

Genome Sequence of *Lactobacillus sakei* subsp. *sakei* LS25, a Commercial Starter Culture Strain for Fermented Sausage

Anette McLeod,^a Dag Anders Brede,^b Ida Rud,^a Lars Axelsson^a

Nofima–Norwegian Institute of Food, Fisheries and Aquaculture Research, Ås, Norway^a; Norwegian University of Life Sciences, Department of Chemistry, Biotechnology and Food Science, Ås, Norway^b

Lactobacillus sakei is a lactic acid bacterium associated primarily with fermented meat and fish. Here, we present the draft genome sequence of *L. sakei* subsp. *sakei* strain LS25, a commercial starter culture strain for fermented sausage.

Received 31 May 2013 Accepted 7 June 2013 Published 11 July 2013

Citation McLeod A, Brede DA, Rud I, Axelsson L. 2013. Genome sequence of *Lactobacillus sakei* subsp. *sakei* LS25, a commercial starter culture strain for fermented sausage. Genome Announc. 1(4):e00475-13. doi:10.1128/genomeA.00475-13.

Copyright © 2013 McLeod et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Lars Axelsson, lars.axelsson@nofima.no.

Lactic acid bacteria (LAB) are important in various fermentation processes, for which some species are described as highly versatile while others are adapted to specific niches (1–3). *Lactobacillus sakei* is associated primarily with fermented meat and fish, and members of this species are commercially used as starter cultures (4–7). Others have potential use in food preservation (8–10). Presently, only one genome, that of strain 23K, has been completely sequenced and annotated (11). A distinction between two genetic subgroups is consistent with the species division into *L. sakei* subsp. *sakei* and *L. sakei* subsp. *carnosus* (12, 13), with the majority of the strains, including 23K, belonging to the latter (14– 16). Here we report the draft genome sequence of *L. sakei* strain LS25, the first strain of the subspecies *sakei* to be sequenced.

L. sakei strain LS25, a commercial starter culture strain, originated from a single colony from the commercial product Bitec LS-25 (Gewürzmüller, GmbH, Stuttgart, Germany) (17). Phenotypic, genotypic, and genomic diversity is present among various *L. sakei* strains, and investigations of metabolic mechanisms underlying the growth performance of strains show that LS25 differs from the majority of strains and exhibits traits well suited for starter culture strains (15, 16, 18, 19).

A genomic library was generated and pair-end sequenced (150 bp) on a MiSeq system (Illumina) by the Norwegian Sequencing Center (University of Oslo). Initial assembly of reads was performed with the CLC Genomics Workbench 6.0 software (CLC Bio), together with mapping to the reference genome of L. sakei 23K (11). A total of 7,752,672 reads were used, resulting in approximately 500-fold average genome coverage. The minimum contig size was 500 bp, and the minimum coverage was 200-fold. This draft assembly meets Human Microbiome Project highquality draft standards (http://www.hmpdacc.org/reference _genomes/reference_genomes.php) and consists of 39 contigs, one of which we identified as a circular plasmid by performing a BLAST search, complementary PCR, and Sanger sequencing. The draft genome consists of 2,015,095 bp and the plasmid 20,510 bp, with GC contents of 40.9% and 37.6%, respectively. Genome annotation was performed using the RAST annotation server (20), and the annotation was curated using a preliminary annotation

from the IGS (Institute for Genome Sciences) annotation engine and the highly homologous genome of *L. sakei* 23K (11). In addition, the annotation of new coding sequences (CDS) was manually curated by using NCBI BLAST comparisons to sequenced genes.

The draft genome has 1,972 predicted protein CDS, 49 genes coding for tRNAs, and 3 pseudo-tRNAs. The average contig coverage suggests 7 rRNA operons. Compared to the *L. sakei* 23K genome (11), 1,618 genes are orthologous (>60% identity; >70% coverage). Of these, 1,420 genes have >95% nucleotide sequence identity. More than 250 protein-encoding genes were unique to LS25, including an array of genes for carbohydrate metabolism, various transporters, and dehydrogenases/oxidoreductases. A cluster of genes related to citrate metabolism is present in LS25; however, this cluster is not orthologous to the one found in 23K.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number ASTI00000000. The version described in this paper is version ASTI01000000.

ACKNOWLEDGMENTS

This work was funded by the Foundation for Research Levy on Agricultural Products (Norway).

We thank Signe Marie Drømtorp for technical assistance and the Institute for Genome Sciences (IGS), University of Maryland, School of Medicine, for automatic genome annotation.

REFERENCES

- Claesson MJ, van Sinderen D, O'Toole PW. 2007. The genus *Lactoba-cillus*—a genomic basis for understanding its diversity. FEMS Microbiol. Lett. 269:22–28.
- 2. Makarova K, Slesarev A, Wolf Y, Sorokin A, Mirkin B, Koonin E, Pavlov A, Pavlova N, Karamychev V, Polouchine N, Shakhova V, Grigoriev I, Lou Y, Rohksar D, Lucas S, Huang K, Goodstein DM, Hawkins T, Plengvidhya V, Welker D, Hughes J, Goh Y, Benson A, Baldwin K, Lee JH, Diaz-Muniz I, Dosti B, Smeianov V, Wechter W, Barabote R, Lorca G, Altermann E, Barrangou R, Ganesan B, Xie Y, Rawsthorne H, Tamir D, Parker C, Breidt F, Broadbent J, Hutkins R, O'Sullivan D, Steele J, Unlu G, Saier M, Klaenhammer T, Richardson P, Kozyavkin S, Weimer B, Mills D. 2006. Comparative genomics of the lactic acid bacteria. Proc. Natl. Acad. Sci. U. S. A. 103:15611–15616.

- 3. Makarova KS, Koonin EV. 2007. Evolutionary genomics of lactic acid bacteria. J. Bacteriol. 189:1199–1208.
- 4. Hammes WP, Bantleon A, Min S. 1990. Lactic acid bacteria in meat fermentation. FEMS Microbiol. Rev. 87:165–174.
- Hammes WP, Hertel C. 1998. New developments in meat starter cultures. Meat Sci. 49:125–138.
- Lyhs U, Björkroth J, Korkeala H. 1999. Characterisation of lactic acid bacteria from spoiled, vacuum-packaged, cold-smoked rainbow trout using ribotyping. Int. J. Food Microbiol. 52:77–84.
- Leroi F, Joffraud JJ, Chevalier F, Cardinal M. 1998. Study of the microbial ecology of cold-smoked salmon during storage at 8 degrees C. Int. J. Food Microbiol. 39:111–121.
- Bredholt S, Nesbakken T, Holck A. 1999. Protective cultures inhibit growth of *Listeria monocytogenes* and *Escherichia coli* O157:H7 in cooked, sliced, vacuum- and gas-packaged meat. Int. J. Food Microbiol. 53:43–52.
- 9. Bredholt S, Nesbakken T, Holck A. 2001. Industrial application of an antilisterial strain of *Lactobacillus sakei* as a protective culture and its effect on the sensory acceptability of cooked, sliced, vacuum-packaged meats. Int. J. Food Microbiol. **66**:191–196.
- Castellano P, Aristoy MC, Sentandreu MA, Vignolo G, Toldra F. 2012. Lactobacillus sakei CRL1862 improves safety and protein hydrolysis in meat systems. J. Appl. Microbiol. 113:1407–1416.
- 11. Chaillou S, Champomier-Vergès M-C, Cornet M, Crutz-Le Coq A-M, Dudez A-M, Martin V BS, Darbon-Rongère E, Bossy R, Loux V, Zagorec M. 2005. The complete genome sequence of the meat-borne lactic acid bacterium *Lactobacillus sakei* 23K. Nat. Biotechnol. 23: 1527–1533.
- 12. Klein G, Dicks LMT, Pack A, Hack B, Zimmerman K, Dellaglio F, Reuter G. 1996. Emended description of *Lactobacillus sake* (Katahiri, Katahara and Fukami) and *Lactobacillus curvatus* (Abo-Elnega Kandler): numerical classification revealed by protein fingerprinting and identification based on biochemical patterns and DNA–DNA hybridizations. Int. J. Syst. Bacteriol. **46**:367–376.

- 13. Torriani S, Van Reenen GA, Klein G, Reuter G, Dellaglio F, Dicks LM. 1996. Lactobacillus curvatus subsp. curvatus subsp. nov. and Lactobacillus curvatus subsp. nov. and Lactobacillus sake subsp. sake subsp. nov. and Lactobacillus sake subsp. nov., new subspecies of Lactobacillus curvatus Abo-Elnaga and Kandler 1965 and Lactobacillus sake Katagiri, Kitahara, and Fukami 1934 (Klein et al. 1996, emended descriptions), respectively. Int. J. Syst. Bacteriol. 46:1158–1163.
- Chaillou S, Daty M, Baraige F, Dudez A-M, Anglade P, Jones R, Alpert C-A, Champomier-Vergés M-C, Zagorec M. 2009. Intraspecies genomic diversity and natural population structure of the meat-borne lactic acid bacterium *Lactobacillus sakei*. Appl. Environ. Microbiol. 75:970–980.
- McLeod A, Nyquist OL, Snipen L, Naterstad K, Axelsson L. 2008. Diversity of *Lactobacillus sakei* strains investigated by phenotypic and genotypic methods. Syst. Appl. Microbiol. 31:393–403.
- 16. Nyquist OL, McLeod A, Brede DA, Snipen L, Aakra A, Nes IF. 2011. Comparative genomics of *Lactobacillus sakei* with emphasis on strains from meat. Mol. Genet. Genomics 285:297–311.
- Hagen BF, Naes H, Holck AL. 2000. Meat starters have individual requirements for Mn²⁺. Meat Sci. 55:161–168.
- McLeod A, Snipen L, Naterstad K, Axelsson L. 2011. Global transcriptome response in *Lactobacillus sakei* during growth on ribose. BMC Microbiol. 11:145.
- McLeod A, Zagorec M, Champomier-Vergés MC, Naterstad K, Axelsson L. 2010. Primary metabolism in *Lactobacillus sakei* food isolates by proteomic analysis. BMC Microbiol. 10:120.
- 20. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75.