



Original article

Proficiency of data interpretation: identification of signaling SNPs/specific loci for coronary artery disease

Asma N. Cheema^{1,2,*}, Samantha L. Rosenthal³ and M. Ilyas Kamboh³

¹Atta-Ur-Rahman School of Applied Biosciences, National University of Sciences & Technology, Islamabad, Pakistan, ²Department of Pathology, University Medical & Dental College, The University of Faisalabad, Faisalabad, Pakistan and ³Department of Human Genetics, University of Pittsburgh, Pittsburgh, PA, USA

*Corresponding author: Tel: +92 41 8869861 62, Ext: 170; Fax: +92 41 8868220; Email: asma.cheema4@gmail.com or dr.asma.naseer@tuf.edu.pk

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Abstract

Coronary artery disease (CAD) is a complex disorder involving both genetic and non-genetic factors. Genome-wide association studies (GWAS) have identified hundreds of single nucleotides polymorphisms (SNPs) tagging over > 40 CAD risk loci. We hypothesized that some non-coding variants might directly regulate the gene expression rather than tagging a nearby locus. We used RegulomeDB to examine regulatory functions of 58 SNPs identified in two GWAS and those SNPs in linkage disequilibrium (LD) ($r^2 \geq 0.80$) with the GWAS SNPs. Of the tested 1200 SNPs, 858 returned scores of 1–6 by RegulomeDB. Of these 858 SNPs, 97 were predicted to have regulatory functions with RegulomeDB score of < 3. Notably, only 8 of the 97 predicted regulatory variants were genome-wide significant SNPs (*LIPA*/rs2246833, RegulomeDB score = 1b; *ZC3HC1*/rs11556924, *CYP17A1-CNNM2-NT5C2*/rs12413409, *APOE-APOC1*/rs2075650 and *UBE2Z*/rs46522, each with a RegulomeDB score = 1f; *ZNF259-APOA5-APOA1*/rs964184, *SMG6*/rs2281727 and *COL4A1-COL4A2*/rs4773144, each with a RegulomeDB score = 2b). The remainder 89 functional SNPs were in linkage disequilibrium with GWAS SNPs. This study supports the hypothesis that some of the non-coding variants are true signals via regulation of gene expression at transcription level. Our study indicates that RegulomeDB is a useful database to examine the putative functions of large number of genetic variants and it may help to identify a true association among multiple tagged SNPs in a complex disease, such as CAD.

Database URLs: <http://www.regulomedb.org/>; <https://www.broadinstitute.org/mpg/snap/>

Background

Most human DNA sequence is non-coding (98%) and hence only small portion (2%) of human genome encodes proteins (1). Although the pathogenesis of monogenic disorders is largely explained, it has been difficult to determine the underlying mechanisms of complex disorders like coronary artery disease (CAD). Before the development of genome-wide association studies (GWAS), only the *APOE*4* allele showed consistent association with the risk of CAD across many populations (2–5).

The hypothesis-free GWAS approach was designed with the assumption that common DNA variants explain the bulk of the variation in common diseases (6). About 90% of GWAS-implicated variants, exert only minimal to modest effect sizes on disease phenotypes, and they are present in non-coding rather than coding regions (7). Highly sensitive molecular and computational techniques have identified different regulatory elements (DNase hypersensitive regions, sequences affecting the binding of transcription factors and promoters or enhancers) in intergenic regions (8). Common variants located in one of these regulatory elements may affect gene expression. To predict the role of these variants in gene regulation and to differentiate between physically tagged and functional single nucleotides polymorphism (SNPs), many databases have been created (9). RegulomeDB is one of such databases that describes the role of these variants in transcriptional regulation.

Similar to many other complex diseases, GWAS have identified hundreds of risk variants associated with CAD that need to be analyzed for their functional role in gene expression (10). Recently, we have used SNAP Webportal and Regulome DB to identify potential regulatory function of variants in associated risk loci for Alzheimer's disease (11). In this study, we have applied the same approach to identify the regulatory nature of GWAS-implicated variants with CAD and those that are in linkage disequilibrium (LD) with these variants.

Objective

The objective of our study was to assess the GWAS-implicated CAD variants and those variants in LD with GWAS variants for their potential regulatory effects on gene transcription using bioinformatics tools.

Materials and methods

SNPs selection

A total of 58 SNPs within 54 CAD loci was selected, including 52 with accepted genome-wide significant

Table 1. Number of SNPs in LD for all published GWAS SNPs for HapMap3 and 1000 genomes populations at tested r^2 threshold

	LD		
	r^2 threshold	0.80	0.90
1000 Genomes	1176	928	480
Hap Map3	210	157	74
Total (overlaps removed)	1200	934	485

threshold ($P < 5 \times 10^{-8}$) and 6 with suggestive associations ($P > 5 \times 10^{-8}$) identified in two GWAS (12, 13). Detailed information on the selected 58 SNPs is provided in [Supplementary Table S1](#).

Linkage disequilibrium

For the LD assessment of the selected 58 SNPs, we used SNAP web portal (<https://www.broadinstitute.org/mpg/snap/>, accessed 13 July 2016) (14) ([Supplementary Table S2](#)). SNAP contains data from the Northern European from Utah (CEU) population derived from the 1000 Genomes Pilot Project 1 and three different releases of the International-Hap Map Project. We used data from both the 1000 Genomes Project and HapMap 3 (release 2) to identify SNPs in strong LD ($r^2 \geq 0.80$) with our SNPs of interest. We did not select an array bound search, and query SNPs were included in the output. We performed the search at three thresholds— $r^2 \geq 0.80$, $r^2 \geq 0.90$ and $r^2 \geq 1.0$ —for both SNP datasets and identified a total of 1,200 SNPs in LD with the 58 published GWAS SNPs, including the GWAS SNPs themselves. As shown in [Table 1](#), the number of proxy SNPs decreased with the increased level of r^2 .

Functional assessment of CAD-associated SNPs

We used RegulomeDB to identify potentially functional SNPs among the 1200 SNPs of interest. Regulome DB is a database that scores SNPs functionality based upon experimental data, such as its existence in a DNase hypersensitive site or transcription factor binding site. These regions have been characterized biochemically, and data are drawn from published literature, Gene Expression Omnibus and ENCODE project that include a total of 962 experimental datasets, covering over 100 tissues and cell lines and representing nearly 60 million annotations. The output data can be mapped to Human genome version 19. It is a user friendly and freely accessible database (<http://www.regulomedb.org/> accessed 17 July 2016) (15). The functional Grades 1–6 of RegulomeDB are given in [Table 2](#). SNPs

Table 2. RegulomeDB category summaries (15)

Category	Description
Likely to affect binding and linked to expression of a gene target	
1b	eQTL + TF binding + any motif + DNase footprint + DNase peak
1c	eQTL + TF binding + matched TF motif + DNase peak
1d	eQTL + TF binding + any motif + DNase peak
1e	eQTL + TF binding + matched TF motif
1f	eQTL + TF binding/DNase peak
Likely to affect binding	
2a	TF binding + matched TF motif + matched DNase footprint + DNase peak
2b	TF binding + any motif + DNase footprint + DNase peak
2c	TF binding + matched TF motif + DNase peak
Less likely to affect binding	
3a	TF binding + any motif + DNase peak
3b	TF binding + matched TF motif
Minimal binding evidence	
4	TF binding + DNase peak
5	TF binding or DNase peak
6	Motif hit

showing the strongest evidence of being regulatory (affecting the binding of transcription factor) are given a score of 1 and SNPs demonstrating the least evidence of being functional are given a score of 6.

Results

Among the 1200 SNPs evaluated with RegulomeDB, 342 had no data (Supplementary Table S3). Of the 858 SNPs for which RegulomeDB provided a score, 97 had a score of <3 (likely to affect the binding) and among these only 8 SNPs were genome-wide significant, including *LIPA*/rs2246833 (RegulomeDB score = 1b; eQTL in monocytes), *ZC3HC1*/rs11556924 (RegulomeDB score = 1f; eQTL in monocytes), *CYP17A1-CNNM2-NT5C2*/rs12413409 (RegulomeDB score = 1f; eQTL in monocytes and lymphoblasts), *APOE-APOC1*/rs2075650, and *UBE2Z*/rs46522 (RegulomeDB score = 1f; eQTL in monocytes), *ZNF259-APOA5-APOA1*/rs964184, *UBE2Z*/rs46522, *SMG6*/rs2281727, *COL4A1-COL4A2*/rs4773144 (RegulomeDB score = 2b; eQTLs in monocytes and lymphoblasts). A flow chart summarizes these results (Figure 1). The remaining 89 SNPs with RegulomeDB scores < 3 were not identified in GWAS but they were in LD ($r^2 \geq 0.80$) with the 29 GWAS reported SNPs. A summary of the regulatory SNPs in LD with GWAS SNPs is provided in Table 3.

Overall, we had 97 functional SNPs (RegulomeDB < 3). Eight of these were GWAS SNPs, and the remaining 89 were in LD ($r^2 \geq 0.80$) with the GWAS SNPs.

Three variants, *FES*/rs1894401, *LIPA*/rs2246833 and *VAMP8*/rs1009, were strongly predicted to be functional

with score of 1b. *FES*/rs1894401 is an intronic SNP that is an eQTL for *FES* in thyroid and transformed lymphoblasts, is present in the binding motif of Pax5, and affects the binding of eleven transcription factors. *LIPA*/rs2246833 (RegulomeDB score = 1b), located in Intron 6 of *LIPA*, in the DNA motif of EWSRCFL11, is a GWAS reported SNP along with 4 other functional SNPs (of 12 tested) in this region and it affects the binding of CTCF. It is an eQTL in the whole blood. *VAMP5-VAMP8-GGCX*/rs1009 is in exon 3 of *VAMP8* and affects the binding of CTCF and HSF1. rs1009 of *VAMP8* is an eQTL in lymphoblasts, skeletal muscles, adipose tissue and thyroid. Of 42 SNPs analyzed in this locus, we found 8 other SNPs with RegulomeDB score < 3 (Table 3).

There were 33 functional SNPs within 15 GWAS identified CAD loci: *ABO* (1 of 10 assessed), *ADAMTS7* (1 of 15 assessed), *CXCL12* (2 of 36 assessed), *HHIPL1* (3 of 17 assessed), *KCNE2* (2 of 18 assessed), *KIAA1462* (1 of 9 assessed), *MIA3* (1 of 27 assessed), *PPAP2B* (2 of 22 assessed), *SORT1* (3 of 9 assessed), *WDR12* (5 of 214 assessed), *IL6R* (3 of 14 assessed), *LPL* (2 of 6 assessed), *PLG* (2 of 41 assessed), *SLC22A4-SLC22A5* (1 of 2 assessed) and *TRIB1* (4 of 16 assessed).

Of 97 SNPs with RegulomeDB score < 3, 25 were in the *CYP17A1-CNNM2-NT5C2* region, and one of them was a GWAS reported SNP (rs12413409). The regional LD plot of this SNP is given in Supplementary Figure S1. rs9633712 (RegulomeDB score = 1e) is located in Intron 3 of *NT5C2* and is an eQTL for *USMG5* in monocytes. This SNP was also found in the motifs of the following transcription factors: PU1, ELF1, Sfpil, PU.1 and c-Ets-1.

Table 3. Functional SNPs (RegluomeDB Score < 3) in LD ($r^2 \geq 0.80$) with published GWAS SNPs

GWAS SNPs	Functional proxy SNPs	Regulome DB score	
<i>LIPA/rs2246833</i>	<i>LIPA/rs1332327</i>	2b	
	<i>LIPA/rs1332328</i>	2b	
	<i>LIPA/rs1412444</i>	1d	
	<i>LIPA/rs2246833^a</i>	1b	
	<i>LIPA/rs2250644</i>	2b	
<i>ZC3HC1/rs11556924</i>	<i>ZC3HC1/rs11556924^a</i>	1f	
<i>CYP17A1-CNNM2-NT5C2/rs12413409</i>	<i>AS3MT/rs11191454</i>	1f	
	<i>BORCS7-ASMT/rs4409766</i>	1f	
	<i>CNNM2/rs10883808</i>	1f	
	<i>MAT2A/rs1446668</i>	2a	
	<i>NT5C2/rs10883832</i>	1f	
	<i>CNNM2/rs11191479</i>	1f	
	<i>NT5C2/rs11191557</i>	1f	
	<i>CNNM2/rs11191499</i>	1f	
	<i>NT5C2/rs11191558</i>	1f	
	<i>CNNM2/rs11191514</i>	1f	
	<i>NT5C2/rs11191580</i>	1f	
	<i>CNNM2/rs11191515</i>	1f	
	<i>NT5C2/rs11191582</i>	1f	
	<i>CNNM2/rs12221064</i>	2b	
	<i>NT5C2/rs12412038</i>	1f	
	<i>CNNM2/rs12411886</i>	1f	
	<i>NT5C2/rs12413046</i>	1f	
	<i>CNNM2/rs12413409^a</i>	1f	
	<i>NT5C2/rs9633712</i>	1e	
	<i>CNNM2/rs17115213</i>	1f	
	<i>NT5C2/rs11191548</i>	1f	
	<i>CNNM2/rs2297787</i>	2a	
	<i>CNNM2/rs3781285</i>	1f	
	<i>CNNM2/rs943037</i>	1f	
	<i>CNNM2/rs12219901</i>	2b	
	<i>APOE-APOC1/TOMM40/rs2075650</i>	<i>APOE-APOC1/rs2075650^a</i>	1f
		<i>GIP/rs2291725</i>	1f
<i>GIP/rs4794004</i>		1d	
<i>SNF8/rs1994970</i>		1f	
<i>SNF8/rs4793992</i>		1f	
<i>UBE2Z/rs12601672</i>		2b	
<i>UBE2Z/rs15563</i>		1f	
<i>UBE2Z/rs3744608</i>		2a	
<i>UBE2Z/rs3848460</i>		1f	
<i>UBE2Z/rs46522^a</i>		1f	
<i>UBE2Z/rs11079844</i>		1f	
<i>ZNF259-APOA5-APOA1/rs964184</i>	<i>ZNF259-APOA5-APOA1/rs964184</i>	1f	
<i>SMG6/rs2281727</i>	<i>SMG6/rs2281727^a</i>	2b	
	<i>SMG6/rs7217687</i>	2b	
	<i>SMG6/rs9908888</i>	2b	
<i>COL4A1-COL4A2/rs4773144</i>	<i>COL4A1-COL4A2/rs4773144^a</i>	2b	
<i>ABO/rs579459</i>	<i>ABO/rs649129</i>	2b	
<i>ADMTS7/rs7173743</i>	<i>LOC105370915/rs5029904</i>	2b	
	<i>PHACTR1/rs4773143</i>	2b	
<i>CXCL12/rs501120</i>	<i>CXCL12/rs518594</i>	2b	
	<i>CXCL12/rs1746052</i>	2b	

(Continued)

Table 3. Continued

GWAS SNPs	Functional proxy SNPs	Regulome DB score
<i>FURIN-FES/rs17514846</i>	<i>FES/rs1894401</i>	1b
<i>HHIPL1/rs2895811</i>	<i>HHIPL1/rs28391527</i>	2b
	<i>HHIPL1/rs4624107</i>	2b
	<i>HHIPL1/rs7145262</i>	2b
<i>IL6R/rs4845625</i>	<i>IL6R/rs7549250</i>	2b
	<i>IL6R/rs7549338</i>	2b
	<i>IL6R/rs7553796</i>	2b
<i>KCNE2/rs9982601</i>	<i>KCNE2/rs28591415</i>	2b
<i>KIAA1462/rs2505083</i>	<i>KIAA1462/rs3739998</i>	2b
<i>LPL/rs264</i>	<i>LPL/rs271</i>	1f
	<i>LPL/rs3779788</i>	2b
<i>MIA3/rs17465637</i>	<i>MIA3/rs17163301</i>	2b
<i>PLG/rs4252120</i>	<i>PLG/rs4252126</i>	1f
	<i>PLG/rs4252135</i>	1f
<i>PPAP2B/rs17114036</i>	<i>LOC101929929/rs72664304</i>	2a
	<i>PLPP3/rs4634932</i>	1f
<i>SLC22A4-SLC22A5/rs273909</i>	<i>SLC22A5/rs17689550</i>	1f
	<i>SMG6/rs7217687</i>	2b
	<i>SMG6/rs9908888</i>	2b
<i>SORT1/rs602633</i>	<i>CELSR2/rs12740374</i>	2b
	<i>CELSR2/rs629301</i>	1f
	<i>CELSR2/rs646776</i>	1f
<i>TRIB1/rs2954029</i>	<i>LOC105375745/rs2980853</i>	2b
	<i>LOC105375745/rs2001844</i>	2b
	<i>LOC105375745/rs6982636</i>	2b
	<i>TRIB1/rs2980856</i>	2b
<i>VAMP5-VAMP8-GGCX/rs1561198</i>	<i>GGCX/rs6738645</i>	1f
	<i>GGCX/rs10187424</i>	1f
	<i>VAMP8/rs1009</i>	1b
	<i>GGCX/rs6547621</i>	1f
	<i>VAMP8/rs1348818</i>	1f
	<i>GGCX/rs2886722</i>	1f
	<i>VAMP8/rs3770098</i>	1f
	<i>VAMP8/rs6757263</i>	1f
<i>WDR12/rs6725887</i>	<i>ICA1L/rs72934715</i>	2b
	<i>NBEAL1/rs2351524</i>	1f
	<i>WDR12/rs72936852</i>	2b
	<i>NBEAL1/rs4675310</i>	1f
	<i>NBEAL1/rs72934512</i>	2b
<i>REST-NOA1/rs17087335</i>	<i>REST/rs2227901</i>	1f
	<i>REST-NOA1/rs7687767</i>	1d
<i>SWAP70/rs10840293</i>	<i>SWAP70/rs93138</i>	1f
	<i>SWAP70/rs360136</i>	1f
<i>SMAD3/rs56062135</i>	<i>SMAD3/rs17293632</i>	2a
	<i>SMAD3/rs1866316</i>	2b
	<i>MTERF1/rs8032739</i>	2b
<i>CDKN2BAS1/rs1333049</i>	<i>CDKN2BAS1/rs4977574</i>	2c

^aGWAS significant SNPs with functional evidence (RegulomeDB score < 3) are bolded.

It appears to affect the binding of SPI1. Twenty SNPs returned a score of 1f (likely to affect the binding), and 18 of them were in intronic regions. *NT5C2/rs11191558* lies in HOXC series of DNA motifs, and *CNNM2/rs3781285* lies between NF-kappaB and P50:50. *NT5C2/rs2297787*

returned a score of 2a, affecting the binding motifs of FOXI1, HNF3-alpha and FOXP1 and the binding of FOXA1. SNP *rs12412038* is located in Intron 10 of *NT5C2* and is in the binding motif of Irx. The remaining two SNPs, *rs12219901* and *rs12221064*, lie in the

CNNM2-NT5C2 intergenic region and upstream of *CNNM2*, respectively. They are located *in* DNA motifs of SRF and MAZR and affect the binding of POLRA2 and CTCF/ETS. Interestingly, rs943037 resides in exon 7 of *CNNM2*. Nineteen of the 25 SNPs in the region of *CYP17A1-CNNM2-NT5C2* are eQTLs for *USMG5* (Table 4).

One SNP rs2075650 lies in Intron 2 of *ApoEapoC1/TOMM40* with a RegulomeDB score of 1f. It is located in RREB1 DNA motif and is an eQTL for *TOMM40* (Table 4).

In total 3 of 107 SMG6 associated SNPs, rs2281727, rs7217687 and rs9908888 had a score of 2b and they affect the binding of EP300. rs2281727 is a genome-wide significant SNP located in Intron 9 of *SMG6*. It is in binding motifs of SRY, Srf and Zfp105 and affects the binding of CREBBP, EP300, STAT3, TRIM28, MYC and RBBP5 (Table 4).

The *UBE2Z* region had 10 functional SNPs, including a GWAS reported SNP, *UBE2Z/rs46522* (RegulomeDB score of 1f). The SNP with the most evidence of regulatory function in this locus is rs4794004 with a score of 1d. It is in DNA motif of Gata5 that alters the expression of *UBE2Z* and *ATP5G1* and affects the binding of NR3C1, IN3AK20, CREB1, TAF12, CTCF, POLR2A, USF1, FOXA1, FOXA2 and RBBP5. The other 5 SNPs in this region have a score of 1f. The remaining two regulatory SNPs, rs3744608 and rs12601672, have scores of 2a and 2b, respectively. rs3744608 is located in Intron 3 of *UBE2Z* and it affects the binding of large number of transcription factors (Table 4).

COL4A1-COL4A2/rs473144 is a GWAS reported SNP, achieving a RegulomeDB score of 2b. This SNP lies in Intron 3 of *COL4A2* between STST3:STAT3 DNA motif and affects the binding of POLR2A and EZH2 (Table 4). *ZNF259-APOA5-APOA1/rs964184* is a GWAS significant SNP with a score of 1f and is an eQTL for TAGLN. This SNP is located downstream of this gene region and is present in FOXJ2 DNA motif. Another GWAS significant SNP, *ZC3HC1/rs11556924* is an exonic variant and the only functional SNP (score = 1f) in this locus; it is also an eQTL for *ZC3HC1* (Table 4).

REST-NOA1/rs17087335 is in LD with two functional SNPs (rs2227901 and rs7687767 with RegulomeDB scores of 1f and 1d, respectively). rs768776 lies in DNA motif of Sox8 and affects the binding of FOXA1. *SWAP70* has two functional SNPs, rs93138 and rs360136, each with a RegulomeDB score of 1f. *SWAP70/rs93138* is an eQTL as evidenced in monocytes.

SMAD3 has three functional SNPs, *SMAD3/rs17293632* and *SMAD3/rs1866316* and *MTERF1/rs8032739* with RegulomeDB scores of 2a, 2a and 2b, respectively. Both are in

LD with a lead GWAS SNP (*SMAD3/rs56062135*). *CDKN2BAS1/rs1333049* has one functional SNP(rs4977574) only with RegulomeDB score of 3c. It is a part of a gene cluster on chromosome 9p21 and it maps to Intron 16 of cyclin dependent kinase, an important regulator of cell cycle.

Discussion

Following the sequencing of human genome, a large number of SNPs have been identified that affect disease phenotypes, but their exact roles remain unclear (16). One possible explanation is that some variation affects disease expression at the transcriptional level other than at the protein level. For example, a base pair change in a transcription factor binding site may affect the binding affinity of transcription factors that consequently may alter the transcription of the related genes. These effects are indirect and may seem subtle, but their interactions with other genetic or environmental factors may result in the pathogenesis of common diseases.

Like other complex disorders, a large number of CAD associated risk variants have been discovered by multiple GWAS (12, 13, 17). ENCODE provides information regarding the functionality of human genome (18). This data requires careful interpretation and helps to define the biological function of previously termed ‘junk DNA’. Using bioinformatics tools, we may generate new hypotheses about the gene regulation of complex disorders. In this study, we have used two bioinformatics tools, SNAP and RegulomeDB, in order to identify the putative roles of CAD-associated SNPs.

We examined a total 1,200 SNPs in 54 loci implicated by GWAS, including 58 genome-wide significant SNPs. Ninety-seven SNPs were predicted to have regulatory functions with a RegulomeDB score of <3, but only 8 of them were genome-wide significant. Interestingly, all 8 genome-wide significant SNPs with suggested regulatory function are located either in intronic or intergenic regions, suggesting that these are true associations that regulate gene expression at the transcriptional level.

Among these eight GWAS reported functional SNPs, the SNP with the top RegulomeDB score was *LIPA/rs2246833* (Regulome DB score = 1b). This variant is located in Intron 6 of lipase A (*LIPA*) and is an eQTL for the same gene which catalyzes intracellular triglyceride and hydrolyses cholesterol ester (19).

ZC3HC1/rs11556924 is a GWAS significant CAD associated SNP that returned a score of 1f. rs11556924 is a coding SNP located in the *ZC3HC1* gene region encoding NIPA (Nuclear Interaction Partner of ALK) protein. This polymorphism is responsible for arginine-histidine amino acid alteration at position 363 (R363H). The SNP has been

Table 4. Putative functional SNPs and corresponding motifs, eQTL and related transcription factors (Regulome DB score < 3)

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
chr15:91429041	rs1894401	1b	<i>FES</i>	<i>Intron 2</i>	FES	Pax5	SPI1 USF1 POLR2A GABPA BHLHE40 CEBPB CTCF MAX RFX5 RUNX3 STAT5A CTCF
chr10:91005853	rs2246833	1b	<i>LIPA</i>	<i>Intron 6</i>	LIPA	EWSR-FLI1 zNF143	CTCF HSF1
chr2:85808736	rs1009	1b	<i>VAMP8</i>	<i>Exon 3</i>	VAMP8 LOC388969		CTCF NR3C1
chr17:47038470	rs4794004	1d	<i>GIP</i>	<i>Intron 4</i>	ATP5G1 UBE2Z	Gata5	IN3AK20 CREB1 TAF1 TCF12 CTCF POLR2A USF1 FOXA1 FOXA2 RBBP5
chr10:91002926	rs1412444	1d	<i>LIPA</i>	<i>Intron3</i>	LIPA	SAP1a ELK1 ELK3 ELK4 MECP2 ERF ERG ETS1 ETV1 ETV2 ETV3 Gabpa	ATF2 FOXM1 SP1 SPI1 MTA3 RUNX3
chr10:104873760	rs9633712	1e	<i>NT5C2</i>	<i>Intron 3</i>	UMG5	PU1 ELF-1 Sfpil PU.1 c-Ets-1	SP11
chr11:116648916	rs964184	1f	<i>ZPR1</i>	<i>Downstream ZRP1</i>	TAGLN	FOXJ2	
chr1:109818529	rs646776	1f	<i>CELSR2</i>	<i>Upstream CELSR2</i>	PSMA5		CTCF HEY1 REST POLR2A ZBTB7A
chr10:104616662	rs4409766	1f	<i>BORCS7-ASMT</i>	<i>Intron 1</i>	C10orf77 USMG5	Tcf3	TAF7 BACH1 MAFF

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
chr17:47008206	rs4793992	1f	<i>SNF8</i>	<i>Intron 7</i>	ATP5G1 UBE2Z		MAFK POLR2A TEAD4
chr6:161152293	rs4252126	1f	<i>PLG</i>	<i>Intron 11</i>	PLG		CTCF RUNX3 TEAD4 RAD21
chr6:161154231	rs4252135	1f	<i>PLG</i>	<i>Intron 12</i>	PLG		CTCF FOXA1 NFKB1 RAD21 ZNF263 SMC3 ZNF143 FOXA2
chr10:104846177	rs11191548	1f	<i>NT5C2 gene region</i>	<i>Downstream NT5C2</i>	USMG5	TEAD1 TEAD3	
chr10:104864613	rs11191557	1f	<i>NT5C2</i>	<i>Intron 5</i>	USMG5		
chr10:104864677	rs11191558	1f	<i>NT5C2</i>	<i>Intron 5</i>	USMG5	HOXC13 Hoxa13 Hoxc13 Hoxd12 HOXA13 HOXD9 HOXC11	
chr10:104871203	rs12413046	1f	<i>NT5C2</i>	<i>Intron 3</i>	USMG5		NR3C1 TRIM28 CTCF ATF2 IKZF1 TCF7L2 ZNF263
chr10:104871278	rs10883832	1f	<i>NT5C2</i>	<i>Intron 3</i>	USMG5		TRIM28 TCF7L2
chr10:104913652	rs11191582	1f	<i>NT5C2</i>	<i>Intron 2</i>	USMG5		EP300 NFIC TCF12 TEAD4 STAT1 ARID3A EP300 JUN RCOR1
chr10:104906210	rs11191580	1f	<i>NT5C2</i>	<i>Intron 2</i>	USMG5		TRIM28 SETDB1 GATA1 GTF2F1 CEBPB FOS JUND ZNF263
chr10:104856161	rs12412038	1f	<i>NT5C2</i>	<i>Intron 10</i>	USMG5	Irx-3 Irx-2	

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
chr2:85807081	rs1348818	1f	VAMP8	<i>Intron 2</i>	GGCX	Irx-4 Irx-6 HMG1Y Mtf1 Srf Zfp105 HMG1Y	EBF1,
chr2:85805366	rs3770098	1f	VAMP8	<i>Intron1</i>	VAMP8 LOC388969		POLR2A BHLHE40 E2F6 KDM5B MAX MXI1 MYC NFIC WRNIP1
chr2:85803541	rs6757263	1f	VAMP8	<i>Upstream VAMP8</i>	GGCX VAMP8 LOC388969		SP1 EP300 NFIC
chr17:46988596	rs46522	1f	UBE2Z	<i>Intron 2</i>	ATP5G1 UBE2Z		NFKB NFYB RUNX3
chr19:45395618	rs2075650	1f	TOMM40	<i>Intron 2</i>	TOMM40	RREB1	
chr1:56996190	rs4634932	1f	PLPP3	<i>Intron 2</i>	PPAP2B		POLR2A
chr2:203880833	rs4675310	1f	NBEAL1	<i>Intron 1</i>	ALS2CR13		
chr10:104681142	rs17115213	1f	CNNM2	<i>Intron 1</i>	USMG5		
chr10:104721125	rs10883808	1f	CNNM2	<i>Intron 1</i>	USMG5		
chr10:104723619	rs11191479	1f	CNNM2	<i>Intron 1</i>	USMG5		GATA1 TAL1 CEBPB
chr10:104773363	rs11191514	1f	CNNM2	<i>Intron 1</i>	USMG5		PAX5
chr10:104776526	rs11191515	1f	CNNM2	<i>Intron 1</i>	USMG5		
chr10:104825664	rs3781285	1f	CNNM2	<i>Intron 4</i>	USMG5	NF-kappaB P50:50	IKZF1
chr10:104835918	rs943037	1f	CNNM2	<i>Exon 7</i>	USMG5	TBX20 Foxj1	
chr8:19813701	rs271	1f	LPL	<i>Intron 6</i>	LPL		
chr17:47039131	rs2291725	1f	GIP	<i>Exon 4</i>	GIP		GATA2 TCF4 FOSL2 EGR1 ELF1 FOS NR3C1 EP300 RXRA CHD2 JUND POLR2A RAD21 FOSL1 REST
chr2:203880991	rs2351524	1f	NBEAL1	<i>5' UTR</i>	ALS2CR13		

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
chr1:109818305	rs629301	1f	<i>CELSR2</i>	3' UTR	PSRC1		CTCF POLR2A
chr2:85774675	rs6547621	1f		3' UTR	GGCX		ELK4 POLR2A
chr10:104660003	rs11191454	1f	<i>AS3MT</i>	<i>Intron10</i>	USMG5		
chr10:104685298	rs12411886	1f	<i>CNNM2</i>	<i>Intron1</i>	USMG5	Zec	
chr10:104719095	rs12413409	1f	<i>CNNM2</i>	<i>Intron1</i>	USMG5		POLR3A
chr10:104764270	rs11191499	1f	<i>CNNM2</i>	<i>Intron1</i>	USMG5		
chr17:47014126	rs1994970	1f	<i>SNF8</i>	<i>Intron4</i>	ATP5G1 UBE2Z	TFII-I	
chr2:85742296	rs2886722	1f	<i>Pseudogene</i>		LOC388969		TCF7L2
chr2:85783127	rs6738645	1f	<i>GGCX</i>	<i>Intron5</i>		Evi-1	POLR2A
chr2:85794296	rs10187424	1f	<i>Pseudogene</i>		GGCX LOC388969		
chr5:131723064	rs17689550	1f			RAPGEF6		
chr7:129663495	rs11556924	1f	<i>ZC3HC1</i>	<i>Exon8</i>	KIAA0265		
chr17:4702833	rs11079844	1f	<i>Pseudogene</i>		ATP5G1		
chr17:47005192	rs15563	1f	<i>UBE2Z</i>	<i>Exon7</i>	ATP5G1	PRDM1	
chr17:47047113	rs3848460	1f	<i>UBE2Z</i>		ATP5G1		CEBPB
chr10:104680136	rs2297787	2a	<i>CNNM2</i>	<i>Intron 1</i>		Freac-7 HFH3(FOXI1) HNF3alpha FOXP1 Elf3 Foxl1 Srf Tcf3 Tcfap2e Zfp105 HFH(FOXI1)	FOXA1 SIN3A ZNF263 HNF4G FOXA1
chr1:56948289	rs72664304	2a	<i>C8B</i>	<i>Intron 6</i>		FOXA1 Foxa2	FOXA1 FOXA2 TCF4 SP1 HNF4G HNF4A HDAC2 JUND EP300
chr17:46993232	rs3744608	2a	<i>UBE2Z</i>	<i>Intron 3</i>		Zfp740 MZF1 MAZR SP1 SP1:SP3 WT1ZNF21 Zfp281 ZFp740 WT1 ZNF219 ZNF740 SP4	SPI1 POLR2A IKZF1 MAX TFAP2A TFAP2C SP1 CEBPB NR3C1 BATF BCL11A MEF2A NFKB1 JUND

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
							EP300 STAT3 IRF4 EBF1 FOSL2 BATF NR3C1 RUNX3 MYC STAT3 CTCF POLR2A TAF1 RFX5 RAD21 HEY1 CDX2 HNF4A ZNF263 NR3C1 CTCF MYC AR MYBL2 TEAD4 MAZ CHD2 SMC3 TBP ZNF143 CDX2 E2F6 MAX NR3C1 SIN3A YY1 REST HMGN3 POLR2A EGR1 SPI1 ELF1
chr2:85764959	rs1446668	2a	MAT2A	<i>nc transcript</i> <i>Upstream MAT2A</i>		CTCF	CTCF POLR2A TAF1 RFX5 RAD21 HEY1 CDX2 HNF4A ZNF263 NR3C1 CTCF MYC AR MYBL2 TEAD4 MAZ CHD2 SMC3 TBP ZNF143 CDX2 E2F6 MAX NR3C1 SIN3A YY1 REST HMGN3 POLR2A EGR1 SPI1 ELF1
chr17:47006492	rs12601672	2b	UBE2Z	<i>Downstream UBE2Z</i>		Zfx	CTCF MYC PAX5 ZNF143 JUND POLR2A TFAP2C MXI1 CEBPB NFYA POLR2A
chr10:30316071	rs3739998	2b	KIAA1462	<i>Exon 2</i>		RELA	CTCF MYC PAX5 ZNF143 JUND POLR2A TFAP2C MXI1 CEBPB NFYA POLR2A
chr8:126476378	rs2980856	2b	<i>TRIB1 gene</i> <i>region</i>	<i>Intergenic region</i> <i>Downstream TRIB1</i>		pax-8 Sox17	CTCF MYC PAX5 ZNF143 JUND POLR2A TFAP2C MXI1 CEBPB NFYA POLR2A
chr9:136154303	rs649129	2b	<i>ABO gene region</i>	<i>Intergenic region</i> <i>Upstream ABO</i>		IRF	CTCF MYC PAX5 ZNF143 JUND POLR2A TFAP2C MXI1 CEBPB NFYA POLR2A

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
							FOS IRF1 NFYB PML POLR2A
chr10:104840966	rs12219901	2b	<i>CNNM2</i> gene region	<i>Intergenic region</i> <i>Downstream CXCL12</i>		SRF	
chr10:44778545	rs1746052	2b	<i>CXCL12</i> gene region	<i>Intergenic region</i> <i>Downstream CXCL12</i>		GATA1	TAL1
chr21:35593826	rs28451064	2b	<i>LINC00310</i> gene region	<i>Intergenic region</i> <i>Downstream LINC00310</i>		PPAR	SP1 FOXA2
chr17:2098271	rs7217687	2b	<i>SMG6</i>	<i>Intron 13</i>	-	NF-1	SIN3A TCF12 MAX YY1 ZNF263 EP300 TEAD4
chr13:110960711	rs4773144	2b	<i>COL4A2</i>	<i>Intron 3</i>		STAT3:STAT3	POLR2A EZH2
chr14:100116251	rs28391527	2b	<i>HHIPL1</i>	<i>Intron 3</i>		MyoD SCRT1 FIGLA	BHLHE40 USF1 FOXA1 MAX
chr1:154404335	rs7549250	2b	<i>IL6R</i>	<i>Intron 3</i>		TBX15	MXI1 FOS JUNB MAX JUND JUN STAT3 FOSL1 MAFK RCOR1 MYC USF2 TEAD4 RCOR1 YY1
chr1:154404379	rs7549338	2b	<i>IL6R</i>	<i>Intron 3</i>		GR AR	FOS JUNB JUND JUN STAT3
chr1:154404405	rs7553796	2b	<i>IL6R</i>	<i>Intron 3</i>		NF-kappaB,	FOS JUND JUN STAT3
chr14:100127439	rs4624107	2b	<i>HHIPL1</i>	<i>Intron 7</i>		Pax5	JUND
chr10:91011457	rs1332328	2b	<i>LIPA</i>	<i>Intron 9</i>		UF1H3BETA	CREBBP ZNF263 CDX2 ELF1

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
							ZEB1 TBP TFAPC2 TBP POLR2A ETS1 GABPA HEY1 CREBBP
chr17:2117944	rs2281727	2b	SMG6	<i>Intron 13</i>		SRY Srf Zfp105	EP300 STAT3 TRIM28 MYC RBBP5
chr8:126479314	rs6982636	2b	LOC105375745	<i>Intron1</i>		MAF	SMARCC1 RFX3 POLR2A GATA2 CHD2 GTF2F1
chr14:100125720	rs7145262	2b	HHIPL1	<i>Intron4</i>		ESR2	SMARC4 ZBTB7A SMARB1 POLR2A EZH2 RAD21 BACH1
chr10:104677125	rs12221064	2b	CNNM2	<i>Upstream CNNM2</i>		MAZR,	CTCF, ETS1 ETS1
chr8:126478349	rs2980853	2b	LOC105375745	<i>Upstream LOC105375745</i>		Pit-1,	RFX3
chr15:79152421	rs5029904	2b	LOC105370915	<i>Upstream LOC105370915</i>		NeuroD	USF1 POLR2A YY1 FOXA1 E2F4 MAX TAF7 TAF1 MXI1
chr8:126478744	rs2001844	2b	LOC105375745	<i>Upstream LOC105375745</i>		HSF1	RFX3
chr10:91011680	rs1332327	2b	LIPA	<i>5' UTR</i>		AP-4,	CREBBP CDX2 ELF1 TBP SPI1 NRF1 ELF1 ETS1 GABPA SPI1 PAX5

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
chr17:2102452	rs9908888	2b	SMG6	<i>Intron10</i>		GR AR	SREBF1 CEBPB
chr10:91008878	rs2250644	2b	LIPA	<i>Intron1</i>		Oct-1 XBP-1 MAfb Mafk MAFB MAFK NRL	RUNX3
chr1:109817589	rs12740374	2b	CELSR2	<i>Exon34</i>		HNF1 HNF1A DUXA	EBF1
chr1:222794090	rs17163301	2b	MIA3	<i>Intron1</i>		HNF1 HNF1A HNF1B DUXA	EBF1
chr21:35644028	rs28591415	2b	<i>pseudogene</i>			PPAR	EP300 FOXA1 HDAC2 NFIC SP1
chr2:203713279	rs72934715	2b	ICA1L	<i>Intron2</i>		HMG1Y	ATF2 NFIC EBF1 EP300 NFKB1 PAX5
chr2:203775474	rs72936852	2b	WDR12	<i>Intron1</i>		AR	MAFF
chr2:203926270	rs72934512	2b	NBEAL1	<i>Intron6</i>		TEAD1 TEAD3	TEAD4
chr8:19803092	rs3779788	2b	LPL	<i>Intron1</i>		TGIF	CEBPB
chr10:44757106	rs518594	2b	CXCL12	<i>Downstream intergenic</i>		E2A NRSE NRSF	FOXM1 NFIC MAX EBF1 TBL1XR1
chr4:57824931	rs7687767	1d	CECR6			Sox8	FOXA1
chr11:5759712	rs93138	1f	SWAP70	<i>Intron8</i>			
	rs360136	1f	SWAP70	<i>Exon13</i>			
chr:457798188	rs2227901	1f	REST-NOA1	<i>Exon6</i>			SPI1
chr:1567442595	rs17293632	2a	SMAD3	<i>Intron4</i>		Bach1 AP-1 JundM2 Pou1f1 Pou3f1 Sox5 JDP2	SIN3A TCF7L2 TFAP2A TFAP2C ZNF217 POLR2A STAT1 MXI1 TAF1 E2F1 POLR2A HDAC1

(Continued)

Table 4. Continued

Coordinate 0-based	SNP ID	RegulomeDB score	Gene/Locus	Position	eQTL	Motif	Protein-binding
							TCF7L2 MYC POLR2A ESR1 EP300 CDX2 HNF4A CEBPB EP300 FOS FOSL1 GATA3 JUNB JUN MYC NR2F2 NR3C1 RAD21 RCOR1 TCF12 JUND MAFK EGR1 MAX
chr15:67441996	rs18866316	2b	SMAD3	Exon4		AIRE Elf3 Srf Tcf3 Tcfap2e	ESR1 NR3C1 POLR2A
chr15:67448898	rs8032739	2b	MTERF	Intron4		SRF	CEBPB FOS MYC STAT3
chr9:22098573	rs4977574	2c	CDKN2BAS1	Intron16		AR NR3C1 NR3C2 Ar Elk-1 c-Ets-1(p54)	AR

associated with essential hypertension in Finnish population (20, 21).

The *CYP17A1-CNNM2-NT5C2* gene region has the highest number of regulatory SNPs, including one GWAS significant SNP, rs12413409. This locus affects diastolic blood pressure, systolic blood pressure and body mass index. All three measures are important risk factors for CAD (22). There are 25 putative regulatory SNPs in LD with rs12413409 that are located across four genes on chromosome 10 (*CNNM2*, *NT5C2*, *AS3MT* and *BORCS7/ASMT*) but affect the expression of same protein

USMG5. These findings suggest that USMG5 should be investigated as an important player for CAD pathogenesis. USMG5 (upregulated during skeletal muscle growth protein 5) is also known as diabetes-associated protein in insulin-sensitive tissues that plays a crucial role in the maintenance of ATP synthase structure in mitochondria (23). Chen et al. (24) have purified this protein from bovine heart mitochondria and suggested its role in cell energy metabolism.

APOE-APOC1/TOMM40/rs2075650 is present in the *TOMM40* gene region near the *APOE-C1* cluster.

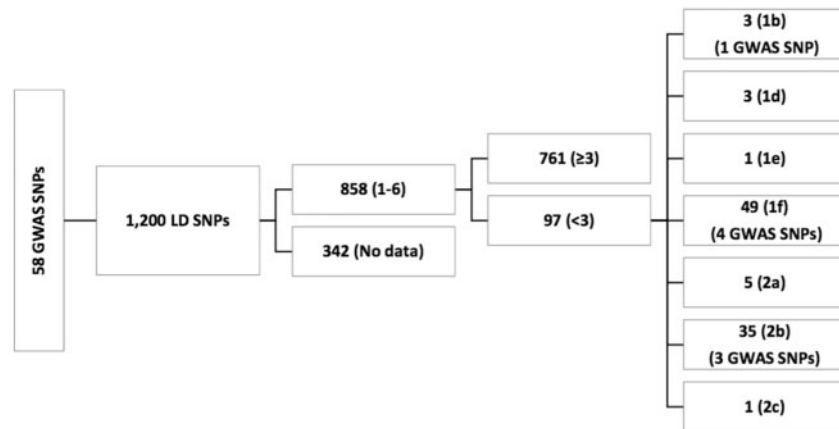


Figure 1. 58 GWAS ANPs in LD with 1200 SNPs. We used SNAP webportal to determine LD SNPs. These 1200 SNPs were further evaluated by RegulomeDB to identify their functional role. RegulomeDB did not provide data for 342 SNPs. A total of 858 SNPs returned the scores of 1–6 by RegulomeDB. Of those 858 SNPs, 97 returned the scores of <3. Among 97 functional SNPs, only 8 were GWAS SNPs. Lower the RegulomeDB score, more evidence of functionality.

TOMM40 encodes TOMM40 protein, which is an important subunit 40 of outer mitochondrial membrane protein complex. rs2075650 risk allele has shown an association with low levels of CRP in CAD patients (25).

rs46522, an intronic SNP in Ubiquitin-conjugating enzyme E2Z (*UBE2Z*) gene region returns a RegulomeDB score of 1f. This SNP is associated with CAD in Iranian and Han Chinese populations (26, 27). The exact mechanism by which genetic alteration in *UBE2Z* can attribute to the CAD risk is not yet clear; however, rs46522 is in strong LD with the causal SNPs in gastric inhibitory peptide (GIP) gene that encodes GIP protein, a protein that modifies the glucose and lipid metabolism potentially mediating known CAD risk factors.

ZNF259-APOA5/APOA1/rs964184 is also an important regulatory SNP. *ZNF259* protein polymorphism has been associated with metabolic syndrome in Chinese population. Aung et al. (28) have also shown its association with lipid levels. *ZNF259* is located close to *APOA5*. Overexpression of *APOA5* in mice reduces plasma triglyceride levels and mice lacking *APOA5* have hypertriglyceridemia (29).

COL4A2/rs4773144 has been identified as functional lead SNP by RegulomeDB (score = 2b). This gene controls collagen proliferation, indicating a potential functional role in atherosclerotic plaque strengthening (30).

SMG6/rs2281727 is an intronic SNP. The potential function of *SMG6* in CAD is not yet established. This gene promotes the endonuclease activity and is responsible for protection of telomere ends of chromosomes (16).

Although regulatory elements are most often found in non-coding regions of the genome, we found 5 loci with exonic regulatory SNPs (*VAMP8/rs1009*, *CNNM2/rs943037*, *GIP/rs2291725*, *KIAA1462/rs3739998* and

UBE2Z/rs15563), indicating the presence of regulatory signals inside the coding sequences as well.

USF1 is an upstream transcription factor whose binding is affected by three SNPs (*GIP/rs4794004*, *FES/rs1894401*, *HHIPL1/rs28391527*), suggests a potential functional link between *FES*, *GIP* and *HHIPL1* (31).

RegulomeDB identified three important functional SNPs affecting CAD phenotype. Among these, *REST-NOA1/rs17087335* is the lead GWAS SNP that encodes a transcription factor which suppresses the voltage gated sodium and potassium channels and it has shown to maintain vascular smooth muscle cells in non-proliferative phase (32). *SWAP70/rs10840293* encodes a signaling molecule that is implicated in cell adhesion and migration and it appears to be a potential regulator of leukocyte migration and their adhesion to endothelial cells (33). *SMAD3* is a major regulator of TGF- β . A study on mice has shown that mutations in this gene lead to decreased connective tissue deposition in response to vascular injury (34).

It should be noted that 342 SNPs had returned ‘No Data’ when queried by RegulomeDB. This suggests that current evidence does not support a functional role for those variants. Our results also showed that some loci harbor markedly more regulatory SNPs as compared with other regions. We caution against interpreting this finding to mean that one region is more functionally relevant, as regions with ‘fewer’ functional SNPs may have yet to be interrogated as thoroughly and thus have fewer annotations.

Since these loci are mostly in Europeans, and only 5 of them are replicated in South Asians (35), the findings may not be as relevant to other populations as they are to Europeans as genetic effects can differ across populations. The cause of this varying association with disease phenotype may be the ethnic admixture resulting in population

stratification. It is also noteworthy that robust associations of variants with different diseases have been reported in Europeans while other populations (Africans, Asians and Hispanics) failed to demonstrate those associations (36, 37).

Though the cellular mechanisms underlying CAD pathogenesis are established, the molecular basis is not yet agreed upon. Comprehending the molecular basis of disease is crucial before pathogenesis is completely described. The study has identified 97 regulatory SNPs associated with CAD. In summary, our results highlight the importance of considering both disease-associated SNPs and those SNPs in LD, as well as the regulatory function of these SNPs to help identify the causal genetic mechanisms of CAD. The methods which we have implemented here can inform planning of more complete and better directed functional genomic studies.

Supplementary data

Supplementary data are available at *Database* Online.

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Conflict of interest. None declared.

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