# A vitamin D analog inhibits Th2 cytokine- and TGFβ -induced periostin production in fibroblasts: a potential role for vitamin D in skin sclerosis

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Abbreviations: LS, localized scleroderma; SSc, systemic sclerosis; 25OH-D3, 25-hydroxyvitamin D3; POSTN, periostin; BLM, bleomycin; OCT, Maxacalcitol (1α, 25-(OH 2-22-oxavitaminD3); NHDFs, normal human dermal fibroblasts.

Scleroderma is an autoimmune disease characterized by extracellular matrix deposition and inflammation. Topical vitamin D analogs have been reported as effective treatments for scleroderma. We previously reported that a matricellular protein, periostin (POSTN), contributes to pathogenesis of scleroderma as POSTN knockout mice were resistant to bleomycin (BLM)-induced scleroderma. We investigated whether a vitamin D analog affects the expression of POSTN in dermal fibroblasts and in a BLM-induced scleroderma model. The vitamin D analog, maxacalcitol (22-oxacalcitriol [OCT]), was applied to dermal fibroblasts and POSTN expression was measured. The effect of OCT on Th2 cytokine- and TGF $\beta$ -induced POTSN and Collagen 1  $\alpha$  1 (Col1A1) expression was also assessed. In vivo, OCT was administered to BLM-induced scleroderma model and outcomes were determined by dermal thickness, collagen density and POSTN expression. Treatment with OCT significantly decreased POSTN expression in dermal fibroblasts. Th2 cytokine- and TGF $\beta$ -induced expression of POSTN and Col1A1 was also suppressed by OCT. In vivo, OCT administration decreased the density of collagen bundles and POSTN expression in a BLM-induced scleroderma model. In addition to the previously reported immunosuppressive effect, the vitamin D analog OCT might be effective to treat scleroderma, in part through inhibition of Th2 cytokine- and TGF $\beta$ -induced POSTN expression.

### Introduction

Scleroderma is an autoimmune disease characterized by extracellular matrix deposition and inflammation. Scleroderma may be further sub-categorized into localized scleroderma (LS), limited cutaneous systemic sclerosis, diffuse cutaneous systemic sclerosis, and systemic sclerosis (SSc).<sup>1</sup> The etiologies of scleroderma are not completely understood. However, a combination of genetics and environmental factors is thought to cause the disease.<sup>2</sup>

Low serum levels of 25-hydroxyvitamin D3 (25OH-D3) have been reported in patients with SSc.<sup>3-6</sup> The average concentration of 25OH-D3 was significantly lower in patients with a high degree of skin involvement (Rodnan skin score > 10) compared with patients with low degree of skin involvement (Rodnan skin score  $\leq 10$ ).<sup>3</sup> The effects of systemic vitamin D supplementation in SSc have been evaluated in some studies, however, the results vary between studies.<sup>7-9</sup>

The effects of topical vitamin D analogs have also been reported to improve scleroderma in LS patients.<sup>10-13</sup> In one study, calcipotriol, a derivative of vitamin D, inhibited the proliferation of fibroblasts derived from LS patients.<sup>14</sup> These data suggest that inhibition of fibroblast proliferation and collagen synthesis is one mechanism by which vitamin D acts to treat LS. Recently, we reported that periostin (POSTN), a matricellular protein, plays an important role in scleroderma.<sup>15</sup> The expression of POSTN was increased in the skin of SSc patients. In addition, POSTN knockout mice were resistant to bleomycin (BLM)induced scleroderma.<sup>15</sup> Serum POSTN levels are also reported to correlate with progressive skin sclerosis in patients with SSc.<sup>16</sup> Furthermore, histamine is reported to induce collagen expression through POSTN.<sup>17</sup> Finally, the Th2 cytokines play an important role as inducers of POSTN in chronic allergic inflammation and in subepithelial fibrosis of bronchial asthma.<sup>18,19</sup>

In this study, we investigated the effect of a vitamin D3 analog, maxacalcitol (22-oxacalcitriol [OCT]), on Th2 cytokine-induced

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**Figure 1.** OCT decreases POSTN and IL-6 expressions in mouse dermal fibroblasts. (**A** and **C**) Mouse dermal fibroblasts were treated with indicated doses of OCT (**A**) or cortisol (GC) (**C**) for 24 h. Expressions of POSTN and IL-6 was measured by RT-PCR. GAPDH served as an internal control. Bars indicate the mean  $\pm$  SD. N = 4; \*P < 0.0001, one-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons. (**B**) Mouse dermal fibroblasts were treated with 10<sup>-7</sup> M of OCT for the indicated lengths of time. Expressions of POSTN and IL-6 was measured by RT-PCR. GAPDH served as an internal control. Bars indicate the mean  $\pm$  SD. N = 4; \*P < 0.0001, one-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons. (**B**) Mouse dermal fibroblasts were treated with 10<sup>-7</sup> M of OCT for the indicated lengths of time. Expressions of POSTN and IL-6 was measured by RT-PCR. GAPDH served as an internal control. Bars indicate the mean  $\pm$  SD. N = 4; \*P < 0.0001, one-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons.

fibrosis. OCT is the most used vitamin D3 analog in Japan, which displays approximately 10 times greater efficacy at suppressing keratinocyte proliferation in vitro compared with calcipotriol

with the approval of the Animal Experiments Committee of Osaka University (#20-003-0). Bleomycin (Nippon Kayaku, Tokyo, Japan) was dissolved in phosphate-buffered saline (PBS)

and tacalcitol.<sup>20</sup> We found that OCT significantly suppressed the expression of IL-4 and IL-13induced POSTN in dermal fibroblasts. We hypothesize that decreased POSTN expression might be one of the mechanisms by which topical treatment with a vitamin D analog is effective in scleroderma, especially in LS.

## **Material and Methods**

## Cell culture

NHDFs were purchased from DS Pharma Biomedical (Osaka, Japan). NHDFs were cultured in Dulbecco's Modified Eagle's Medium (DMEM) containing 10% fetal bovine serum (FBS). Isolation and culture of mouse fibroblasts were carried out as previously described.<sup>21</sup> Fullthickness skin harvested from day 2 to day 4 newborn mice was treated with 4 mg/ml of dispase (Gibco; Invitrogen, Paisley, UK) for 1 h at 37°C. Next, the epidermis was peeled from the dermis. The dermis was placed in DMEM+0.05% type-1 colla-(Sigma- Aldrich, St genase Louis, MO, USA) and incubated at 37°C for 30 min with vigorous agitation to prepare single cells. After filtration, cells were centrifuged at 200 g for 10 min, resuspended in DMEM+10% FBS, and then incubated at 37°C and 5% CO<sub>2</sub>. The fibroblasts used for experiments came from the first or second passages.

## Mice

Eight-week-old male Hos: HR-1 mice (hairless mice) were obtained from Japan SLC, Inc. (Osaka, Japan). Animal care was in strict accordance with the institutional guidelines of Osaka University. All of the animal experiments were carried out at a concentration of 1 mg/ml. Injections of 100 µl of BLM or PBS were administered to subcutaneously to the dorsal area 3 times per week for 3 weeks. One day after final BLM injection, 100  $\mu$ l of 10<sup>-7</sup> M OCT diluted in ethanol, which was further diluted with PBS (1:1000), were subcutaneously injected to BLM-induced sclerotic skin per day for continuous 5 d. As a vehicle control, ethanol diluted in PBS (1:1000) was injected. OCT was kindly provided by Chugai Pharma, Ltd (Tokyo, Japan).

### Histopathological analysis

The dorsal skin was removed one day after the final injection of OCT. The skin pieces were fixed in 10% formaldehyde for 24 h, then embedded in paraffin for microtome sectioning. Sections (4 µm thick) were stained with hematoxylin and eosin (H&E). The amount of collagen was evaluated after tissue staining with the Masson's trichrome technique. For immunohistochemical analysis, sections were hydrated by passage through xylene and graded ethanol. Next, slides were blocked with 2% bovine serum albumin for 10 min, and then stained with primary antibody for 60 min (rabbit anti-POSTN 1:2000 dilution, Abcam, Cambridge, UK). After washing with TBS containing 0.05% Triton-X100 (TBST), slides were developed using the Vectastain ABC kit (Vector Laboratories, Burlingame, CA, USA) followed by counterstaining with hematoxylin. Rabbit IgG was used as the isotype control.

### Western blot analysis

Cells or skin samples crushed in liquid nitrogen were solubilized at 4°C in lysis buffer (0.5% sodium deoxycholate, 1% Nonidet P40, 0.1% sodium dodecyl



Figure 2. OCT causes downregulation of IL-4-, IL-13-, and TGF $\beta$ -induced POSTN and Col1A1 expressions in fibro*blasts.* (**A**) Mouse dermal fibroblasts were pre-treated with  $10^{-7}$  M of OCT for 2 h and treated with IL-4 (10 ng/ ml), IL-13 (10 ng/ml), or TGF $\beta$  (10 ng/ml) for additional 24 h. Bars indicate the mean  $\pm$  SD. N = 4; \*P < 0.01, \*\*P < 0.0001, 2-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons. (B) Mouse dermal fibroblasts were treated as in Figure 2A. The expression of POSTN in cell lysates was measured by western blot analysis.  $\beta$ -actin served as an internal control. (C) Bars show the results of densitometric analysis of POSTN relative to  $\beta$ -actin. Mean  $\pm$  SD of each group are shown. N = 3; \*P < 0.0001, 2-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons. (D) Mouse dermal fibroblasts were treated as in Figure 2A. Expression of Col1A1 and Col1A2 in culture medium was measured by protein gel blot analysis.  $\beta$ -actin served as an internal control. (**E**) Bars show the densitometric analysis of Col1A1 relative to  $\beta$ -actin. The mean  $\pm$  SD is shown for each group. N = 3; \*P < 0.05, \*\*P < 0.001, 2-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons. (F) Normal Human dermal fibroblasts were pre-treated with 10<sup>-7</sup> M of OCT for 2 h and treated with IL-4 (10 ng/ml), IL-13 (10 ng/ml), or TGFB (10 ng/ml) for an additional 24 h. Expression of POSTN was measured in cell lysates by western blot analysis.  $\beta$ -actin served as an internal control. (G) Bars show the densitometric analysis of POSTN relative to  $\beta$ -actin. The mean  $\pm$  SD of each group is shown. N = 3; \*P < 0.0001, 2-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons.



Figure 3. For figure legend, see page 5.

sulfate, 100 µg/ml phenylmethylsulphonyl fluoride, and protease inhibitor cocktail). Five micrograms of protein were separated on SDS-polyacrylamide gels and transferred onto polyvinylidine fluoride membranes (Bio-Rad, Hercules, CA USA). Non-specific antibody binding was blocked by incubating the membranes in 5% w/v non-fat milk powder in TBS-T (50 mM Tris-HCl, pH 7.6, 150 mM NaCl, and 0.1% v/v Tween-20). The membranes were incubated with rabbit anti-POSTN antibody (Abcam, Cambridge, UK) diluted 1:1000 overnight at 4°C or with mouse anti-B-actin antibody (Sigma- Aldrich, St Louis, MO, USA) diluted 1:10,000 overnight at 4°C. Then, the membranes were washed 3 times in TBS-T for 5 min. Finally, the membranes were incubated with either HRP-conjugated anti-mouse or antirabbit at a dilution of 1:10,000 for 60 min at room temperature. Protein bands were detected using the ECL Plus kit (Thermo Scientific, Rockford, IL USA). The intensity of the bands was quantified using NIH image J software.

## RNA isolation and quantitative real-time polymerase chain reaction (qRT-PCR)

Total RNA was isolated from cells using the SV Total RNA Isolation System (Promega). The product was reverse-transcribed into first-strand cDNA (cDNA). qRT-PCR was performed on a ABI 7000 Prism (Applied Biosystems) using the Power SYBR Green PCR Master Mix (Applied Biosystems, Foster City, CA, USA) according to the manufacturer's protocol. The cycling conditions were as follows: 40 cycles of denaturation at 92°C for 15 sec and annealing at 60°C for 60 sec. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as a housekeeping gene control. Sequence-specific primers were designed as follows: mouse Col1A1, sense 5'-gagccctcgcttccgtactc-3', antisense 5'tgttccctactcagccgtctgt-3'; mouse POSTN, sense 5'-aaccaaggacctgaaacacg-3', antisense 5'- caaagagcgtgaagtgacca-3'; mouse IL-6, sense 5'- ctgatgctggtgacaaccac-3', antisense 5'- cagaattgccattgcacaac-3'; human POSTN, sense 5'- gtgatccatggagagccaat-3', antisense 5'- aacttcctcacgggtgtgtc-3'; mouse GAPDH, sense 5'tgtcatcatacttggcaggtttct-3', antisense 5'-catggccttccgtgttccta-3'; human GAPDH, sense 5'- gcaacaatatccactttaccagagttaa-3', antisense 5'-ggagtcaacggatttggtcgta-3'.

### Statistical analysis

The data are expressed as mean values  $\pm$  standard deviations (SD). The Student's *t*-test was used to determine the level of significance of differences between the 2 groups. Analysis of variance for the groups was performed by ANOVA, followed by the Bonferroni-Dunn for multiple comparisons to allow pairwise testing

for significant differences between groups. Statistical significance was defined as P < 0.05.

### Results

## OCT decreased the expressions of POSTN and IL-6 in mouse dermal fibroblasts in a dose-dependent manner

First, we evaluated the expressions of POSTN and IL-6 in mouse dermal fibroblasts treated with OCT. The doses of OCT treatment ranged from  $10^{-9}$  M to  $10^{-6}$  M. We observed a dose-dependent downregulation of POSTN and IL-6 in response to OCT. (Fig. 1A). Additionally, treatment with  $10^{-7}$  M of OCT caused a time dependent decrease in the expression of POSTN from 24 h and IL-6 from 6 h (Fig. 1B). In contrast, treatment with cortisol ranging in doses from  $10^{-9}$  M to  $10^{-6}$  M did not alter the expression of POSTN (Fig. 1C). In contrast, cortisol at dose of  $10^{-6}$  M decreased the expression of IL-6 (Fig. 1C).

## OCT inhibits IL-4-, IL-13-, TGF $\beta$ -induced expression of POSTN and Collagen 1 $\alpha$ 1 in mouse dermal fibroblasts

IL-4, IL-13 and TGF $\beta$  are known to induce the expression of POSTN and reported to associate with fibrosis and remodeling.<sup>18</sup> We examined whether OCT could inhibit IL-4-, IL-13-, TGF $\beta$ -induced POSTN expression. Indeed, the addition of IL-4, IL-13 and TGF $\beta$  increased the expression of POSTN as previously reported (Fig. 2A). When cells were treated with OCT prior to any of these inducing agents, POSTN expression was significantly decreased, as shown by RT-PCR (Fig. 2A) and western blot (Figs. 2B and C). Moreover, Collagen 1  $\alpha$  1 (Col1A1) expression in the culture supernatant was decreased in groups treated with OCT (Figs. 2D and E).

## OCT inhibits IL-4-, IL-13-, TGF $\beta$ -induced POSTN expression in NHDFs

Next, we evaluated the effects of OCT in normal human dermal fibroblasts (NHDF). Similar to mouse dermal fibroblasts, prior treatment with OCT decreased the IL-4-, IL-13-, TGF $\beta$ induced POSTN expression in NHDF (**Figs. 2F and G**). However, in contrast to mouse dermal fibroblasts, OCT treatment did not affect basal POSTN level in NHDFs (**Fig. 2F**).

### OCT inhibits BLM-induced scleroderma

Finally, we evaluated the effects of OCT in BLM-induced skin sclerosis model. Subcutaneous injection of BLM induced dermal thickening and thickening of collagen bundles (Figs. 3A

**Figure 3** (See previous page). *OCT inhibits BLM-induced scleroderma and POSTN induction*. (**A**) Representative H&E staining of back skin treated with PBS or BLM. Bar = 100  $\mu$ m. (**B**) Representative Masson's trichrome staining (low magnification and high magnification) of back skin treated with PBS or BLM. Bar = 100  $\mu$ m. (**C**) Dermal area of skin sections treated with PBS or BLM. The dermal area in each mouse was measured using Image J. Bars show the mean dermal area  $\pm$  SD (N = 6). (**D**) Collagen area of skin sections treated with PBS of BLM. The collagen area in each mouse was measured using Olympus cellSens Dimension software. Bars show the mean dermal area  $\pm$  SD (N = 6) (**E**) Representative POSTN staining of back skin treated with PBS of BLM. Arrows show matricellular staining of POSTN. (**F**) Western blot analysis of POSTN expression in mouse skin treated with either PBS or BLM. (**G**) Bars show the densitometric analysis of POSTN relative to the  $\beta$ -actin loading control. Mean  $\pm$  SD of each group are shown. N = 4; \**P* < 0.0001, one-way ANOVA followed by the Bonferroni-Dunn test for multiple comparisons.

and B). We calculated the dermal area as a means of evaluating dermal thickness. While treatment with OCT did not cause a significant decrease in dermal area, collagen bundles were thicker and more densely packed in vehicle-treated mice compared with OCT-treated mice, as observed by Masson's trichrome staining (Figs. 3B and C). Accordingly, collagen area was significantly lower in OCT-treated group compared with vehicle-treated group (Fig. 3D).

### OCT inhibits BLM-induced POSTN induction

The expression of POSTN was induced in BLM-treated scleroderma skin as previously reported.<sup>15</sup> We found that the expression of POSTN was significantly decreased in mice treated with OCT as shown by immunohistochemistry (**Fig. 3E**) and protein gel blot (**Fig. 3F and G**).

### Discussion

The effect of vitamin D in BLM-induced dermal sclerosis has been reported previously.<sup>22</sup> In that study, metabolites of vitamin D, such as 20S-hydroxyvitamin D, markedly suppressed BLM-induced fibrogenesis in mice.<sup>22</sup> TGF $\beta$ -induced collagen and hyaluronan production was inhibited by 20(OH)D3 and 1,25 (OH)<sub>2</sub>D3 *in vitro*.<sup>22</sup> This study, as well as others, implicates vitamin D as an effective treatment for sclerodermas.

In addition to TGF $\beta$ , the Th2 cytokines play an important role in fibrogenesis in SSc. Th1 and Th17 cells are predominant in the early stages of inflammation, and Th2 cells are predominant in the late fibrosing stage,.<sup>23</sup> IL-4, IL-13, and IL-6, which are secreted by Th2 cells, work as profibrotic mediators.<sup>24</sup> IL-6 produced from dermal fibroblasts also contribute to fibrosis by activating fibroblasts to myofibroblasts, which are known to produce collagen.<sup>25</sup> In this study, we found that OCT significantly suppressed the expression of not only TGF $\beta$ -induced POSTN expression, but also IL-4 and IL-13-induced POSTN expression in dermal fibroblasts. As POSTN is an essential matricellular protein in skin sclerosis of SSc patients and in a BLM-induced scleroderma model,<sup>15</sup> we hypothesize that OCT suppresses dermal sclerosis in part by downregulating the expression of POSTN.

In BLM-induced skin sclerosis model, OCT did not markedly affected the thickness of the dermis, however, thinning of the collagen fiber bundles represented by decreased collagen area was observed. These histological findings are very similar with

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previous findings in scleroderma patients treated with tocilizumab.<sup>26</sup> These patients showed softening of the skin with reductions of Vesmeter hardness, and similar histological changes.<sup>26</sup> Thickness of the collagen fiber bundles but not dermal thickness may be important in skin softening in scleroderma.

Besides inhibiting fibroblast proliferation and collagen synthesis, vitamin D has immunosuppressive effects. Vitamin D decreases expression of toll-like receptors (TLRs) and suppresses TLR-mediated inflammation in monocytes.<sup>27</sup> In peripheral blood mononuclear cells, vitamin D decreases proinflammatory cytokine release.<sup>28</sup> Vitamin D also inhibits TNF $\alpha$ - and /or IFN $\gamma$ -induced IL-8 and IL-6 production in keratinocytes.<sup>29,30</sup> Thus, in addition to its effect of suppressing POSTN, topical OCT might be effective on local scleroderma by inhibiting inflammatory cytokine expression.

In this study, we also evaluated the effect of cortisol, the frequently used topical immunosuppressive ointment to treat local sclerosis, on POSTN expression in dermal fibroblasts (Fig. 1C). In contrast to OCT, cortisol did not alter the expression of POSTN in dermal fibroblasts, suggesting that effect of cortisol on sclerosis might not mediate POSTN.

In conclusion, we found that the vitamin D3 analog, OCT, suppressed expression of POSTN that is normally induced by Th2 cytokines and TGF $\beta$  in dermal fibroblasts. Because POSTN plays a key role in scleroderma, the effectiveness of OCT in scleroderma might be due to suppression of POSTN expression. We hypothesis that this suppressive effect on POSTN may work synergistically with previously reported immunosuppressive and anti-collagen synthesis effects of vitamin D analogs to improve fibrogenesis in scleroderma.

### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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