# CORRESPONDENCE

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# A novel genotype of "Anaplasma capra" in wildlife and its phylogenetic relationship with the human genotypes

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Anaplasma spp. are gram-negative obligate intracellular bacteria of remarkable importance in both human and veterinary health. Six Anaplasma species have currently been classified in the genus Anaplasma and are known as causative agents of tick-borne diseases<sup>1</sup>. The impacts of the diseases caused by Anaplasma on the productivity and health of animals have been recognized for over a century, and it is still a significant threat to livestock industries<sup>2</sup>. A zoonotic role has recently been uncovered for Anaplasma phagocytophilum, resulting in a rising interest in these organisms<sup>3</sup>. Anaplasma phagocytophilum has been considered the main cause of human anaplasmosis worldwide in recent years. Recently, a potential novel tick-transmitted Anaplasma species was identified in asymptomatic goats in China, and it has been recognized as a cause of human infection; the name "Anaplasma capra" was proposed provisionally<sup>4</sup>. Sequence and phylogenetic analyses of "Anaplasma capra" based on different gene loci revealed that this organism was distinct from all known Anaplasma species<sup>4</sup>. Although "A. capra" has not yet been formally recognized, human cases of "A. capra" infection have been recorded in northeastern China, and subsequent reports have also shown that "A. capra" seems to be widely distributed in China<sup>4-7</sup>, suggesting this agent may pose a substantial public health concern and may be responsible for anaplasmosis cases of unknown cause.

Wild animals are important reservoirs for tick-borne pathogens. The zoonotic *A. phagocytophilum* has been identified in a great number of wild animal species<sup>2</sup>. In previous reports, specific DNA of "*A. capra*" has been detected in domestic and some species of wild animals<sup>5,7–9</sup>, and "*A. capra*"-like bacteria were identified in ticks<sup>10</sup>, indicating "*A. capra*" might have a broad host range and genetic diversity.

This study was conducted in wild animals from Tangjiahe National Nature Reserve, which is a national protected area located in Sichuan Province, China (Supplementary Figure S1). This reserve is one of the biodiversity hotspots in the world and is an important natural habitat for a great number of endangered wildlife. From March 2016 to April 2017, 13 dead freeranging wild animals were found in the field, including the first-grade nationally protected takin (Budorcas taxicolor) (five) and forest musk deer (Moschus berezovskii) (one); the second-grade nationally protected Himalayan goral (Naemorhedus goral) (three); and two nonendangered protected species, Reeves's muntjac (Muntiacus reevesi) (three), and wild boar (one). Blood or liver samples were collected individually from animals, frozen at -20 °C, and transported in a freezer pack to a laboratory immediately. The total DNA was extracted using a QIAamp DNA Mini Kit according to the protocol. This study was approved by the Animal Ethics Committee of Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Sciences.

AllDNA samples were screened for the presence of "*A. capra*" by nested PCR targeting on *gltA* gene as previously described<sup>4,10</sup>. Positive (DNA extracted from a goat infected by "*A. capra*", KX417336) and negative controls (double-distilled water) were included in each

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assay. "Anaplasma capra" was detected in six wild animals, including three takins, two Reeves's muntjacs, and one forest musk deer (Supplementary Table S1). To further characterize the "A. capra" strains identified in this study, the 16S rRNA and msp4 genes were amplified from the *gltA* gene-positive samples<sup>5,10</sup>. The PCR products of the partial 16S rRNA (1261 bp), gltA(594 bp), and msp4 (656 bp) gene were purified using the AxyPrep DNA gel extraction kit (Axygen, USA), cloned (pGEM-T Easy vector, Promega, USA) and sequenced (GenScript, Nanjing, China). Sequence analysis revealed that the 16S rRNA sequences (MH188286) obtained in this study were 100% identical to each other and to those of "A. capra" isolates identified from sheep (MF066918), H. qinghaiensis ticks (KX673825) and Japanese serows (AB509223); and they were 99.8% identical to those of the isolates identified from humans (HLJ-14, KM206273) and *H. longicornis* ticks (KP314237). Phylogenetic analysis based on the 16S rRNA gene demonstrated that the isolates were clustered within the clade inside the"*A. capra*" but distinct from other, well-defined *Anaplasma* species (Fig. 1a).

According to the *gltA* sequence alignment, five sequence variants (MH192359–MH192363) with 2–6 bp nucleotide substitutions were obtained and showed 98.4–100% identity to those of "*A. capra*" isolates from sheep (MF071308 and MF071309) and *H. qinghaiensis* ticks (KX685885 and KX685886), but they were 87.2–88.3% identical to the isolate HLJ-14 from human (KM206274). Moreover, the *msp4*sequences of these isolates showed 88.4% identity to "*A. capra*" isolate HLJ-14 (KM206277). Phylogenetic analysis of both *gltA* and *msp4* gene sequences revealed that the "*A. capra*" strains

were clustered independently from *Anaplasma* species identified previously and formed two distinct subclades with high bootstrap values. The isolates identified in this study were closely related to "*A. capra*" isolates from *H. qinghaiensis* and sheep but clearly distinct from human isolate HLJ-14 (Fig. 1b, c).

Anaplasma circulation in nature involves tick vectors and a great number of vertebrates that act as hosts and sources of infection for ticks, animals, and humans<sup>2</sup>. Since Anaplasma spp. are transmitted by ixodid ticks transstadially rather than transovarially, the reservoir hosts play a critical role in the maintenance and dispersal of these bacteria<sup>11</sup>. Numerous wild animal species have been considered competent reservoir hosts for Anaplasma, especially wild ruminants<sup>11</sup>, such as red deer (Cervus elaphus), roe deer (Capreolus capreolus), white-tailed deer (Odocoileus virginianus), mouflon (Ovis musimon), and chamois (*Rupicapra rupicapra*)<sup>11</sup>. In the present study, the novel tick-transmitted zoonotic "A. capra" was first identified in free-ranging wild animals (takin, Reeves's muntjac, and forest musk deer) from a national nature reserve, where contact between wild and domestic animals rarely occurs. These results suggest that "A. capra" is endemic in this protected area and that those wild animals can act as competent sylvatic reservoirs for this organism.

The 16S rRNA gene sequence of "A. capra" was first described in cattle from Japan in 2001<sup>12</sup>, and the agent was mistakenly assumed to be A. centrale (Aomori strain, AF283007) (Fig. 1a)<sup>13</sup>. More recently, this novel Anaplasma species was recorded in goats from northeastern China, and it was subsequently identified as a human pathogen in an active surveillance of patients who sought treatment after a tick bite<sup>4</sup>. "Anaplasma capra" has been detected in several tick species, including Ixodes persulcatus, Haemaphysalis longicornis, and Haemaphysalis qinghaiensis<sup>4,6,10</sup>; however, the vector competence of these ticks for transmission of "A. capra" is still unclear and needs to be further evaluated. The infection of "A. capra" has also been reported in sheep and goats from several provinces of China<sup>5</sup>, in deer (Anaplasma sp. NS104, AB454075) and free-living serows (Anaplasma sp. Kamoshika17, AB509223) from Japan<sup>8</sup>, and in cattle (MG910977-9) from Malaysia<sup>9</sup>. Those findings together with the results obtained in this study suggested that additional tick species and/or domestic and wild animals might be involved in the transmission and maintenance of "A. capra", which warrants further investigation.

Sequence and phylogenetic analysis based on the 16S rRNA gene demonstrated that "*A. capra*" isolates obtained from ticks, domestic, and wild animals as well as humans exhibit high sequence similarities to each other (99.8–100%) and cluster in a separate clade within the genus *Anaplasma*, but they are clearly distinct from the other members of *Anaplasma*, indicating the novelty of this causative agent.

Obviously, these isolates of "A. capra" were a single species according to the classification criteria of bacteria<sup>1,13</sup> (>99% 16S rRNA gene sequence similarity). However, they were classified into two different groups with low sequence similarity and a divergent phylogenetic relationship based on the gltA and msp4 genes (87.2-88.3% for gltA and 88.4% for msp4), indicating two genotypes of "A. capra" in ticks and reservoir hosts. The "A. capra" genotype 1 identified in I. persulcatus, H. longicornis, sheep, goat, and human significantly differs from the genotype 2 identified in H. qinghaiensis, takin, Reeves's muntjac and forest musk deer, suggesting a high degree of genetic diversity and host tropisms of "A. capra", as has been documented in A. phagocytophilum<sup>2</sup>. However, it is unclear whether these two genotypes of "A. capra" have pathogenicity variation for animals and human, which should be clarified in the future.

In the past three decades, an increasing number of novel tick-associated microbes with zoonotic potential continue to be discovered, representing a considerable impact on public health worldwide. Since A. phagocytophilum was recognized as the causative agent of human granulocytic anaplasmosis, the cases have constantly increased in many countries<sup>14</sup>. However, numerous anaplasmosis cases of undetermined cause remain, indicating additional Anaplasma species are responsible for human anaplasmosis<sup>2,15</sup>. However, clinical cases of anaplasmosis might be generally neglected because the symptoms of the illness are notoriously nonspecific, making it likely to be confused with other illnesses<sup>14</sup>. The identification of the novel ticktransmitted zoonotic "A. capra" calls for clinicians to be aware of the possible infection from this causative agent in patients suspected of anaplasmosis or other tick-borne diseases, especially in areas where anaplasmosis can occur.

In summary, "A. capra" infection was first documented in three species of free-ranging wild ruminants from a national nature reserve of China, and a novel genotype of "A. capra" was identified that clearly differed from the genotype identified from human. Further studies should be conducted to fully elucidate the host range, vector ticks, pathogenicity, and geographic distribution of this organism.

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#### Conflict of interest

The authors declare that they have no conflict of interest.

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## References

- Dumler, J. S. et al. Reorganization of genera in the families Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: unification of some species of Ehrlichia with Anaplasma, Cowdria with Ehrlichia and Ehrlichia with Neorickettsia, descriptions of six new species combinations and designation of Ehrlichia equi and 'HGE agent' as subjective synonyms of Ehrlichia phagocytophila. Int. J. Syst. Evol. Microbiol. 51, 2145–2165 (2001).
- Battilani, M., De Arcangeli, S., Balboni, A. & Dondi, F. Genetic diversity and molecular epidemiology of Anaplasma. *Infect. Genet. Evol.* 49, 195–211 (2017).
- Chen, S.-M., Dumler, J. S., Bakken, J. S. & Walker, D. H. Identification of a granulocytotropic *Ehrlichia* species as the etiologic agent of human disease. *J. Clin. Microbiol.* **32**, 589–595 (1994).
- Li, H. et al. Human infection with a novel tick-bome Anaplasma species in China: a surveillance study. Lancet Infect. Dis. 15, 663–670 (2015).
- Yang, J. F. et al. A novel zoonotic *Anaplasma* species is prevalent in small ruminants: potential public health implications. *Parasit. Vectors* **10**, 264 (2017).

- Sun, X. F., Zhao, L., Wen, H. L., Luo, L. M. & Yu, X. J. Anaplasma species in China. Lancet Infect. Dis. 15, 1263–1264 (2015).
- Peng, Y. et al. Detection and phylogenetic characterization of *Anaplasma capra*: an emerging pathogen in sheep and goats in China. *Front. Cell. Infect. Microbiol.* 8, 283 (2018).
- Sato, M. et al. Phylogenetic analysis of the 16S rRNA gene of Anaplasma species detected from Japanese serows (*Capricornis crispus*). J. Vet. Med. Sci. 71, 1677–1679 (2009).
- Koh, F. X., Panchadcharam, C., Sitam, F. T. & Sun, T. T. Molecular investigation of *Anaplasma* spp. in domestic and wildlife animals in Peninsular Malaysia. *Vet. Parasitol. Regional Studies Rep.* **13**, 141–147 (2018).
- Yang, J. et al. Molecular survey and characterization of a novel Anaplasma species closely related to Anaplasma capra in ticks, northwestern China. Parasit. Vectors 9, 603 (2016).
- Rar, V. & Golovljova, I. Anaplasma, Ehrlichia, and "Candidatus Neoehrlichia" bacteria: pathogenicity, biodiversity, and molecular genetic characteristics, a review. Infect. Genet. Evol. 11, 1842–1861 (2011).
- Inokuma, H., Terada, Y., Kamio, T., Raoult, D. & Brouqui, P. Analysis of the 16S rRNA gene sequence of *Anaplasma centrale* and its phylogenetic relatedness to other ehrlichiae. *Clin. Diagn. Lab. Immunol.* 8, 241–244 (2001).
- Khumalo, Z. T. H. et al. Evidence confirming the phylogenetic position of Anaplasma centrale (ex Theiler 1911) Ristic and Kreier 1984. Int. J. Syst. Evol. Microbiol. 68, 2682–2691 (2018).
- Sanchez, E., Vannier, E., Wormser, G. P. & Hu, L. T. Diagnosis, treatment, and prevention of lyme disease, human granulocytic Anaplasmosis, and Babesiosis: a review. JAMA 315, 1767–1777 (2016).
- Beyer, A. R. & Carlyon, J. A. Of goats and men: rethinking anaplasmoses as zoonotic infections. *Lancet Infect. Dis.* 15, 619–620 (2015).