



# **Prevention of Colorectal Cancer by Targeting Obesity-Related Disorders and Inflammation**

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Abstract: Colorectal cancer is a major healthcare concern worldwide. Many experimental and clinical studies have been conducted to date to discover agents that help in the prevention of this disease. Chronic inflammation in colonic mucosa and obesity, and its related metabolic abnormalities, are considered to increase the risk of colorectal cancer. Therefore, treatments targeting these factors might be a promising strategy to prevent the development of colorectal cancer. Among a number of functional foods, various phytochemicals, including tea catechins, which have anti-inflammatory and anti-obesity properties, and medicinal agents that ameliorate metabolic disorders, might also be beneficial in the prevention of colorectal cancer. In this review article, we summarize the strategies for preventing colorectal cancer by targeting obesity-related disorders and inflammation through nutraceutical and pharmaceutical approaches, and discuss the mechanisms of several phytochemicals and medicinal drugs used in basic and clinical research, especially focusing on the effects of green tea catechins.

Keywords: colorectal cancer; chemoprevention; inflammation; obesity; green tea

### 1. Introduction

Colorectal cancer (CRC) is considered as a heterogeneous disease characterized by multiple genetic mutations and epigenetic alterations in genes that regulate cell growth and differentiation [1]. In most cases, CRC is non-hereditary (sporadic) because of the sequential accumulation of mutations in multiple genes. Numerous molecular genetic studies have identified several essential gene defects associated with sporadic CRC [2]. CRC is known to be a common malignant disease with a high mortality rate, and its clinical incidence has increased gradually over the past decade [3]. Therefore, more attention should be focused on the prevention and screening methods in patients with a high risk of CRC.

Chronic inflammation is a key predisposing factor to CRC development [4], which is one of the major complications in inflammatory bowel disease (IBD), including ulcerative colitis and Crohn's disease [5,6]. Obesity is considered an important health issue, and has become more prevalent in the recent years worldwide [7]. Recent epidemiological and experimental evidence has indicated that obesity and related metabolic abnormalities, especially diabetes mellitus, are associated with the development of various malignancies, including CRC [8,9]. Several pathophysiological mechanisms of interaction between obesity and CRC have been studied, including insulin resistance, adipocytokine imbalances, alterations in the insulin-like growth factor (IGF)-1/IGF-1 receptor (IGF-1R) axis, chronic inflammation, and oxidative stress [8–12]. These studies suggest that targeting the pathophysiological

interactions using nutritional and/or pharmaceutical interventions could be a promising strategy to prevent colorectal tumorigenesis.

Numerous studies have reported the beneficial effects of green tea catechins (GTCs) on improving the metabolic abnormalities such as obesity, thus preventing the development of malignancies [13–16]. Another plant-derived substance, curcumin, which is a component of turmeric, and a form of carotenoid, astaxanthin, have also been demonstrated to have preventive effects against colorectal carcinogenesis [17–21]. Branched-chain amino acid (BCAA) supplements, containing essential amino acids such as leucine, isoleucine, and valine could alleviate protein malnutrition and exert anti-cancer properties by ameliorating insulin resistance [22]. Pharmaceutical approaches using the 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitor pitavastatin, anti-hypertensive drugs, histamine receptor antagonists, and an anti-inflammatory agent pentoxifylline have been investigated and reported to attenuate chronic inflammation and reduce oxidative stress, leading to the prevention of colonic neoplastic lesion development [17,20,23–28].

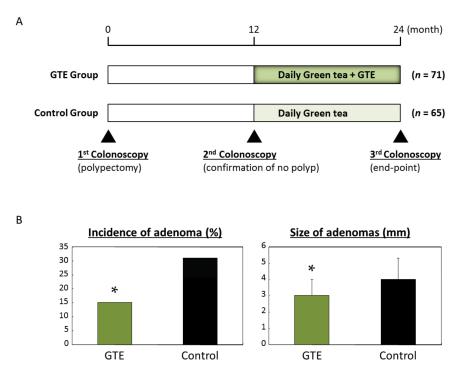
The current review summarizes the most promising strategies for the prevention of CRC by targeting obesity-related disorders and inflammation through nutritional and/or pharmaceutical approaches with several of the phytochemicals and medicinal drugs described above, because these agents have been closely studied in obesity-associated CRC models. In addition, this review article also further discusses the mechanisms of several phytochemical, especially GTCs, and medicinal agents (used in basic and clinical research) responsible for the chemoprevention of CRC.

#### 2. Preventive Effects of Green Tea and Its Constituents on CRC Development

Several population-based studies have indicated that the consumption of green tea provides protective effects against CRC development [29–31]. A prospective cohort study investigating the effects of green tea intake on CRC incidence and mortality has demonstrated that green tea consumption lowers the risk of CRC-related mortality with a moderate dose-response relationship [32]. A meta-analysis study discussing the association between green tea intake and the risk of CRC development reported several case-control studies showing an inverse correlation between green tea consumption and CRC risk, while many other studies reported no correlation [33].

Only a few interventional clinical trials have examined the chemopreventive effects of green tea on CRC development. In a pilot study, we investigated the effects of green tea extracts (GTEs) on the development of colorectal adenoma, a pre-cancerous lesion in the colorectum [34]. Patients who had undergone polypectomy for the removal of colorectal adenomas participated in the trial (Figure 1A). We have found that the administration of 1.5 g of GTEs per day for one year successfully inhibited the development of metachronous colorectal adenoma in comparison with the control group (Figure 1B). The study also demonstrated that the size of recurrent adenomas in the GTE-administered group was significantly smaller than that of the untreated control group, and no adverse events were observed in the treatment group.

The anti-cancer activity of green tea and its constituents has been demonstrated by in vitro studies and in chemically- or genetically-induced animal models of various tumors, including the lungs, skin, esophagus, stomach, liver, pancreas, bladder, small and large intestines, and prostate [35–37]. A number of studies have also investigated the effects of green tea and its constituents on CRC development. Chen et al. [38] have reported that the treatment of human colon cancer cells with (–)-epigallocatechin-3-gallate (EGCG), a tea catechin and a major biologically active component in green tea, inhibits the growth of the cancer cells. Our research group has shown that both EGCG and standardized polyphenol polyphenon E (PolyE), which contains 65% EGCG, 25% other catechins, and 0.6% caffeine, can preferentially inhibit the growth of various human colon cancer cells [39]. We have also found that the growth of human CRC xenografts was markedly reduced by the administration of EGCG [40]. Another in vivo experiment using a chemically induced rat CRC model has demonstrated that the consumption of green tea significantly suppresses the development of premalignant aberrant crypt foci (ACF) lesions in the colorectum [41].



**Figure 1.** Protocol of a pilot study to investigate chemopreventive effects of green tea extracts on metachronous adenomas in the colorectum after polypectomy. (**A**) The study included 136 participants who underwent endoscopic resection of colorectal adenomas. In 12 months, the participants received a second colonoscopy to confirm the absence of detectable adenoma. The participants were then randomized into two groups: the GTE group (n = 71) was given three green tea extracts (GTEs) tablets per day for 12 months and the control group (n = 65) received no supplementation; (**B**) After 12 months of GTE administration, the end-point colonoscopy was performed in 125 patients to check for the presence of new colonic adenomas. Administration of 1.5 g/day of GTEs for 12 months successfully inhibits the development of colorectal adenoma compared to the control group. \* p < 0.05.

Previous studies have demonstrated that receptor tyrosine kinases (RTKs) are one of the important targets of EGCG to inhibit cancer cell growth. EGCG inhibits the activation of subclass I proteins of the RTK superfamily, including EGFR, HER2, and HER3, in various cancer cells [39,42]. Activities of other RTK superfamily proteins, such as IGF-1R and vascular endothelial growth factor (VEGF) receptors, are also shown to be inhibited by EGCG. Hence, the RTK-associated cell signaling, such as the Ras/MAPK and PI3K/Akt pathways, is thought to be down-regulated in cancer cells by EGCG, leading to the modulation of the target gene expression, which is associated with the induction of apoptosis and cell cycle arrest. The molecular mechanisms which explain how EGCG affects RTK signaling have been studied in detail by Adachi et al. [43–45]. The studies indicate a target of EGCG for anti-cancer mechanisms associated with RTKs, particularly detergent-insoluble ordered plasma membrane domains "lipid rafts", which are important as signal processing hubs of RTKs. EGCG alters the lipid organization on the plasma membrane and induces the EGFR internalization of endosomes, which prevents ligands from binding to receptors. The degradation of EGFR due to internalization appeared to be induced by phosphorylation of the receptor, which is associated with the activation of p38 MAPK by EGCG. This suggested mechanism may be able to explain the ubiquitous effects of EGCG on various types of RTKs, because most RTKs function on lipid rafts. Among RTKs, IGF-1R is thought to be one of the most critical targets for the inhibition of obesity-related carcinogenesis by tea catechins, although the direct alteration of catechins on IGF-1R needs to be clarified. For more details on the effects of EGCG on RTKs and other anti-neoplastic efficacy, please refer to the review articles by Shimizu et al. [13,14] and to Figure 2, which summarizes the properties.

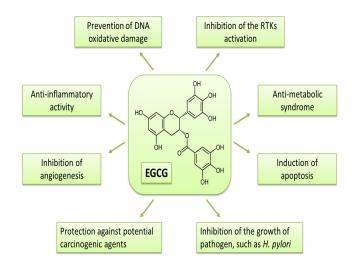


Figure 2. Proposed mechanisms of action of EGCG against malignancy.

Chronic inflammation plays a vital role in carcinogenesis, including CRC [4], which is known as one of the most serious complications of IBD [5,6]. Persistent inflammation, characterized by the production of pro-inflammatory cytokines, causes oxidative damage to DNA, mutations in oncogenes and tumor suppressor genes, including adenomatous polyposis coli (APC), p53, and K-ras, and genomic instability, leading to colitis-associated tumor development. While it is considered that inflammation does not initiate sporadic CRC, chronic inflammation is also known to facilitate tumor promotion, progression, and metastasis in the pathogenesis of colitis-associated and sporadic CRC [46]. Tanaka et al. [47] have developed an experimental mouse model of inflammation-related colon carcinogenesis induced by the administration of azoxymethane (AOM) and dextran sodium sulfate, which mimics the chronic intestinal inflammation that occurs in IBD. Employing this rodent model, we demonstrated the suppressive effects of EGCG and PolyE on inflammation-related colon carcinogenesis [48]. In this study, EGCG or Poly E significantly suppressed the multiplicity and volume of colonic neoplasms. In addition, treatment with EGCG or Poly E decreased the protein and mRNA expression levels of cyclooxygenase (COX)-2 and the mRNA expression of inflammatory cytokines, including tumor necrosis factor (TNF)- $\alpha$ , interferon- $\gamma$ , interleukin (IL)-6, IL-12, and IL-18 in the colonic mucosa. Previous studies have indicated that EGCG or green tea extract reduces the expression of TNF-a and IL-6 via attenuating NF- $\kappa$ B activity [49]. These results suggest that tea catechins can ameliorate colonic inflammation and have beneficial effects for inhibiting the development of cancer in the inflamed colon.

Recent epidemiological and experimental evidence has indicated that obesity is related to the incidence of CRC [8–10,50]. Insulin resistance and hyperinsulinemia, metabolic disorders associated with obesity, are considered important risk factors for CRC development [51]. It is reported that insulin and its regulated signal transduction network play important roles in carcinogenesis [52–54]. Many studies have shown that the IGF-1/IGF-1R axis plays a key role in the carcinogenesis of various cancers, including CRC [52–54]. In addition, insulin resistance and an increased fat mass induce oxidative stress in tissues and increase the expression of various pro-inflammatory cytokines, including TNF- $\alpha$ and IL-6, which further lead to the growth and progression of malignancies [55–57]. Oxidative stress induces DNA damage and activates the PI3K/Akt signaling pathway, both of which are thought to promote cancer development [58,59]. Therefore, insulin resistance, inflammation, and oxidative stress can be considered as important factors in the development of obesity-related CRC [60,61]. This imbalance is usually caused by enhanced fat storage, increased levels of leptin, and decreased levels of adiponectin in the serum [60,61]. Leptin induces the production of TNF- $\alpha$  and IL-6 [62,63], and thus stimulates CRC cell growth [64]. Moreover, an epidemiological study has reported a positive correlation between the circulating leptin levels and CRC development [65]. These findings suggest that obesity-associated abnormalities cooperatively increase the risk of CRC in obese individuals.

A genetically-modified C57BLKS/J-+Lepr<sup>db</sup>/+Lepr<sup>db</sup> (db/db) mouse exhibiting the characteristics of obesity and type 2 diabetes was recognized as a useful model for investigating various types metabolic disorders [66]. Hirose et al. [67] have shown that db/db mice have hyperlipidemia, hyperinsulinemia, and hyperleptinemia, and are susceptible to the colonic carcinogen AOM. We used a db/db mouse and investigated the effects of EGCG on AOM-induced colon carcinogenesis [68], and observed that EGCG markedly decreases the total number of ACF and  $\beta$ -catenin accumulated crypts (BCACs), both of which are premalignant lesions in the colorectum. Additionally, we found decreased IGF-1 and restored IGF binding protein-3 (IGFBP-3) levels in serum and down-regulated levels of COX-2, cyclin D1, and the activated form of IGF-1R in colonic mucosa upon EGCG administration. With regard to the IGF/IGF-1R axis, treatment with EGCG showed decreased levels of IGF-1 and reduced IGF-1R activation, whereas the levels of IGFBP-3 were found to be increased in colon cancer cells [69].

#### 3. Prevention of CRC through a Nutraceutical Approach

The colorectal mucosa of a db/db mouse expresses higher levels of the activated form of IGF-1R,  $\beta$ -catenin, and COX-2 than the control [67]. In accordance with the study demonstrating the effects of EGCG on AOM-induced colon premalignant lesions in db/db mice [68], dietary supplementation with other types of phytochemicals was also found to suppress the development of pre-cancerous lesions in the db/db mice [70,71]. In addition, we have used this rodent model to investigate the chemopreventive effects of curcumin, a yellow pigment in the rhizome of the spice turmeric with known anti-inflammatory properties [72,73], on obesity-related carcinogenesis. Kubota et al. [18] have demonstrated that the administration of curcumin successfully prevents the development of colonic premalignant lesions in AOM-injected db/db mice by inhibiting the NF-κB activity and down-regulating the expression of TNF- $\alpha$ , IL-6, and COX-2, further ameliorating the adipokine imbalance. Moreover, a type of carotenoid, astaxanthin, inhibited the development of colonic premalignant lesions in the same carcinogenesis model by reducing leptin levels, inhibiting NF-κB activation, and attenuating chronic inflammation and oxidative stress in the colonic mucosa [17]. Furthermore, supplementation with amino acid-preparation BCAA caused a significant decrease in the number of ACF and BCAC in the same colon tumorigenesis model [22]. The test group administered with BCAA demonstrated reduced levels of COX-2, cyclin D1, Akt, and the activated form of IGF-1R in mucosa and decreased serum levels of insulin, IGF-1, IGF-2, triglycerides, total cholesterol, and leptin [22].

These observations suggest that supplementation with certain kinds of phytochemicals and carotenoids or BCAA effectively suppresses the development of premalignant lesions of CRC by attenuating chronic inflammation, down-regulating the IGF/IGF-1R axis, improving dyslipidemia, ameliorating hyperleptinemia, and/or inhibiting the expression of COX-2, which appears to be a promising target for the prevention of CRC [74,75].

#### 4. Prevention of CRC through a Pharmaceutical Approach

There are several reports of clinical trials examining the effects of non-steroidal anti-inflammatory drugs such as celecoxib, aspirin, and metformin on the development of CRC or its precursor lesion adenomatous polyp in patients, where these agents appear to be promising [76–80]. The recent randomized and placebo-controlled clinical trials are summarized in Table 1. Recently, we reported that pentoxifylline, which is a methylxanthine derivative and known to possess anti-inflammatory effects, attenuated chronic inflammation and oxidative stress, leading to the prevention of colonic tumorigenesis in an obesity-related colon cancer model [23]. In addition, our research group also demonstrated that histamine and histamine receptors appeared to be critical molecules during inflammation and carcinogenesis in the colorectum, and that several histamine receptor antagonists might be potential chemopreventive agents for inflammation-related CRC development [26].

Reference	Agent	Target Lesion	No. of Subjects	<b>Observation Period</b>	Preventive Effects
2006 Bertagnolli [71]	Celecoxib (200 or 400 mg twice a day)	Sporadic colorectal adenomas	2035 subjects; placebo (679) or 200 mg (685) or 400 mg (671) of celecoxib group	Either one and three years	The estimated cumulative incidence of adenomas by year 3 was lower in those receiving 200 mg (risk ratio 0.67 [95% CI: 0.59–0.77]) and 400 mg celecoxib (risk ratio 0.55 [95% CI: 0.48–0.64]).
2006 Arber [72]	Celecoxib (400 mg/day)	Sporadic colorectal adenomatous polyps	1561 subjects (628 in the placebo and 933 in the celecoxib group)	Either one and three years	The cumulative rate of adenomas detected through year 3 was lower in the celecoxib group; relative risk 0.64 (95% CI: 0.56–0.75).
2013 Ishikawa [73]	Aspirin (100 mg/day)	Polyps in patients with familial adenomatous polyposis (FAP)	34 subjects with FAP (17 each in the aspirin and placebo groups)	Six-ten months	The increase in mean diameter of polyps tended to be greater in the placebo group compared to the aspirin group.
2014 Ishikawa [74]	Aspirin (100 mg/day)	Colorectal adenomas and adenocarcinomas	311 subjects (159 in the placebo and 152 in the aspirin group)	Two years	The subjects treated with aspirin displayed reduced colorectal tumourigenesis; adjusted OR 0.60 (95% CI: 0.36–0.98).
2016 Higurashi [75]	Metformin (250 mg/day)	Sporadic colorectal polyps	151 subjects (72 in the placebo and 79 in the metformin group)	One year	The prevalence of total polyps and adenomas in the metformin group was significantly lower; (total polyps) risk ratio 0.67 (95% CI: 0.47–0.97), (adenomas) risk ratio 0.60 (95% CI: 0.39–0.92).

Table 1. The recent randomized and placebo-controlled clinical trials using medicinal agents for the prevention of CRC.

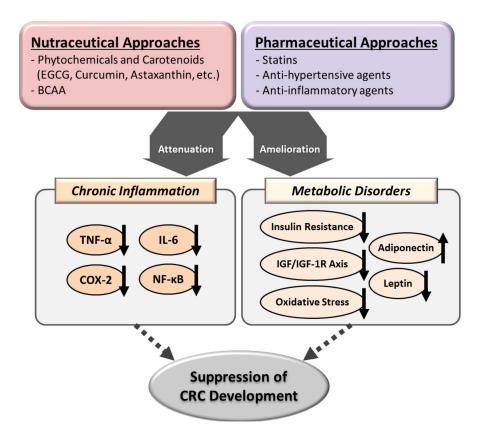
Several studies have indicated the anti-cancer properties of drugs related to metabolic disorders. Statins and HMG-CoA reductase inhibitors are widely recognized as effective agents against dyslipidemia. In addition, statins have been shown to possess anti-cancer properties [81]. Statins induce apoptosis in CRC cells, attenuate colonic inflammation, and suppress inflammation-related colorectal carcinogenesis in mice [28,82]. Several epidemiological studies have also demonstrated the chemopreventive effects of statins on various malignant diseases, including CRC [81,83]. In our previous study, we demonstrated the cancer preventive effects of a lipophilic statin, pitavastatin, on AOM-induced colorectal carcinogenesis in a db/db mouse model [27]. We found that pitavastatin administration significantly reduced the number of pre-neoplastic BCAC lesions, which may have been caused by the inhibition of the proliferation and decrease in the expression levels of COX-2 and pro-inflammatory cytokines, such as TNF- $\alpha$  and IL-6, in the colonic mucosa. Pitavastatin also elevated the serum levels of adiponectin, while reducing the serum levels of leptin, TNF- $\alpha$ , and IL-6 [27].

Hypertension and dyslipidemia are thought to be involved in obesity-related diseases [9,84]. The activation of the renin-angiotensin system (RAS) has been shown to contribute to high blood pressure, obesity, and metabolic syndrome [85]. RAS has been demonstrated to be frequently up-regulated in malignancies attributed to systemic oxidative stress and hypoxia, which are thought to trigger a state of chronic inflammation [86]. We investigated the effects of anti-hypertensive agents on the prevention of colorectal premalignant lesions in an obesity-related CRC model [25]. The employed agents were an angiotensin-converting enzyme inhibitor, captopril, and an angiotensin-II type 1 receptor blocker, telmisartan, both of which have the ability to inhibit the RAS, and are widely used in clinical practice. The development of colorectal lesions, ACF and BCAC, was significantly inhibited by the treatment with either captopril or telmisartan. These agents markedly decreased the expression levels of TNF- $\alpha$  in the colonic mucosa, and also reduced oxidative stress in the body [25]. Captopril was also reported to prevent the development of ACF by a similar mechanism in diabetic and hypertensive rats [24].

The findings discussed above suggest that both lipid-lowering and anti-hypertensive agents can suppress obesity-associated colorectal carcinogenesis by improving hyperleptinemia and dyslipidemia, and by attenuating chronic inflammation in the colorectum. Therefore, the pharmaceutical approach appears to be one of the potential strategies for the prevention of obesity-related CRC because these drugs are in clinical use and have known pharmacological effects against the obesity-related metabolic disorders, in addition to their cancer chemopreventive effects.

#### 5. Concluding Remarks

In this review article, we have discussed the use of nutraceutical and pharmaceutical approaches as promising strategies to prevent CRC development by targeting chronic inflammation and ameliorating metabolic disorders (Figure 3). Moreover, GTCs are easily available and are considered safe based on the long history of their global use. Several interventional studies on humans have also demonstrated that the consumption of GTCs, even in relatively high doses, has no serious adverse reactions [34,87,88], while clinical trials have reported that drugs such as celecoxib can increase the risk of cardiovascular events [76]. In addition, BCAA, statins, and anti-hypertensive drugs are widely used and have beneficial effects on various metabolic disorders. Hence, active intervention using these agents may be a promising strategy for the chemoprevention of CRC.



**Figure 3.** Proposed mechanisms of action of several nutraceuticals and pharmaceuticals in the suppression of colorectal carcinogenesis. The upward and downward arrows indicate up-regulation and down-regulation, respectively.

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## Abbreviations

aberrant crypt foci			
azoxymethane			
branched-chain amino acid			
β-catenin accumulated crypt			
confidence interval			
colorectal cancer			
(–)-epigallocatechin-3-gallate			
familial adenomatous polyposis			
green tea catechin			
green tea extract			
3-hydroxy-3-methylglutaryl coenzyme A			
inflammatory bowel disease			
insulin like growth factor			
odds ratio			
Polyphenon E			
renin-angiotensin system			
receptor tyrosine kinae			
vascular endothelial growth factor			

## References

- 1. Jemal, A.; Siegel, R.; Ward, E.; Hao, Y.; Xu, J.; Thun, M.J. Cancer statistics, 2009. *CA Cancer J. Clin.* 2009, *59*, 225–249. [CrossRef] [PubMed]
- 2. Fearon, E.R. Molecular genetics of colorectal cancer. Annu. Rev. Pathol. 2011, 6, 479–507. [CrossRef] [PubMed]
- 3. Cui, C.; Feng, H.; Shi, X.; Wang, Y.; Feng, Z.; Liu, J.; Han, Z.; Fu, J.; Fu, Z.; Tong, H. Artesunate down-regulates immunosuppression from colorectal cancer Colon26 and RKO cells in vitro by decreasing transforming growth factor β and interleukin-10. *Int. Immunopharmacol.* **2015**, *27*, 110–121. [CrossRef] [PubMed]
- 4. Itzkowitz, S.H.; Yio, X. Inflammation and cancer IV. Colorectal cancer in inflammatory bowel disease: The role of inflammation. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2004**, *287*, G7–G17. [CrossRef] [PubMed]
- 5. Choi, P.M.; Zelig, M.P. Similarity of colorectal cancer in Crohn's disease and ulcerative colitis: Implications for carcinogenesis and prevention. *Gut* **1994**, *35*, 950–954. [CrossRef] [PubMed]
- Rutter, M.; Saunders, B.; Wilkinson, K.; Rumbles, S.; Schofield, G.; Kamm, M.; Williams, C.; Price, A.; Talbot, I.; Forbes, A. Severity of inflammation is a risk factor for colorectal neoplasia in ulcerative colitis. *Gastroenterology* 2004, 126, 451–459. [CrossRef] [PubMed]
- 7. Khandekar, M.J.; Cohen, P.; Spiegelman, B.M. Molecular mechanisms of cancer development in obesity. *Nat. Rev. Cancer* **2011**, *11*, 886–895. [CrossRef] [PubMed]
- 8. Aleksandrova, K.; Nimptsch, K.; Pischon, T. Obesity and colorectal cancer. *Front. Biosci.* **2013**, *5*, 61–77. [CrossRef]
- 9. Calle, E.E.; Rodriguez, C.; Walker-Thurmond, K.; Thun, M.J. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. *N. Engl. J. Med.* **2003**, *348*, 1625–1638. [CrossRef] [PubMed]
- 10. Giovannucci, E.; Michaud, D. The role of obesity and related metabolic disturbances in cancers of the colon, prostate, and pancreas. *Gastroenterology* **2007**, *132*, 2208–2225. [CrossRef] [PubMed]
- 11. Ramos-Nino, M.E. The role of chronic inflammation in obesity-associated cancers. *ISRN Oncol.* **2013**, 2013, 697521. [CrossRef] [PubMed]
- Shirakami, Y.; Shimizu, M.; Kubota, M.; Araki, H.; Tanaka, T.; Moriwaki, H.; Seishima, M. Chemoprevention of colorectal cancer by targeting obesity-related metabolic abnormalities. *World J. Gastroenterol.* 2014, 20, 8939–8946. [PubMed]
- 13. Shimizu, M.; Adachi, S.; Masuda, M.; Kozawa, O.; Moriwaki, H. Cancer chemoprevention with green tea catechins by targeting receptor tyrosine kinases. *Mol. Nutr. Food Res.* **2011**, *55*, 832–843. [CrossRef] [PubMed]
- 14. Shimizu, M.; Shirakami, Y.; Moriwaki, H. Targeting receptor tyrosine kinases for chemoprevention by green tea catechin, EGCG. *Int. J. Mol. Sci.* 2008, *9*, 1034–1049. [CrossRef] [PubMed]
- Shimizu, M.; Weinstein, I.B. Modulation of signal transduction by tea catechins and related phytochemicals. *Mut. Res.* 2005, 591, 147–160. [CrossRef] [PubMed]
- 16. Yang, C.S.; Wang, X.; Lu, G.; Picinich, S.C. Cancer prevention by tea: Animal studies, molecular mechanisms and human relevance. *Nat. Rev. Cancer* **2009**, *9*, 429–439. [CrossRef] [PubMed]
- 17. Kochi, T.; Shimizu, M.; Sumi, T.; Kubota, M.; Shirakami, Y.; Tanaka, T.; Moriwaki, H. Inhibitory effects of astaxanthin on azoxymethane-induced colonic preneoplastic lesions in C57/BL/KsJ-db/db mice. *BMC Gastroenterol.* **2014**, *14*, 212. [CrossRef] [PubMed]
- Kubota, M.; Shimizu, M.; Sakai, H.; Yasuda, Y.; Terakura, D.; Baba, A.; Ohno, T.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Preventive effects of curcumin on the development of azoxymethane-induced colonic preneoplastic lesions in male C57BL/KsJ-db/db obese mice. *Nutr. Cancer* 2012, *64*, 72–79. [CrossRef] [PubMed]
- 19. Murakami, A.; Furukawa, I.; Miyamoto, S.; Tanaka, T.; Ohigashi, H. Curcumin combined with turmerones, essential oil components of turmeric, abolishes inflammation-associated mouse colon carcinogenesis. *Biofactors* **2013**, *39*, 221–232. [CrossRef] [PubMed]
- 20. Yasui, Y.; Hosokawa, M.; Mikami, N.; Miyashita, K.; Tanaka, T. Dietary astaxanthin inhibits colitis and colitis-associated colon carcinogenesis in mice via modulation of the inflammatory cytokines. *Chem. Biol. Interact.* **2011**, *193*, 79–87. [CrossRef] [PubMed]
- 21. Tanaka, T.; Shnimizu, M.; Moriwaki, H. Cancer chemoprevention by carotenoids. *Molecules* **2012**, *17*, 3202–3242. [CrossRef] [PubMed]

- 22. Shimizu, M.; Shirakami, Y.; Iwasa, J.; Shiraki, M.; Yasuda, Y.; Hata, K.; Hirose, Y.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Supplementation with branched-chain amino acids inhibits azoxymethane-induced colonic preneoplastic lesions in male C57BL/KsJ-db/db mice. *Clin. Cancer Res.* **2009**, *15*, 3068–3075. [CrossRef] [PubMed]
- 23. Fukuta, K.; Shirakami, Y.; Maruta, A.; Obara, K.; Iritani, S.; Nakamura, N.; Kochi, T.; Kubota, M.; Sakai, H.; Tanaka, T.; et al. Preventive Effects of Pentoxifylline on the Development of Colonic Premalignant Lesions in Obese and Diabetic Mice. *Int. J. Mol. Sci.* **2017**, *18*, 413. [CrossRef] [PubMed]
- 24. Kochi, T.; Shimizu, M.; Ohno, T.; Baba, A.; Sumi, T.; Kubota, M.; Shirakami, Y.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Preventive effects of the angiotensin-converting enzyme inhibitor, captopril, on the development of azoxymethane-induced colonic preneoplastic lesions in diabetic and hypertensive rats. *Oncol. Lett.* **2014**, *8*, 223–229. [CrossRef] [PubMed]
- 25. Kubota, M.; Shimizu, M.; Sakai, H.; Yasuda, Y.; Ohno, T.; Kochi, T.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Renin-angiotensin system inhibitors suppress azoxymethane-induced colonic preneoplastic lesions in C57BL/KsJ-db/db obese mice. *Biochem. Biophys. Res. Commun.* **2011**, *410*, 108–113. [CrossRef] [PubMed]
- Tanaka, T.; Kochi, T.; Shirakami, Y.; Mori, T.; Kurata, A.; Watanabe, N.; Moriwaki, H.; Shimizu, M. Cimetidine and Clobenpropit Attenuate Inflammation-Associated Colorectal Carcinogenesis in Male ICR Mice. *Cancers* 2016, *8*, 25. [CrossRef] [PubMed]
- 27. Yasuda, Y.; Shimizu, M.; Shirakami, Y.; Sakai, H.; Kubota, M.; Hata, K.; Hirose, Y.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Pitavastatin inhibits azoxymethane-induced colonic preneoplastic lesions in C57BL/KsJ-db/db obese mice. *Cancer Sci.* **2010**, *101*, 1701–1707. [CrossRef] [PubMed]
- Yasui, Y.; Suzuki, R.; Miyamoto, S.; Tsukamoto, T.; Sugie, S.; Kohno, H.; Tanaka, T. A lipophilic statin, pitavastatin, suppresses inflammation-associated mouse colon carcinogenesis. *In. J. Cancer* 2007, 121, 2331–2339. [CrossRef] [PubMed]
- 29. Arab, L.; Il'yasova, D. The epidemiology of tea consumption and colorectal cancer incidence. *J. Nutr.* **2003**, *133*, 3310S–3318S. [PubMed]
- 30. Marques-Vidal, P.; Ravasco, P.; Ermelinda Camilo, M. Foodstuffs and colorectal cancer risk: A review. *Clin. Nutr.* **2006**, *25*, 14–36. [CrossRef] [PubMed]
- 31. Tavani, A.; La Vecchia, C. Coffee, decaffeinated coffee, tea and cancer of the colon and rectum: A review of epidemiological studies, 1990–2003. *Cancer Causes Control* **2004**, *15*, 743–757. [CrossRef] [PubMed]
- 32. Suzuki, E.; Yorifuji, T.; Takao, S.; Komatsu, H.; Sugiyama, M.; Ohta, T.; Ishikawa-Takata, K.; Doi, H. Green tea consumption and mortality among Japanese elderly people: The prospective Shizuoka elderly cohort. *Ann. Epidemiol.* **2009**, *19*, 732–739. [CrossRef] [PubMed]
- 33. Sun, C.L.; Yuan, J.M.; Koh, W.P.; Yu, M.C. Green tea, black tea and colorectal cancer risk: A meta-analysis of epidemiologic studies. *Carcinogenesis* **2006**, *27*, 1301–1309. [CrossRef] [PubMed]
- 34. Shimizu, M.; Fukutomi, Y.; Ninomiya, M.; Nagura, K.; Kato, T.; Araki, H.; Suganuma, M.; Fujiki, H.; Moriwaki, H. Green tea extracts for the prevention of metachronous colorectal adenomas: A pilot study. *Cancer Epidemiol. Biomark. Prev.* **2008**, *17*, 3020–3025. [CrossRef] [PubMed]
- 35. Butt, M.S.; Sultan, M.T. Green tea: Nature's defense against malignancies. *Crit. Rev. Food Sci. Nutr.* **2009**, *49*, 463–473. [CrossRef] [PubMed]
- 36. Khan, N.; Afaq, F.; Saleem, M.; Ahmad, N.; Mukhtar, H. Targeting multiple signaling pathways by green tea polyphenol (–)-epigallocatechin-3-gallate. *Cancer Res.* **2006**, *66*, 2500–2505. [CrossRef] [PubMed]
- 37. Shukla, Y. Tea and cancer chemoprevention: A comprehensive review. *Asian Pac. J. Cancer Prev.* 2007, *8*, 155–166. [PubMed]
- 38. Chen, C.; Shen, G.; Hebbar, V.; Hu, R.; Owuor, E.D.; Kong, A.N. Epigallocatechin-3-gallate-induced stress signals in HT-29 human colon adenocarcinoma cells. *Carcinogenesis* **2003**, *24*, 1369–1378. [CrossRef] [PubMed]
- Shimizu, M.; Deguchi, A.; Lim, J.T.; Moriwaki, H.; Kopelovich, L.; Weinstein, I.B. (–)-Epigallocatechin gallate and polyphenon E inhibit growth and activation of the epidermal growth factor receptor and human epidermal growth factor receptor-2 signaling pathways in human colon cancer cells. *Clin. Cancer Res.* 2005, 11, 2735–2746. [CrossRef] [PubMed]
- 40. Shimizu, M.; Shirakami, Y.; Sakai, H.; Yasuda, Y.; Kubota, M.; Adachi, S.; Tsurumi, H.; Hara, Y.; Moriwaki, H. (–)-Epigallocatechin gallate inhibits growth and activation of the VEGF/VEGFR axis in human colorectal cancer cells. *Chem. Biol. Interact.* **2010**, *185*, 247–252. [CrossRef] [PubMed]

- 41. Jia, X.; Han, C. Effects of green tea on colonic aberrant crypt foci and proliferative indexes in rats. *Nutr. Cancer* **2001**, *39*, 239–243. [CrossRef] [PubMed]
- Shimizu, M.; Deguchi, A.; Joe, A.K.; McKoy, J.F.; Moriwaki, H.; Weinstein, I.B. EGCG inhibits activation of HER3 and expression of cyclooxygenase-2 in human colon cancer cells. *J. Exp. Ther. Oncol.* 2005, *5*, 69–78. [PubMed]
- Adachi, S.; Nagao, T.; Ingolfsson, H.I.; Maxfield, F.R.; Andersen, O.S.; Kopelovich, L.; Weinstein, I.B. The inhibitory effect of (–)-epigallocatechin gallate on activation of the epidermal growth factor receptor is associated with altered lipid order in HT29 colon cancer cells. *Cancer Res.* 2007, *67*, 6493–6501. [CrossRef] [PubMed]
- 44. Adachi, S.; Nagao, T.; To, S.; Joe, A.K.; Shimizu, M.; Matsushima-Nishiwaki, R.; Kozawa, O.; Moriwaki, H.; Maxfield, F.R.; Weinstein, I.B. (–)-Epigallocatechin gallate causes internalization of the epidermal growth factor receptor in human colon cancer cells. *Carcinogenesis* **2008**, *29*, 1986–1993. [CrossRef] [PubMed]
- Adachi, S.; Shimizu, M.; Shirakami, Y.; Yamauchi, J.; Natsume, H.; Matsushima-Nishiwaki, R.; To, S.; Weinstein, I.B.; Moriwaki, H.; Kozawa, O. (–)-Epigallocatechin gallate downregulates EGF receptor via phosphorylation at Ser1046/1047 by p38 MAPK in colon cancer cells. *Carcinogenesis* 2009, 30, 1544–1552. [CrossRef] [PubMed]
- 46. Terzic, J.; Grivennikov, S.; Karin, E.; Karin, M. Inflammation and colon cancer. *Gastroenterology* **2010**, *138*, 2101–2114 e5. [CrossRef] [PubMed]
- Tanaka, T.; Kohno, H.; Suzuki, R.; Yamada, Y.; Sugie, S.; Mori, H. A novel inflammation-related mouse colon carcinogenesis model induced by azoxymethane and dextran sodium sulfate. *Cancer Sci.* 2003, *94*, 965–973. [CrossRef] [PubMed]
- Shirakami, Y.; Shimizu, M.; Tsurumi, H.; Hara, Y.; Tanaka, T.; Moriwaki, H. EGCG and Polyphenon E attenuate inflammation-related mouse colon carcinogenesis induced by AOM plus DDS. *Mol. Med. Rep.* 2008, *1*, 355–361. [CrossRef] [PubMed]
- Sueoka, N.; Suganuma, M.; Sueoka, E.; Okabe, S.; Matsuyama, S.; Imai, K.; Nakachi, K.; Fujiki, H. A new function of green tea: Prevention of lifestyle-related diseases. *Ann. N. Y. Acad. Sci.* 2001, *928*, 274–280. [CrossRef] [PubMed]
- 50. Frezza, E.E.; Wachtel, M.S.; Chiriva-Internati, M. Influence of obesity on the risk of developing colon cancer. *Gut* **2006**, *55*, 285–291. [CrossRef] [PubMed]
- 51. Chang, C.K.; Ulrich, C.M. Hyperinsulinaemia and hyperglycaemia: Possible risk factors of colorectal cancer among diabetic patients. *Diabetologia* **2003**, *46*, 595–607. [CrossRef] [PubMed]
- 52. Clayton, P.E.; Banerjee, I.; Murray, P.G.; Renehan, A.G. Growth hormone, the insulin-like growth factor axis, insulin and cancer risk. *Nat. Rev. Endocrinol.* **2011**, *7*, 11–24. [CrossRef] [PubMed]
- 53. Giovannucci, E. Insulin, insulin-like growth factors and colon cancer: A review of the evidence. *J. Nutr.* **2001**, *131*, 3109S–3120S. [PubMed]
- 54. Pollak, M. Insulin and insulin-like growth factor signalling in neoplasia. *Nat. Revs. Cancer* **2008**, *8*, 915–928. [CrossRef] [PubMed]
- 55. Esposito, K.; Nappo, F.; Marfella, R.; Giugliano, G.; Giugliano, F.; Ciotola, M.; Quagliaro, L.; Ceriello, A.; Giugliano, D. Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: Role of oxidative stress. *Circulation* **2002**, *106*, 2067–2072. [CrossRef] [PubMed]
- 56. Flores, M.B.; Rocha, G.Z.; Damas-Souza, D.M.; Osorio-Costa, F.; Dias, M.M.; Ropelle, E.R.; Camargo, J.A.; de Carvalho, R.B.; Carvalho, H.F.; Saad, M.J.; et al. Obesity-induced increase in tumor necrosis factor-α leads to development of colon cancer in mice. *Gastroenterology* **2012**, *143*, 741–753. [CrossRef] [PubMed]
- 57. Szlosarek, P.; Charles, K.A.; Balkwill, F.R. Tumour necrosis factor-alpha as a tumour promoter. *Eur. J. Cancer* **2006**, *42*, 745–750. [CrossRef] [PubMed]
- 58. Leslie, N.R. The redox regulation of PI 3-kinase-dependent signaling. *Antioxid. Redox Signal.* 2006, *8*, 1765–1774. [CrossRef] [PubMed]
- 59. Valko, M.; Izakovic, M.; Mazur, M.; Rhodes, C.J.; Telser, J. Role of oxygen radicals in DNA damage and cancer incidence. *Mol. Cell. Biochem.* **2004**, *266*, 37–56. [CrossRef] [PubMed]
- 60. Barb, D.; Williams, C.J.; Neuwirth, A.K.; Mantzoros, C.S. Adiponectin in relation to malignancies: A review of existing basic research and clinical evidence. *Am. J. Clin. Nutr.* **2007**, *86*, s858–s866. [PubMed]

- 61. Considine, R.V.; Sinha, M.K.; Heiman, M.L.; Kriauciunas, A.; Stephens, T.W.; Nyce, M.R.; Ohannesian, J.P.; Marco, C.C.; McKee, L.J.; Bauer, T.L.; et al. Serum immunoreactive-leptin concentrations in normal-weight and obese humans. *N. Engl. J. Med.* **1996**, *334*, 292–295. [CrossRef] [PubMed]
- Fenton, J.I.; Hursting, S.D.; Perkins, S.N.; Hord, N.G. Interleukin-6 production induced by leptin treatment promotes cell proliferation in an Apc <sup>Min/+</sup> colon epithelial cell line. *Carcinogenesis* 2006, 27, 1507–1515. [CrossRef] [PubMed]
- 63. Molina, A.; Vendrell, J.; Gutierrez, C.; Simon, I.; Masdevall, C.; Soler, J.; Gomez, J.M. Insulin resistance, leptin and TNF-alpha system in morbidly obese women after gastric bypass. *Obes. Surg.* **2003**, *13*, 615–621. [CrossRef] [PubMed]
- 64. Amemori, S.; Ootani, A.; Aoki, S.; Fujise, T.; Shimoda, R.; Kakimoto, T.; Shiraishi, R.; Sakata, Y.; Tsunada, S.; Iwakiri, R.; et al. Adipocytes and preadipocytes promote the proliferation of colon cancer cells in vitro. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2007**, *292*, G923–G929. [CrossRef] [PubMed]
- 65. Stattin, P.; Lukanova, A.; Biessy, C.; Soderberg, S.; Palmqvist, R.; Kaaks, R.; Olsson, T.; Jellum, E. Obesity and colon cancer: Does leptin provide a link? *Int. J. Cancer* **2004**, *109*, 149–152. [CrossRef] [PubMed]
- 66. Lee, G.H.; Proenca, R.; Montez, J.M.; Carroll, K.M.; Darvishzadeh, J.G.; Lee, J.I.; Friedman, J.M. Abnormal splicing of the leptin receptor in diabetic mice. *Nature* **1996**, *379*, 632–635. [CrossRef] [PubMed]
- 67. Hirose, Y.; Hata, K.; Kuno, T.; Yoshida, K.; Sakata, K.; Yamada, Y.; Tanaka, T.; Reddy, B.S.; Mori, H. Enhancement of development of azoxymethane-induced colonic premalignant lesions in C57BL/KsJ-db/db mice. *Carcinogenesis* **2004**, *25*, 821–825. [CrossRef] [PubMed]
- Shimizu, M.; Shirakami, Y.; Sakai, H.; Adachi, S.; Hata, K.; Hirose, Y.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. (–)-Epigallocatechin gallate suppresses azoxymethane-induced colonic premalignant lesions in male C57BL/KsJ-db/db mice. *Cancer Prev. Res.* 2008, *1*, 298–304. [CrossRef] [PubMed]
- 69. Shimizu, M.; Deguchi, A.; Hara, Y.; Moriwaki, H.; Weinstein, I.B. EGCG inhibits activation of the insulin-like growth factor-1 receptor in human colon cancer cells. *Biochem. Biophys. Res. Commun.* **2005**, *334*, 947–953. [CrossRef] [PubMed]
- Hayashi, K.; Suzuki, R.; Miyamoto, S.; Shin-Ichiroh, Y.; Kohno, H.; Sugie, S.; Takashima, S.; Tanaka, T. Citrus auraptene suppresses azoxymethane-induced colonic preneoplastic lesions in C57BL/KsJ-db/db mice. *Nutr. Cancer* 2007, *58*, 75–84. [CrossRef] [PubMed]
- Miyamoto, S.; Yasui, Y.; Ohigashi, H.; Tanaka, T.; Murakami, A. Dietary flavonoids suppress azoxymethane-induced colonic preneoplastic lesions in male C57BL/KsJ-db/db mice. *Chem. Biol. Interact.* 2010, 183, 276–283. [CrossRef] [PubMed]
- 72. Aggarwal, B.B.; Kumar, A.; Bharti, A.C. Anticancer potential of curcumin: Preclinical and clinical studies. *Anticancer Res.* **2003**, *23*, 363–398. [PubMed]
- 73. Mukhopadhyay, A.; Banerjee, S.; Stafford, L.J.; Xia, C.; Liu, M.; Aggarwal, B.B. Curcumin-induced suppression of cell proliferation correlates with down-regulation of cyclin D1 expression and CDK4-mediated retinoblastoma protein phosphorylation. *Oncogene* **2002**, *21*, 8852–8861. [CrossRef] [PubMed]
- 74. Guillem-Llobat, P.; Dovizio, M.; Alberti, S.; Bruno, A.; Patrignani, P. Platelets, cyclooxygenases, and colon cancer. *Semin. Oncol.* **2014**, *41*, 385–396. [CrossRef] [PubMed]
- 75. Wang, D.; Dubois, R.N. The role of COX-2 in intestinal inflammation and colorectal cancer. *Oncogene* **2010**, *29*, 781–788. [CrossRef] [PubMed]
- 76. Bertagnolli, M.M.; Eagle, C.J.; Zauber, A.G.; Redston, M.; Solomon, S.D.; Kim, K.; Tang, J.; Rosenstein, R.B.; Wittes, J.; Corle, D.; et al. Celecoxib for the prevention of sporadic colorectal adenomas. *N. Engl. J. Med.* 2006, 355, 873–884. [CrossRef] [PubMed]
- 77. Arber, N.; Eagle, C.J.; Spicak, J.; Racz, I.; Dite, P.; Hajer, J.; Zavoral, M.; Lechuga, M.J.; Gerletti, P.; Tang, J.; et al. Celecoxib for the prevention of colorectal adenomatous polyps. *N. Engl. J. Med.* **2006**, *355*, 885–895. [CrossRef] [PubMed]
- 78. Ishikawa, H.; Wakabayashi, K.; Suzuki, S.; Mutoh, M.; Hirata, K.; Nakamura, T.; Takeyama, I.; Kawano, A.; Gondo, N.; Abe, T.; et al. Preventive effects of low-dose aspirin on colorectal adenoma growth in patients with familial adenomatous polyposis: Double-blind, randomized clinical trial. *Cancer Med.* 2013, 2, 50–56. [CrossRef] [PubMed]
- 79. Ishikawa, H.; Mutoh, M.; Suzuki, S.; Tokudome, S.; Saida, Y.; Abe, T.; Okamura, S.; Tajika, M.; Joh, T.; Tanaka, S.; et al. The preventive effects of low-dose enteric-coated aspirin tablets on the development of colorectal tumours in Asian patients: A randomised trial. *Gut* **2014**, *63*, 1755–1759. [CrossRef] [PubMed]

- 80. Higurashi, T.; Hosono, K.; Takahashi, H.; Komiya, Y.; Umezawa, S.; Sakai, E.; Uchiyama, T.; Taniguchi, L.; Hata, Y.; Uchiyama, S.; et al. Metformin for chemoprevention of metachronous colorectal adenoma or polyps in post-polypectomy patients without diabetes: A multicentre double-blind, placebo-controlled, randomised phase 3 trial. *Lancet Oncol.* **2016**, *17*, 475–483. [CrossRef]
- 81. Gauthaman, K.; Fong, C.Y.; Bongso, A. Statins, stem cells, and cancer. J. Cell. Biochem. 2009, 106, 975–983. [CrossRef] [PubMed]
- Cho, S.J.; Kim, J.S.; Kim, J.M.; Lee, J.Y.; Jung, H.C.; Song, I.S. Simvastatin induces apoptosis in human colon cancer cells and in tumor xenografts, and attenuates colitis-associated colon cancer in mice. *Int. J. Cancer* 2008, 123, 951–957. [CrossRef] [PubMed]
- Poynter, J.N.; Gruber, S.B.; Higgins, P.D.; Almog, R.; Bonner, J.D.; Rennert, H.S.; Low, M.; Greenson, J. K.; Rennert, G. Statins and the risk of colorectal cancer. *N. Engl. J. Med.* 2005, 352, 2184–2192. [CrossRef] [PubMed]
- 84. Kochi, T.; Shimizu, M.; Ohno, T.; Baba, A.; Sumi, T.; Kubota, M.; Shirakami, Y.; Tsurumi, H.; Tanaka, T.; Moriwaki, H. Enhanced development of azoxymethane-induced colonic preneoplastic lesions in hypertensive rats. *Int. J. Mol. Sci.* **2013**, *14*, 14700–14711. [CrossRef] [PubMed]
- 85. De Kloet, A.D.; Krause, E.G.; Woods, S.C. The renin angiotensin system and the metabolic syndrome. *Physiol. Behav.* **2010**, *100*, 525–534. [CrossRef] [PubMed]
- 86. Smith, G.R.; Missailidis, S. Cancer, inflammation and the AT1 and AT2 receptors. *J. Inflamm.* **2004**, *1*, 3. [CrossRef] [PubMed]
- 87. Bettuzzi, S.; Brausi, M.; Rizzi, F.; Castagnetti, G.; Peracchia, G.; Corti, A. Chemoprevention of human prostate cancer by oral administration of green tea catechins in volunteers with high-grade prostate intraepithelial neoplasia: A preliminary report from a one-year proof-of-principle study. *Cancer Res.* **2006**, *66*, 1234–1240. [CrossRef] [PubMed]
- 88. Chow, H.H.; Cai, Y.; Alberts, D.S.; Hakim, I.; Dorr, R.; Shahi, F.; Crowell, J.A.; Yang, C.S.; Hara, Y. Phase I pharmacokinetic study of tea polyphenols following single-dose administration of epigallocatechin gallate and polyphenon E. *Cancer Epidemiol. Biomark. Prev.* **2001**, *10*, 53–58.



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