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# Chromosomal mapping of rRNA genes, core histone genes and telomeric sequences in *Brachidontes puniceus* and *Brachidontes rodriguezii* (Bivalvia, Mytilidae)

Concepción Pérez-García, Jorge Guerra-Varela, Paloma Morán, Juan J Pasantes\*

## Abstract

**Background:** Chromosome rearrangements are an important part of the speciation process in many taxa. The study of chromosome evolution in bivalves is hampered by the absence of clear chromosomal banding patterns and the similarity in both chromosome size and morphology. For this reason, obtaining good chromosome markers is essential for reliable karyotypic comparisons. To begin this task, the chromosomes of the mussels *Brachidontes puniceus* and *B. rodriguezii* were studied by means of fluorochrome staining and fluorescent *in situ* hybridization (FISH).

**Results:** *Brachidontes puniceus* and *B. rodriguezii* both have  $2n = 32$  chromosomes but differing karyotype composition. Vertebrate-type telomeric sequences appear at both ends of every single chromosome. *B. puniceus* presents a single terminal major rRNA gene cluster on a chromosome pair while *B. rodriguezii* shows two. Both mussels present two 5S rDNA and two core histone gene clusters intercalary located on the long arms of two chromosome pairs. Double and triple-FISH experiments demonstrated that one of the 5S rDNA and one of the major rDNA clusters appear on the same chromosome pair in *B. rodriguezii* but not in *B. puniceus*. On the other hand, the second 5S rDNA cluster is located in one of the chromosome pairs also bearing one of the core histone gene clusters in the two mussel species.

**Conclusion:** Knowledge of the chromosomal distribution of these sequences in the two species of *Brachidontes* is a first step in the understanding of the role of chromosome changes on bivalve evolution.

## Background

Over recent years, an increasing number of molecular studies have focused on the elucidation of the phylogenetic relationships among species of bivalves in order to further reveal their evolutionary history [1]. Nevertheless, cytogenetic comparisons between species remain quite rare mainly due to difficulties in obtaining good chromosome banding patterns [2]. This fact, together with the small differences in chromosome size and morphology in many species of bivalves, makes the unequivocal identification of each chromosome pair very difficult and therefore, complicates cytogenetic comparisons.

Fluorescent *in situ* hybridization (FISH) allows combining cytogenetic and molecular techniques, improving our understanding of genome organization. The chromosomal mapping of different types of DNA sequences in bivalves has greatly increased over the last years [2-4]. Although the information gained so far is of great importance in the analysis of chromosome evolution in bivalves, the data obtained are still scarce and focused on a limited number of DNA sequences, mainly in species with commercial interest.

Most FISH reports in bivalves have been concentrated on the chromosomal localization of major rRNA genes rather than on other gene families such as 5S rDNAs and histone genes [2,5]. Major rDNA has been located in about 50 species of bivalves [2,5], including seven species of the family Mytilidae [5-11]. 5S rDNA has

\* Correspondence: pasantes@uvigo.es  
Dpto. Bioquímica, Xenética e Inmunoloxía. Universidade de Vigo. Vigo, Spain

been located in a small number of species belonging to the families Cardiidae [12], Mytilidae [5,11], Ostreidae [13,14] and Pectinidae [15-20]. The organization of the histone genes has been characterized in two species of mussels [21-24], one scallop [25] and three species of venerid clams [26] but their chromosomal locations are only known in two species of mussels [5,22,24] and four species of Pectinidae [27]. Telomeric sequences have been characterized and located in a few species of bivalves, including two species of Mytilidae [5,6,28].

*Brachidontes (senegalensis) puniceus* (Gmelin 1791) is a small mussel species native to the African Atlantic coast, from Mauritania to Angola. *B. rodriguezii* (d'Orbigny 1846) is a medium-sized mussel distributed from Rio Grande do Sul (Brazil) to the northern part of Patagonia (Argentina) on the Atlantic coast of South America. Karyological data on the genus *Brachidontes* is limited to the knowledge of the karyotypes and the chromosomal location of the major rDNA in *B. rodriguezii* [8] and *B. pharaonis* [10]. In order to expand the cytogenetic information on the family Mytilidae, we analyzed the chromosomes of *B. puniceus* and *B. rodriguezii* by means of 4',6-diamidino-2-phenylindole (DAPI)/propidium iodide (PI) and chromomycin A3 (CMA)/DAPI fluorescence staining and fluorescent *in situ* hybridization (FISH) with 28S rDNA, 5S rDNA, core histone genes and telomeric probes.

## Methods

### Animals

Samples of *Brachidontes puniceus* were collected at São Vicente (Cape Verde) and São Tomé (São Tomé and Príncipe). *B. rodriguezii* specimens were collected at Mar del Plata (Argentina). Mussels were maintained in the laboratory in tanks of 5 L of aerated, filtered seawater at  $18 \pm 1^\circ\text{C}$  and fed on a mixture of microalgae cells (*Isochrysis galbana* and *Tetraselmis suecica*) for at least 15 days in order to promote both somatic growth and gonadic maturation.

### Chromosome preparation and fluorochrome staining

Chromosome preparations were obtained following the technique described previously [6]. Juvenile specimens were housed in 0.5 L beakers and exposed to colchicine (0.005%) for 12 h. Gill and mantle tissues were excised and immersed in 50% and 25% seawater for 90 min and fixed with ethanol/acetic acid for 1 h. Chromosome spreads were obtained by dissociating small pieces of tissue in 60% acetic acid, pipetting suspension drops onto slides heated to  $55^\circ\text{C}$  and air-drying.

Some chromosome preparations were stained for 2 h with CMA ( $0.25 \text{ mg mL}^{-1}$ ) and counterstained with DAPI ( $0.14 \text{ } \mu\text{g mL}^{-1}$ ) for 8 min. Once washed with distilled water, slides were air-dried and mounted with

antifade (Vectashield, Vector). After visualization and photography, preparations were washed and re-stained with a combination of DAPI ( $0.14 \text{ } \mu\text{g mL}^{-1}$ ) and PI ( $0.07 \text{ } \mu\text{g mL}^{-1}$ ). The slides were then washed in distilled water, air-dried, mounted in antifade and photographed again.

Chromosome counts and karyotype analysis were performed in 10 specimens (5 males, 5 females) of each species. In order to determine the karyotype of *Brachidontes puniceus*, ten good metaphase plates were used to construct karyotypes. Chromosome and arm lengths were carefully measured and relative lengths and centromeric indices were calculated. Chromosome nomenclature follows Levan et al. [29].

### DNA extraction, PCR amplification and probe labeling

Total DNA was extracted following the method of Estoup et al. [30] with minor modifications. About 3 mg of adductor muscle tissue was homogenized in 0.4 mL of a pre-warmed ( $60^\circ\text{C}$ ) 10% Chelex 100 (BioRad) solution. After adding pronase ( $1.4 \text{ mg mL}^{-1}$ ) and incubating for 1 h at  $60^\circ\text{C}$  in agitation, the extracted DNA was stored at  $4^\circ\text{C}$ .

FISH probes were obtained by PCR. Amplifications were performed in 20  $\mu\text{l}$  of a solution containing 50 ng DNA,  $1\times$  PCR buffer, 0.5 mM each dNTP, 2.5 mM  $\text{MgCl}_2$ , 1  $\mu\text{M}$  each primer and 1 U BIOTAQ DNA polymerase (Bioline). The combinations of primers used in the amplification appear in Table 1. Universal primers (*LR10R*, *LR12*) were used to amplify a fragment of the 28S rRNA gene of the major rDNA repeat [31]. For the 5S rDNA amplification, primers were designed from the sequence of the 5S rRNA of *Mytilus edulis* [32]. The amplifications of the H2B/H2A and the H3 histone genes were performed using primers designed from the sequences of the histone genes of *Mytilus edulis* [23] and those described by Giribet and Distel [1], respectively. After 5 min denaturation at  $95^\circ\text{C}$ , 30 cycles of amplification were performed using the conditions that are shown in Table 1. A final extension step of 7 min at  $72^\circ\text{C}$  was applied. All reactions were performed in a GeneAmp PCR system 9700 (Applied Biosystem) and PCR products were examined by electrophoresis on a 2% agarose gel. Single products were obtained after amplification using each set of primers. 28S rDNA was labeled with biotin 16-UTP and/or digoxigenin-UTP ( $10\times$  DIG Labeling Mix, Roche) by nick translation. 5S rDNA and histone genes were directly labeled by PCR either with biotin-16-UTP or digoxigenin-UTP. The labeled PCR products were precipitated before FISH.

### Fluorescent *in situ* hybridization (FISH)

Single and double FISH experiments were performed following the methods published previously [33] with

**Table 1 Primers and parameters used in the PCR**

Gene	Sequences of the primers	Denaturation	Annealing	Elongation
28S	LR10R: 5'GACCCTGTTGAGCTTGA3' LR12: 5'GACTTAGAGGCGTTCAG3'	95°C, 20 s	48°C, 20 s	72°C, 30 s
5S	F: 5'CAACGTGATATGGTCGTAGAC3' R: 5'AACACCGGTTCTCGTCCGATC3'	95°C, 30 s	44°C, 30 s	72°C, 30 s
h2ba	F: 5'TCCCGAGATGTGATGGTAGA3' R: 5'AGTACAGCCTGGATGTTTGGTAA3'	95°C, 30 s	45°C, 30 s	72°C, 40 s
h3	F: 5'ATGGCTCGTACCAAGCAGACVGC3' R: 5'ATATCCTTRGGCATRATRGTGAC3'	95°C, 15 s	48°C, 15 s	72°C, 15 s

minor modifications. Preparations were denatured at 69°C for 2 min and hybridized overnight at 37°C. Signal detection was carried out with fluorescein avidin and biotinylated anti-avidin for the biotinylated probes and with mouse antidigoxigenin, goat anti-mouse rhodamine and rabbit anti-goat rhodamine for the digoxigenin-labeled probes. Slides were counterstained with DAPI and mounted in antifade.

In order to map three probes on the same metaphase plates, two consecutive FISH were performed. In the first one, a histone gene probe labeled with biotin and a 5S rDNA probe labeled with digoxigenin were employed. After visualization and photography, the preparations were re-hybridized using a 28S rDNA probe simultaneously labeled by nick translation with both biotin and digoxigenin and the same metaphase plates were visualized and photographed again. In addition, we also performed FISH with a telomeric (CCCTAA)<sub>3</sub> peptide nucleic acid (PNA) probe (Applied Biosystems) following the protocol indicated by the supplier.

Slide visualization and photography were performed with a Nikon Eclipse-800 microscope equipped with an epifluorescence system. Separated images for each fluorochrome were obtained with a Sensys CCD camera (Photometrics) connected to the microscope. Camera control and digital image acquisition employed a Power Macintosh computer. Pseudocoloring and merging of the images was performed with Adobe Photoshop.

## Results

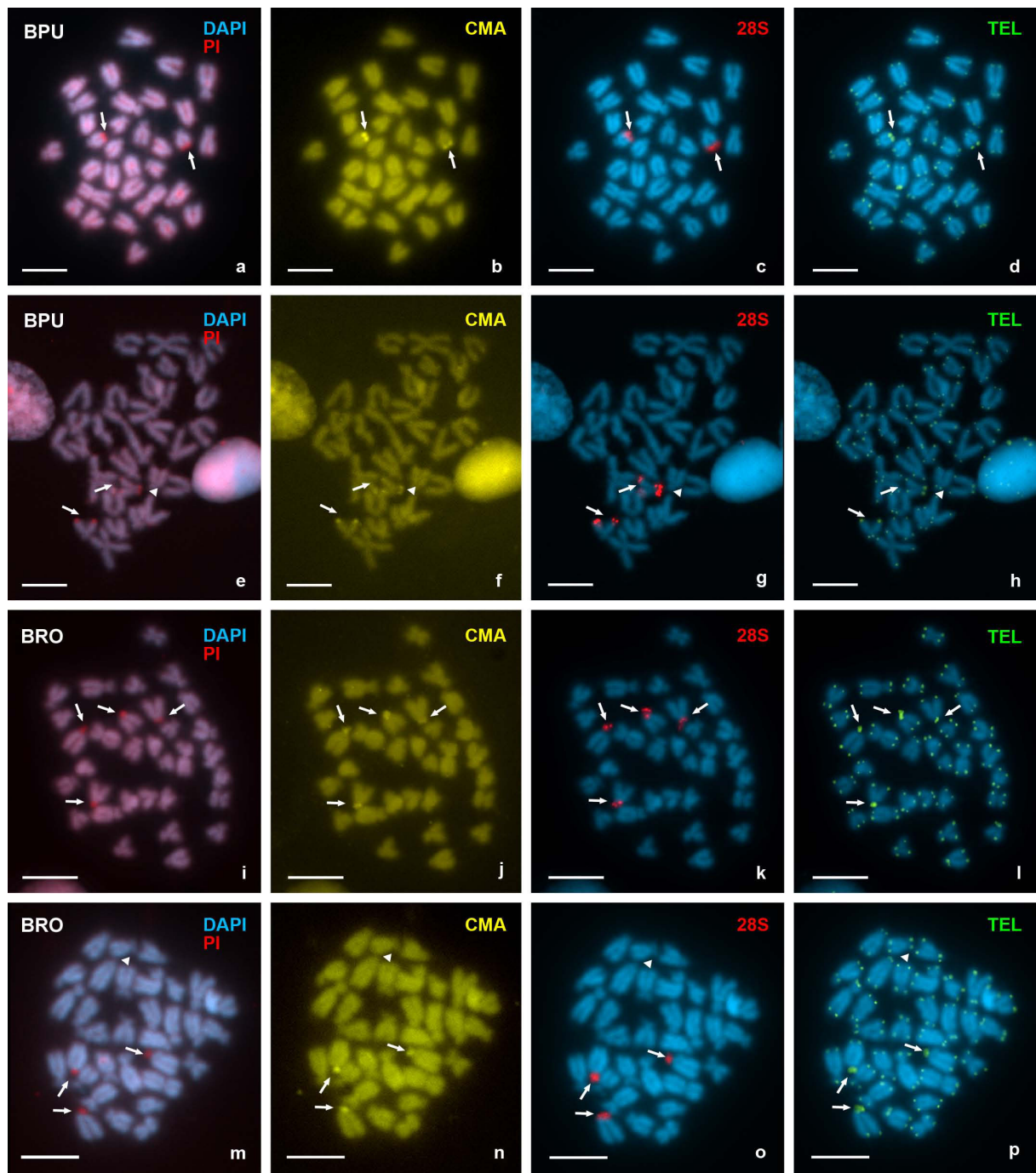
Coinciding with previously reported data [8], *Brachidontes rodriguezii* presents a diploid chromosome number of 2n = 32 (Figure 1i-p) and its karyotype is composed of 2 metacentric, 12 subtelocentric and 2 telo/subtelocentric chromosome pairs (Figure 2j). *B. puniceus* also shows a diploid complement of 2n = 32 chromosomes (Figure 1a-h). The karyotype is composed of 1 metacentric, 3 submetacentric, 4 submeta/subtelocentric, 4 subtelocentric and 4 telocentric chromosome pairs (Figure 2i). A summary of the results obtained after measuring the DAPI-stained chromosomes of 10 of the best complete metaphase plates is given in Table 2.

No differences were observed between the São Vicente and São Tomé populations.

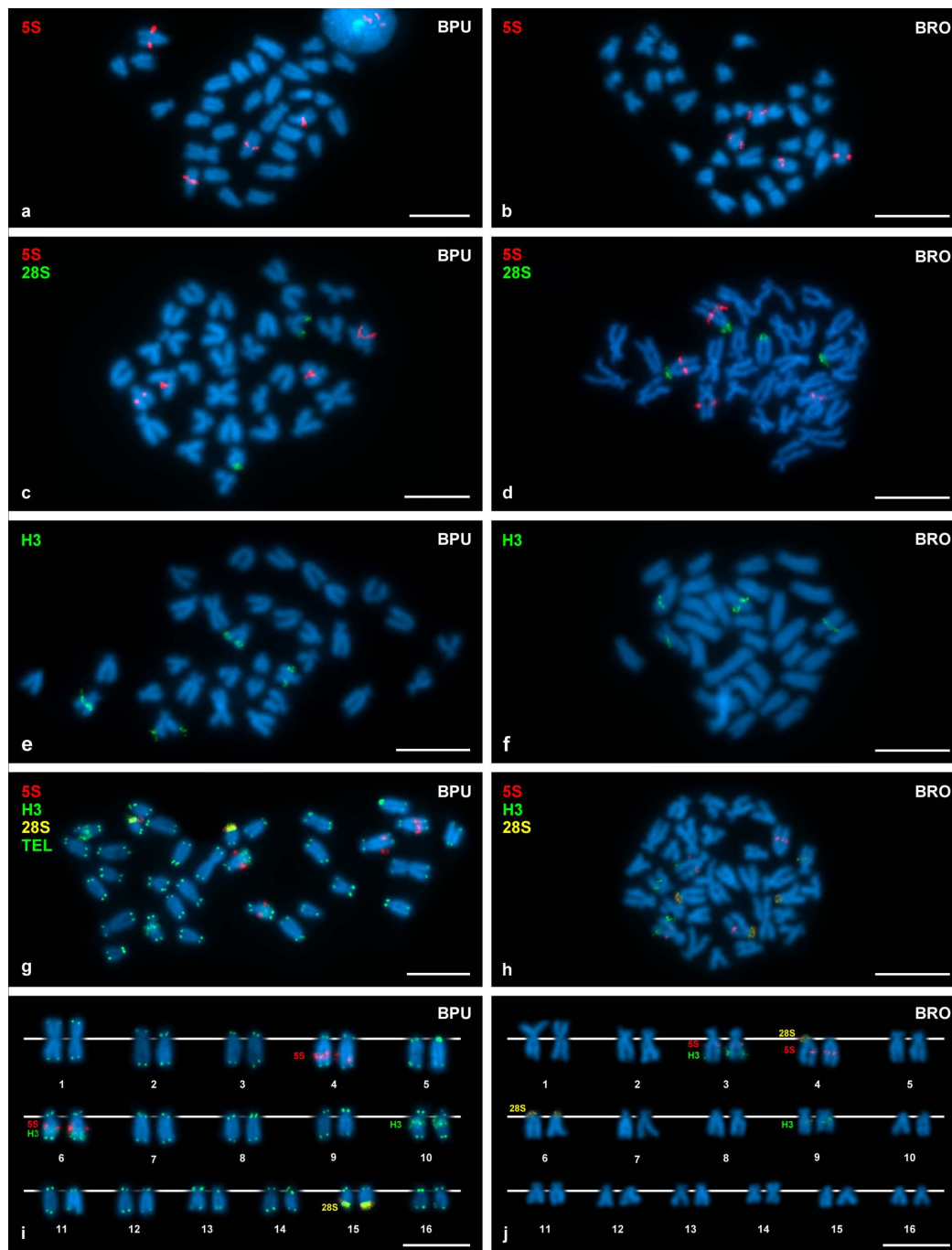
Telomeric sequences were detected using a vertebrate telomeric (CCCTAA)<sub>3</sub> PNA probe. Single distinct terminal signals appear at the ends of both sister chromatids of every mitotic chromosome in the two species (Figure 1d, h, l, p). No additional interstitial telomeric sequences were observed. Although the sizes of the telomeric signals do not show a high variability, consistently bigger signals appear at terminal regions of the long arm of one of the members of one chromosome pair in *Brachidontes puniceus* (Figure 1d) and of the short arms of two chromosome pairs in *B. rodriguezii* (Figure 1l). One of the *B. puniceus* mussels presents an additional bigger telomeric signal (Figure 1h) and one of the *B. rodriguezii* specimens shows only three bigger telomeric signals (Figure 1p).

DAPI/PI staining of mitotic metaphase plates reveals the presence of a DAPI negative (DAPI-) terminal region on the long arm of the subtelocentric chromosome pair # 15 in *Brachidontes puniceus* (Figure 1a, e). One of the mussels shows an additional DAPI- band on the short arm of the chromosome pair also bearing the standard band (Figure 1e). CMA staining of the same metaphase plates (Figure 1b, f) shows that these regions are CMA bright (CMA+). *B. rodriguezii* presents DAPI-/CMA+ regions on the short arms of the two telo/subtelocentric chromosome pairs, # 4 and # 6 (Figure 1i, j, m, n), but one of the individuals analyzed shows only three DAPI-/CMA+ positive regions in every single cell (Figure 1m, n).

FISH signals for the major ribosomal genes were analyzed in 130 complete metaphase plates in five *Brachidontes puniceus* mussels and in 150 metaphases in five specimens of *B. rodriguezii*. *B. puniceus* presents two major rDNA signals on a terminal region of the long arm of the subtelocentric chromosome pair # 15 (Figure 1c). These signals coincide with the DAPI-/CMA+ regions (Figure 1a, b). The mussel showing an additional DAPI-/CMA+ region (Figure 1e, f) also presents a third 28S rDNA signal at the same position (Figure 1g). Most *B. rodriguezii* mussels present four major ribosomal signals, coincident with the DAPI-/CMA+ regions, on the



**Figure 1** Sequential DAPI/PI and CMA staining followed by FISH using 28S rDNA (28S) and telomeric (TEL) probes on *Brachidontes puniceus* (BPU) and *Brachidontes rodriguezii* (BRO) chromosomes. Sequential staining of the same metaphase plates with DAPI/PI (a, e) and CMA (b, f) shows the presence of two GC-rich (DAPI dull/CMA bright) regions located at the ends of the long arms on one chromosome pair in *B. puniceus* (arrows in a, b, e, f). One of the *B. puniceus* specimens shows an additional GC-rich region on the short arm of one of the members of the chromosome pair bearing the standard band (arrowhead in e, f). In *B. rodriguezii* four GC-rich regions appear on the short arms of two chromosome pairs (arrows in i, j) but one of the mussels shows only three of these bands (arrows in m, n) lacking the fourth (arrowhead in m, n). FISH using a 28S rDNA probe (digoxigenin, rhodamine, red) gives signals at the CMA+ positions in both *B. puniceus* (arrows in c, g, arrowhead in g) and *B. rodriguezii* (arrows in k, o). Note the absence of the fourth major rDNA signal in o (arrowhead). FISH using a PNA-telomeric probe (fluorescein, green) gives signals at both ends of every sister chromatid. Telomeric signals situated on the GC-rich NORs are consistently bigger than the rest of the telomeric signals and their sizes are similar to the sizes of the 28S signals (arrows in d, h, l, p). Scale bars, 5  $\mu$ m.



**Figure 2 Chromosomal location of 5S rDNA, major rDNA and core histone genes on *Brachidontes puniceus* and *Brachidontes rodriguezii*.** FISH using a 5S rDNA probe (digoxigenin, rhodamine, red) reveals the presence of two clusters of 5S rDNA on two chromosome pairs in both *B. puniceus* (BPU, **a**) and *B. rodriguezii* (BRO, **b**). Double-FISH experiments using a 5S rDNA probe (digoxigenin, rhodamine, red) and a 28S rDNA probe (biotin, fluorescein, green) show that major and minor gene clusters in *B. puniceus* (**c**) are to be found on different chromosome pairs but that in *B. rodriguezii* (**d**) one of the chromosome pairs bearing a 5S rDNA signal also bears one of the major rDNA signals. FISH using a histone H3 gene probe (biotin, fluorescein, green) shows two clusters of core histone genes on the long arms of two chromosome pairs in both *B. puniceus* (**e**) and *B. rodriguezii* (**f**). Triple hybridization (**g**, **h**) using a digoxigenin labeled 5S rDNA probe (red), a biotin labeled histone H3 gene probe (green) and a double (biotin and digoxigenin) labeled 28S rDNA probe (yellow, as a result of the mixture of green and red) reveals that the histone gene clusters are independent of the major rDNA clusters in *B. puniceus* (**g**, **i**) and in *B. rodriguezii* (**h**, **j**). In both species, one of the chromosome pairs bearing a 5S rDNA signal also carries one of the core histone gene signals (**g**, **h**, **i**, **j**). The metaphase plate in **g**, **i** was also re-hybridized using a PNA telomeric probe (fluorescein, green). Scale bars, 5  $\mu$ m.

**Table 2 Relative lengths and centromeric indices of *Brachidontes puniceus* chromosomes**

Pair	Relative length		Centromeric index		Type
	Mean	SE	Mean	SE	
1	8.72	0.53	43.48	0.78	m
2	7.99	0.50	25.48	1.09	sm/st
3	7.20	0.27	16.21	0.90	st
4	7.05	0.34	20.56	0.77	st
5	6.96	0.54	8.59	0.55	t
6	6.56	0.40	24.66	1.20	sm/st
7	6.46	0.51	7.11	0.45	t
8	6.16	0.46	7.96	0.78	t
9	6.11	0.24	24.38	0.87	sm/st
10	5.84	0.28	27.94	0.84	sm
11	5.83	0.43	8.10	0.64	t
12	5.52	0.29	24.75	0.82	sm/st
13	5.26	0.27	29.39	1.01	sm
14	5.13	0.23	21.76	0.91	St
15	4.74	0.31	22.02	0.61	St
16	4.49	0.33	30.43	0.76	Sm

m: metacentric, sm: submetacentric, st: subtelocentric; t: telocentric; sm/st: submeta/subtelocentric

short arms of the telo/subtelocentric chromosome pairs # 4 and # 6 (Figure 1i, j, k), but the one showing only three DAPI-/CMA+ regions (Figure 1m, n) also presents only three major rDNA signals (Figure 1o). The consistently bigger telomeric signals appear at the DAPI-/CMA+ NORs.

FISH signals corresponding to the minor ribosomal genes were studied in 111 metaphases in *B. puniceus* and 150 in *B. rodriguezii*. As shown in Figure 2a, 5S rDNA signals are interstitially located on the long arms of chromosome pairs # 4 and # 6 in *B. puniceus*. In *B. rodriguezii* 5S rDNA signals are also intercalary located on the long arms of two chromosome pairs, # 3 and # 4 (Figure 2b). The number of minor rDNA signals was always 4 in *B. puniceus* but 2 or 4 in different metaphase plates in *B. rodriguezii*. Double-color FISH experiments (Figure 2c, d) confirmed the presence of major and minor rDNA clusters on chromosome pair # 4 in *B. rodriguezii* and therefore allowed the study of the distribution of the variable number of 5S rDNA signals (Table 3). A 66.67% of the metaphase plates showed signals only on the NOR-bearing chromosome pair # 4. The remaining metaphases showed 4 signals on pairs # 3 and # 4 (32%), being always weaker those in chromosome # 3 (Figure 2b, d, h, j), or only 2 signals on pair # 3 (1.33%).

Core histone genes were mapped by FISH using both histone H3 and histone H2B/H2A probes. FISH signals were analyzed in at least 20 complete metaphases per mussel in 5 individuals in each species. Core histone

**Table 3 Number and proportion of metaphases showing 2 or 4 5S rDNA signals on chromosome pairs # 3, not bearing 28S rDNA signals, and # 4, also bearing 28S rDNA signals, on five specimens of *Brachidontes rodriguezii***

Mussel	2 signals, # 3		2 signals, # 4		4 signals, # 3, # 4		Total
	n	%	n	%	n	%	
1	0	0.00	15	60.00	10	40.00	25
2	1	2.33	29	67.44	13	30.23	43
3	1	4.17	8	33.33	15	62.50	24
4	0	0.00	24	82.76	5	17.24	29
5	0	0.00	24	82.76	5	17.24	29
Total	2	1.33	100	66.67	48	32.00	150

gene signals are interstitial to the long arms of chromosome pairs # 6 and # 10 in *B. puniceus* (Figure 2e). The signal in pair # 10 is closer to the centromere. *B. rodriguezii* also shows intercalary signals on two chromosome pairs, # 3 and # 9 (Figure 2f). In this species the signal on chromosome pair # 9 is closer to the centromere. Double-color FISH experiments using H3 and H2B/H2A histone gene probes labeled differently always showed coincident signals (not shown). In order to confirm the location of the histone gene clusters in relation to the minor and major rDNA clusters, double and triple-color FISH experiments were performed. One of the core histone gene clusters and one of the 5S rDNA clusters are on the same chromosome pair (Figure 2g, h) both in *B. puniceus* (pair # 6) and *B. rodriguezii* (# 3). The relative positions of the major rDNA, 5S rDNA and histone gene clusters are indicated on the karyotypes that appear on Figure 2i, j.

## Discussion

The chromosomal characterization of the mussels of the family Mytilidae includes the knowledge of mitotic chromosome numbers and karyotypes in 32 species. Six of these species show the same diploid chromosome number ( $2n = 32$ ) present in both *Brachidontes puniceus* and *B. rodriguezii* [8]. On the contrary, the only other species of this genus in which the karyotype has been described, *B. pharaonis*, shows  $2n = 28$  chromosomes [10]. The variability of chromosome numbers between species of the same genus is widespread in the family Mytilidae; with the exception of *Mytilus*, all genera of mussels in which karyotypes have been determined for more than one species show differences in chromosome numbers [34]. On the other hand, the coincidence in chromosome numbers in *B. puniceus* and *B. rodriguezii* is accompanied by considerable differences in karyotype composition.

The detection of the vertebrate telomeric (TTAGGG)<sub>n</sub> repeat at chromosome ends in *Brachidontes puniceus*

and *B. rodriguezii* agrees with results obtained in chromosomes of other species of bivalves [2], including the mussels *Mytilus galloprovincialis* [6,28] and *Perumytilus purpuratus* [5]. On the contrary, the interstitial telomeric sequences clearly detected by FISH in *P. purpuratus* [5] and presumably appearing in *M. galloprovincialis* [28] were not detected in these two species of *Brachidontes*. Although in bivalves telomeric sequences have only been isolated and characterized in *Donax trunculus* [28], the hybridization results obtained so far indicate that the bivalve telomeres are composed of tandem repeats of the hexanucleotide which also constitutes the vertebrate telomeric sequence.

The presence of major rDNA signals on CMA bright regions terminally located on a single chromosome pair in *Brachidontes puniceus* and on two chromosome pairs in *B. rodriguezii* is concordant with results obtained in other bivalves. NORs have been located in around 50 species of bivalves [2,5] and are located at terminal positions on one to three chromosome pairs. In the family Mytilidae, the position of major rRNA genes is known in seven species. One NOR-bearing chromosome pair has been detected in *B. pharaonis* [10], two in *Mytilus galloprovincialis* [6], *M. edulis* [11], *B. rodriguezii* [8] and *Perumytilus purpuratus* [5], and three in *M. trossulus* and *M. californianus* [9]. The species of the genus *Brachidontes* show differences both in number and chromosomal location of the NORs. The single major ribosomal gene cluster appears at a terminal position on the long arm of one small subtelocentric chromosome pair in both *B. puniceus* and *B. pharaonis* [10]. However, the two major ribosomal gene clusters of *B. rodriguezii* are spread over the whole short arms of the two subtelocentric/telocentric chromosomes pairs [8].

The occurrence of individuals showing a different number of rDNA clusters to those usually found in the species has been described in many taxa, including humans [35]. In this sense, the presence of an additional NOR in one of the *B. puniceus* specimens is not remarkable and could be the result of a translocation process as proposed for human ectopic NORs [35]. The absence of one of the major rDNA clusters in one chromosome in *B. rodriguezii* can be attributed to unequal crossover events [36].

The mostly coincident FISH signal sizes obtained at the terminal NORs after using 28S rDNA and telomeric probes in both *B. rodriguezii* and *B. puniceus* might indicate that some telomeric repeats are interspersed within the major rDNA repeats; fiber-FISH and molecular analysis are necessary to confirm or discard this hypothesis. Although not common, interspersion of telomeric and major ribosomal DNA sequences has been observed in bivalves, such as the mussel *Mytilus galloprovincialis* [6] and the scallop *Patinopecten yessoensis* [20], and in

other organisms such as fishes [37] and mammals [38]. The meaning of such organization is not well understood but a functional role in nucleolus organization for tandem repeats has been proposed [39].

The chromosomal location of 5S rDNA clusters is only known in 15 species of bivalves [2,5]. In most of these species, minor rDNAs usually appear at interstitial loci [18-20]. The presence of two 5S rDNA clusters in both *Brachidontes puniceus* and *B. rodriguezii* differs from the results obtained in *Mytilus edulis* and *M. galloprovincialis* [11], in which four 5S rDNA clusters appear on three chromosome pairs, and *Perumytilus purpuratus* [5], showing three 5S rDNA clusters on two chromosome pairs. Regarding the other species of bivalves in which the location of these sequences is known, the Ostreidae [13,14] and two species of Pectinidae [15,17] also show two 5S rDNA clusters. On the contrary, *Cerastoderma edule* (Cardiidae) presents five clusters of 5S rDNA on five different chromosome pairs [12] and most species of Pectinidae show a single 5S rDNA cluster [16,18,20].

The variation in number of 5S rDNA FISH signals detected in *Brachidontes rodriguezii* is mostly due to the presence or absence of signals in chromosome pair # 3. The small size of these signals probably indicates that the number of 5S rDNA repeats at this locus is close to the minimum necessary for the signal to be detected. Similar variations in the number of 5S rDNA signals have been detected in two other species of Mytilidae, *Mytilus galloprovincialis* and *M. edulis* [11].

Our results demonstrate that major and minor rDNA clusters are on different chromosome pairs in *Brachidontes puniceus* but that in *B. rodriguezii* one of the NOR-bearing chromosome pairs also carries one of the 5S rDNA clusters. This is also the case in *Perumytilus purpuratus* [5] and *Chlamys farreri* [16] but differs from the presence of both major and 5S rDNA clusters on different chromosome pairs in the rest of the bivalve species studied so far.

Histone genes are usually organized as tandem repeats in invertebrate genomes [40]. These clusters can be composed by a copy of each one of the core histone genes or both core and linker histone genes. Histone gene arrangement in bivalves has been studied in the mussels *Mytilus edulis* [21,23] and *M. galloprovincialis* [22,24], and the scallop *Chlamys farreri* [25]. All of them present clusters of core histone genes ordered in the same way thus probably indicating a conserved arrangement. Our results showing hybridization signals at the same positions after FISH using H3 gene and H2B/H2A gene probes point also to this situation in *Brachidontes puniceus* and *B. rodriguezii*. Histone genes are known to be located in two chromosome pairs in the mussel *M. galloprovincialis* [24] and the scallop

*Patinopecten yessoensis* [27] but forming a single cluster in other mussel species, *Perumytilus purpuratus* [5], and in the scallops *Argopecten irradians*, *Chlamys farreri* and *C. nobilis* [27]. Therefore, in the family Mytilidae, the presence of two histone gene clusters in these two *Brachidontes* species coincides with the situation in *M. galloprovincialis* [24] but differs from the one in *P. purpuratus* [5].

One of the histone gene clusters and one of the 5S rDNA clusters appear on the long arm of the same chromosome pair in both *Brachidontes puniceus* and *B. rodriguezii*. In addition, the relative positions of these clusters with respect to the centromere are similar in both species being the histone genes distal to the 5S rDNA. Although these facts seem to indicate that this chromosome is conserved, further analyses are necessary in order to determine their homology. The only other species of bivalves in which the position of these two sequences was investigated together is *Perumytilus purpuratus* [5]. In contrast with *B. puniceus* and *B. rodriguezii*, in *P. purpuratus* the single histone gene cluster and the 5S rDNA clusters are located in different chromosome pairs.

The similarities in the chromosomal distribution of major rDNA, 5S rDNA and core histone gene clusters found in *Brachidontes puniceus*, *B. rodriguezii* and *Perumytilus purpuratus* [5] are not shared by *Mytilus galloprovincialis* [11,22,24] and confirm the closeness of *P. purpuratus* to the species of *Brachidontes*, therefore keeping open the possibility of its assignation to this genus but discarding that *P. purpuratus*, *B. rodriguezii* and *B. darwinianus* belongs to the same taxon [41].

## Conclusions

The FISH results obtained in this work after using major rDNA, 5S rDNA and histone gene probes allow unequivocally identifying four of the 16 chromosome pairs that compose the karyotypes of *Brachidontes puniceus* and *B. rodriguezii*. Additional research is in progress in order to find more markers that allow chromosome identification in Mytilidae and therefore to gain insights on chromosome evolution in this family.

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## Authors' contributions

CPG undertook most of the cytogenetic procedures and collaborated on the molecular work, the bibliographic review, and the writing of this paper. JGV participated in the first steps of the amplification, cloning, characterization and mapping of the 5S rDNA.

PM participated in developing the molecular techniques and helped in the writing. JJP coordinated the study, helped in developing the laboratory techniques and cytogenetic analyses and coordinated the writing of the manuscript. All authors read and approved the final manuscript.

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