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Design, synthesis, and biological evaluation of novel iso-flavones derivatives as H₃R antagonists

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ABSTRACT

Histamine H₃ receptor (H₃R), a kind of G-protein coupled receptor (GPCR), is expressed mainly in the central nervous system (CNS) and plays a vital role in homoeostatic control. This study describes the design and synthesis of a series of novel H₃R antagonists based on the iso-flavone scaffold. The results of the bioactivity evaluation show that four compounds (**1c**, **2c**, **2h**, and **2o**) possess significant H₃R inhibitory activities. Molecular docking indicates that a salt bridge, π–π T-shape interactions, and hydrophobic interaction all contribute to the interaction between compound **2h** and H₃R.

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H₃R antagonist; iso-flavone; molecular docking

Introduction

Histamine, a distinctly important neurotransmitter, exerts as a modulator in the brain and dominates several homoeostatic functions such as thermoregulation, fluid balance, and energy metabolism¹. Apart from that, histamine is also involved in numerous processes, for instance, circadian rhythms, the sleep–wake cycle, attention, memory, learning, and neuroendocrine regulation². According to recent studies, the biosynthesis and release of histamine in central nervous system (CNS) are modulated by four different G-protein coupled receptors (GPCRs) subtypes, namely histamine H₁ receptor (H₁R), histamine H₂ receptor (H₂R), histamine H₃ receptor (H₃R), histamine H₄ receptor (H₄R). Unlike H₁R and H₂R, H₃R shows higher homology to H₄R³ and is highly expressed in brain⁴, such as basal ganglia and globus pallidus, which could couple with G i/o protein and then activate mitogen-activated protein kinase (MAPK) and phosphatidylinositol 3-kinase (PI3K) pathways⁵. Subsequently, the phospholipase A₂ (PLA₂) is induced to recruit Ca²⁺ from intracellular stores⁶, reduces cAMP formation⁷, and enhances phosphorylation². Moreover, H₃R is recognised as an auto- and hetero-receptor on non-histaminergic neurons controlling the release of many other important neurotransmitters^{8,9}, such as acetylcholine, norepinephrine, dopamine, and serotonin¹⁰. A clinical study revealed that neurotransmitters could trigger the postsynaptic signalling pathways bound to cognition which supported the hypothesis that H₃R is a drug target for cognitive disorders^{3,6,11,12}, especially for Alzheimer Disease (AD), schizophrenia and epilepsy^{13–16}. Because of the unique functions of H₃R, a wide variety of selective H₃R antagonists have been developed and some of them have shown promising effects^{4,12,17–21}.

Flavone and iso-flavone, which are regarded as privileged structures, exhibit variety of pharmacological activities, such as anti-cancer, antimicrobial, anti-inflammatory, and also are used in neurodegenerative disorders, for example, Alzheimer's disease^{22–24}. Our previous study had confirmed the iso-flavone and flavone compounds possessed

moderate inhibitory activity against H₃R²⁵. Particularly, the optimization at the 8-position of the flavones and 7-position of iso-flavone provided satisfactory bioactivity (compound **A**, **B**, and **C**, Figure 1), which enlightened us to modify 8-position of iso-flavone to enhance the H₃R inhibitory effect. In addition, we also want to modify the 6-position of isoflavones to see whether compounds with better antagonistic activity can be obtained. In this current work, two series of novel iso-flavone derivatives were designed and synthesised based on our previous study. After screening the H₃R inhibitory activities at a fixed concentration, compounds that possessed good H₃R inhibitory activity were further tested to determine the IC₅₀ values. In addition, molecular docking studies were performed to investigate the interaction between H₃R and the most potent antagonist.

Materials and methods

Chemistry

Unless otherwise indicated, all solvents and organic reagents were obtained from commercially available sources and were used without further purification. The reaction process was monitored using thin layer chromatography (TLC) with silica gel plates (thickness = 0.20 mm, GF254) under UV light. Column chromatography was performed using a ZCX-II (200–300 mesh), to purify the final products. All final products were found to have purities ≥95% analysed by HPLC. Melting points were determined using a YRT-3 apparatus (Tian Jin Optical Instrument Factory, Tianjin, China) and were presented as uncorrected values. ¹H NMR spectra were recorded on a Varian Mercury-300 MHz instrument, whereas ¹³C NMR was recorded at 400 MHz on a Varian Mercury using DMSO-d₆ as a solvent and tetramethylsilane (TMS) as an internal standard (¹H NMR and ¹³C NMR were recorded in different time). Mass spectra were obtained using a Waters Acquity UPLC-SQD mass spectrometer (Waters, Milford, MA). High-resolution mass spectra

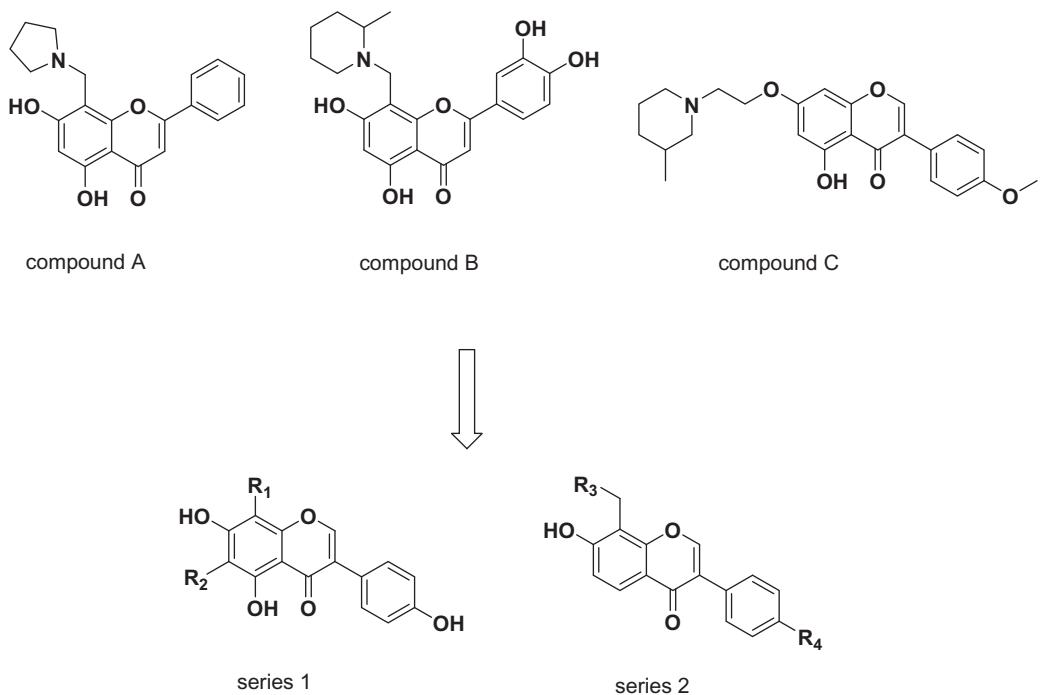
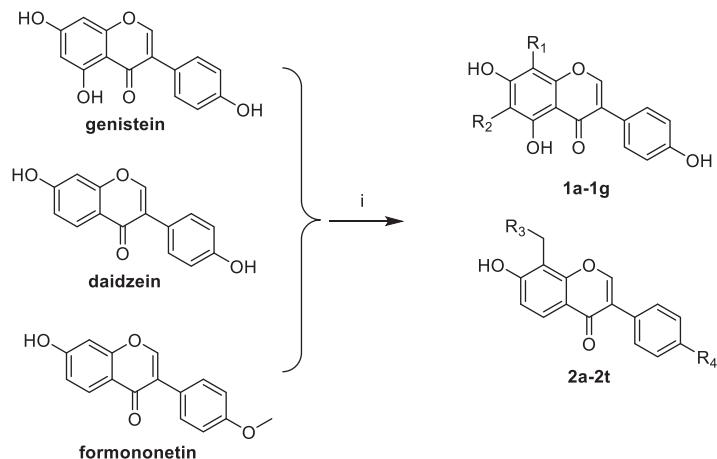


Figure 1. The structures of previously reported H_3R antagonists and two novel series of compounds.



Scheme 1. Synthesis of compounds 1a–1g, 2a–2t. Reagents and conditions: (i) 37% formalin, aliphatic amines, 25 °C, 24 h.

(HRMS) were recorded on an Agilent Technologies LC/MSD TOF spectrometer (Agilent Technologies Co. Ltd., Santa Clara, CA).

The synthetic route of novel compounds is depicted in **Scheme 1**. All title compounds were synthesised through Mannich reactions using iso-flavone, 37% formalin, and aliphatic amines as starting materials. Compounds 1a–1g, 2a–2i, and 2j–2t were synthesised from genistein, daidzein, and formononetin, respectively. The use of DMF-methanol as a solvent for formononetin and daidzein never resulted in the formation of 6-substituted products, but only 8-position substituted products were obtained.

General procedure for the synthesis of compounds 1a–1g

Genistein (0.50 g, 1.85 mmol), 37% formalin (0.30 g, 3.70 mmol), aliphatic amines (0.225 g, 2.780 mmol), and methanol (30 ml) were added into a three-necked flask (100 ml) and stirred at 25 °C for

24 h. After reactions completed monitored by TLC (DCM:MeOH = 10:1), the solvent was removed under reduced pressure. The residue was purified by column chromatography using a mixture of dichloromethane and methanol (30:1) as the eluent to give the target compounds in yields ranging from 41% to 91%.

The similar procedure was followed for the synthesis of compounds 2a–2t.

Title compounds were characterised as follows:

5,7-dihydroxy-3-(4-hydroxyphenyl)-8-(pyrrolidin-1-ylmethyl)-4H-chromen-4-one (1a)

White solid, yield: 24%; mp 218–220 °C; 1H NMR (300 MHz, DMSO- d_6) δ 8.15 (s, 1H), 7.35 (d, J = 8.7 Hz, 2H), 6.80 (d, J = 8.4 Hz, 2H), 6.10 (s, 1H), 3.96 (s, 2H), 2.83 (m, 4H), 1.83 (m, 4H). ^{13}C NMR (100 MHz, DMSO- d_6) δ 179.9, 170.4, 161.6, 159.5, 157.8, 153.5,

130.7, 123.8, 122.2, 115.5, 104.7, 102.2, 94.7, 53.3, 50.0, 23.6. HR-MS (ESI) Calcd for $C_{20}H_{19}NO_5$ [M + H]⁺, 354.1341, found: 354.1368.

8-((4-benzylpiperazin-1-yl)methyl)-5,7-dihydroxy-3-(4-hydroxyphenyl)-4H-chromen-4-one (1b)

White solid, yield: 24%; mp 191–193 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.35–7.28 (m, 7H), 6.81 (d, *J* = 8.4 Hz, 2H), 6.17 (s, 1H), 3.81 (s, 2H), 3.46 (s, 2H), 2.56 (m, 4H), 2.40 (m, 4H). ¹³C NMR (100 MHz, DMSO-d₆) δ 180.9, 165.5, 161.1, 157.9, 155.6, 154.2, 138.5, 130.7, 129.4, 128.7, 127.5, 122.7, 121.7, 115.6, 104.7, 100.1, 99.5, 62.3, 52.8, 52.5, 51.7. HR-MS (ESI) Calcd for $C_{27}H_{26}N_2O_5$ [M + H]⁺, 459.1920, found: 459.1939.

5,7-dihydroxy-3-(4-hydroxyphenyl)-8-((3-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (1c)

White solid, yield: 18%; mp 209–211 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.29 (s, 1H), 7.36 (d, *J* = 8.7 Hz, 2H), 6.81 (d, *J* = 8.4 Hz, 2H), 6.10 (s, 1H), 3.84 (s, 2H), 2.89 (brs, 2H), 2.15 (t, *J* = 10.8 Hz, 1H), 1.89 (t, *J* = 10.8 Hz, 1H), 1.65–1.48 (m, 4H), 0.94 (m, 1H), 0.82 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 180.4, 168.0, 163.5, 157.9, 157.4, 154.1, 130.7, 122.5, 121.9, 115.6, 104.0, 103.5, 94.5, 60.0, 53.1, 52.7, 32.0, 30.9, 24.9, 19.6. HR-MS (ESI): Calcd for $C_{22}H_{23}NO_5$ [M + H]⁺, 382.1654, found: 382.1682.

6-((3R,5S)-3,5-dimethylmorpholino)methyl)-5,7-dihydroxy-3-(4-hydroxyphenyl)-4H-chromen-4-one (1d)

White solid, yield: 20%; mp >250 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 13.02 (s, 1H), 8.36 (s, 1H), 7.37 (d, *J* = 8.4 Hz, 2H), 6.81 (d, *J* = 8.4 Hz, 2H), 6.22 (s, 1H), 3.73 (s, 2H), 3.54 (t, *J* = 9 Hz, 2H), 2.81 (d, *J* = 12 Hz, 2H), 1.78 (t, *J* = 10.8 Hz, 2H), 1.04 (d, *J* = 6.3 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 181.0, 177.9, 176.8, 164.5, 157.9, 155.8, 154.3, 130.7, 121.7, 115.6, 104.9, 100.5, 99.4, 71.5, 58.6, 51.2, 19.4. HR-MS (ESI) Calcd for $C_{22}H_{23}NO_6$ [M + H]⁺, 391.1604, found: 398.1633.

5,7-dihydroxy-8-((4-hydroxymethyl)piperidin-1-yl)methyl)-3-(4-hydroxyphenyl)-4H-chromen-4-one (1e)

White solid, yield: 19%; mp 223–225 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.29 (s, 1H), 7.36 (d, *J* = 8.4 Hz, 2H), 6.81 (d, *J* = 8.7 Hz, 2H), 6.09 (s, 1H), 3.86 (s, 2H), 3.57 (brs, 2H), 2.84 (t, *J* = 6.6 Hz, 2H), 2.38 (t, *J* = 10.2 Hz, 2H), 1.68 (d, *J* = 12.9 Hz, 2H), 1.12 (brs, 1H), 1.23–1.11 (m, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 180.6, 167.4, 161.4, 157.9, 155.4, 153.8, 130.7, 122.6, 121.8, 115.6, 104.1, 99.9, 99.2, 65.9, 52.7, 52.6, 38.1, 28.6. HR-MS (ESI) Calcd for $C_{22}H_{23}NO_6$ [M + H]⁺: 398.1604, found: 398.1584.

5,7-dihydroxy-3-(4-hydroxyphenyl)-6-(morpholinomethyl)-4H-chromen-4-one (1f)

White solid, yield: 16%; mp 210–212 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 13.03 (s, 1H), 8.36 (s, 1H), 7.38 (d, *J* = 8.4 Hz, 2H), 6.82 (d, *J* = 8.4 Hz, 2H), 6.24 (s, 1H), 3.74 (s, 2H), 3.58 (m, 4H), 2.49 (m, 4H). ¹³C NMR (100 MHz, DMSO-d₆) δ 180.9, 164.6, 161.4, 157.9, 154.4, 143.3, 130.7, 122.7, 121.7, 115.6, 104.9, 100.7, 99.3, 66.6, 53.14, 51.4. HR-MS (ESI) Calcd for $C_{20}H_{19}NO_6$ [M + H]⁺, 370.1291, found: 370.1320.

5,7-dihydroxy-3-(4-hydroxyphenyl)-8-((4-methylpiperazin-1-yl)methyl)-4H-chromen-4-one (1g)

White solid, yield: 19%; mp 231–233 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.34 (s, 1H), 7.37 (d, *J* = 8.4 Hz, 2H), 6.82 (d, *J* = 8.7 Hz, 2H), 6.18 (s, 1H), 3.80 (s, 2H), 3.55 (m, 4H), 2.35 (m, 4H), 2.07 (s, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 180.7, 166.4, 161.5, 158.0, 155.6, 154.2, 130.7, 122.7, 121.7, 115.6, 104.7, 100.1, 99.6, 54.9, 52.4, 51.7, 46.0. HR-MS (ESI) Calcd for $C_{21}H_{22}N_2O_5$ [M + H]⁺, 383.1607, found: 383.1609.

7-hydroxy-8-((4-(2-hydroxyethyl)piperazin-1-yl)methyl)-3-(4-hydroxyphenyl)-4H-chromen-4-one (2a)

White solid, yield: 20%; mp 230–232 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.30 (s, 1H), 7.91 (d, *J* = 8.7 Hz, 1H), 7.37 (d, *J* = 8.7 Hz, 2H), 6.88 (d, *J* = 9.0 Hz, 1H), 6.77 (d, *J* = 8.4 Hz, 2H), 3.95 (s, 2H), 3.49 (t, *J* = 6.3 Hz, 2H), 2.57–2.48 (m, 8H), 2.38 (t, *J* = 6.3 Hz, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.3, 163.4, 157.7, 155.5, 153.0, 130.6, 126.3, 123.8, 123.0, 116.9, 115.5, 115.5, 108.7, 60.5, 59.0, 53.4, 52.7, 52.4. HR-MS (ESI) Calcd for $C_{22}H_{24}N_2O_5$ [M + H]⁺, 397.1763, found: 397.1767.

7-hydroxy-8-((4-(hydroxymethyl)piperidin-1-yl)methyl)-3-(4-hydroxyphenyl)-4H-chromen-4-one (2b)

White solid, yield: 24%; mp 244–246 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.28 (s, 1H), 7.89 (d, *J* = 8.7 Hz, 1H), 7.37 (d, *J* = 8.4 Hz, 2H), 6.84–6.77 (m, 3H), 3.98 (s, 2H), 3.26 (d, *J* = 6.3 Hz, 2H), 2.95 (d, *J* = 11.1 Hz, 2H), 2.21 (t, *J* = 11.1 Hz, 2H), 1.73 (d, *J* = 13.2 Hz, 2H), 1.43 (brs, 1H), 1.19 (m, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.3, 164.2, 157.7, 155.5, 152.9, 130.6, 126.1, 123.8, 123.0, 116.6, 115.7, 115.5, 108.4, 66.00, 53.3, 52.9, 38.2, 28.9. HR-MS (ESI) Calcd for $C_{22}H_{23}NO_5$ [M + H]⁺, 382.1654, found: 382.1648.

7-hydroxy-3-(4-hydroxyphenyl)-8-((4-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (2c)

White solid, yield: 14%; mp 250–252 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.28 (s, 1H), 7.89 (d, *J* = 8.7 Hz, 1H), 7.37 (d, *J* = 8.7 Hz, 2H), 6.84–6.77 (m, 3H), 3.97 (s, 2H), 2.95 (d, *J* = 11.1 Hz, 2H), 2.22 (t, *J* = 11.4 Hz, 2H), 1.68 (d, *J* = 13.2 Hz, 2H), 1.42 (brs, 1H), 1.17 (m, 2H), 0.91 (d, *J* = 6.6 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.3, 164.2, 157.7, 155.5, 152.9, 130.6, 126.1, 123.8, 123.0, 116.6, 115.7, 115.5, 108.5, 53.1, 34.1, 30.2, 22.0. HR-MS (ESI) Calcd for $C_{22}H_{23}NO_4$ [M + H]⁺, 366.1705, found: 366.1749.

8-((3R,5S)-3,5-dimethylmorpholino)methyl)-7-hydroxy-3-(4-hydroxyphenyl)-4H-chromen-4-one (2d)

White solid, yield: 27%; mp 230–232 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.32 (s, 1H), 7.92 (d, *J* = 9 Hz, 1H), 7.38 (d, *J* = 9 Hz, 2H), 6.93 (d, *J* = 9 Hz, 1H), 6.80 (d, *J* = 8.4 Hz, 2H), 3.88 (s, 2H), 3.56 (t, *J* = 8.4 Hz, 2H), 2.83 (d, *J* = 10.8 Hz, 2H), 1.90 (t, *J* = 11.1 Hz, 2H), 1.05 (d, *J* = 6.3 Hz, 6H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.4, 162.8, 157.8, 155.8, 153.1, 130.6, 126.4, 123.8, 123.00, 117.1, 115.5, 115.4, 109.1, 71.5, 58.7, 51.8, 19.4. HR-MS (ESI) Calcd for $C_{22}H_{23}NO_5$ [M + H]⁺, 382.1654, found: 382.1664.

7-hydroxy-3-(4-hydroxyphenyl)-8-((3-hydroxypiperidin-1-yl)methyl)-4H-chromen-4-one (2e)

White solid, yield: 15%; mp 226–228 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.28 (s, 1H), 7.89 (d, *J* = 9 Hz, 1H), 7.37 (d, *J* = 8.4 Hz, 2H),

6.84–6.77 (m, 3H), 3.96 (s, 2H), 2.87 (d, $J = 7.2$ Hz, 2H), 2.13 (t, $J = 11.1$ Hz, 1H), 1.87 (t, $J = 10.8$ Hz, 1H), 1.69–1.42 (m, 4H), 0.90 (m, 1H), 0.84 (d, $J = 6.3$ Hz, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.3, 164.2, 157.6, 155.4, 152.9, 130.6, 126.2, 123.8, 123.0, 116.6, 115.7, 115.5, 108.3, 60.6, 53.4, 53.2, 32.3, 31.2, 25.2, 19.7. HR-MS (ESI) Calcd for C₂₂H₂₃NO₄ [M + H]⁺, 366.1705, found: 366.1716.

7-hydroxy-3-(4-hydroxyphenyl)-8-(pyrrolidin-1-ylmethyl)-4H-chromen-4-one (2f).

White solid, yield: 23%; mp 177–179 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.27 (s, 1H), 7.88 (d, $J = 8.7$ Hz, 1H), 7.37 (d, $J = 8.4$ Hz, 2H), 6.84–6.77 (m, 3H), 4.08 (s, 2H), 2.67 (m, 4H), 1.77 (m, 4H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.3, 164.7, 157.7, 155.4, 152.8, 130.6, 126.2, 123.8, 123.1, 116.1, 115.9, 115.5, 109.1, 53.6, 50.0, 23.7. HR-MS (ESI) Calcd for C₂₀H₁₉NO₄ [M + H]⁺, 338.1392, found: 338.1413.

(S)-7-hydroxy-8-((2-(hydroxymethyl)pyrrolidin-1-yl)methyl)-3-(4-hydroxyphenyl)-4H-chromen-4-one (2g)

White solid, yield: 35%; mp 205–207 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.27 (s, 1H), 7.88 (d, $J = 8.7$ Hz, 1H), 7.37 (d, $J = 8.7$ Hz, 2H), 6.85–6.77 (m, 3H), 4.34–4.01 (s, 2H), 3.51 (brs, 2H), 2.92–2.83 (d, $J = 27.6$ Hz, 2H), 2.40 (d, $J = 8.1$ Hz, 1H), 1.89 (m, 1H), 1.67 (m, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.3, 172.8, 157.8, 155.2, 152.6, 138.2, 130.6, 126.1, 123.8, 123.0, 120.0, 115.5, 109.6, 65.6, 62.8, 54.6, 49.5, 27.6, 23.1. HR-MS (ESI) Calcd for C₂₁H₂₁NO₅ [M + H]⁺, 368.1498, found: 368.1482.

7-hydroxy-3-(4-hydroxyphenyl)-8-((2-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (2h)

White solid, yield: 12%; mp 228–230 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.28 (s, 1H), 7.89 (d, $J = 8.7$ Hz, 1H), 7.37 (d, $J = 8.4$ Hz, 2H), 6.79–6.77 (m, 3H), 4.32–4.27 (d, $J = 15$, 1H), 3.9–3.85 (d, $J = 15$ Hz, 1H), 2.83 (d, $J = 12$ Hz, 1H), 2.66 (brs, 1H), 2.30 (t, $J = 9.3$ Hz, 1H), 1.62–1.35 (m, 6H), 1.15 (d, $J = 6.3$ Hz, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.3, 164.6, 157.7, 155.2, 152.9, 130.6, 125.9, 123.8, 123.1, 116.4, 115.9, 115.4, 108.7, 57.8, 56.3, 50.4, 36.3, 33.8, 25.6, 22.5. HR-MS (ESI) Calcd for C₂₂H₂₃NO₄ [M + H]⁺, 366.1705, found: 366.1731.

7-hydroxy-3-(4-hydroxyphenyl)-8-((4-methylpiperazin-1-yl)methyl)-4H-chromen-4-one (2i)

White solid, yield: 18%; mp 215–217 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.28 (s, 1H), 7.90 (d, $J = 8.7$ Hz, 1H), 7.37 (d, $J = 8.7$ Hz, 2H), 6.88–6.77 (m, 3H), 3.93 (s, 2H), 2.56 (m, 4H), 2.34 (m, 4H), 2.15 (s, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.4, 163.3, 157.7, 155.6, 153.1, 137.2, 130.6, 126.3, 123.9, 123.0, 116.9, 115.5, 108.9, 55.0, 52.6, 52.3, 46.1. HR-MS (ESI) Calcd for C₂₁H₂₂N₂O₄ [M + H]⁺, 367.1658, found: 367.1646.

7-hydroxy-3-(4-methoxyphenyl)-8-((4-methylpiperazin-1-yl)methyl)-4H-chromen-4-one (2j)

White solid, yield: 23%; mp 202–204 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.36 (s, 1H), 7.92 (d, $J = 8.7$ Hz, 1H), 7.50 (d, $J = 9$ Hz, 2H), 6.99 (d, $J = 8.7$ Hz, 2H), 6.87 (d, $J = 9$ Hz, 1H), 3.95 (s, 2H), 3.77 (s, 3H), 2.57 (m, 4H), 2.36 (m, 4H), 2.16 (s, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.3, 163.2, 159.5, 155.6, 153.4, 130.6, 126.3, 124.7, 123.5, 116.9, 115.5, 114.1, 109.0, 55.7, 55.0, 52.6, 52.3, 46.1. HR-MS (ESI) Calcd for C₂₂H₂₄N₂O₄ [M + H]⁺, 381.1814, found: 381.1814.

7-hydroxy-3-(4-methoxyphenyl)-8-(morpholinomethyl)-4H-chromen-4-one (2k)

White solid, yield: 28%; mp 235–237 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.31 (s, 1H), 7.92 (d, $J = 8.7$ Hz, 1H), 7.38 (d, $J = 8.7$ Hz, 2H), 6.93 (d, $J = 9$ Hz, 1H), 6.77 (d, $J = 8.4$ Hz, 2H), 3.88 (s, 3H), 3.59 (m, 6H), 2.48 (m, 4H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.4, 162.6, 157.7, 155.9, 153.2, 130.6, 126.4, 123.8, 123.0, 117.1, 115.5, 115.3, 109.3, 66.6, 53.2, 52.0. HR-MS (ESI) Calcd for C₂₀H₁₉NO₅ [M + H]⁺, 354.1341, found: 354.1315.

7-hydroxy-3-(4-methoxyphenyl)-8-((4-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (2l)

White solid, yield: 21%; mp 208–210 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.90 (d, $J = 8.7$ Hz, 1H), 7.50 (d, $J = 9$ Hz, 2H), 6.95 (d, $J = 11.7$ Hz, 2H), 6.82 (d, $J = 9$ Hz, 1H), 3.98 (s, 2H), 3.77 (s, 3H), 2.96 (d, $J = 11.4$ Hz, 2H), 2.22 (t, $J = 10.8$ Hz, 2H), 1.68 (d, $J = 12.3$ Hz, 2H), 1.42 (brs, 1H), 1.17 (m, 2H), 0.91 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.3, 159.5, 155.5, 153.2, 130.6, 126.2, 124.7, 123.5, 116.6, 115.8, 114.1, 108.4, 55.7, 53.2, 53.1, 34.1, 30.2, 22.0. HR-MS (ESI) Calcd for C₂₃H₂₅NO₄ [M + H]⁺, 380.1862, found: 380.1881.

7-hydroxy-3-(4-methoxyphenyl)-8-(pyrrolidin-1-ylmethyl)-4H-chromen-4-one (2m)

White solid, yield: 12%; mp 173–175 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.32 (s, 1H), 7.89 (d, $J = 8.7$ Hz, 1H), 7.50 (d, $J = 8.7$ Hz, 2H), 6.98 (d, $J = 8.7$ Hz, 2H), 6.82 (d, $J = 8.7$ Hz, 1H), 4.09 (s, 2H), 3.77 (s, 3H), 2.68 (m, 4H), 1.77 (m, 4H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.5, 159.5, 155.4, 153.1, 130.6, 126.2, 124.8, 123.4, 116.2, 115.9, 114.1, 109.3, 55.6, 53.6, 49.9, 23.7. HR-MS (ESI) Calcd for C₂₁H₂₁NO₄ [M + H]⁺, 352.1568, found: 352.1568.

7-hydroxy-8-((4-hydroxypiperidin-1-yl)methyl)-3-(4-methoxyphenyl)-4H-chromen-4-one (2n)

White solid, yield: 21%; mp 205–207 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.90 (d, $J = 8.7$ Hz, 1H), 7.50 (d, $J = 8.7$ Hz, 2H), 6.98 (d, $J = 9$ Hz, 2H), 6.83 (d, $J = 9$ Hz, 1H), 3.96 (s, 2H), 3.77 (s, 3H), 3.56 (brs, 1H), 2.80 (t, $J = 7.2$ Hz, 2H), 2.35 (t, $J = 10.8$ Hz, 2H), 1.75 (d, $J = 12.9$ Hz, 2H), 1.45 (q, $J = 6.9$ Hz, 2H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.1, 159.5, 155.4, 153.2, 131.5, 125.9, 124.3, 123.4, 116.5, 115.3, 113.9, 109.4, 55.6, 55.4, 52.9, 50.6, 34.4. HR-MS (ESI) Calcd for C₂₂H₂₃NO₅ [M + H]⁺, 382.1654, found: 382.1669.

7-hydroxy-3-(4-methoxyphenyl)-8-((3-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (2o)

White solid, yield: 21%; mp 165–167 °C; ^1H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.90 (d, $J = 9$ Hz, 1H), 7.50 (d, $J = 8.7$ Hz, 2H), 6.98 (d, $J = 9$ Hz, 2H), 6.82 (d, $J = 9$ Hz, 1H), 3.96 (s, 2H), 3.77 (s, 3H), 2.87 (d, $J = 7.5$ Hz, 2H), 2.17 (t, $J = 11.7$ Hz, 1H), 1.83 (t, $J = 10.8$ Hz, 1H), 1.65–1.49 (m, 4H), 0.93 (m, 1H), 0.82 (d, $J = 6.6$ Hz, 3H). ^{13}C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.3, 159.5, 155.5, 153.2, 130.6, 126.2, 124.7, 123.5, 116.6, 115.7, 114.1, 108.4, 60.6, 55.7, 53.4, 53.2, 32.3, 31.2, 25.2, 19.7. HR-MS (ESI) Calcd for C₂₃H₂₅NO₄ [M + H]⁺, 380.1862, found: 380.1897.

Table 1. Structures and activities of compounds 1a–1g.

Compound	1a–1g		Inhibit rate (%) at 10 μ M	IC_{50} (μ M)
	R ₁	R ₂		
1a		H	–7.61	
1b		H	–10.31	
1c		H	38.14	17.83 ± 0.06
1d	H		–61.00	
1e		H	–7.54	
1f	H		–19.15	
1g		H	–43.72	
Thioperamide			72.34	1.03 ± 0.01

Bold values indicates that the compound has a high inhibit rate (%) at 10 μ M and is able to posses an IC_{50} .

7-hydroxy-8-((3-hydroxypiperidin-1-yl)methyl)-3-(4-methoxyphenyl)-4H-chromen-4-one (2p)

White solid, yield: 25%; mp 188–190 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.91 (d, *J* = 9 Hz, 1H), 7.50 (d, *J* = 8.7 Hz, 2H), 6.98 (d, *J* = 9 Hz, 2H), 6.84 (d, *J* = 8.7 Hz, 1H), 3.95 (s, 2H), 3.77 (s, 3H), 3.58 (brs, 1H), 2.85 (d, *J* = 9.3 Hz, 1H), 2.66 (d, *J* = 11.1 Hz, 1H), 2.27–2.13 (m, 2H), 1.71 (d, *J* = 10.5 Hz, 2H), 1.45 (m, 1H), 1.24 (m, 1H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.0, 159.5, 155.5, 153.2, 130.6, 126.2, 124.7, 123.5, 116.5, 115.8, 114.1, 108.4, 66.00, 55.7, 53.3, 52.9, 38.2, 28.9. HR-MS (ESI) Calcd for C₂₃H₂₅NO₅ [M + H]⁺, 396.1811, found: 396.1806.

J = 11.4 Hz, 2H), 1.73 (d, *J* = 12.9 Hz, 2H), 1.23 (brs, 1H), 1.11 (m, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.3, 159.5, 155.5, 153.2, 130.6, 126.2, 124.7, 123.5, 116.5, 115.8, 114.1, 108.4, 66.00, 55.7, 53.3, 52.9, 38.2, 28.9. HR-MS (ESI) Calcd for C₂₃H₂₅NO₅ [M + H]⁺, 396.1811, found: 396.1806.

7-hydroxy-8-((4-(2-hydroxyethyl)piperazin-1-yl)methyl)-3-(4-methoxyphenyl)-4H-chromen-4-one (2s)

White solid, yield: 27%; mp 196–198 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.35 (s, 1H), 7.91 (d, *J* = 8.7 Hz, 1H), 7.50 (d, *J* = 8.7 Hz, 2H), 6.98 (d, *J* = 9 Hz, 2H), 6.89 (d, *J* = 9 Hz, 1H), 3.94 (s, 2H), 3.77 (s, 3H), 3.49 (t, *J* = 6.3 Hz, 3H), 2.57–2.40 (m, 8H), 2.36 (t, *J* = 7.2 Hz, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.2, 163.4, 159.5, 155.6, 153.3, 130.6, 126.3, 124.7, 123.3, 116.9, 115.5, 114.1, 108.8, 60.5, 59.0, 55.7, 53.5, 52.7, 52.4. HR-MS (ESI) Calcd for C₂₃H₂₆N₂O₅ [M + H]⁺, 411.1920, found: 411.1904.

7-hydroxy-3-(4-methoxyphenyl)-8-((2-methylpiperidin-1-yl)methyl)-4H-chromen-4-one (2t)

White solid, yield: 17%; mp 141–143 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.32 (s, 1H), 7.87 (d, *J* = 9.9 Hz, 1H), 7.50 (d, *J* = 9.9 Hz, 2H), 6.98 (d, *J* = 9 Hz, 2H), 6.80 (d, *J* = 9.9 Hz, 1H), 4.31–4.26 (d, *J* = 15 Hz, 1H), 3.90–3.85 (d, *J* = 15 Hz, 1H), 3.77 (s, 3H), 2.83 (d, *J* = 12.3 Hz, 1H), 2.66 (brs, 1H), 2.33 (t, *J* = 9.6 Hz, 1H), 1.48–1.35 (m, 6H), 1.15 (d, *J* = 6.3 Hz, 3H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.1, 164.8, 159.5, 155.2, 153.1, 130.6, 125.9, 124.8, 123.5, 116.4, 115.9, 114.1, 108.6, 56.6, 55.6, 51.7, 50.4, 33.8, 26.9, 25.7, 22.6. HR-MS (ESI) Calcd for C₂₃H₂₅NO₄ [M + H]⁺, 380.1862, found: 380.2130.

7-hydroxy-8-((4-(hydroxymethyl)piperidin-1-yl)methyl)-3-(4-methoxyphenyl)-4H-chromen-4-one (2r)

White solid, yield: 27%; mp 195–197 °C; ¹H NMR (300 MHz, DMSO-d₆) δ 8.33 (s, 1H), 7.90 (d, *J* = 8.7 Hz, 1H), 7.50 (d, *J* = 8.7 Hz, 2H), 6.98 (d, *J* = 9 Hz, 2H), 6.82 (d, *J* = 9 Hz, 1H), 3.98 (s, 2H), 3.77 (s, 3H), 3.27 (d, *J* = 6 Hz, 2H), 2.99 (d, *J* = 11.1 Hz, 2H), 2.25 (t,

J = 11.4 Hz, 2H), 1.73 (d, *J* = 12.9 Hz, 2H), 1.23 (brs, 1H), 1.11 (m, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 175.2, 164.3, 159.5, 155.5, 153.2, 130.6, 126.2, 124.7, 123.5, 116.5, 115.8, 114.1, 108.4, 66.00, 55.7, 53.3, 52.9, 38.2, 28.9. HR-MS (ESI) Calcd for C₂₃H₂₅NO₄ [M + H]⁺, 380.1862, found: 380.2130.

Table 2. Structures and activities of compounds 2a–2t.

Compound	R ₃	R ₄	Inhibit rate (%) at 10 μM	IC ₅₀ (μM)	Compd.	R ₃	R ₄	Inhibit rate (%) at 10 μM	IC ₅₀ (μM)
2a		OH	-1.85		2k		OCH ₃	-23.17	
2b		OH	-32.59		2l		OCH ₃	-1.51	
2c		OH	39.59	14.24 ± 0.08	2m		OCH ₃	14.56	
2d		OH	-57.52		2n		OCH ₃	1.72	
2e		OH	-0.82		2o		OCH ₃	66.77	4.71 ± 0.01
2f		OH	18.72		2p		OCH ₃	-9.49	
2g		OH	-8.12		2q		OCH ₃	-9.56	
2h		OH	81.83	3.84 ± 0.04	2r		OCH ₃	-55.09	
2i		OH	-20.42		2s		OCH ₃	-42.75	
2j		OCH ₃	-9.69		2t		OCH ₃	2.53	

Bold values indicates that the compound has a high inhibit rate (%) at 10 μM and is able to posses an IC₅₀.

Bioassay studies

Cell lines and cell culture

The cell-based histamine receptor 3 (H₃R) assay was carried out based on β-lactamase complementation technology. The H3-bla U2OS cells (Invitrogen, Invitrogen, Waltham, Massachusetts) stably expressed two fusion proteins, as well as a β-lactamase reporter gene under the control of a UAS response element. The first fusion protein was human H₃R linked to a Gal4-VP16 transcription factor through the TEV protease site, and the other was the β-arrestin/TEV protease fusion protein. H₃-bla U2OS cells were cultured in McCoy's 5 A Medium supplemented with 10% foetal bovine serum (FBS; Gibco, Shanghai, China) at 37 °C in a humidified atmosphere with 5% CO₂. To each well in a 384-well plate was seeded exponentially growing cells in a density of 6.5 × 10³ cells/mL in 32 μL of media. The plate was incubated at 37 °C, 18–24 h, 5% CO₂ for cell adherence.

Fluorescent H₃R assay

Stock solutions of test compounds (10 mM) were prepared in DMSO and then diluted 100 times in media. Cells were exposed

to 4 μL of test compounds and the control compound thioperamide (Sigma-Aldrich, St. Louis, Missouri) for 30 min and then stimulated with 4 μL of methylhistamine at 400 nM (Sigma-Aldrich) for 5 h. Then, 8 μL of LiveBLazer-FRET B/G Substrate (CCF4-AM; Invitrogen) was added and incubation continued for 2 h. Plates were subjected to the fluorescence reading with a Spectra Max M5 microplate reader (BioTek, Winooski, Vermont); equipped with 410 nm excitation and 460 nm and 530 nm emission filters. The inhibition percentage was calculated based on the fluorescence according to the following equation: % inhibition = (Model_{Response ratio} - Compound_{Response ratio}) / Model_{Response ratio}. And IC₅₀ values were determined from log concentration – inhibition curves. At least three separate tests were carried out.

Molecular docking

We chose the most active compounds for molecular docking studies to predict how molecules and proteins work. A homology modelling of H₃R was built as our previous report²⁵. The 3D structure of compound **2h** was built using DS MODELER (Discovery Studio 2016, BIOVIA Inc, San Diego, CA) and evaluated the model

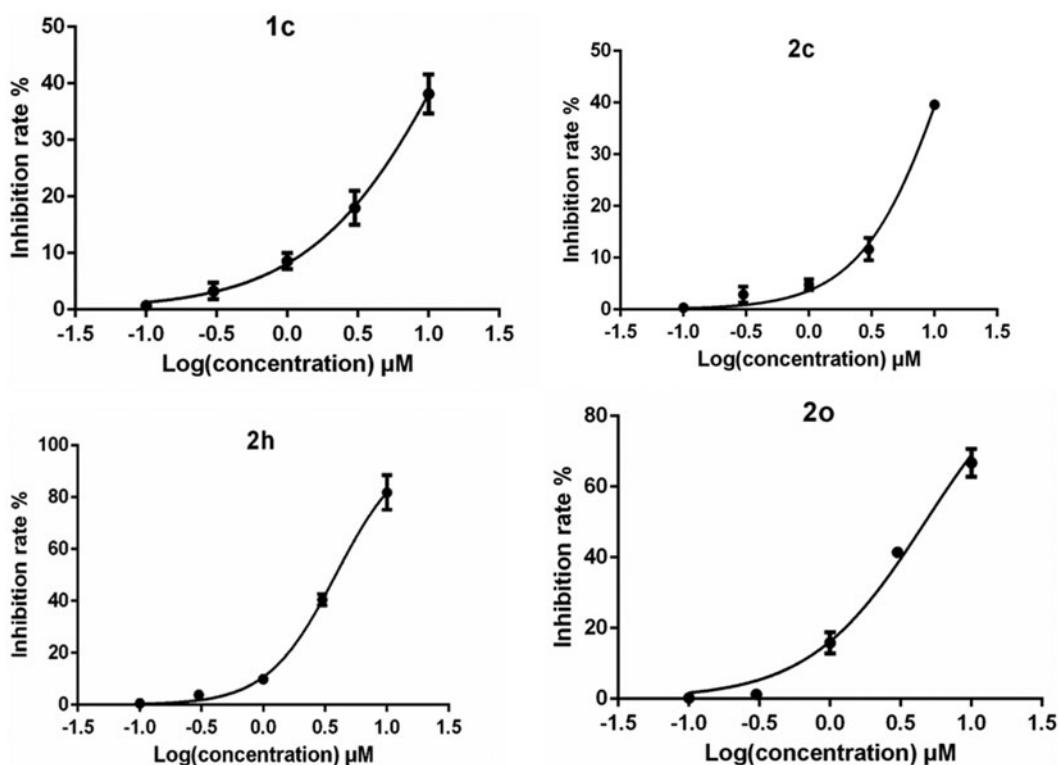


Figure 2. The IC₅₀ of the four compounds (**1c**, **2c**, **2h**, and **2o**) showed good H₃R inhibitory activity.

according to the PDF Total Energy and the Profile-3D procedure. Flexible Docking was used for the docking procedure. The 3D model of H₃R with the lowest PDF Total Energy was chosen for docking. Water and the cognate ligand (doxepin) were removed from the model, and hydrogen atoms were added to amino acid residues. The binding mode was shown by DS visualizer.

Results and discussion

Structure–activity relationship

The compounds were initially evaluated for inhibition rate on H₃R at a fixed concentration of 10 μM (Tables 1 and 2). Of the 27 compounds evaluated, four compounds (**1c**, **2c**, **2h**, **2o**) performed satisfactory inhibitory effect (Figure 2). According to reports in the literature, H₃R inhibitory activities were increased by the introduction of pyrrolidine and piperidine to the iso-flavone scaffold¹⁰. Thus, we introduced various pyrrolidine, piperidine, piperazine and morpholine moieties onto 6- or 8-position of iso-flavone. The results for series 1 are shown in Table 1. The advantage of piperidine groups outweighed pyrrolidine moieties. As for substituted piperazine and morpholine moieties, the subsequent data did not give satisfactory results. Then, we modified daidzein and formononetin with substituted piperidine and pyrrolidine fragments. It should be noted that further steric modification on piperidine was detrimental for the inhibitory activities. For example, 4-hydroxy-methyl, 3-hydroxy piperidine (compound **2b**, **2e**) attached to the structure of daidzein led the inhibitory activity to decrease. However, the 2-methyl piperidine group (compound **2h**) showed very strong inhibition. Interestingly, for formononetin, 3-methyl piperidine (compound **2o**) and pyrrolidine (compound **2m**) fragments showed unexpected inhibitory effect. Structurally,

substituted piperidine (such as methyl- and hydroxyl-) or pyrrolidine groups could improve bioactivity but bulky substitutions may hinder binding H₃ pockets, namely, binding affinity would loss¹⁰. Comparing different iso-flavone structures, even though 4'-hydroxy or 4'-methoxy benzene ring in 4-position of iso-flavone scaffold showed significant fluctuation in bioactivity level according to the data shown in Table 2, in most cases, daidzein derivatives have advantages over formononetin as H₃R antagonists, for example, compound **2c** vs **2l**; **2h** vs **2t**.

Binding modes of compound **2h**

The results showed that compound **2h** bound with H₃R through multiple sites (Figure 3). The protonated amine of the pyridine group interacted with Glu206 through a salt bridge. The Tyr-115 and Phe-198 bound to the aromatic ring structural on one side of compound **2h** by π–π T-shape interactions. In addition to this, compound **2h** also formed hydrophobic interaction, π-sigma and π-alkyl interaction with the protein.

Conclusions

In this work, two series of iso-flavone derivatives were synthesised and evaluated for their H₃R inhibitory activity. Ultimately, we identified compound **1c**, **2c**, **2h**, **2o** which possessed favourable H₃R inhibitory activity. The structure–activity relationship (SAR) study identified the piperazine group in the 8-position of iso-flavone was essential for the H₃R inhibitory activity (compound **2h**). Molecular docking showed 2'-methyl piperidine substituent of **2h** formed a salt bridge and hydrophobic interactions with the protein. In this paper, we creatively modified the iso-flavone derivatives and determined this scaffold possessing the potential H₃R

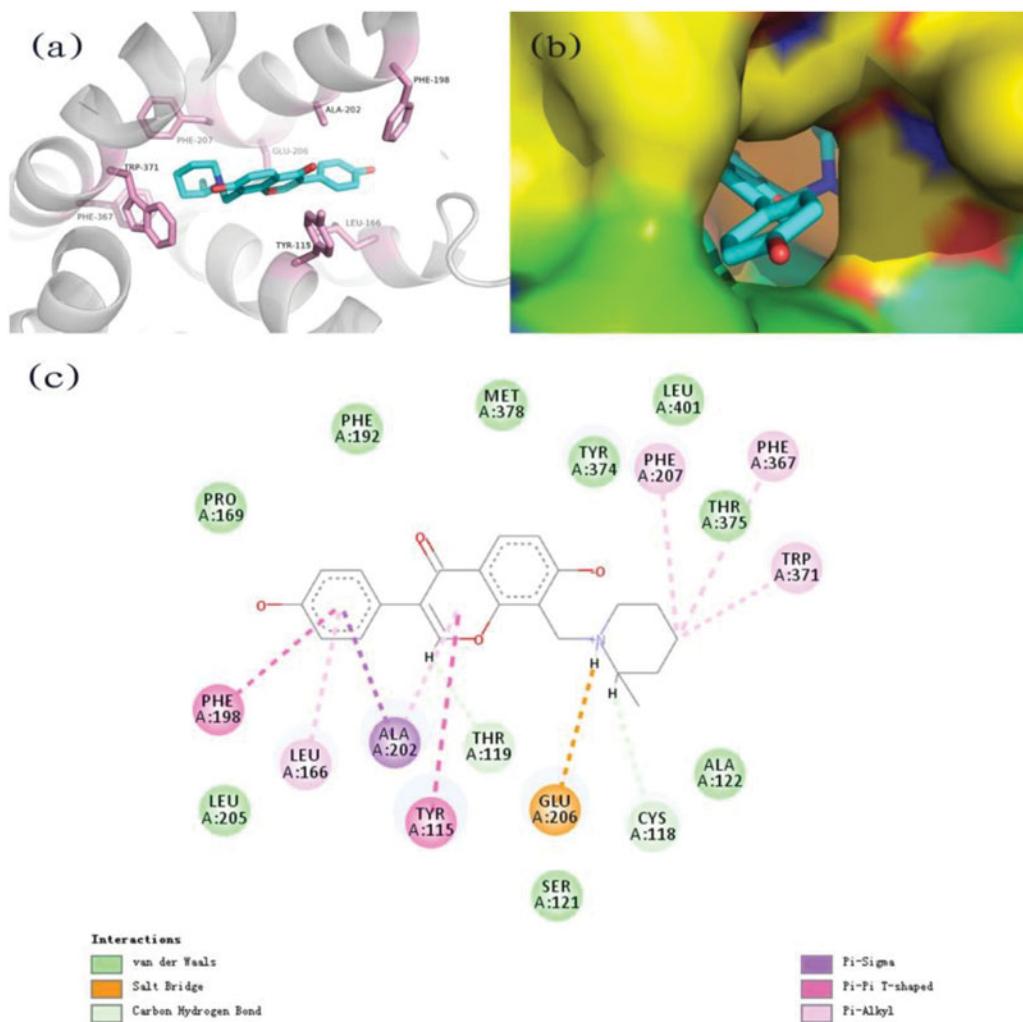


Figure 3. (a) The predicted binding mode of compound 2h with H₃R; (b) the binding pocket of H₃R by the surface representation; (c) 2D schematic diagram of potential interactions between compound 2h and H₃R.

inhibitory activity. Moreover, these results also provided clues for the development of novel H₃R antagonists.

Disclosure statement

No potential conflict of interest was reported by the authors.

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