



The Controversial Role of Irisin in Clinical Management of Coronary Heart Disease

Wen-Lu Ou-Yang¹, Bei Guo¹, Feng Xu¹, Xiao Lin², Fu-Xing-Zi Li¹, Su-Kang Shan¹, Feng Wu³, Yi Wang¹, Ming-Hui Zheng¹, Qiu-Shuang Xu¹ and Ling-Qing Yuan^{1*}

¹ Department of Metabolism and Endocrinology, National Clinical Research Center for Metabolic Diseases, Hunan Provincial Key Laboratory of Metabolic Bone Diseases, The Second Xiangya Hospital, Central South University, Changsha, China, ² Department of Radiology, The Second Xiangya Hospital, Central South University, Changsha, China, ³ Department of Pathology, The Second Xiangya Hospital, Central South University, Changsha, China

Irisin, a PGC1 α -dependent myokine, was once believed to have beneficial effects induced by exercise. Since its first discovery of adipose browning in 2012, multiple studies have been trying to explore the metabolic functions of irisin, such as glucose and lipid metabolism. However, recently many studies with irisin concentration measuring were doubt for methodological problems, which may account for the continuous inconsistencies. New tools like recombinant irisin and gene-knockout mice are required to reconfirm the questioned functions of irisin. In this paper, we make a critical introduction to the latest researches concerning the relationship between irisin and coronary heart disease, which includes atherosclerosis, stable angina pectoris and acute coronary syndromes. These studies provided various controversial evidence of short and longterm monitoring and therapeutic effect from molecular cellular mechanisms, *in vivo* experiments and epidemiological investigation. But with ambiguities, irisin still has a long way to go to identify its functions in the clinical management.

OPEN ACCESS

Edited by:

Rita De Matteis, University of Urbino Carlo Bo, Italy

Reviewed by:

Michael Lichtenauer, Paracelsus Medical University, Austria Vincenzo Calderone, University of Pisa, Italy

*Correspondence:

Ling-Qing Yuan allenylq@csu.edu.cn

Specialty section:

This article was submitted to Cellular Endocrinology, a section of the journal Frontiers in Endocrinology

Received: 09 March 2021 Accepted: 17 June 2021 Published: 01 July 2021

Citation:

Ou-Yang W-L, Guo B, Xu F, Lin X, Li F-X-Z, Shan S-K, Wu F, Wang Y, Zheng M-H, Xu Q-S and Yuan L-Q (2021) The Controversial Role of Irisin in Clinical Management of Coronary Heart Disease. Front. Endocrinol. 12:678309. doi: 10.3389/fendo.2021.678309 Keywords: irisin, myokine, coronary heart disease, infarction, atherosclerosis

INTRODUCTION

Irisin

Irisin is a relatively newly discovered myokine, a small protein with 112 amino acids. Its precursor, fibronectin type III domain-containing protein 5 (FNDC5), is regulated by the transcription factor PGC1 α (1). In human, irisin is mainly secreted by cardiomyocytes and skeletal muscle cells, and a small amount is also distributed in adipose tissue, brain, liver, spleen, testis and other tissue (2). Irisin is highly conserved among different species, homology of which between mouse and human irisin is almost 100% (1). However, Raschke et al. found that start codon of FNDC5 gene is different, and doubted whether it's appropriate to translate all the beneficial effect observed in mice to humans (3).

The production level of irisin is affected by many factors. Exercise, cold, age, BMI, glucose, lipid metabolism level, and other cytokines are all related to its serum concentration (4-8). It was initially widely considered as an exercise-induced myokine, whose circulating level was affected by type and frequency of physical activity (1, 9, 10). Jedrychowski's study (11) validated it by quantitation mass spectrometry (MS). However, inconsistencies emerged to doubt whether the stimulation of exercise

1

worked. Browning of white adipose tissue (WAT), a major effect of irisin, was observed only in inguinal WAT of training mice and attenuated in FNDC5 knockout mice (12, 13), while cold exposure was effective for all adipose depots in murine (14). When it comes to human, current evidence are not qualified enough to confirm the irisin's responses to exercise (15–20). The disagreement may blame to the insufficient detection accuracy considering the minor amounts of irisin's expression (0.3 ng/mL in mice (21) and 3-4 ng/mL in humans (11) of serum level measured by MS) (15, 22–24).

In recent years, thousands of papers are trying to find out irisin's engages in multiple biological processes of our body (Figure 1). First of all, browning the white adipose tissue was recognized at the same time irisin was discovered in 2012 (1). However, as the discussion above, exercise has little effect on the stimulation of irisin's browning function in human (15-20). Despite the disputes on stimulators like exercise, primary mouse adipocytes were observed browning effect when directly incubated with recombinant C-terminal FNDC5 peptide (25). Then in brown adipose tissue, the uncoupling protein 1 (UCP1) could release the storage of energy generated by oxidation in the form of heat and reduce the production of ATP simultaneously (26). These metabolic effects could be achieved by activating p38 MAPK and ERK pathways (27). The discovery of the PGC1\alpha-FNDC5-irisin axis was the theoretical basis for the latter studies concerning energy metabolism mechanisms, which made irisin closely related to obesity, T2DM, and other metabolic syndromes (1, 28-30) (Figure 2). While the browning function is now facing challenges, similar questions would also be raised in related area such as fat storage, energy consumption and insulin sensitivity.

In addition, FNDC5 are associated with cognitive function of the brain through brain-derived neurotrophic factor (BDNF). BDNF is the main mediator of exercise effect on the hippocampus, which can promote the growth, differentiation and repair of neurons (31). Wrann showed a positive correlation between FNDC5 and BDNF in exercising mice (31). Increasing the brain level of FNDC5/irisin can enhance synaptic plasticity and memory in Alzheimer's disease mouse models (32), and stimulate the STAT3 signal transduction pathway required for sensory neuron development (33). However, the studies didn't explain the mechanism of serum irisin to cross the blood-brain barrier, and applied much higher concentration of irisin than physiological level, which later showed no obvious effects on mouse hippocampal neuronal cells (33). Thus, evidence is not enough to demonstrate the direct relationship of irisin and BDNF. Besides, in terms of bone metabolism, irisin were reported to activate proliferation and differentiation of osteoblast through p38 and ERK (34-36), and reduce the extent of osteoporosis and muscle atrophy (37, 38). But according to H Kim, the increase of trabecular bone volume and BMD were observed in female FNDC5-KO mice (39). So, we still couldn't figure out whether irisin helps in bone formation or not. Also, the increase of irisin can be detected in a variety of tumors. In cell assays, irisin showed its potential to inhibit epithelial-mesenchymal transition and cancer cell invasion via PI3K/Akt pathway, enhance the activity of caspase-3 and caspase-7 to induce cancer cell apoptosis, and inhibit the activity of NF-kb to reduce inflammation (40, 41). Moreover, irisin produced by cardiomyocytes is much more than that of skeletal muscle cells (42). Thus, studies on treatment of irisin

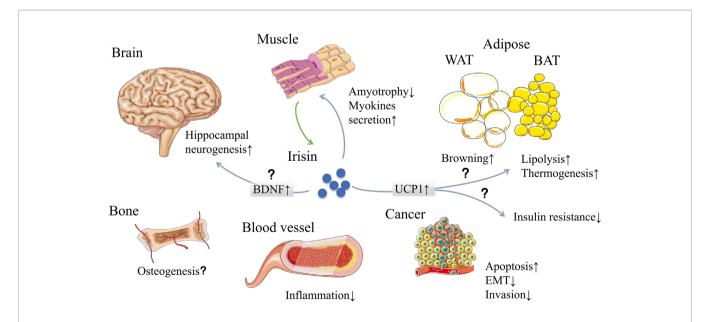
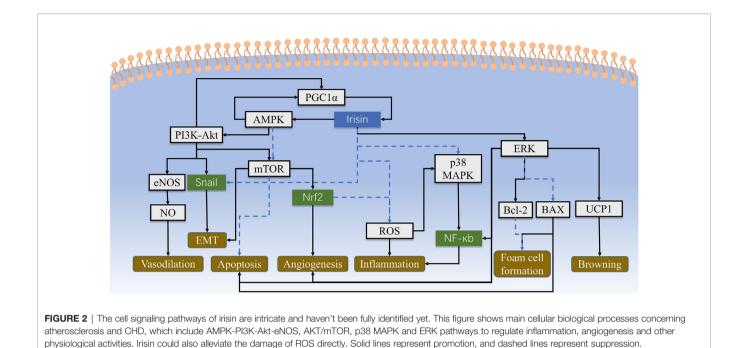


FIGURE 1 | Irisin, mainly secreted by muscle, may have effects on multiple organs and tissues. The classic functions of irisin are browning white adipose tissue and generating heat. Other effects include muscle feedback, neurogenesis, osteogenesis, inflammation reduction, cancer suppression and so on. However, some of the functions are now facing challenges due to the measuring problems and interspecies genetic differences. WAT, white adipose tissue; BAT, brown adipose tissue; BDNF, brain-derived neurotrophic factor; UCP1, uncoupling protein 1; EMT, epithelial-mesenchymal transition. [The elements were produced using Servier Medical Art (https://smart.servier.com/) with some adaption].



showed its effects on cardiac hypertrophy, mainly through mTOR activation to prompt protective autophagy (43–45). In the development of cardiovascular disease, irisin may participate in this process through various non-metabolic ways, which will be discussed in the latter part.

However, there has been a big shadow cast on the measurement of irisin. In 2015, Albrecht E showed that the antibodies used in commercial ELISA tests are unspecific with a huge variation in plasma concentrations (46), which could explain the huge difference between the early rough estimate by ELISA kits (1) and recent measurement by quantitative MS from the same group (21). Calibrated MS was regarded as a "gold standard" to detect irisin (47), but Albrecht further doubt the repeatability of LC/MS method for detection of extremely low plasma concentrations (48). Additionally, due to the extremely low amounts of irisin in samples (11, 21), it's hard to purify it for gain-of-function experiments. Yet new approaches like recombinant irisin and FNDC5 knockout mice could provide more direct evidence to verify the functions of irisin. Most studies creating FNDC5 knockout mice found no obvious difference of multiple biological process under normal conditions (21, 43, 49, 50). But they would suffer a lot more in several abnormal conditions compared to wild type, such as fasting and high-fat diet (13, 43, 49, 51). Thus, we would put more focus on the studies with new approaches to draw a rather reliable picture of irisin.

Coronary Heart Disease (CHD)

CHD is one of the most common cardiovascular diseases. The prevalence and fatality rate has been increasing consecutively year after year, seriously endangering people's health (52). Due to coronary artery spasm or atherosclerosis, myocardial cells suffer

from ischemia and hypoxia, leading to functional or structural pathological changes of heart. As early as irisin was first discovered, some scholars (53) suggested that there may be certain crosstalk between irisin and the cardiovascular system. As time passes by, a number of evidences accumulated to support this speculation. Irisin has participated in the development of CHD in a variety of ways, which is likely to lay a theoretical foundation for the clinical application of irisin as a biomarker or treatment.

IRISIN IN ATHEROSCLEROSIS

Atherosclerosis, a chronic inflammatory disease, is the most common pathogenesis of CHD and a variety of vascular diseases. The etiology has not been clear yet, usually considered to be closely associated with lifestyle and caused by multiple factors such as lipid metabolism disorder and endothelial injury (54). Irisin participates in the regulation of this process through direct or indirect ways (such as glucose and lipid metabolism).

Controversies on Irisin's Expression of Atherosclerosis

In many studies conducted in different sample sizes and different populations, the relationship between serum irisin levels and atherosclerosis is not consistent. Deng W et al. (55) conducted a cross-sectional study on the serum irisin concentration and coronary atherosclerosis index (CAI) of 350 coronary artery disease (CAD) patients and 214 healthy participants, finding that the serum irisin concentration was significantly different in the two groups of subjects. The lower the serum irisin concentration in CAD patients, the higher the CAI. Parallel to this result, in the study of cardiovascular complications, Khorasani (56) and Saadeldin (57) respectively confirmed that the serum irisin concentration was negatively correlated with the degree of coronary atherosclerosis in patients with type 2 diabetes. Besides, in some special populations [such as patients with dialysis (58) or Behcet's Syndrome (59)], the detection of carotid artery intima-media thickness (IMT) was also found in line with this pattern. Irisin's regulation on glucose and lipid metabolism is speculated to be the main contributor to this negative relationship, since hyperglycemia, insulin resistance and obesity are closely associated with atherosclerosis (55, 59, 60). Another possible explanation is the cross-talks between irisin and other molecules such as sclerostin or adipocytokines (57, 58).

However, some studies stood on the opposite side. As early as 2014, in a study of 192 European ancestry healthy people, it was found that irisin was positively correlated with the subjects' carotid artery IMT (60). Moreover, a 25-year cohort study by Kwaniewska M (61) and a cross-sectional study of men with HIV infection (62) supported this result. In this regard, they explained that irisin may indirectly participate in the formation of atherosclerosis through fat or insulin (60, 62). And as the disease advances, the target cell's sensitivity to irisin or insulin decreases, so that muscle/fat tissue "compensatively" releases more irisin to achieve the same effect as before. There is also an opinion, arguing that the difference in baseline between different studies leads to the opposite result: the various degrees of obesity of the study subjects may interfere with the results. As in nonobese subject, the serum irisin is mainly derived from muscle, while in the obese subject the proportion of fat source increases (63). Similar to the degrees of obesity, Moreno found that the high value of plasminogen activator inhibitor-1 (PAI-1) at baseline could also have a significant impact on the results, which could even count as an independent biomarker related to the thickness of the carotid artery IMT in patients (62). In a study based on a Japanese male population (64), the authors found that if cardiometabolic risk factors are taken into account, irisin had nothing to do with the prevalence of coronary artery calcification, but was related to progression. They believed that irisin may affect other unknown ways to cause this result except for traditional cardiometabolic risk factors such as fat and blood glucose. The inaccuracy measurement of irisin due to the questioned commercial ELISA may also count for the inconsistencies (46).

In short, the current studies lack consistency in the relationship between the level of irisin and the degree of atherosclerosis. And the correlation in multivariate analysis as well as the measurement is not sufficiently valid, which makes it difficult for irisin to be a predictive or clinical indicator of CHD.

The Physiopathological Function of Irisin in Atherosclerosis

The cellular biological processes related to atherosclerosis mainly involves vascular endothelial damage and foam cell formation, which then leads to wall thickening, endothelial narrowing, and eventually fibrous cap rupture (54).

As mentioned above, irisin is surmised to participate in atherosclerosis by some indirect ways such as influencing insulin release. In recent years, a series of studies have found that irisin can also directly regulate endothelial function though suppressing the inflammation and oxidation and encouraging the proliferation of cells. On one hand, Lu J (65) proved that irisin can activate AMPK-PI3K-Akt-eNOS signaling pathway in diabetic mice, inhibit high glucose-induced apoptosis in human umbilical vein endothelial cells (HUVECs), and increase the expression of antioxidant enzymes to reduce inflammation and oxidative stress of endothelial cells. Furthermore, Han F et al. (66) conducted similar experiments on healthy obese mice fed by a high-cholesterol diet and found that the activation of AMPKeNOS signaling pathway by irisin can be dependent or independent of adiponectin. On the other hand, Zhang Y et al. (67) elaborated in mice that irisin could reduce the vascular damage induced by oxLDL. In animal experiment, irisin can regulate the p38 MAPK/NF-KB signaling pathway by inhibiting the generation of reactive oxygen species (ROS) and the nuclear translocation of NF-KB. It can also inhibit PKC-B/NADPH oxidase, down-regulate inflammatory factors to reduce oxidative/nitrative stresses, thus alleviating vascular endothelial inflammation (67, 68). In addition, Zhang M et al. found that irisin can increase cell viability, cell migration and capillary formation in apolipoprotein E (ApoE) deficient mice. This may be achieved by the up-regulation of microRNA126-5p in the ERK signaling pathway so as to maintain the stability of endothelial cells and promote proliferation (69). In an in vitro experiment exploring diabetes complicated with vascular diseases, it reported that irisin can inhibit ROS-NLRP3 inflammasome signaling, thereby slowing down the process of diabetes-related endothelial inflammation and other impairment (70). Protective functions of irisin are also found to lessen endothelial damage at an early stage. Irisin could suppress the expression of TNFa-induced VCAM-1 in HUVECs (71). A recent survey in obese children found that irisin can inhibit the expression of hsCPR, ICAM-1 and E-selectin in endothelial cells, indicating that irisin may have an effect on cardiovascular disease before symptoms show up (72).

Besides endothelial cell, irisin could also act on macrophages to slow down the formation of plaques. By up-regulating Bcl-2, down-regulating the expression of Bax and caspase-3, irisin could prevent macrophages from turning into foam cells, and inhibit cell apoptosis (67). Zheng G et al. (73) further found lipid accumulation in macrophages cultured in vitro and oxLDLrelated apoptosis were reduced under the interference of irisin, which may be related to the inhibition of PERK/eIF2α/CHOP and ATF6/CHOP endoplasmic reticulum stress signaling pathways. All of these demonstrated that irisin does engage in multiple mechanisms in atherosclerosis, either by acting on endotheliocyte or on macrophage. However, some of the experiments were conducted on mice, which may not be qualified enough to translate the protective effects to human. And it must be noted that the serum irisin level is much lower than the applied dose in experiment, thus challenging the physiological functions of irisin under normal circumstances.

Therapeutic Effect of Irisin in Atherosclerosis

Various cell experiments have confirmed that irisin has a positive effect on vascular endothelial cells, while the therapeutic effect of irisin have shine a light on atherosclerosis in animal experiments. In apolipoprotein E-Null diabetic mice (69) or obese mice induced by high-cholesterol diet (66), the endotheliumdependent dilation function (EDV) was improved with the injection of irisin, while the endothelial cell apoptosis and the area of atherosclerotic plaque were pleasantly found to have a significant reduction. In the carotid artery partial ligation model (67, 69), systemic application of irisin can also inhibit the formation of the new carotid artery intima, for the reason that irisin may promote endothelial cell proliferation and inhibit monocyte recruitment and lipid deposition. These evidences show that irisin has potentiality to contribute to the therapy of atherosclerosis, while further researches are compulsory to provide sufficient clinical support.

IRISIN IN CHD

CHD can be divided into two categories: stable angina pectoris (SAP) and acute coronary syndromes (CHD) according to its pathogenesis and clinical manifestations. Based on current studies, there is controversy over the role of irisin in the pathogenesis and repair of myocardial cells, and the specific mechanism is still unclear. But when it comes to treatment, most studies tend to support the protective effects of irisin on CHD.

Fluctuation of Irisin During CHD Limited Research of Irisin in SAP

SAP is the most common clinical type of CHD and is at the initial stage of CHD development. For many reasons, the insufficient blood supply of the coronary arteries leads to transient ischemia and hypoxia in the myocardium, often accompanied by cardiac dysfunction, but myocardial necrosis rarely occurs (74). There are not enough studies conducted on irisin in SAP, the quality of which are at different levels. In a small cross-sectional study, Efe et al. (75) found that the serum irisin level of stable coronary artery disease patients with a higher SYNTAX score (\geq 23) was significantly decreased compared to the group with low SYNTAX score or the healthy control group. They believed that serum irisin level can be used as an independent indicator to observe the severity of SAP, which is related to the degree of coronary stenosis in patients (76). Some previous researches (42, 77) also partly agreed with it. However, 24 hours after SAP patients having percutaneous coronary intervention (PCI), the serum irisin concentration was found to be lower than that of the control group (76). And the results of Park et al. (77) showed that compared with other groups, SAP patients with low preoperative serum irisin levels had a significantly higher proportion of no CHD events within 12 months after PCI. It should be noted that the measurement of irisin in those studies were all conducted by commercial ELISA, which reduced credibility of the potential

connection between irisin and SAP, emphasizing the need for more clinical research.

Irisin's Engagement in Acute Coronary Syndromes (ACS)

ACS happens after plaque rupture or erosion on the basis of coronary atherosclerosis, which leads to vascular embolism, a sharp decrease in oxygen supply to the myocardium, and then damage. The degree of risk and prognosis are related to the location, area and speed of the establishment of collateral circulation (78–80). Many studies believe that irisin is involved in the occurrence of ACS and can be used as a therapeutic drug.

Several studies reported that the concentration fluctuation of irisin had a common pattern in different stages of myocardial infarction (MI). Kuloglu et al. (81) used isoproterenol on rats to induce myocardial infarction. The serum irisin level of MI rats decreased significantly in the first 2 hours, and slowly increased afterwards, but never returned to the baseline within 24 hours during monitoring. In the meanwhile, the synthesis level in tissue like skeletal muscle, liver and kidney decreased within 1-4 hours after MI, and recovered after 6 hours. Further monitoring of myocardial infarction rats showed that the serum irisin level was highly positively correlated with QRS duration, amplitude and TAS, while highly negatively correlated with ST-elevation, QTc, CK-MB, troponin and MDA (82). Similar changing pattern can be observed in human that the concentration of irisin fluctuates during ACS, but the point of time was delayed (83, 84). Similarly, the serum irisin measured 6 hours after PCI also decreased from baseline (76). However, during coronary artery bypass surgery, the serum irisin concentration increased from induction to the removal of the crossclamp, and then gradually decreased as the patient rewarmed (85).

According to this phenomenon, Kuloglu (81) believed that it had something to do with the uncoupling effect of irisin. and may have a protective effect on cardiomyocytes. In addition, excessive irisin will lead to increased mitochondrial respiration and fatty acid oxidation, which leads to increased oxygen consumption and ROS, and up-regulates the level of caspase 9 which enhances cell apoptosis (1). On the contrary, the appropriate amount of irisin can activate Opa1-induced mitochondrial autophagy, alleviate oxidative stress and maintain the vitality of cardiomyocytes after myocardial infarction (86). Irisin can also lead to bradycardia through effects on the nucleus ambiguous (87), which helps reduce the energy consumption of the heart.

However, different perspectives interpret the possible mechanisms behind the changes of irisin during ACS. One point is that the reduction of irisin is a passive performance rather than an active regulation. When cardiomyocytes were treated with TNF α or IL-1 β , the expression of FNDC5 protein decreased significantly, indicating that inflammatory factors could inhibit the secretion of irisin (6). And MI can restrain the expression of PPAR- α and PPAR- γ (nuclear receptors), thereby preventing the interaction of PGC1- α with a variety of transcription factors, resulting in a decrease in the synthesis of FNDC5 (irisin precursor) (88). Another explanation suggested that the decrease in irisin level is the reason rather than the result of the decrease in coronary blood flow. Since previous animal experiments have proven that irisin can

regulate endothelial function and induce vasodilation (89), low levels of irisin may lead to decreased vasodilatation then reduced blood flow, forming a vicious circle.

It should be noticed that the commercial ELISA kits used to detect irisin in those studies were lack of specificity (46). Besides the shadow of questioned measuring methods, the mechanism of this fluctuation still remains controversial. Sharp decrease of irisin may protect mitochondria and allow cardiomyocytes to save energy in such an ischemic environment, which seems to be a self-protection mechanism of our body. But it may be also a consequence of damage and even assist in the progression of ACS. While there is still no reliable method of irisin measurement, irisin is blocked to become a marker showing the process of ACS.

Invalid Prognostic Indicator

For the long-term stage after the occurrence of ACS, the level and function of circulating irisin are inconclusive. In terms of surgery, the level of irisin at 6 months after PCI was higher than the baseline (53). The relatively high level of irisin was positively correlated with the incidence of postoperative major adverse cardiovascular events (MACE), especially angina, the risk of which increased by almost 4 times (hazard ratio=3.96) (83). On the contrary, a meta-analysis involving 867 patients and 700 controls showed that patients with cardiovascular disease or atherosclerosis had significantly lower levels of irisin (90). In 2014, a cross-sectional study by Emanuele et al. (91) found that serum irisin in young subjects who had suffered from MI was significantly lower than healthy controls. However, in an agerelated study (53), there was no significant difference in irisin levels among people in their sixties, regardless of whether they had had ACS. The measurement problem may be the main reason to rationale the difference (46). Moreover, The circulating level of irisin in the body is influenced by fat, exercise, diet and so many other factors, and the level of irisin tends to increase randomly due to aging (92). Meanwhile, systematic errors such as inconsistent experimental design and detection levels cannot be ruled out.

There are also studies concerning the distribution of irisin genotypes. The G allele of rs3480 and the A allele of rs726344 are significantly related to cardiometabolic risk factors. Besides, the two SNPs are associated with each other (93). Dyslipidemia is a major initiator of atherosclerosis, and ApoE is particularly important to maintain the normal metabolism of lipoprotein (94, 95). Fuku et al. reported that FNDC5 rs16835198 was associated with APOE $\epsilon 2/\epsilon 4$ allele (96), while direct evidence towards atherosclerosis needed to be further explored. Hence future investigations are needed to explore genetic risk factors for myocardial infarction.

Nevertheless, limited researches still couldn't provide sufficient materials to make a conclusion, which suggests that a more critical experimental design and use of validated measurement are required to draw a rounded picture for prognosis of irisin.

Irisin's Potential in Treatment After Infarction

There is an interesting phenomenon in the distribution of irisin during myocardial infarction. Although serum irisin level decreases in the acute phase, the expression of irisin contrarily increases in connective tissues around the heart (81, 97), suggesting that irisin may participate in the follow-up process of MI such as restoration. As described above (67), we have mentioned that irisin can reduce the damage of vascular endothelial cells by inhibiting the production of ROS. Similar events may also occur in cardiomyocytes. Peng Q et al. (98) found that irisin can promote cell proliferation while reducing the production of HO-induced ROS and cell apoptosis in cardiomyocytes, which may be achieved by the miR-19b/AKT/mTOR signaling pathway. Zhao et al. (99) tried to find the connection between irisin and HDAC4 in vitro. HDAC4 is produced by H9c2 cardioblasts and is up-regulated in oxidative stress. Its overexpression can lead to mitochondrial dysfunction and even cell death (100). And irisin can induce HDAC4 degradation through ubiquitination modification to protect cardiomyocytes. Partially different from the results above (99), Moscoso et al. (97) only observed the protective effect of irisin under hypoxic conditions. They believed that under the condition of myocardial infarction, hypoxia and the compensatory adrenergic response would cause a lipotoxic environment and create conditions for apoptosis (101). Consequently, irisin could activate the Akt signaling pathway to resist apoptosis induced by lipotoxicity.

When the blood flow of myocardial tissue recanalized after ischemic injury, ischemia/reperfusion injury (I/R) often occurs. Irisin has shown its ability to reduce I/R injury *in vitro* and *in vivo* experiments. Mitochondrial dysfunction leads to the release of a large amount of ROS, and this oxidative stress response is one of the core steps of I/R. Irisin could inhibit the opening of mPTP (mitochondrial permeability transition pore), reduce mitochondrial swelling and increase the activity of superoxide dismutase 2 (SOD2) by upregulating mitochondrial ubiquitin ligase (MITOL/MARCH5), so as to protect mitochondria and reduce the oxidative stress response induced by mitochondria *in vitro* (99, 102, 103).

Studies on the therapeutic effect of irisin are still ongoing. Irisin was used for treatment in mice with coronary artery obstruction caused by ligation. It indicated that irisin could activate the ERK signaling pathway and promote endothelial cell migration, thereby accelerating angiogenesis at the infarct border zone, reducing infarct size and fibrosis and improving ventricular diastolic function in animal model (104, 105), which may involve the AKT/mTOR/S6K1/Nrf2 pathway (106). The lack of those protective effects was also observed in FNDC5knockout mice (13). Irisin can also assist the cardiac repair process. Deng et al. (107) used bone marrow mesenchymal stem cells (BM-MSCs) pretreated with FNDC5 to transplant into myocardial infarction mice. They reported that compared with non-pretreatment transplantation, irisin promoted the effect of BM-MSCs in inducing proliferation of myocardial cell and vascular, and significantly reduced myocardial remodeling and fibrosis as well as apoptosis signals. In addition, they also observed an increase in the survival rate of BM-MSCs and the secretion of exosomes. Chen et al. (100) believed that the repair effect may be related to HDAC4 degradation and p38 activation. This may show the possibility that irisin would improve the

success rate of cell transplantation and provide a new direction for the treatment of MI.

However, the concentration used in experiments were much higher than physiological state, suggesting that irisin may not function in the restoration of heart itself, yet might be promising in medical treatment to reduce the damage of cardiomyocytes after infarction and help with cell transplantation of heart. New tools like recombinant irisin should be encouraged for higher credibility. More importantly, since the current researches concerning treatment are all conducted on cells or animals, whether irisin helps in human remains unknown.

DISCUSSION

Since irisin was discovered, people have conducted a lot of indepth research on its relationship with metabolic diseases due to its most widely recognized uncoupling effect. In recent years, as the anti-inflammatory, anti-oxidant and anti-apoptotic effects of irisin have been exposed, more and more researchers are trying to explore the maximum value of this newfound molecule. In this review, studies have explored its potential in cardiovascular system (**Table 1**). Some *in vitro* and *in vivo* experiments have shown that irisin can regulate endothelial cell function and reduce the damage of atherosclerosis through various ways, which could be a sign of therapeutic use. During ACS, it was believed that an appropriate amount of irisin's serum concentration can protect the myocardium from damage. The serum concentration drops first and then rises after MI, which may be related to the reduction of energy consumption and the protection of mitochondrial function. Nevertheless, irisin would enhance the repairing effect in the process of cell transplantation of heart, which showed its therapeutic potential in a new way.

Yet there has been a big limitation of irisin's development in recent years. With the questioning of traditional measuring methods, lots of findings of irisin call for reconfirmation. The huge range of irisin level caused by inaccurate measurement led to the inconsistency of cross-sectional studies, making it hard for irisin to become a detection indicator (90). Besides, the restoration function of irisin in an infarcted heart is also facing challenges. The effective concentration applied to cells or mice was 10- to 8000-fold higher than physiological level in the gain-of-function experiments,

TABLE 1 | Functions of Irisin in CHD.

Effect	Experimental Subjects	Detail	Reference
Reducing endothelial inflammation and	HUVECs	Activating AMPK-PI3K-Akt-enos signaling pathway	(65, 66)
oxidative stress	HUVECs	Inhibiting p38 MAPK/NF-kB pathway	(67)
	HUVECs	Inhibiting PKC-β/NADPH oxidase	(68)
	HUVECs	Inhibiting ROS-NLRP3 inflammasome signaling	(70)
	HUVECs	Suppressing TNFα-induced VCAM-1	(71)
Promoting endothelial proliferation	HUVECs	Up-regulating microRNA126-5p	(69)
	HUVECs	Up-regulating ERK signaling pathway and suppressing high glucose- induced apoptosis	(109)
Reducing lipid accumulation in macrophages	RAW264.7 macrophages	Regulating Bcl-2, Bax and caspase-3	(67)
	HCAECs	Inhibiting the expression of hsCPR, ICAM-1 and E-selectin	(72)
	RAW264.7 macrophages	Inhibiting PERK/eif2a/CHOP and ATF6/CHOP ER pathways	(73)
Protecting cardiomyocytes	H9c2 cells, mouse ESCs	Inducing HDAC4 degradation to protect mitochondria	(97, 100)
	Primary cardiomyocytes	Activating mitochondrial autophagy	(86)
	H9c2 cells	Reducing the production of HO-induced ROS	(98)
	H9c2 cells	Inhibit the opening of mPTP	(99)
	H9c2 cells	Increasing the activity of SOD2	(102)
	H9c2 cells	anti-oxidation by the AKT/GSK3β/FYN/Nrf2 axis	(49)
	H9c2 cells	Activating AMPK pathway	(110)
	Sprague–Dawley rats	Causing bradycardia to reduce the energy consumption	(87)
Indicating the process of ACS	Rats or mice	Decreasing at acute phase after MI	(81, 82)
	Patients	Decreasing at acute phase after MI	(76, 83– 85)
Inconsistencies in long-term monitoring of	Patients	Increasing at 6 months after PCI	(53)
ACS	Patients	Lower in MI patients	(91)
	Patients	Positively correlating to CHD events after PCI	(77)
	Patients	A positive predictor of MACE after surgery	(83)
	Patients	Irisin genotypes be considered as genetic risk factors of MI	(93)
Treating atherosclerosis	C57BL/6 mice, apoE-deficient mice	Improving EDV and reducing the area of atherosclerotic plaque	(66, 69)
	apoE-deficient mice	Inhibiting the new carotid artery intima's formation	(67, 69)
Treating ACS	BM-MSCs	Enhancing the repairing effect in the process of cell transplantation of heart	(107)
	C57BL/6 mice	Accelerating the restoration of infarcted area	(104, 107)
	FNDC5 knockout mice	Accelerating the restoration of infarcted area	(105)

HUVEC, human coronary artery endothelial cell; ROS, reactive oxygen species; mPTP, mitochondrial permeability transition pore; SOD2, superoxide dismutase 2; MI, myocardial infarction; CHD, coronary heart disease; PCI, percutaneous coronary intervention; MACE, major adverse cardiovascular events; EDV, endothelium-dependent dilation function; apoE, apolipoprotein E; BM-MSC, bone marrow-derived mesenchymal stem cell.

which is not convincing enough to prove the physiological functions of irisin (108). In addition, due to the extremely low amount of irisin, purification is not easy to meet the demand.

With all those setbacks and a relative scarcity of preclinical and clinical data, we failed to draw a consistent conclusion of irisin's function in CHD. Thus, future researches need to be careful of selecting measuring techniques as well as observation subjects in order to control the baseline. New methods like recombinant irisin and gene-knockout mice would be the development direction in function experiments. In addition, the studies have been too scattered, each team working own its own perspective. It would push forward a lot if a general framework would form. And the experiments at present still remained at the stage of small-scale animal experiments, indicating that higher levels need to be performed.

In brief, while irisin hasn't been qualified enough yet as an indicator of CHD, it is quite promising in treatment based on current

REFERENCES

- Bostrom P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, et al. A PGC1-Alpha-Dependent Myokine That Drives Brown-Fat-Like Development of White Fat and Thermogenesis. *Nature* (2012) 481(7382):463–8. doi: 10.1038/nature10777
- Aydin S, Kuloglu T, Aydin S, Kalayci M, Yilmaz M, Cakmak T, et al. A Comprehensive Immunohistochemical Examination of the Distribution of the Fat-Burning Protein Irisin in Biological Tissues. *Peptides* (2014) 61:130– 6. doi: 10.1016/j.peptides.2014.09.014
- Raschke S, Elsen M, Gassenhuber H, Sommerfeld M, Schwahn U, Brockmann B, et al. Evidence Against a Beneficial Effect of Irisin in Humans. *PloS One* (2013) 8(9):e73680. doi: 10.1371/journal.pone.0073680
- Dong J, Dong Y, Dong Y, Chen F, Mitch WE, Zhang L. Inhibition of Myostatin in Mice Improves Insulin Sensitivity via Irisin-Mediated Cross Talk Between Muscle and Adipose Tissues. Int J Obes (Lond) (2016) 40 (3):434–42. doi: 10.1038/ijo.2015.200
- Rodriguez A, Becerril S, Mendez-Gimenez L, Ramirez B, Sainz N, Catalan V, et al. Leptin Administration Activates Irisin-Induced Myogenesis via Nitric Oxide-Dependent Mechanisms, But Reduces its Effect on Subcutaneous Fat Browning in Mice. Int J Obes (Lond) (2015) 39(3):397–407. doi: 10.1038/ ijo.2014.166
- Matsuo Y, Gleitsmann K, Mangner N, Werner S, Fischer T, Bowen TS, et al. Fibronectin Type III Domain Containing 5 Expression in Skeletal Muscle in Chronic Heart Failure-Relevance of Inflammatory Cytokines. J Cachexia Sarcopenia Muscle (2015) 6(1):62–72. doi: 10.1002/jcsm.12006
- Kurdiova T, Balaz M, Vician M, Maderova D, Vlcek M, Valkovic L, et al. Effects of Obesity, Diabetes and Exercise on Fndc5 Gene Expression and Irisin Release in Human Skeletal Muscle and Adipose Tissue: *In Vivo* and *In Vitro* Studies. *J Physiol* (2014) 592(5):1091–107. doi: 10.1113/jphysiol. 2013.264655
- Tang S, Zhang R, Jiang F, Wang J, Chen M, Peng D, et al. Circulating Irisin Levels Are Associated With Lipid and Uric Acid Metabolism in a Chinese Population. *Clin Exp Pharmacol Physiol* (2015) 42(9):896–901. doi: 10.1111/ 1440-1681.12439
- Fox J, Rioux BV, Goulet EDB, Johanssen NM, Swift DL, Bouchard DR, et al. Effect of an Acute Exercise Bout on Immediate Post-Exercise Irisin Concentration in Adults: A Meta-Analysis. Scand J Med Sci Sports (2018) 28(1):16–28. doi: 10.1111/sms.12904
- Qiu S, Cai X, Sun Z, Schumann U, Zugel M, Steinacker JM. Chronic Exercise Training and Circulating Irisin in Adults: A Meta-Analysis. Sports Med (2015) 45(11):1577–88. doi: 10.1007/s40279-014-0293-4
- Jedrychowski MP, Wrann CD, Paulo JA, Gerber KK, Szpyt J, Robinson MM, et al. Detection and Quantitation of Circulating Human Irisin by Tandem Mass Spectrometry. *Cell Metab* (2015) 22(4):734–40. doi: 10.1016/j.cmet.2015.08.001

studies. Only with more considerations in experimental design and larger-scale studies will irisin show its true colors in the near future.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

FUNDING

This work was supported by funding from the National Natural Science Foundation of China (Nos. 81770881 and 82070910) and Key R&D Plan of Hunan Province (2020SK2078).

- Lehnig AC, Dewal RS, Baer LA, Kitching KM, Munoz VR, Arts PJ, et al. Exercise Training Induces Depot-Specific Adaptations to White and Brown Adipose Tissue. *iScience* (2019) 11:425–39. doi: 10.1016/j.isci.2018.12.033
- Xiong Y, Wu Z, Zhang B, Wang C, Mao F, Liu X, et al. Fndc5 Loss-of-Function Attenuates Exercise-Induced Browning of White Adipose Tissue in Mice. FASEB J (2019) 33(5):5876–86. doi: 10.1096/fj.201801754RR
- Vitali A, Murano I, Zingaretti MC, Frontini A, Ricquier D, Cinti S. The Adipose Organ of Obesity-Prone C57BL/6J Mice Is Composed of Mixed White and Brown Adipocytes. J Lipid Res (2012) 53(4):619–29. doi: 10.1194/jlr.M018846
- Norheim F, Langleite TM, Hjorth M, Holen T, Kielland A, Stadheim HK, et al. The Effects of Acute and Chronic Exercise on PGC-1alpha, Irisin and Browning of Subcutaneous Adipose Tissue in Humans. *FEBS J* (2014) 281 (3):739–49. doi: 10.1111/febs.12619
- Camera DM, Anderson MJ, Hawley JA, Carey AL. Short-Term Endurance Training Does Not Alter the Oxidative Capacity of Human Subcutaneous Adipose Tissue. *Eur J Appl Physiol* (2010) 109(2):307–16. doi: 10.1007/ s00421-010-1356-3
- Stinkens R, Brouwers B, Jocken JW, Blaak EE, Teunissen-Beekman KF, Hesselink MK, et al. Exercise Training-Induced Effects on the Abdominal Subcutaneous Adipose Tissue Phenotype in Humans With Obesity. J Appl Physiol (1985) (2018) 125(5):1585–93. doi: 10.1152/japplphysiol.00496.2018
- Tsiloulis T, Carey AL, Bayliss J, Canny B, Meex RCR, Watt MJ. No Evidence of White Adipocyte Browning After Endurance Exercise Training in Obese Men. Int J Obes (Lond) (2018) 42(4):721–7. doi: 10.1038/ijo.2017.295
- Vosselman MJ, Hoeks J, Brans B, Pallubinsky H, Nascimento EB, van der Lans AA, et al. Low Brown Adipose Tissue Activity in Endurance-Trained Compared With Lean Sedentary Men. *Int J Obes (Lond)* (2015) 39(12):1696– 702. doi: 10.1038/ijo.2015.130
- Pillon NJ, Gabriel BM, Dollet L, Smith JAB, Sardon Puig L, Botella J, et al. Transcriptomic Profiling of Skeletal Muscle Adaptations to Exercise and Inactivity. *Nat Commun* (2020) 11(1):470. doi: 10.1038/s41467-019-13869-w
- Kim H, Wrann CD, Jedrychowski M, Vidoni S, Kitase Y, Nagano K, et al. Irisin Mediates Effects on Bone and Fat *via* alphaV Integrin Receptors. *Cell* (2019) 178(2):507–8. doi: 10.1016/j.cell.2019.06.028
- 22. Bettini S, Favaretto F, Compagnin C, Belligoli A, Sanna M, Fabris R, et al. Resting Energy Expenditure, Insulin Resistance and UCP1 Expression in Human Subcutaneous and Visceral Adipose Tissue of Patients With Obesity. *Front Endocrinol (Lausanne)* (2019) 10:548. doi: 10.3389/fendo.2019.00548
- Lim J, Park HS, Kim J, Jang YJ, Kim JH, Lee Y, et al. Depot-Specific UCP1 Expression in Human White Adipose Tissue and Its Association With Obesity-Related Markers. *Int J Obes (Lond)* (2020) 44(3):697–706. doi: 10.1038/s41366-020-0528-4
- Timmons JA, Baar K, Davidsen PK, Atherton PJ. Is Irisin a Human Exercise Gene? *Nature* (2012) 488(7413):E9–10; discussion E-1. doi: 10.1038/ nature11364

- Shan T, Liang X, Bi P, Kuang S. Myostatin Knockout Drives Browning of White Adipose Tissue Through Activating the AMPK-PGC1alpha-Fndc5 Pathway in Muscle. FASEB J (2013) 27(5):1981–9. doi: 10.1096/fj.12-225755
- Ouellet V, Labbe SM, Blondin DP, Phoenix S, Guerin B, Haman F, et al. Brown Adipose Tissue Oxidative Metabolism Contributes to Energy Expenditure During Acute Cold Exposure in Humans. J Clin Invest (2012) 122(2):545–52. doi: 10.1172/JCI60433
- Zhang Y, Xie C, Wang H, Foss RM, Clare M, George EV, et al. Irisin Exerts Dual Effects on Browning and Adipogenesis of Human White Adipocytes. *Am J Physiol Endocrinol Metab* (2016) 311(2):E530–41. doi: 10.1152/ ajpendo.00094.2016
- Moreno-Navarrete JM, Ortega F, Serrano M, Guerra E, Pardo G, Tinahones F, et al. Irisin Is Expressed and Produced by Human Muscle and Adipose Tissue in Association With Obesity and Insulin Resistance. J Clin Endocrinol Metab (2013) 98(4):E769–78. doi: 10.1210/jc.2012-2749
- Yang M, Chen P, Jin H, Xie X, Gao T, Yang L, et al. Circulating Levels of Irisin in Middle-Aged First-Degree Relatives of Type 2 Diabetes Mellitus -Correlation With Pancreatic Beta-Cell Function. *Diabetol Metab Syndr* (2014) 6(1):133. doi: 10.1186/1758-5996-6-133
- Chang CL, Huang SY, Soong YK, Cheng PJ, Wang CJ, Liang IT. Circulating Irisin and Glucose-Dependent Insulinotropic Peptide Are Associated With the Development of Polycystic Ovary Syndrome. J Clin Endocrinol Metab (2014) 99(12):E2539–48. doi: 10.1210/jc.2014-1180
- Wrann CD, White JP, Salogiannnis J, Laznik-Bogoslavski D, Wu J, Ma D, et al. Exercise Induces Hippocampal BDNF Through a PGC-1alpha/FNDC5 Pathway. *Cell Metab* (2013) 18(5):649–59. doi: 10.1016/j.cmet.2013.09.008
- Lourenco MV, Frozza RL, de Freitas GB, Zhang H, Kincheski GC, Ribeiro FC, et al. Exercise-Linked FNDC5/irisin Rescues Synaptic Plasticity and Memory Defects in Alzheimer's Models. *Nat Med* (2019) 25(1):165–75. doi: 10.1038/s41591-018-0275-4
- Moon HS, Dincer F, Mantzoros CS. Pharmacological Concentrations of Irisin Increase Cell Proliferation Without Influencing Markers of Neurite Outgrowth and Synaptogenesis in Mouse H19-7 Hippocampal Cell Lines. *Metabolism* (2013) 62(8):1131–6. doi: 10.1016/j.metabol.2013.04.007
- 34. Ma Y, Qiao X, Zeng R, Cheng R, Zhang J, Luo Y, et al. Irisin Promotes Proliferation But Inhibits Differentiation in Osteoclast Precursor Cells. *FASEB J* (2018) 32(11): 5813–23. doi: 10.1096/fj.201700983RR
- 35. Qiao X, Nie Y, Ma Y, Chen Y, Cheng R, Yin W, et al. Irisin Promotes Osteoblast Proliferation and Differentiation *via* Activating the MAP Kinase Signaling Pathways. *Sci Rep* (2016) 6:18732. doi: 10.1038/srep21053
- Colaianni G, Cuscito C, Mongelli T, Oranger A, Mori G, Brunetti G, et al. Irisin Enhances Osteoblast Differentiation In Vitro. *Int J Endocrinol* (2014) 2014:902186. doi: 10.1155/2014/902186
- Colaianni G, Mongelli T, Cuscito C, Pignataro P, Lippo L, Spiro G, et al. Irisin Prevents and Restores Bone Loss and Muscle Atrophy in Hind-Limb Suspended Mice. Sci Rep (2017) 7(1):2811. doi: 10.1038/s41598-017-02557-8
- 38. Anastasilakis AD, Polyzos SA, Makras P, Gkiomisi A, Bisbinas I, Katsarou A, et al. Circulating Irisin Is Associated With Osteoporotic Fractures in Postmenopausal Women With Low Bone Mass But Is Not Affected by Either Teriparatide or Denosumab Treatment for 3 Months. Osteoporos Int (2014) 25(5):1633–42. doi: 10.1007/s00198-014-2673-x
- Kim H, Wrann CD, Jedrychowski M, Vidoni S, Kitase Y, Nagano K, et al. Irisin Mediates Effects on Bone and Fat *via* alphaV Integrin Receptors. *Cell* (2018) 175(7):1756–68 e17. doi: 10.1016/j.cell.2018.10.025
- Shao L, Li H, Chen J, Song H, Zhang Y, Wu F, et al. Irisin Suppresses the Migration, Proliferation, and Invasion of Lung Cancer Cells via Inhibition of Epithelial-to-Mesenchymal Transition. *Biochem Biophys Res Commun* (2017) 485(3):598–605. doi: 10.1016/j.bbrc.2016.12.084
- Gannon NP, Vaughan RA, Garcia-Smith R, Bisoffi M, Trujillo KA. Effects of the Exercise-Inducible Myokine Irisin on Malignant and non-Malignant Breast Epithelial Cell Behavior In Vitro. *Int J Cancer* (2015) 136(4):E197– 202. doi: 10.1002/ijc.29142
- Park SE, Park CY, Sweeney G. Biomarkers of Insulin Sensitivity and Insulin Resistance: Past, Present and Future. *Crit Rev Clin Lab Sci* (2015) 52(4):180– 90. doi: 10.3109/10408363.2015.1023429
- 43. Li RL, Wu SS, Wu Y, Wang XX, Chen HY, Xin JJ, et al. Irisin Alleviates Pressure Overload-Induced Cardiac Hypertrophy by Inducing Protective Autophagy via mTOR-Independent Activation of the AMPK-ULK1

Pathway. J Mol Cell Cardiol (2018) 121:242-55. doi: 10.1016/j.yjmcc. 2018.07.250

- 44. Yu Q, Kou W, Xu X, Zhou S, Luan P, Xu X, et al. FNDC5/Irisin Inhibits Pathological Cardiac Hypertrophy. *Clin Sci (Lond)* (2019) 133(5):611–27. doi: 10.1042/CS20190016
- Zhou X, Xu M, Bryant JL, Ma J, Xu X. Exercise-Induced Myokine FNDC5/ irisin Functions in Cardiovascular Protection and Intracerebral Retrieval of Synaptic Plasticity. *Cell Biosci* (2019) 9:32. doi: 10.1186/s13578-019-0294-y
- Albrecht E, Norheim F, Thiede B, Holen T, Ohashi T, Schering L, et al. Irisin

 a Myth Rather Than an Exercise-Inducible Myokine. *Sci Rep* (2015) 5:8889.
 doi: 10.1038/srep08889
- Polyzos SA, Mathew H, Mantzoros CS. Irisin: A True, Circulating Hormone. Metabolism (2015) 64(12):1611–8. doi: 10.1016/j.metabol.2015.09.001
- Albrecht E, Schering L, Buck F, Vlach K, Schober HC, Drevon CA, et al. Irisin: Still Chasing Shadows. *Mol Metab* (2020) 34:124–35. doi: 10.1016/ j.molmet.2020.01.016
- Liu TY, Xiong XQ, Ren XS, Zhao MX, Shi CX, Wang JJ, et al. FNDC5 Alleviates Hepatosteatosis by Restoring AMPK/mTOR-Mediated Autophagy, Fatty Acid Oxidation, and Lipogenesis in Mice. *Diabetes* (2016) 65(11):3262–75. doi: 10.2337/db16-0356
- Trevellin E, Scorzeto M, Olivieri M, Granzotto M, Valerio A, Tedesco L, et al. Exercise Training Induces Mitochondrial Biogenesis and Glucose Uptake in Subcutaneous Adipose Tissue Through eNOS-Dependent Mechanisms. *Diabetes* (2014) 63(8):2800–11. doi: 10.2337/db13-1234
- Zhou B, Ling L, Zhang F, Liu TY, Zhou H, Qi XH, et al. Fibronectin Type III Domain-Containing 5 Attenuates Liver Fibrosis Via Inhibition of Hepatic Stellate Cell Activation. *Cell Physiol Biochem* (2018) 48(1):227–36. doi: 10.1159/000491722
- Virani SS, Alonso A, Benjamin EJ, Bittencourt MS, Callaway CW, Carson AP, et al. Heart Disease and Stroke Statistics-2020 Update: A Report From the American Heart Association. *Circulation* (2020) 141(9):e139–596. doi: 10.1161/CIR.00000000000757
- Aronis KN, Moreno M, Polyzos SA, Moreno-Navarrete JM, Ricart W, Delgado E, et al. Circulating Irisin Levels and Coronary Heart Disease: Association With Future Acute Coronary Syndrome and Major Adverse Cardiovascular Events. *Int* J Obes (Lond) (2015) 39(1):156–61. doi: 10.1038/ijo.2014.101
- Hopkins PN. Molecular Biology of Atherosclerosis. *Physiol Rev* (2013) 93 (3):1317–542. doi: 10.1152/physrev.00004.2012
- Deng W. Association of Serum Irisin Concentrations With Presence and Severity of Coronary Artery Disease. *Med Sci Monit* (2016) 22:4193–7. doi: 10.12659/MSM.897376
- Khorasani ZM, Bagheri RK, Yaghoubi MA, Chobkar S, Aghaee MA, Abbaszadegan MR, et al. The Association Between Serum Irisin Levels and Cardiovascular Disease in Diabetic Patients. *Diabetes Metab Syndr* (2019) 13(1):786–90. doi: 10.1016/j.dsx.2018.11.050
- 57. Saadeldin MK, Elshaer SS, Emara IA, Maged M, Abdel-Aziz AK. Serum Sclerostin and Irisin as Predictive Markers for Atherosclerosis in Egyptian Type II Diabetic Female Patients: A Case Control Study. *PloS One* (2018) 13 (11):e0206761. doi: 10.1371/journal.pone.0206761
- Lee MJ, Lee SA, Nam BY, Park S, Lee SH, Ryu HJ, et al. Irisin, a Novel Myokine Is an Independent Predictor for Sarcopenia and Carotid Atherosclerosis in Dialysis Patients. *Atherosclerosis* (2015) 242(2):476–82. doi: 10.1016/j.atherosclerosis.2015.08.002
- 59. Icli A, Cure E, Cumhur Cure M, Uslu AU, Balta S, Arslan S, et al. Novel Myokine: Irisin May Be An Independent Predictor for Subclinic Atherosclerosis in Behcet's Disease. J Investig Med (2016) 64(4):875–81. doi: 10.1136/jim-2015-000044
- 60. Sesti G, Andreozzi F, Fiorentino TV, Mannino GC, Sciacqua A, Marini MA, et al. High Circulating Irisin Levels Are Associated With Insulin Resistance and Vascular Atherosclerosis in a Cohort of Nondiabetic Adult Subjects. *Acta Diabetol* (2014) 51(5):705–13. doi: 10.1007/s00592-014-0576-0
- Kwasniewska M, Kostka T, Jegier A, Dziankowska-Zaborszczyk E, Leszczynska J, Rebowska E, et al. Regular Physical Activity and Cardiovascular Biomarkers in Prevention of Atherosclerosis in Men: A 25-Year Prospective Cohort Study. *BMC Cardiovasc Disord* (2016) 16:65. doi: 10.1186/s12872-016-0239-x
- 62. Moreno-Perez O, Reyes-Garcia R, Munoz-Torres M, Merino E, Boix V, Reus S, et al. High Irisin Levels in Nondiabetic HIV-Infected Males Are

Associated With Insulin Resistance, Nonalcoholic Fatty Liver Disease, and Subclinical Atherosclerosis. *Clin Endocrinol (Oxf)* (2018) 89(4):414–23. doi: 10.1111/cen.13800

- Perakakis N, Triantafyllou GA, Fernandez-Real JM, Huh JY, Park KH, Seufert J, et al. Physiology and Role of Irisin in Glucose Homeostasis. Nat Rev Endocrinol (2017) 13(6):324–37. doi: 10.1038/nrendo.2016.221
- 64. Hisamatsu T, Miura K, Arima H, Fujiyoshi A, Kadota A, Kadowaki S, et al. Relationship of Serum Irisin Levels to Prevalence and Progression of Coronary Artery Calcification: A Prospective, Population-Based Study. *Int J Cardiol* (2018) 267:177–82. doi: 10.1016/j.ijcard.2018.05.075
- 65. Lu J, Xiang G, Liu M, Mei W, Xiang L, Dong J. Irisin Protects Against Endothelial Injury and Ameliorates Atherosclerosis in Apolipoprotein E-Null Diabetic Mice. *Atherosclerosis* (2015) 243(2):438–48. doi: 10.1016/ j.atherosclerosis.2015.10.020
- 66. Han F, Zhang S, Hou N, Wang D, Sun X. Irisin Improves Endothelial Function in Obese Mice Through the AMPK-eNOS Pathway. Am J Physiol Heart Circ Physiol (2015) 309(9):H1501–8. doi: 10.1152/ajpheart.00443.2015
- 67. Zhang Y, Mu Q, Zhou Z, Song H, Zhang Y, Wu F, et al. Protective Effect of Irisin on Atherosclerosis *via* Suppressing Oxidized Low Density Lipoprotein Induced Vascular Inflammation and Endothelial Dysfunction. *PloS One* (2016) 11(6):e0158038. doi: 10.1371/journal.pone.0158038
- Zhu D, Wang H, Zhang J, Zhang X, Xin C, Zhang F, et al. Irisin Improves Endothelial Function in Type 2 Diabetes Through Reducing Oxidative/ Nitrative Stresses. J Mol Cell Cardiol (2015) 87:138–47. doi: 10.1016/ j.yjmcc.2015.07.015
- 69. Zhang Y, Song H, Zhang Y, Wu F, Mu Q, Jiang M, et al. Irisin Inhibits Atherosclerosis by Promoting Endothelial Proliferation Through Microrna126-5p. J Am Heart Assoc (2016) 5(9):e004031. doi: 10.1161/JAHA.116.004031
- Deng X, Huang W, Peng J, Zhu TT, Sun XL, Zhou XY, et al. Irisin Alleviates Advanced Glycation End Products-Induced Inflammation and Endothelial Dysfunction *via* Inhibiting ROS-NLRP3 Inflammasome Signaling. *Inflammation* (2018) 41(1):260–75. doi: 10.1007/s10753-017-0685-3
- Shimba Y, Togawa H, Senoo N, Ikeda M, Miyoshi N, Morita A, et al. Skeletal Muscle-Specific PGC-1alpha Overexpression Suppresses Atherosclerosis in Apolipoprotein E-Knockout Mice. *Sci Rep* (2019) 9(1):4077. doi: 10.1038/ s41598-019-40643-1
- 72. Yin C, Hu W, Wang M, Lv W, Jia T, Xiao Y. Irisin as a Mediator Between Obesity and Vascular Inflammation in Chinese Children and Adolescents. *Nutr Metab Cardiovasc Dis* (2020) 30(2):320–9. doi: 10.1016/ j.numecd.2019.09.025
- 73. Zheng G, Li H, Zhang T, Yang L, Yao S, Chen S, et al. Irisin Protects Macrophages From Oxidized Low Density Lipoprotein-Induced Apoptosis by Inhibiting the Endoplasmic Reticulum Stress Pathway. *Saudi J Biol Sci* (2018) 25(5):849–57. doi: 10.1016/j.sjbs.2017.08.018
- Kannel WB, Feinleib M. Natural History of Angina Pectoris in the Framingham Study. Prognosis and Survival. Am J Cardiol (1972) 29 (2):154–63. doi: 10.1016/0002-9149(72)90624-8
- Efe TH, Acar B, Ertem AG, Yayla KG, Algul E, Yayla C, et al. Serum Irisin Level Can Predict the Severity of Coronary Artery Disease in Patients With Stable Angina. *Korean Circ J* (2017) 47(1):44–9. doi: 10.4070/kcj.2016.0079
- 76. Anastasilakis AD, Koulaxis D, Kefala N, Polyzos SA, Upadhyay J, Pagkalidou E, et al. Circulating Irisin Levels Are Lower in Patients With Either Stable Coronary Artery Disease (CAD) or Myocardial Infarction (MI) Versus Healthy Controls, Whereas Follistatin and Activin A Levels Are Higher and can Discriminate MI From CAD With Similar to CK-MB Accuracy. *Metabolism* (2017) 73:1–8. doi: 10.1016/j.metabol.2017.05.002
- Park KH, Zaichenko L, Brinkoetter M, Thakkar B, Sahin-Efe A, Joung KE, et al. Circulating Irisin in Relation to Insulin Resistance and the Metabolic Syndrome. *J Clin Endocrinol Metab* (2013) 98(12):4899–907. doi: 10.1210/jc.2013-2373
- Arbustini E, Burke A, Dal Bello B, Morbini P, Specchia G, Virmani R. Plaque Erosion Is a Major Substrate for Coronary Thrombosis in Acute Myocardial Infarction. J Am Coll Cardiol (1998) 31(3):269. doi: 10.1136/hrt.82.3.269
- 79. Higuma T, Soeda T, Abe N, Yamada M, Yokoyama H, Shibutani S, et al. A Combined Optical Coherence Tomography and Intravascular Ultrasound Study on Plaque Rupture, Plaque Erosion, and Calcified Nodule in Patients With ST-Segment Elevation Myocardial Infarction: Incidence, Morphologic Characteristics, and Outcomes After Percutaneous Coronary Intervention. JACC Cardiovasc Interv (2015) 8(9):1166–76. doi: 10.1016/j.jcin.2015.02.026

- Davies MJ, Thomas A. Thrombosis and Acute Coronary-Artery Lesions in Sudden Cardiac Ischemic Death. N Engl J Med (1984) 310(18):1137–40. doi: 10.1056/NEJM198405033101801
- Kuloglu T, Aydin S, Eren MN, Yilmaz M, Sahin I, Kalayci M, et al. Irisin: A Potentially Candidate Marker for Myocardial Infarction. *Peptides* (2014) 55:85–91. doi: 10.1016/j.peptides.2014.02.008
- Bashar SM, Samir El-Sherbeiny SM, Boraie MZ. Correlation Between the Blood Level of Irisin and the Severity of Acute Myocardial Infarction in Exercise-Trained Rats. J Basic Clin Physiol Pharmacol (2018) 30(1):59–71. doi: 10.1515/jbcpp-2018-0090
- Hsieh IC, Ho MY, Wen MS, Chen CC, Hsieh MJ, Lin CP, et al. Serum Irisin Levels Are Associated With Adverse Cardiovascular Outcomes in Patients With Acute Myocardial Infarction. *Int J Cardiol* (2018) 261:12–7. doi: 10.1016/j.ijcard.2017.11.072
- 84. Aydin S, Kobat MA, Kalayci M, Eren MN, Yilmaz M, Kuloglu T, et al. Decreased Saliva/Serum Irisin Concentrations in the Acute Myocardial Infarction Promising for Being a New Candidate Biomarker for Diagnosis of This Pathology. *Peptides* (2014) 56:141–5. doi: 10.1016/j.peptides .2014.04.002
- Aydin S, Catak Z, Eren MN, Topal AE, Aydin S. Irisin in Coronary Bypass Surgery. *Cardiovasc Hematol Disord Drug Targets* (2018) 18(3):208–14. doi: 10.2174/1871529X18666180511141151
- Xin T, Lu C. Irisin Activates Opa1-Induced Mitophagy to Protect Cardiomyocytes Against Apoptosis Following Myocardial Infarction. *Aging (Albany NY)* (2020) 12(5):4474–88. doi: 10.18632/aging.102899
- Brailoiu E, Deliu E, Sporici RA, Brailoiu GC. Irisin Evokes Bradycardia by Activating Cardiac-Projecting Neurons of Nucleus Ambiguus. *Physiol Rep* (2015) 3(6):e12419. doi: 10.14814/phy2.12419
- Tao L, Bei Y, Lin S, Zhang H, Zhou Y, Jiang J, et al. Exercise Training Protects Against Acute Myocardial Infarction *via* Improving Myocardial Energy Metabolism and Mitochondrial Biogenesis. *Cell Physiol Biochem* (2015) 37(1):162–75. doi: 10.1159/000430342
- Jiang M, Wan F, Wang F, Wu Q. Irisin Relaxes Mouse Mesenteric Arteries Through Endothelium-Dependent and Endothelium-Independent Mechanisms. *Biochem Biophys Res Commun* (2015) 468(4):832–6. doi: 10.1016/j.bbrc.2015.11.040
- Guo W, Zhang B, Wang X. Lower Irisin Levels in Coronary Artery Disease: A Meta-Analysis. *Minerva Endocrinol* (2020) 45(1):61–9. doi: 10.23736/ S0391-1977.17.02663-3
- Emanuele E, Minoretti P, Pareja-Galeano H, Sanchis-Gomar F, Garatachea N, Lucia A. Serum Irisin Levels, Precocious Myocardial Infarction, and Healthy Exceptional Longevity. *Am J Med* (2014) 127(9):888–90. doi: 10.1016/j.amjmed.2014.04.025
- Belviranli M, Okudan N. Exercise Training Increases Cardiac, Hepatic and Circulating Levels of Brain-Derived Neurotrophic Factor and Irisin in Young and Aged Rats. *Horm Mol Biol Clin Investig* (2018) 36 (3):20180053. doi: 10.1515/hmbci-2018-0053
- 93. Badr EA, Mostafa RG, Awad SM, Marwan H, Abd El-Bary HM, Shehab HE, et al. A Pilot Study on the Relation Between Irisin Single-Nucleotide Polymorphism and Risk of Myocardial Infarction. *Biochem Biophys Rep* (2020) 22:100742. doi: 10.1016/j.bbrep.2020.100742
- Martins IJ, Hone E, Foster JK, Sunram-Lea SI, Gnjec A, Fuller SJ, et al. Cholesterol Metabolism, Diabetes, and the Convergence of Risk Factors for Alzheimer's Disease and Cardiovascular Disease. *Mol Psychiatry* (2006) 11 (8):721–36. doi: 10.1038/sj.mp.4001854
- Heeren J, Beisiegel U, Grewal T. Apolipoprotein E Recycling: Implications for Dyslipidemia and Atherosclerosis. *Arterioscler Thromb Vasc Biol* (2006) 26(3):442–8. doi: 10.1161/01.ATV.0000201282.64751.47
- Fuku N, Diaz-Pena R, Arai Y, Abe Y, Zempo H, Naito H, et al. Epistasis, Physical Capacity-Related Genes and Exceptional Longevity: FNDC5 Gene Interactions With Candidate Genes FOXOA3 and APOE. *BMC Genomics* (2017) 18(Suppl 8):803. doi: 10.1186/s12864-017-4194-4
- Moscoso I, Cebro-Marquez M, Rodriguez-Manero M, Gonzalez-Juanatey JR, Lage R. FNDC5/Irisin Counteracts Lipotoxic-Induced Apoptosis in Hypoxic H9c2 Cells. J Mol Endocrinol (2019) 63(2):151–9. doi: 10.1530/JME-19-0123
- Peng Q, Wang X, Wu K, Liu K, Wang S, Chen X. Irisin Attenuates H2O2-Induced Apoptosis in Cardiomyocytes via microRNA-19b/AKT/mTOR Signaling Pathway. Int J Clin Exp Pathol (2017) 10(7):7707–17.

- Zhao YT, Wang H, Zhang S, Du J, Zhuang S, Zhao TC. Irisin Ameliorates Hypoxia/ Reoxygenation-Induced Injury Through Modulation of Histone Deacetylase 4. *PloS One* (2016) 11(11):e0166182. doi: 10.1371/journal.pone.0166182
- 100. Chen HP, Denicola M, Qin X, Zhao Y, Zhang L, Long XL, et al. HDAC Inhibition Promotes Cardiogenesis and the Survival of Embryonic Stem Cells Through Proteasome-Dependent Pathway. J Cell Biochem (2011) 112 (11):3246–55. doi: 10.1002/jcb.23251
- Wende AR, Abel ED. Lipotoxicity in the Heart. *Biochim Biophys Acta* (2010) 1801(3):311–9. doi: 10.1016/j.bbalip.2009.09.023
- 102. Wang Z, Chen K, Han Y, Zhu H, Zhou X, Tan T, et al. Irisin Protects Heart Against Ischemia-Reperfusion Injury Through a SOD2-Dependent Mitochondria Mechanism. J Cardiovasc Pharmacol (2018) 72(6):259–69. doi: 10.1097/FJC.00000000000608
- 103. Lu L, Ma J, Tang J, Liu Y, Zheng Q, Chen S, et al. Irisin Attenuates Myocardial Ischemia/Reperfusion-Induced Cardiac Dysfunction by Regulating ER-Mitochondria Interaction Through a Mitochondrial Ubiquitin Ligase-Dependent Mechanism. *Clin Transl Med* (2020) 10(5): e166. doi: 10.1002/ctm2.166
- 104. Liao Q, Qu S, Tang LX, Li LP, He DF, Zeng CY, et al. Irisin Exerts a Therapeutic Effect Against Myocardial Infarction via Promoting Angiogenesis. Acta Pharmacol Sin (2019) 40(10):1314–21. doi: 10.1038/s41401-019-0230-z
- 105. Xin C, Zhang Z, Gao G, Ding L, Yang C, Wang C, et al. Irisin Attenuates Myocardial Ischemia/Reperfusion Injury and Improves Mitochondrial Function Through AMPK Pathway in Diabetic Mice. *Front Pharmacol* (2020) 11:565160. doi: 10.3389/fphar.2020.565160
- 106. Zhang M, Xu Y, Jiang L. Irisin Attenuates Oxidized Low-Density Lipoprotein Impaired Angiogenesis Through AKT/mTOR/S6K1/Nrf2 Pathway. J Cell Physiol (2019) 234(10):18951–62. doi: 10.1002/jcp.28535

- 107. Deng J, Zhang N, Wang Y, Yang C, Wang Y, Xin C, et al. FNDC5/irisin Improves the Therapeutic Efficacy of Bone Marrow-Derived Mesenchymal Stem Cells for Myocardial Infarction. *Stem Cell Res Ther* (2020) 11(1):228. doi: 10.1186/s13287-020-01746-z
- 108. Rabiee F, Lachinani L, Ghaedi S, Nasr-Esfahani MH, Megraw TL, Ghaedi K. New Insights Into the Cellular Activities of Fndc5/Irisin and Its Signaling Pathways. *Cell Biosci* (2020) 10:51. doi: 10.1186/s13578-020-00413-3
- 109. Song H, Wu F, Zhang Y, Zhang Y, Wang F, Jiang M, et al. Irisin Promotes Human Umbilical Vein Endothelial Cell Proliferation Through the ERK Signaling Pathway and Partly Suppresses High Glucose-Induced Apoptosis. *PloS One* (2014) 9(10):e110273. doi: 10.1371/journal. pone.0110273
- 110. Fan J, Zhu Q, Wu Z, Ding J, Qin S, Liu H, et al. Protective Effects of Irisin on Hypoxia-Reoxygenation Injury in Hyperglycemia-Treated Cardiomyocytes: Role of AMPK Pathway and Mitochondrial Protection. J Cell Physiol (2020) 235(2):1165–74. doi: 10.1002/jcp.29030

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Ou-Yang, Guo, Xu, Lin, Li, Shan, Wu, Wang, Zheng, Xu and Yuan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.