic factors for OS

The median OS for the entire cohort ($N = 215$) was 11.1 months (95% CI, 8.982–13.218). Among the 26 variables in the univariate analysis, 15 variables were identified to be statistically significantly prognostic factors (gender, weight loss, liver metastasis, bone metastasis, number of metastatic sites, first-line chemotherapy, second-line chemotherapy, palliative radiation history, white blood cell count, neutrophil count, lymphocyte count, NLR, PLR, CYFRA2-11, SCC; $P < 0.05$).

The 17 factors identified in the above-described univariate analysis ($P < 0.1$) were used to construct the multivariate cox proportional hazards model for survival. The following eight factors remaining in the model were considered independent prognostic factors: gender ($P = 0.000$), weight loss ($P = 0.000$), liver metastasis

($P = 0.026$), first-line chemotherapy ($P = 0.000$), second-line chemotherapy ($P = 0.000$), surgery history ($P = 0.000$), palliative radiation history ($P = 0.000$), and pretreatment lymphopenia ($P = 0.021$). The results of univariate and multivariate analyses are shown in Table 2.

Relationship between pretreatment lymphopenia and patient characteristics

Pretreatment lymphopenia was significantly correlated with liver metastasis ($P = 0.005$), bone metastasis ($P = 0.019$), and number of metastatic sites ($P = 0.000$). As for the other clinicopathological data (age, sex, KPS, weight loss, primary tumor location, lung metastasis, and distant lymph node metastasis), no significant differences were detected between the groups. Moreover, pretreatment lymphopenia was also significantly correlated with some laboratory data,

Table 2. Univariate and multivariate analysis of the characteristics associated with the overall survival.

| Characteristics | mOS (month) | Univariate analysis | | Multivariate analysis | |
|--|-------------|---------------------|----------------|-----------------------|----------------|
| | | Chi-square | <i>P</i> value | HR (95% CI) | <i>P</i> value |
| Clinicopathological characteristics | | | | | |
| Age ($\leq 65 / > 65$) | 11.1/12.0 | 0.001 | 0.970 | | |
| Gender (Male/Female) | 10.3/28.9 | 10.651 | 0.001 | 0.298 (0.162–0.549) | 0.000 |
| KPS ($> 80 / \leq 80$) | 11.1/11.2 | 0.229 | 0.632 | | |
| Weight loss ($\leq 5\% / > 5\%$) | 13.5/9.0 | 16.641 | 0.000 | 1.991 (1.363–2.909) | 0.000 |
| Grade (well or moderately/poorly or undifferentiated) | 12.0/11.1 | 0.206 | 0.650 | | |
| Primary tumor location (Cervical or upper/Middle or lower) | 13.5/10.7 | 0.937 | 0.333 | | |
| Liver metastasis (Yes/No) | 8.7/12.9 | 14.178 | 0.000 | 1.559 (1.053–2.307) | 0.026 |
| Lung metastasis (Yes/No) | 10.6/13.0 | 0.021 | 0.886 | | |
| Bone metastasis (Yes/No) | 9.0/11.3 | 4.123 | 0.042 | | |
| Distant lymph node metastasis (Yes/No) | 10.5/13.1 | 0.086 | 0.770 | | |
| Number of metastatic sites ($< 3 / \geq 3$) | 12.0/8.7 | 6.745 | 0.009 | | |
| Treatment characteristics | | | | | |
| First-line chemotherapy (PTX-based regimen/non-PTX-based regimens) | 13.0/8.1 | 20.616 | 0.000 | 0.366 (0.244–0.549) | 0.000 |
| Second-line chemotherapy (Yes/No) | 15.8/9.5 | 5.416 | 0.020 | 0.474 (0.315–0.713) | 0.000 |
| Surgery history (Yes/No) | 13.5/10.6 | 3.185 | 0.074 | 0.486 (0.330–0.717) | 0.000 |
| Radical radiation history (Yes/No) | 12.0/11.1 | 0.322 | 0.571 | | |
| Palliative radiation history (Yes/No) | 13.9/8.6 | 13.264 | 0.000 | 0.475 (0.332–0.680) | 0.000 |
| Pretreatment laboratory characteristics | | | | | |
| White blood cell count ($\leq 10 / > 10$) $\times 10^9/L$ | 11.3/8.7 | 6.374 | 0.012 | 1.711 (0.934–3.134) | 0.082 |
| Neutrophil count ($\leq 8 / > 8$) $\times 10^9/L$ | 11.3/5.2 | 8.281 | 0.004 | | |
| Lymphocyte count ($< 1 / \geq 1$) $\times 10^9/L$ | 12.7/8.2 | 5.435 | 0.020 | 0.586 (0.373–0.922) | 0.021 |
| Platelet count ($\leq 300 / > 300$) $\times 10^9/L$ | 11.2/8.7 | 3.186 | 0.074 | | |
| NLR ($\leq 3.0 / > 3.0$) | 13.5/8.7 | 13.337 | 0.000 | | |
| PLR ($\leq 153.0 / > 153.0$) | 13.6/9.5 | 7.879 | 0.005 | | |
| Albumin ($\leq 40 / > 40$) g/L | 11.3/11.1 | 0.641 | 0.423 | | |
| CEA ($\leq 5 / > 5$) ng/mL | 11.3/10.5 | 0.400 | 0.527 | | |
| CYFRA2-11 ($\leq 3.3 / > 3.3$) ng/mL | 13.7/9.8 | 8.946 | 0.003 | | |
| SCC ($\leq 1.5 / > 1.5$) ng/mL | 13.7/9.8 | 13.705 | 0.000 | 1.422 (0.972–2.081) | 0.069 |

mOS, median overall survival; HR, hazard ratio; KPS, Karnofsky Performance Status; PTX, Paclitaxel; NLR, neutrophil-lymphocyte ratio; PLR, platelet-lymphocyte ratio. Bold values indicate a statistically difference in univariate analysis ($P < 0.1$) and statistically significant difference in multivariate analysis ($P < 0.05$).

including white blood cell count, neutrophil count, NLR, and PLR. There were no significant differences in treatment-related data between the groups except for surgery history ($P = 0.001$). The relationships between the pretreatment lymphocyte counts and patients characteristics are shown in Table 3.

Tumor response and pretreatment lymphocyte counts

Of the total 215 patients, 183 patients were evaluated for response to first-line chemotherapy. The ORR was 35.5% (65 of 183 patients). Pretreatment lymphopenia was significantly associated with lower ORR to chemotherapy. Eight of the 36 patients with pretreatment lymphopenia responded to chemotherapy versus 57 of the 147 patients without pretreatment lymphopenia (22.2% vs. 38.8%, respectively; $P = 0.045$). The details are summarized in Table 3.

Treatment-related toxicities and pretreatment lymphocyte counts

The most common hematological toxicities were leukopenia (67.4%) and neutropenia (61.9%). Similarly, leukopenia and neutropenia were also the most common grade 1–2 and grade 3–4 toxicities (45.6% and 30.2%, respectively). Moreover, the total hematological toxicity for all patients was 22.3% (48 patients) for grade 3 and 11.6% (25 patients) for grade 4. The most prevalent nonhematological toxicity was nausea (67.0%), with 62.3% of cases being grade 1–2 and 4.7% being grade 3–4.

The relationship between pretreatment lymphopenia and grade 3–4 toxicity was assessed in 215 patients (Table 4). Grade 3–4 hematological toxicity was observed in 19 of 41 patients with pretreatment lymphopenia (46.3%), and in 54 of 174 patients (31.0%) with lymphocyte count $\geq 1 \times 10^9/L$ ($P = 0.048$). Pretreatment lymphopenia was not correlated with grade 3–4 nonhematological toxicity

Table 3. Comparison of the characteristics and treatment efficacy of patients with or without pretreatment lymphopenia.

| | Patients with lymphocyte count | | Chi-square | P value |
|--|--------------------------------|----------------------------------|------------|--------------|
| | $< 1 \times 10^9/L$ (n = 41) | $\geq 1 \times 10^9/L$ (n = 174) | | |
| Clinicopathological characteristics | | | | |
| Age ($\leq 65 / > 65$) | 28/13 | 133/41 | 1.170 | 0.188 |
| Gender (Male/Female) | 38/3 | 146/28 | 2.070 | 0.133 |
| KPS ($> 80 / \leq 80$) | 29/12 | 122/52 | 0.006 | 0.551 |
| Weight loss ($\leq 5\% / > 5\%$) | 25/16 | 107/67 | 0.004 | 0.543 |
| Grade (well or moderately/poorly or undifferentiated) | 23/16 | 89/77 | 0.366 | 0.366 |
| Primary tumor location (cervical or upper/middle or lower) | 7/34 | 29/145 | 0.004 | 0.555 |
| Liver metastasis (Yes/No) | 19/22 | 42/132 | 8.049 | 0.005 |
| Lung metastasis (Yes/No) | 16/25 | 58/116 | 0.476 | 0.303 |
| Bone metastasis (Yes/No) | 8/33 | 12/162 | 6.259 | 0.019 |
| Distant lymph node metastasis (Yes/No) | 31/10 | 140/34 | 0.480 | 0.31 |
| Number of metastatic sites ($< 3 / \geq 3$) | 24/17 | 151/23 | 17.481 | 0.000 |
| Treatment characteristics | | | | |
| First-line chemotherapy (PTX-based regime/non-PTX-based regimes) | 27/14 | 133/41 | 1.952 | 0.117 |
| Second-line chemotherapy (Yes/No) | 7/35 | 41/133 | 0.931 | 0.227 |
| Surgery history (Yes/No) | 24/17 | 52/122 | 11.919 | 0.001 |
| Radical radiation history (Yes/No) | 4/37 | 6/168 | 2.977 | 0.100 |
| Palliative radiation history (Yes/No) | 23/18 | 72/102 | 2.915 | 0.063 |
| Pretreatment laboratory characteristics | | | | |
| White blood cell count ($\leq 10 / > 10$) $\times 10^9/L$ | 41/0 | 155/19 | 4.793 | 0.016 |
| Neutrophil count ($\leq 8 / > 8$) $\times 10^9/L$ | 41/0 | 160/14 | 3.444 | 0.050 |
| Platelet count ($\leq 300 / > 300$) $\times 10^9/L$ | 35/6 | 146/28 | 0.053 | 0.517 |
| NLR ($\leq 3.0 / > 3.0$) | 3/38 | 104/70 | 36.518 | 0.000 |
| PLR ($\leq 153.0 / > 153.0$) | 2/39 | 105/69 | 40.835 | 0.000 |
| Albumin ($\leq 40 / > 40$) g/L | 10/31 | 32/142 | 0.760 | 0.252 |
| CEA ($\leq 5 / > 5$) ng/mL | 29/12 | 139/33 | 2.020 | 0.115 |
| CYFRA2-11 ($\leq 3.3 / > 3.3$) ng/mL | 14/22 | 72/80 | 0.843 | 0.233 |
| SCC ($\leq 1.5 / > 1.5$) ng/mL | 24/15 | 89/76 | 0.737 | 0.249 |
| Treatment efficacy | | | | |
| CR + PR/SD + PD | 8/28 | 57/90 | 3.460 | 0.045 |
| ORR[(CR + PR)%] | 22.2% | 38.8% | | |

KPS, Karnofsky Performance Status; PTX, Paclitaxel; NLR, neutrophil-lymphocyte ratio; PLR, platelet-lymphocyte ratio; CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; ORR, overall response rate. Bold values indicate statistically significant difference ($P \leq 0.05$) in chi-square test.

Table 4. Comparison of the grade 3–4 hematological and nonhematological toxicities of patients with or without pretreatment lymphopenia ($N = 215$).

| Grade 3–4 toxicity | Patients with lymphocyte count, n (%) | | Chi-square | P value |
|------------------------|---|--------------------------------------|------------|--------------|
| | $<1 \times 10^9/L$ ($n = 41$) | $\geq 1 \times 10^9/L$ ($n = 174$) | | |
| Hematological toxicity | 19 (46.3) | 54 (31.0) | 3.467 | 0.048 |
| Fatigue | 1 (2.4) | 5 (2.9) | 0.023 | 0.687 |
| Nausea | 1 (2.4) | 9 (5.2) | 0.559 | 0.399 |
| Vomiting | 1 (2.4) | 8 (4.6) | 0.386 | 0.461 |
| Diarrhea | 0 | 2 (1.1) | 0.476 | 0.654 |
| Hair loss | 0 | 10 (5.7) | 2.471 | 0.115 |
| Muscle/joint pain | 0 | 1 (0.6) | 0.237 | 0.809 |
| Liver damage | 1 (2.4) | 6 (3.4) | 0.107 | 0.602 |

Bold values indicate statistically significant difference ($P \leq 0.05$) in chi-square test.

(including fatigue, nausea, vomiting, diarrhea, hair loss, muscle/joint pain, and liver damage).

Discussion

In this study, we established a prognostic model for metastatic ESCC patients receiving first-line chemotherapy consisting of eight factors: pretreatment lymphopenia, gender, weight loss, liver metastasis, first-line chemotherapy regimens, second-line chemotherapy, primary tumor surgery, and palliative radiation. Furthermore, pretreatment lymphopenia was shown to be not only as a prognostic factor for short-term survival but also as a predictive factor for tumor response and treatment-related hematological toxicity in these patients.

To the best of our knowledge, the host immune system plays a central role in cancer suppression [9]. Various factors are related to the body's immune status, such as lymphocytes in the peripheral blood, tumor-infiltrating lymphocytes (TILs) [19–21], and treatment-related lymphopenia [12, 22, 23]. Lymphocytes in the peripheral blood are considered to be crucial components of the immune system and to reflect immune responsiveness. Lymphopenia, which was associated with an immunosuppressed state, was common in patients with solid tumors [10–12, 24]. Previous studies have demonstrated that lymphopenia is related to the short-term survival of various cancers, such as lung cancer, glioma, colorectal cancer, pancreatic cancer, and nasopharyngeal cancer [10–14, 18]. Feng et al. [15] also found that preoperative lymphopenia is an independent poor prognostic factor in ESCC patients who had undergone esophagectomy without preoperative neoadjuvant chemotherapy and/or radiotherapy. Similarly, our study showed that patients with metastatic ESCC and pre-treatment lymphopenia had a shorter survival time than those without lymphopenia (8.2 months vs. 12.7 months; $P = 0.020$).

We also analyzed the relationship between lymphopenia and tumor response to first-line chemotherapy. The result showed that lymphopenia was significantly associated with

lower ORR to chemotherapy (22.2% vs. 38.8%; $P = 0.045$). The same result was found in several other tumors. Lissoni et al. [18] conducted a study that included 183 patients (lung cancer: 89 cases; colorectal cancer: 63 cases; breast cancer: 31 cases) and found that the ORR in lymphocytopenic patients was significantly lower than in patients with normal pretreatment lymphocyte counts (10% vs. 43%, $P < 0.001$). Ceze et al. [10] also found that the objective response rate was significantly lower in lymphocytopenic patients than in other colorectal cancer patients receiving chemotherapy (12.5% vs. 40.2%; $P = 0.004$).

There are several distinct mechanisms that chemotherapeutic agents can modify the interactions between tumor cells and host immune system [25]. Chemotherapy succeeds in triggering tumor cell death, which restore or enhance the expression of tumor antigens and increase their susceptibility to immune attack [25, 26]. In some cases, the immune system may contribute to make chemotherapy optimally efficient. Total lymphocyte counts play a key role in immune response. Lymphopenia might reflect a state of immune depression, which decreased the effect of immune attack causing the lower effect of therapy [18]. However, the underlying mechanism between lymphopenia and decreased tumor response in ESCC patients is complicated and not fully revealed. Future studies need to be done to clarified the mechanisms in patients with metastatic ESCC.

Chemotherapy-related toxicity was impacted by many factors, such as KPS, nutritional status, genetic polymorphism, and drug administration schedule. Our study showed that pretreatment lymphopenia was also related to increased risk of grade 3–4 hematological toxicity in ESCC patients. Several studies had demonstrated that in many tumors lymphopenia was associated with febrile neutropenia [17], platelet transfusion [27], and severe anemia [28]. It was also found that early lymphopenia (day 5 after chemotherapy) was also a risk factor for chemotherapy-induced febrile neutropenia [29]. Furthermore, the subset of pretreatment lymphocytes, such

as CD4+T cells, was considered to be an independent risk factor for febrile neutropenia [30].

Because pretreatment lymphocytes play a role in treatment-related toxicity, as well as ORR and OS after first-line chemotherapy, it was necessary to consider the influence of lymphocytes when selecting a treatment plan. For the patients with pretreatment lymphopenia, which indicated a low ORR and a high toxicity for chemotherapy, it was necessary to determine whether we could decrease the ration of chemotherapy and increase the ration of immunotherapy or targeted therapy.

The cause of pretreatment lymphopenia has not been fully clarified, and the explanations are likely to be multifactorial. Some studies demonstrated that tumors could directly induce T lymphocyte apoptosis using the Fas/FasL pathway [31]. Overexpressed tumor-derived antigens could cause the persistent polyclonal activation of lymphocytes, leading to their apoptosis [32]. In addition to tumor-related lymphopenia, host-related factors could also lead to the decrease in lymphocytes. Altered lymphocyte homeostasis [33], such as a progressive decline of IL-2 [34] in the blood concentrations, appears to be associated with lymphopenia. Cachexia syndrome could exhibit lymphopenia, which might be due to the effect of cytokines, such as TNF- α [35].

Previous studies [36–38] have demonstrated that surgery could induce immune suppression, by causing changes of lymphocyte numbers and subtypes. It was found that surgery and the associated neuroendocrine and paracrine responses could increase the secretion of immune suppressing hormones, which decreased numbers and activity of natural killer cells, Th1 and cytotoxic T lymphocyte cells and increased regulatory T cells, so as to reduce interleukin-12 and interferon- γ expression [37]. However, those studies only focus on the immune response a short period of time after the surgery, the long-term effects of surgery on host immune response in cancer patients are still uncertain. In our study, people who underwent primary tumor surgery had a higher proportion of lymphopenia than patients who did not undergo surgery (31.5% vs. 12.2%, $P = 0.001$). Due to the small numbers of patients with primary tumor surgery history in this retrospective study, we could not draw any definite conclusion; however, it is not possible to exclude that surgery can induce long-term immune suppression and lymphopenia in some patients. Further prospective studies in metastatic ESCC patients are required to verify the results. Our results also showed that live metastasis, bone metastasis, and inflammatory factors (white blood cells, neutrophils, NLR, and PLR) were likely to be contributing factors because they were correlated with lymphopenia. These findings also indicated that lymphopenia was related to both tumor factors and host factors.

Based on these theories, some studies put forward the ideas of reversing lymphopenia or increasing the lymphocyte counts, expecting to improve survival and tumor response. Interventions in cancer patients, such as using hematopoietic stimulating factors, Toll-like receptor (TLR) agonists, nutritional support, have showed promising results [39–41].

Our study also has several limitations. The retrospective approach may lead to inherent biases. Only those patients with collected pretreatment lymphocyte counts receiving first-line chemotherapy at our institution were eligible for inclusion, possibly causing a selection bias. In the future, we will observe both the total and subset of peripheral blood lymphocyte counts, and the TILs can be evaluated in future studies.

In conclusion, pretreatment blood lymphocyte count was an easily detectable predictive factor for stage IV ESCC patients receiving first-line chemotherapy. Pretreatment lymphopenia indicated short-term survival, as well as a low tumor response and high treatment-induced toxicity. Our results, combined with future analyses of subtype and the dynamic change in lymphocyte counts, may lead to individualized treatment and provide evidence for future immunotherapy of ESCC.

Conflict of Interest

None declared.

References

1. Jemal, A., F. Bray, M. M. Center, J. Ferlay, E. Ward, and D. Forman. 2011. Global cancer statistics. *CA Cancer J. Clin.* 61:69–90.
2. Bleiberg, H., T. Conroy, B. Paillot, A. J. Lacave, G. Blijham, J. H. Jacob, et al. 1997. Randomised phase II study of cisplatin and 5-fluorouracil (5-FU) versus cisplatin alone in advanced squamous cell oesophageal cancer. *Eur. J. Cancer* 33:1216–1220.
3. Hayashi, K., N. Ando, H. Watanabe, H. Ide, K. Nagai, N. Aoyama, et al. 2001. Phase II evaluation of protracted infusion of cisplatin and 5-fluorouracil in advanced squamous cell carcinoma of the esophagus: a Japan Esophageal Oncology Group (JEOG) Trial (JCOG9407). *Jpn. J. Clin. Oncol.* 31:419–423.
4. Nakajima, Y., T. Suzuki, S. Haruki, K. Ogiya, K. Kawada, T. Nishikage, et al. 2008. A pilot trial of docetaxel and nedaplatin in cisplatin-pretreated relapsed or refractory esophageal squamous cell cancer. *Hepatogastroenterology* 55:1631–1635.
5. Shirakawa, T., K. Kato, K. Nagashima, A. Nishikawa, R. Sawada, N. Takahashi, et al. 2014. A retrospective study of docetaxel or paclitaxel in patients with advanced or recurrent esophageal squamous cell carcinoma who previously received fluoropyrimidine- and

- platinum-based chemotherapy. *Cancer Chemother. Pharmacol.* 74:1207–1215.
6. Song, Z., and Y. Zhang. 2014. Second-line docetaxel-based chemotherapy after failure of fluorouracil-based first-line treatment for advanced esophageal squamous cell carcinoma. *OncoTargets Ther.* 7:1875–1881.
 7. Crosby, T., C. N. Hurt, S. Falk, S. Gollins, S. Mukherjee, J. Staffurth, et al. 2013. Chemoradiotherapy with or without cetuximab in patients with oesophageal cancer (SCOPE1): a multicentre, phase 2/3 randomised trial. *Lancet Oncol.* 14:627–637.
 8. Dutton, S. J., D. R. Ferry, J. M. Blazeby, H. Abbas, A. Dahle-Smith, W. Mansoor, et al. 2014. Gefitinib for oesophageal cancer progressing after chemotherapy (COG): a phase 3, multicentre, double-blind, placebo-controlled randomised trial. *Lancet Oncol.* 15:894–904.
 9. Dunn, G. P., A. T. Bruce, H. Ikeda, L. J. Old, and R. D. Schreiber. 2002. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat. Immunol.* 3:991–998.
 10. Ceze, N., G. Thibault, G. Goujon, J. Viguier, H. Watier, E. Dorval, et al. 2011. Pre-treatment lymphopenia as a prognostic biomarker in colorectal cancer patients receiving chemotherapy. *Cancer Chemother. Pharmacol.* 68:1305–1313.
 11. Grossman, S. A., X. Ye, G. Lesser, A. Sloan, H. Carraway, S. Desideri, et al. 2011. Immunosuppression in patients with high-grade gliomas treated with radiation and temozolomide. *Clin. Cancer Res.* 17:5473–5480.
 12. Balmanoukian, A., X. Ye, J. Herman, D. Laheru, and S. A. Grossman. 2012. The association between treatment-related lymphopenia and survival in newly diagnosed patients with resected adenocarcinoma of the pancreas. *Cancer Invest.* 30:571–576.
 13. He, J. R., G. P. Shen, Z. F. Ren, H. Qin, C. Cui, Y. Zhang, et al. 2012. Pretreatment levels of peripheral neutrophils and lymphocytes as independent prognostic factors in patients with nasopharyngeal carcinoma. *Head Neck* 34:1769–1776.
 14. Kobayashi, N., S. Usui, S. Kikuchi, Y. Goto, M. Sakai, M. Onizuka, et al. 2012. Preoperative lymphocyte count is an independent prognostic factor in node-negative non-small cell lung cancer. *Lung Cancer* 75:223–227.
 15. Feng, J. F., J. S. Liu, and Y. Huang. 2014. Lymphopenia predicts poor prognosis in patients with esophageal squamous cell carcinoma. *Medicine* 93:e257.
 16. Ray-Coquard, I., H. Ghesquiere, T. Bachelot, C. Borg, P. Biron, C. Sebban, et al. 2011. Identification of patients at risk for early death after conventional chemotherapy in solid tumours and lymphomas. *Br. J. Cancer* 85:816–822.
 17. Ray-Coquard, I., C. Borg, T. Bachelot, C. Sebban, I. Philip, G. Clapisson, et al. 2003. Baseline and early lymphopenia predict for the risk of febrile neutropenia after chemotherapy. *Br. J. Cancer* 88:181–186.
 18. Lissoni, P., F. Brivio, L. Fumagalli, G. Messina, V. Ghezzi, L. Frontini, et al. 2004. Efficacy of cancer chemotherapy in relation to the pretreatment number of lymphocytes in patients with metastatic solid tumors. *Int. J. Biol. Markers* 19:135–140.
 19. Rosenberg, S. A. 1996. The immunotherapy of solid cancers based on cloning the genes encoding tumor-rejection antigens. *Annu. Rev. Med.* 47:481–491.
 20. Bates, G. J., S. B. Fox, C. Han, R. D. Leek, J. F. Garcia, A. L. Harris, et al. 2006. Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. *J. Clin. Oncol.* 24:5373–5380.
 21. Schreiber, R. D., L. J. Old, and M. J. Smyth. 2011. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science* 331:1565–1570.
 22. Campian, J. L., X. Ye, M. Brock, and S. A. Grossman. 2013. Treatment-related lymphopenia in patients with stage III non-small-cell lung cancer. *Cancer Invest.* 31:183–188.
 23. Campian, J. L., G. Sarai, X. Ye, S. Marur, and S. A. Grossman. 2014. Association between severe treatment-related lymphopenia and progression-free survival in patients with newly diagnosed squamous cell head and neck cancer. *Head Neck* 36:1747–1753.
 24. De Giorgi, U., K. Rihawi, M. Aieta, G. Lo Re, T. Sava, C. Masini, et al. 2014. Lymphopenia and clinical outcome of elderly patients treated with sunitinib for metastatic renal cell cancer. *J. Geriatr. Oncol.* 5:156–163.
 25. Zitvogel, L., O. Kepp, G. Kroemer. 2011. Immune parameters affecting the efficacy of chemotherapeutic regimens. *Nat. Rev. Clin. Oncol.* 8:151–160.
 26. Godet, Y., M. Dosset, C. Borg, and O. Adotevi. 2012. Is preexisting antitumor CD4 T cell response indispensable for the chemotherapy induced immune regression of cancer? *Oncoimmunology* 1:1617–1619.
 27. Blay, J. Y., A. Le Cesne, C. Mermet, C. Maugard, A. Ravaud, C. Chevreau, et al. 1988. A risk model for thrombocytopenia requiring platelet transfusion after cytotoxic chemotherapy. *Blood* 92:405–410.
 28. Marec-Berard, P., J. Y. Blay, M. Schell, M. Buclon, C. Demaret, and I. Ray-Coquard. 2003. Risk model predictive of severe anemia requiring RBC transfusion after chemotherapy in pediatric solid tumor patients. *J. Clin. Oncol.* 21:4235–4238.
 29. Choi, C. W., H. J. Sung, K. H. Park, S. Y. Yoon, S. J. Kim, S. C. Oh, et al. 2003. Early lymphopenia as a risk factor for chemotherapy-induced febrile neutropenia. *Am. J. Hematol.* 73:263–266.
 30. Borg, C., I. Ray-Coquard, I. Philip, G. Clapisson, N. Bendriss-Vermare, C. Menetrier-Caux, et al. 2004. CD4

- lymphopenia as a risk factor for febrile neutropenia and early death after cytotoxic chemotherapy in adult patients with cancer. *Cancer* 101:2675–2680.
31. Hoffmann, T. K., G. Dworacki, T. Tsukihira, N. Meidenbauer, W. Gooding, J. T. Johnson, et al. 2002. Spontaneous apoptosis of circulating T lymphocytes in patients with head and neck cancer and its clinical importance. *Clin. Cancer Res.* 8:2553–2562.
 32. Lenardo, M., K. M. Chan, F. Hornung, H. McFarland, R. Siegel, J. Wang, et al. 1999. Mature T lymphocyte apoptosis—immune regulation in a dynamic and unpredictable antigenic environment. *Annu. Rev. Immunol.* 17:221–253.
 33. Dworacki, G., N. Meidenbauer, I. Kuss, T. K. Hoffmann, W. Gooding, M. Lotze, et al. 2001. Decreased zeta chain expression and apoptosis in CD3+ peripheral blood T lymphocytes of patients with melanoma. *Clin. Cancer Res.* 7(3 Suppl):947s–957s.
 34. Lissoni, P., S. Barni, F. Rovelli, and G. Tancini. 1991. Lower survival in metastatic cancer patients with reduced interleukin-2 blood concentrations. Preliminary report. *Oncology* 48:125–127.
 35. Aggarwal, S., S. Gollapudi, and S. Gupta. 1999. Increased TNF-alpha-induced apoptosis in lymphocytes from aged humans: changes in TNF-alpha receptor expression and activation of caspases. *J. Immunol.* 162:2154–2161.
 36. Barta, I., R. Melamed, K. Greenfeld, S. Atzil, A. Glasner, V. Domankevich, et al. 2010. Immune perturbations in patients along the perioperative period: alterations in cell surface markers and leukocyte subtypes before and after surgery. *Brain Behav. Immun.* 24:376–386.
 37. Kadosawa, T., and A. Watabe. 2015. The effects of surgery-induced immunosuppression and angiogenesis on tumour growth. *Vet. J.* 205:175–179.
 38. Greenfeld, K., R. Avraham, M. Benish, Y. Goldfarb, E. Rosenne, Y. Shapira, et al. 2007. Immune suppression while awaiting surgery and following it: dissociations between plasma cytokine levels, their induced production, and NK cell cytotoxicity. *Brain Behav. Immun.* 21:503–513.
 39. Fumagalli, L. A., J. Vinke, W. Hoff, E. Ypma, F. Brivio, and A. Nespoli. 2003. Lymphocyte counts independently predict overall survival in advanced cancer patients: a biomarker for IL-2 immunotherapy. *J. Immunother.* 26:394–402.
 40. Lissoni, P., F. Brivio, L. Fumagalli, G. Di Fede, and G. Brera. 2005. Enhancement of the efficacy of chemotherapy with oxaliplatin plus 5-fluorouracil by pretreatment with IL-2 subcutaneous immunotherapy in metastatic colorectal cancer patients with lymphocytopenia prior to therapy. *In Vivo* 19:1077–1080.
 41. Voo, K. S., L. Bover, M. L. Harline, J. Weng, N. Sugimoto, and Y. J. Liu. 2014. Targeting of TLRs inhibits CD4+ regulatory T cell function and activates lymphocytes in human peripheral blood mononuclear cells. *J. Immunol.* 193:627–634.