



Draft Genome Sequences of Two Cyanobacteria *Leptolyngbya* spp. Isolated from Microbial Mats in Miravalles Thermal Spring, Costa Rica

L. Brenes-Guillén,^a D. Vidaurre-Barahona,^a M. Mora-López,^a L. Uribe-Lorío^{a,b}

^aCenter for Research in Cellular and Molecular Biology, Universidad de Costa Rica, San José, Costa Rica ^bSchool of Agronomy, Universidad de Costa Rica, San José, Costa Rica

ABSTRACT We report the draft genome sequences of *Leptolyngbya* sp. strain 7M and *Leptolyngbya* sp. strain 15MV, isolated from Miravalles Thermal Spring, Costa Rica. The thermophilic cyanobacteria exhibit unique diversity features that provide insight into the adaptation and evolution of phototrophic microorganisms in geothermal habitats.

eptolyngbya sp. is one of the most common filamentous cyanobacteria found in microbial mats in thermal springs (1, 2). Nevertheless, there are few genomes from tropical and thermal environments deposited in any databases. Leptolyngbya sp. strain 7M and Leptolyngbya sp. strain 15MV were isolated from a biofilm in Miravalles Thermal Spring, Costa Rica, in 2004 and 2012, respectively. They were cultivated in BG11 medium (3) and maintained at 25°C. The location, sampling methods, and isolation protocol details were the same as those provided in reference 4. Genomic DNA (gDNA) was isolated using the Plant II kit (Macherey-Nagel) and purified using the NucleoSpin gDNA cleanup kit (Macherey-Nagel), according to the manufacturer's protocol. The total DNA was processed using a Nextera XT DNA library prep kit and MiSeg paired-end sequencing (MiSeq reagent kit v2; 2×250 bp; Illumina, Inc.) at the Center for Research in Cellular and Molecular Biology (CIBCM) of the University of Costa Rica. All the bioinformatics analyses were performed using the Kabré supercomputer, National High Technology Center (CeNAT), Costa Rica. Default parameters were used for all software unless otherwise specified. The sequence reads were filtered using Trimmomatic v0.36 (5) (SLIDINGWINDOW:4:20 and MINLEN of 100 bp), resulting in 753,167 and 544,121 high-quality reads. De novo genome assembly was performed using Unicycler v0.4.8 (6). The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) was used to annotate the contigs for deposit at GenBank. The average nucleotide identity (ANI) was obtained using ChunLab's online ANI calculator (7).

A core phylogeny was constructed using the protein sequences of the ortholog singlecopy genes of *Leptolyngbya* sp. strains 7M and 15MV and published genome sequences derived from hot springs in Yellowstone National Park (8), Tolbo Lake, West Mongolia (9), a thermal spring in Aso-Kuju National Park, Japan (10), soil in Nitzana, Israel, mesophilic freshwater in California, and plankton in the Woods Hole region, MA, USA, using OrthoFinder v2.5.2 (11) (Fig. 1). The tree was inferred using Multiple Sequence Alignments (MSA) with the option "-M msa" (12). We used *Multiple Sequence Comparison by Log-Expectation* (MUSCLE) to perform the multiple alignments of orthogroup sequences. FastTree was used for the maximum likelihood (ML) tree inference (13).

The draft genome sequence of *Leptolyngbya* sp. strain 7M was assembled into nonoverlapping scaffolds totaling 6,990,850 bp, with a G+C content of 49.92%, and it contains 7,020 coding sequences and 88 non-protein-coding genes; the final coverage was $10 \times$. According to the 16S rRNA gene-based sequencing approach (4), isolate 7M was 98.59% identical to the siderophilic *Oscillatoriales* cyanobacterium JSC1.

Citation Brenes-Guillén L, Vidaurre-Barahona D, Mora-López M, Uribe-Lorío L. 2021. Draft genome sequences of two cyanobacteria *Leptolyngbya* spp. isolated from microbial mats in Miravalles Thermal Spring, Costa Rica. Microbiol Resour Announc 10:e00553-21. https://doi.org/10.1128/MRA.00553-21.

Editor J. Cameron Thrash, University of Southern California

Copyright © 2021 Brenes-Guillén et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to L. Brenes-Guillén, laura.brenesguillen@ucr.ac.cr.

Received 27 May 2021 Accepted 24 September 2021 Published 14 October 2021



FIG 1 Phylogeny of 22 cyanobacteria based on a concatenated alignment of the protein sequences of 1,073 ortholog single-copy genes. Bootstrap values (%) are given at the nodes. The scale bar indicates the number of substitutions per site.

The draft genome sequence of *Leptolyngbya* sp. strain 15MV was assembled into nonoverlapping scaffolds totaling 5,089,511 bp, with a G+C content of 66.85%, and it contains 5,299 coding sequences and 86 non-protein-coding genes; the final coverage was $10 \times$.

The average nucleotide identity (ANI)-based comparisons between the two strains was 96.80%; these genomes have low identity (<94% identity, query cover >95%) compared with genomes of *Leptolyngbya* spp. found in thermal springs and other environments. Based on the phylogeny, *Leptolyngbya* sp. strain 7M and *Leptolyngbya* sp. strain 15MV could be new species within the genera.

Data availability. The genome sequences of *Leptolyngbya* sp. strain 7M and *Leptolyngbya* sp. strain 15MV were deposited in DDBJ/ENA/GenBank under accession numbers CP070897.1 and CP071923.1, respectively, and BioProject accession number PRJNA702142; the raw data are publicly available under SRA accession numbers SRR14062498 and SRR14062499, respectively.

ACKNOWLEDGMENT

This work was supported by the Vicerrectoría de Investigación, University of Costa Rica, under project VI-801-C0054.

REFERENCES

- Mackenzie R, Pedrós-Alió C, Díez B. 2013. Bacterial composition of microbial mats in hot springs in Northern Patagonia: variations with seasons and temperature. Extremophiles 17:123–136. https://doi.org/10.1007/ s00792-012-0499-z.
- Rozanov AS, Bryanskaya AV, Ivanisenko TV, Malup TK, Peltek SE. 2017. Biodiversity of the microbial mat of the Garga hot spring. BMC Evol Biol 17: 254. https://doi.org/10.1186/s12862-017-1106-9.
- Rippka R, Cohen-Bazire G. 1983. The Cyanobacteriales: a legitimate order based on the type strain Cyanobacterium stanieri? Ann Microbiol (Paris) 1348:21–36. https://doi.org/10.1016/s0769-2609(83)80094-5.
- Brenes-Guillén L, Vidaurre-Barahona D, Morales S, Mora-López M, Sittenfeld A, Uribe-Lorío L. 2021. Novel cyanobacterial diversity found in Costa Rican thermal springs associated with Rincon de la Vieja and Miravalles volcanoes: a polyphasic approach. J Phycol 57:183–198. https://doi.org/10.1111/jpy.13077.

- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. https://doi.org/10 .1093/bioinformatics/btu170.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Yoon S-H, Ha S-M, Lim J, Kwon S, Chun J. 2017. A large-scale evaluation of algorithms to calculate average nucleotide identity. Antonie Van Leeuwenhoek 110:1281–1286. https://doi.org/10.1007/s10482-017-0844-4.
- Brown II, Bryant DA, Casamatta D, Thomas-Keprta KL, Sarkisova SA, Shen G, Graham JE, Boyd ES, Peters JW, Garrison DH, McKay DS. 2010. Polyphasic characterization of a thermotolerant siderophilic filamentous cyanobacterium that produces intracellular iron deposits. Appl Environ Microbiol 76: 6664–6672. https://doi.org/10.1128/AEM.00662-10.
- 9. Mironov KS, Leusenko PA, Ustinova VV, Bolatkhan K, Zayadan BK, Kupriyanova EV, Shumskaya M, Sinetova MA, Los DA. 2019. Draft genome sequences of a

putative prokaryotic consortium (IPPAS B-1204) consisting of a cyanobacterium (Leptolyngbya sp.) and an alphaproteobacterium (Porphyrobacter sp.). Microbiol Resour Announc 11:e01637-18. https://doi.org/10.1128/MRA.01637-18.

- Nakamori H, Yatabe T, Yoon K-S, Ogo S. 2014. Purification and characterization of an oxygen-evolving photosystem II from Leptolyngbya sp. strain O-77. J Biosci Bioeng 118:119–124. https://doi.org/10.1016/j.jbiosc.2014 .01.009.
- Emms DM, Kelly S. 2019. OrthoFinder: phylogenetic orthology inference for comparative genomics. Genome Biol 20:238. https://doi.org/10.1186/ s13059-019-1832-y.
- 12. Emms DM, Kelly S. 2018. STAG: species tree inference from all genes. bio-Rxiv 267914. https://doi.org/10.1101/267914.
- Price MN, Dehal PS, Arkin AP. 2010. FastTree 2—approximately maximum-likelihood trees for large alignments. PLoS One 5:e9490. https://doi .org/10.1371/journal.pone.0009490.