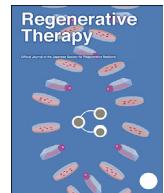




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## Review

### Targeting cancer stem cells in refractory cancer

Norikatsu Miyoshi <sup>a,f,\*</sup>, Naotsugu Haraguchi <sup>b</sup>, Tsunekazu Mizushima <sup>a</sup>, Hideshi Ishii <sup>c</sup>, Hirofumi Yamamoto <sup>a,d</sup>, Masaki Mori <sup>e</sup><sup>a</sup> Department of Gastroenterological Surgery, Graduate School of Medicine, Osaka University, Yamadaoka 2-2-E2, Suita City, Osaka, 565-0871, Japan<sup>b</sup> Department of Gastroenterological Surgery, Osaka International Cancer Institute, 3-1-69, Ohtemae, Chuo-ku, Osaka, 541-8567, Japan<sup>c</sup> Department of Cancer Profiling Discovery/Medical Data Science, Graduate School of Medicine, Osaka University, Suita, Osaka, Japan<sup>d</sup> Department of Molecular Pathology, Division of Health Sciences, Graduate School of Medicine, Osaka University, Yamadaoka 1-7, Suita City, Osaka 565-0871, Japan<sup>e</sup> Department of Surgery and Science, Graduate School of Medical Science, Kyusyu University, 3-1-1, Maidashi, Higashi-ku, Fukuoka City, Fukuoka 812-8582, Japan<sup>f</sup> Department of Innovative Oncology Research and Regenerative Medicine, Osaka International Cancer Institute, 3-1-69, Ohtemae, Chuo-ku, Osaka, 541-8567, Japan

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#### ABSTRACT

Although common cancer therapies, such as chemotherapy and radiation therapy, have recently improved and yielded good results, evaluated as tumor shrinkage, disease recurrence is still a common event for most cancer patients. This is termed refractory cancer. This tumor regrowth following therapy is generally thought to be caused by a small, specific population of tumor cells called cancer stem cells (CSCs). Similar to other stem cells, CSCs have the capacity for self-renewal and multipotent differentiation, and they have been identified in many tumor types based on cell surface protein expression. This specific cell population has stemness characteristics as examined by serial transplantation in animal models. Previous studies have developed a specific signature of cell surface markers and biological functions that can identify CSCs in many solid tumors. In this review, we summarize the characterization of CSCs using new techniques for identifying and quantifying them *in situ*. These techniques and concepts could be valuable for evaluating the effects of therapies on this cell population. Finally, we conclude by discussing several unique preclinical treatment strategies to targets CSCs, such as reprogramming CSCs or inducing attack by immune cells. Therapeutic and diagnostic methodologies that can target and quantify CSCs will be valuable tools for eradicating refractory cancer.

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**Abbreviations:** CSC, cancer stem cells; ODC, ornithine decarboxylase; LGR5, Leucine-rich repeat-containing G-protein coupled receptor; KLF5, Kruppel-like factor 5; iPS cell: induced pluripotent stem cell, iPSC cell: induced pluripotent cancer cell; miRNA, microRNA; PGE2, prostaglandin E2.

\* Corresponding author. Fax: +81-6-6879-3259.

E-mail addresses: [nmiyoshi@gesurg.med.osaka-u.ac.jp](mailto:nmiyoshi@gesurg.med.osaka-u.ac.jp) (N. Miyoshi), [haraguchi-na@mc.pref.osaka.jp](mailto:haraguchi-na@mc.pref.osaka.jp) (N. Haraguchi), [tmizushima@gesurg.med.osaka-u.ac.jp](mailto:tmizushima@gesurg.med.osaka-u.ac.jp) (T. Mizushima), [hishii@gesurg.med.osaka-u.ac.jp](mailto:hishii@gesurg.med.osaka-u.ac.jp) (H. Ishii), [hyamamoto@sahs.med.osaka-u.ac.jp](mailto:hyamamoto@sahs.med.osaka-u.ac.jp) (H. Yamamoto), [m\\_mori@surg2.med.kyushu-u.ac.jp](mailto:m_mori@surg2.med.kyushu-u.ac.jp) (M. Mori).

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## 1. Introduction

Cancer is a disease caused by genetic and epigenetic factors that lead to uncontrolled proliferation. It is one of the leading causes of death worldwide [1,2]. Clinically, many cancers show good response to conventional treatments such as chemotherapy and radiotherapy. However, many tumors later recur, even if they initially showed a good response to treatment. This is called refractory cancer. One of the causes of refractory cancer is cellular heterogeneity, which is produced by small populations of cancer stem cells (CSCs). CSCs were first noted in acute myeloid leukemia [3,4] and have since been detected by various *in vitro* assays and *in vivo* animal experiments [5]. As with other types of stem cells, CSCs have both self-renewal and multipotent properties [6,7]. The possible involvement of CSCs in refractory cancer has since been shown in several solid tumors [8–13]. Recent studies have shown that many cancers possess a differentiation hierarchy that arises from malignant CSCs that undergo uncontrolled proliferation and produce daughter cells [6,7] (Fig. 1). It is thought that CSCs are the cause of resistance to conventional treatments [7,14–16] (Fig. 2). Although CSCs account for only a small proportion of tumor cells,

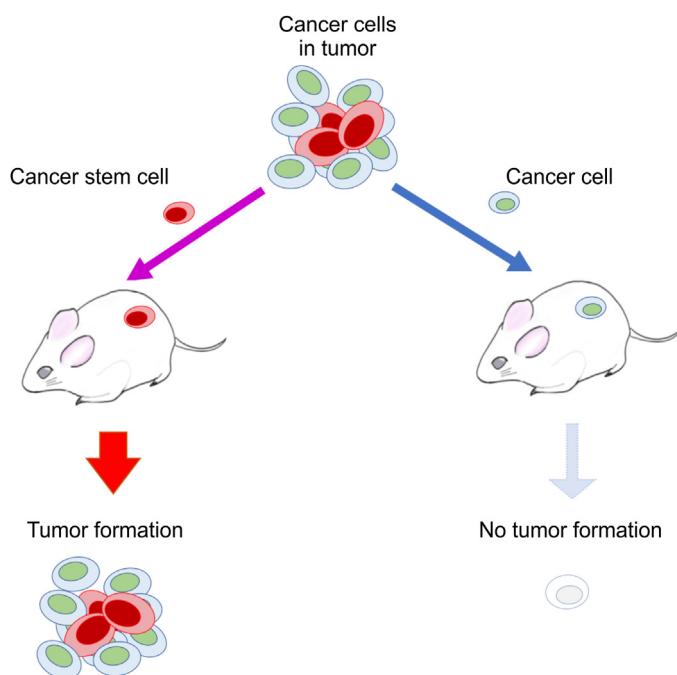
they possess persistent proliferative potential. Because of this uncontrolled proliferative potential and the dysregulation of their mechanisms of differentiation, malignant tumors have been proposed to derive from CSCs [3,5].

Here, we review the recent progress made in CSC research, discuss how CSCs maintain their malignant potential, and describe new strategies for targeting CSCs.

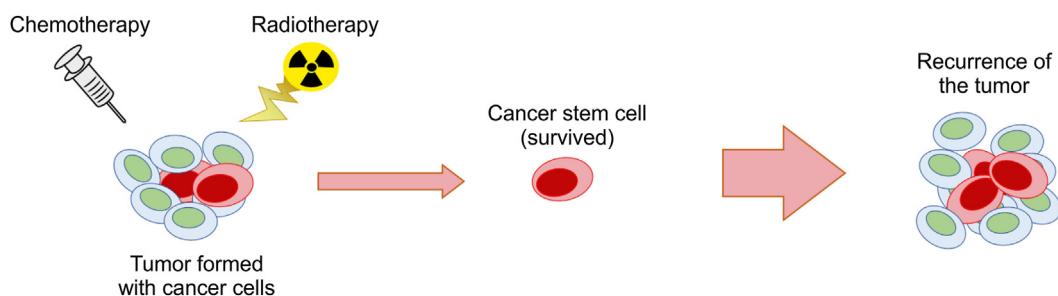
## 2. Detecting CSCs

Previous studies that have isolated CSCs provide great insights into how they promote resistance to treatment [17–20]. CSCs have multiple properties that protect against cytotoxic drugs and DNA damage. Several reports have shown that cancers with higher expression of stem cell markers are associated with an increased risk of recurrence and poorer prognosis [21–23]. CSCs identified from solid tumors usually express the specific markers of the site of origin. In breast cancer, CSCs tend to be CD44<sup>+</sup>CD24<sup>-/low</sup>Lin<sup>-</sup> [10], and as few as 100 cells with this phenotype are able to form tumors in transplanted mice, whereas tens of thousands of cells with alternate phenotypes fail to form tumors. Furthermore, in serial passaging experiments in which the extracted cells from one mouse are transplanted into another, this tumorigenic subpopulation could generate new tumors containing CD44<sup>+</sup>CD24<sup>-/low</sup>Lin<sup>-</sup> cells within each passage. These results indicate that these tumorigenic cells behave like CSCs. CD44 is an adhesion molecule, and the variant isoforms containing v8-v10 (CD44v9) have been identified as CSC surface markers [24]. Similar results have also been reported in brain tumors, and brain tumor stem cells were exclusively isolated using the neural stem cell surface marker CD133 [9,25,26]. In gastrointestinal cancers, several CSC surface markers have been reported [27–31]. Although CSCs are a small proportion of cancer cells, they have low cell turnover, which allows for isolation of CSCs for gene expression profile analysis. Additionally, this population has the ability to extrude dye, which allows for the isolation of CSCs [32,33].

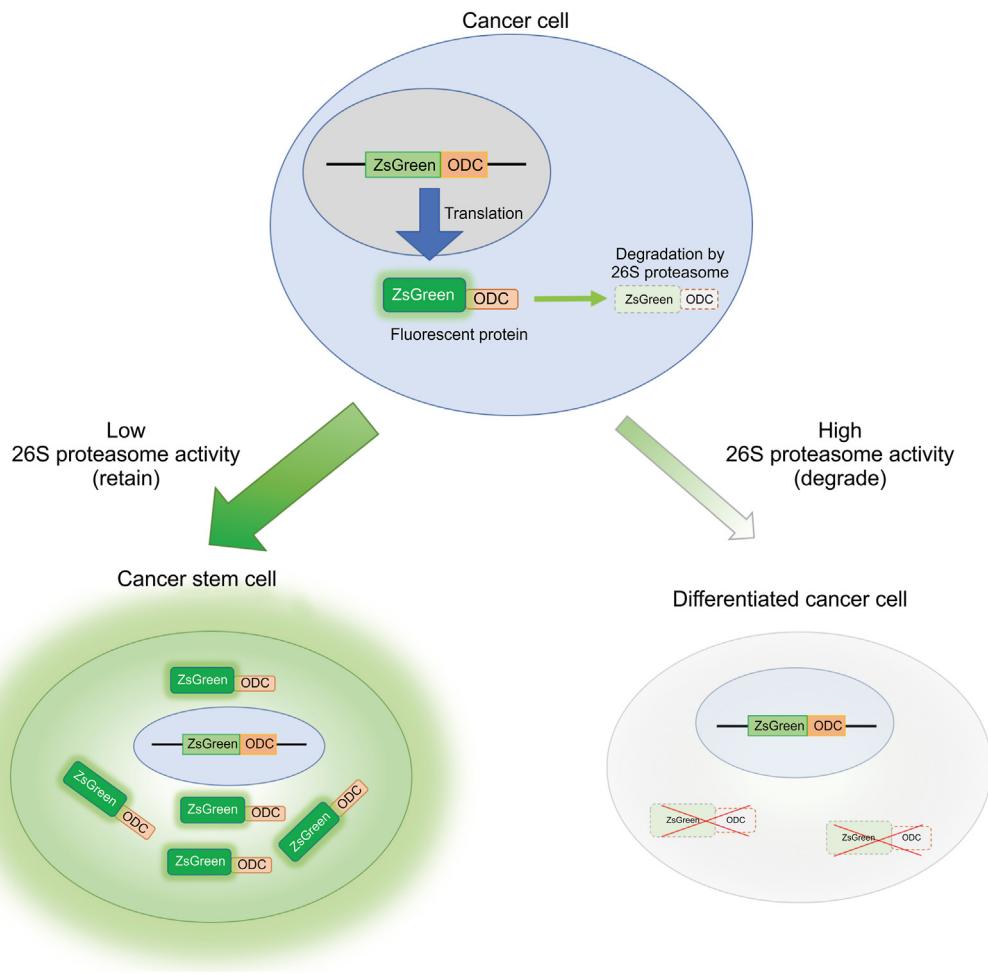
Isolation of CSCs is performed using flow cytometry focusing on cell surface markers, which allows for the study of tumor profiles. Although this procedure can detect the CSC population and target specific clusters, the manipulation stresses them and may alter their biology. Therefore, visualizing CSCs *in situ* would be preferable, and it is necessary to develop a system to analyze CSC behaviors within their natural environment. Focusing on the characteristics of CSCs, such as their dormancy, low protein turnover rate, and decreased 26S proteasome activity, has great advantages for cancer research [34]. A fusion of green fluorescent protein (ZsGreen) and the C-terminal degron of ornithine decarboxylase (ODC) is retained (green fluorescence-positive) in CSCs with low 26S proteasome activity due to decreased protein degradation. In several solid tumors, these fluorescent (ZsGreen-ODC-positive) cells demonstrate features of stemness, such as



**Fig. 1.** Cancer stem cells evaluated with xenograft model. Cancers possess a differentiation hierarchy producing heterogeneity. Cancer stem cells are able to form tumors when transplanted into immunodeficient mice.



**Fig. 2.** Conventional cancer treatments and the stem cells.



**Fig. 3.** Visualization of cancer stem cells. The technique is based on the decreased activity of the 26S proteasome in cancer stem cells. Ornithine decarboxylase (ODC) is normally destroyed by the proteasome. Using a vector encoding a fusion of green fluorescence protein (ZsGreen) and the C-terminal degron of ODC, cancer stem cells with low 26S proteasome activity can be visualized.

tumor formation in xenotransplant models and asymmetric cell division (Fig. 3) [18–20]. The fluorescent cells are also more chemo- and radioresistant than non-fluorescent cells [19,20]. The ZsGreen-ODC system has been used in various solid tumors [35,36]. Visualizing CSCs using this system not only allows for stem cell research but also for drug screening to search for novel agents.

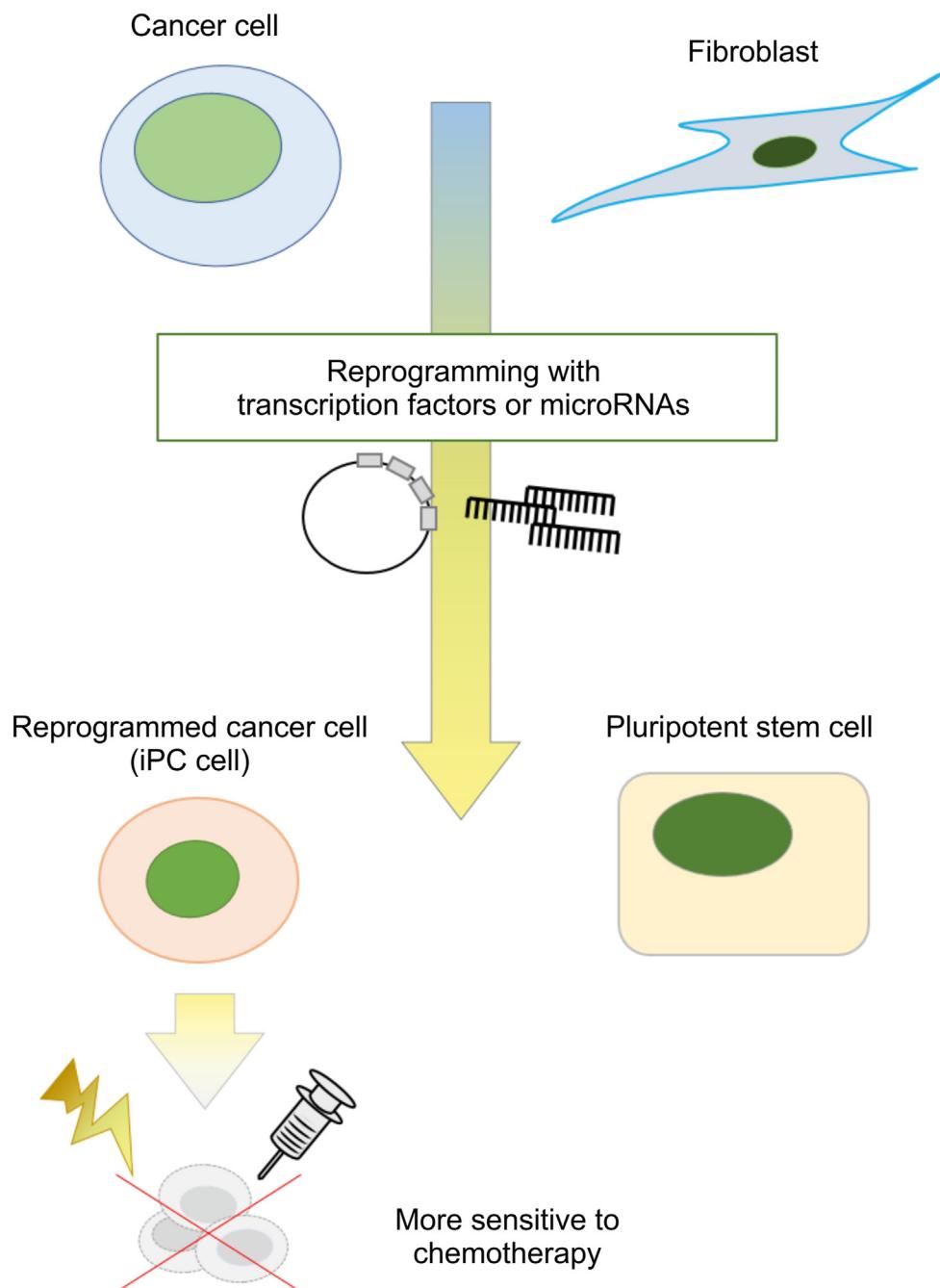
A recent report proposed marking revived stem cells based on high clusterin expression, which is found in cells with damage-induced quiescence [37]. The revived stem cells undergo yes-associated protein 1-dependent transient expansion and reconstitute the leucine-rich repeat-containing G-protein coupled receptor (LGR5)-positive crypt-base columnar cells, regenerating a functional intestine. Another report shows the plasticity of LGR5-negative cancer cells, which drive metastasis in colorectal cancer [38]. It is necessary to think about CSCs as unique entities, not relying solely on the cell surface markers of the original stem cells, because of their plasticity, particularly when focusing on the treatment course of the refractory cancer.

Morimoto and colleagues reported a novel target in CSCs, Kruppel-like factor 5 (KLF5) [39]. In that study, therapeutic microRNA (miRNA) was used to suppress the cancer stemness, specifically focusing on KLF5 expression [39]. KLF5 acts as a core regulator in the process of intestinal oncogenesis, which has been demonstrated in a genetically manipulated mouse model and

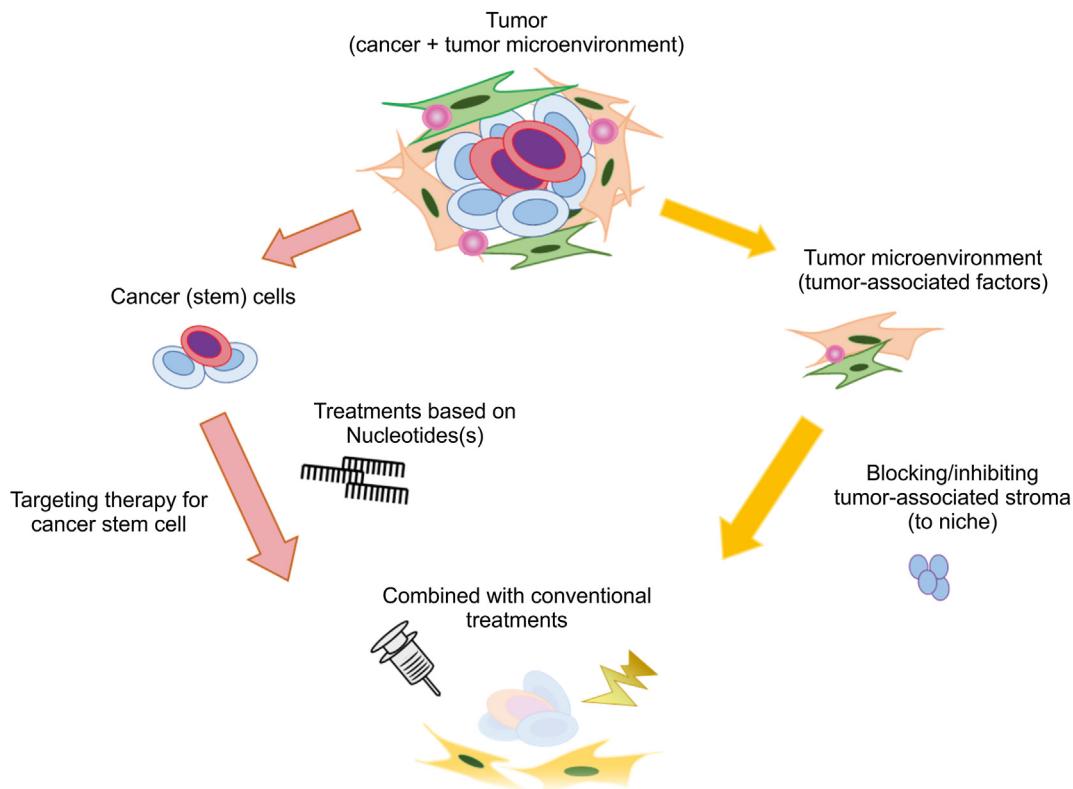
LGR5-positive intestinal stem cells [40]. KLF5 controls the stemness of embryonic stem cells [41].

### 3. Treatments based on cancer cell reprogramming

The investigation of embryonic stem cell development, from zygote to blastodermic vesicle, has helped elucidate the molecular mechanisms specifying pluripotent differentiation [42,43]. In multipotent stem cells, several transcription factors cooperate to regulate pluripotency through strict epigenetic regulation [44–47]. In our previous study, we found that the transcription factors necessary to create induced pluripotent stem (iPS) cells have similar effects as cancer-related oncogenes and tumor suppressor genes. The transcription factors OCT3/4, SOX2, KLF4, and c-MYC were introduced into cancer cell lines. The cells generated in our study were similar to iPS cells in morphology and displayed embryonic stem cell-like gene expression and epigenetic modifications [44–47]. The introduction of these transcription factors into gastrointestinal cancer cells resulted in the reprogramming of the cells to the pluripotent state and also sensitized them to differentiation [48]. The reprogrammed cancer cells could be differentiated into various cells, such as epithelial, mesenchymal, neural, or adipose lineages, under varying conditions [48]. As tumor suppressor genes extend the lifespan of embryonic stem cells, their repression



**Fig. 4.** Strategy of cancer treatment based on reprogramming. Cancer cells can be reprogrammed into induced pluripotent cancer (iPC) cells by specific transcription factors or microRNAs, similar to the manner in which normal fibroblasts can be reprogrammed into induced pluripotent stem cells. The iPC cells have an increased sensitivity to conventional chemotherapy.



**Fig. 5.** Treatment strategy for cancer focusing on the stem cells and the tumor microenvironment. The heterogenous tumor and its microenvironment protect the tumor from anticancer therapies. Stem cell-specific markers or a visualizing system can be used to detect the cancer stem cells. Then, targeted therapy and reprogramming-based treatments can improve the efficacy of conventional treatments. Furthermore, targeting the tumor microenvironment can enhance the treatment strategy, leading to the reduction of recurrence and eradication of the cancer.

increases the induction efficiency of iPS cells and maintains their immortalized state [49–51].

Although these reprogrammed cells have not been fully characterized, because they are distinct from their parental cells, it is possible that the generation of induced pluripotent cancer (iPC) cells will allow for so-called differentiation therapy *via* the induction of drug susceptibility in cancer cells. This reprogramming approach in cancer cells supports that transduction will cause cell differentiation into unique lineages. The initial goal is to exploit drug discoveries with the aim of producing therapeutic and diagnostic reagents. We also demonstrated that iPC cells have the capacity for multipotent differentiation. Originally, we hypothesized that cells differentiated from iPC cells would revert to their original phenotypes. However, we found that the differentiated iPC cells lost the ability to form tumors after transplantation into mice. Furthermore, the differentiated iPC cells became more sensitive to chemotherapy. These findings suggest that reprogramming and epigenetic modifications are promising strategies to understanding CSCs and identifying novel cancer treatments, regardless of any genetic mutations in the parental cells.

A recent report showed that G9a activity is essential for the maintenance of the embryonic-like transcriptional signature that promotes self-renewal, tumorigenicity, and the undifferentiated state. Therefore, this might be useful when transforming pluripotent cells as a surrogate model for CSCs [52].

With a view toward using reprogramming to eradicate CSCs clinically, we developed a method of reprogramming differentiated fibroblasts into pluripotent stem cells *via* a specific combination of

miRNAs [53]. miRNAs are small noncoding RNAs that regulate gene expression to regulate development and differentiation. Specific miRNAs have been characterized in relation to pluripotency [54–56]; therefore, we searched for miRNAs that could reprogram differentiated cells into pluripotent stem cells. Candidate miRNAs that were highly expressed in iPS and embryonic stem cells were transfected into somatic cells derived from transgenic mice with green fluorescent protein driven by the *Nanog* promoter [57]. By examining the fluorescence (a surrogate for *Nanog* activation), we identified a combination of miRNAs that could reprogram mammalian cells into pluripotent stem cells and demonstrated the ability of the miRNA-induced reprogrammed cells to differentiate into cells of different lineages. As with iPS cells created using the four transcription factors, cancer cell lines reprogrammed with miRNAs also demonstrated decreased tumor-initiating capacity and became sensitive to chemotherapy [58]. In an *in vivo* model examining the feasibility, safety, and effectiveness of the miRNAs, the miRNA combination suppressed tumorigenesis, suggesting that this therapy could be useful for both preventing and treating cancer (Fig. 4). In glioblastoma, reprogramming transcription factors and miRNAs drive a stem-like phenotype [59].

However, one limitation of miRNA-based treatment is the presence of RNases in the blood, which makes miRNA-based treatments unsuitable for systemic therapy. To prevent RNA degradation, a drug delivery system was designed using super carbonate apatite nanoparticles. Because of their preference for the low pH conditions found in tumor microenvironments, these nanoparticles deliver RNAs selectively to tumors [60]. We are

currently exploring the use of synthetic miRNAs to increase the stability and efficacy of this miRNA-based therapy.

#### 4. Targeting the tumor microenvironment

Although the refractory nature of the tumor itself is born of tumor heterogeneity, the tumor microenvironment also plays an important role by providing various self-protection properties. These include mechanisms that enable dynamic interactions with surrounding epithelial cells, infiltrating immune cells, cytokines, and chemokines, which maintain refractory cancers regulated by CSC proliferation and self-renewal. The tumor microenvironment maintains CSCs by the induction of specific features [61]. The microenvironment plays a key role in regulating the CSC population by direct cell-cell contacts in which various paracrine factors are secreted. The stemness of CSCs is maintained by microenvironmental factors through pathways that promote self-renewal, such as the Wnt/β-catenin, Notch, and Hedgehog pathways. For example, Wnt activity regulates the self-renewal of CSCs and drives transit amplifying cell proliferation and differentiation [62]. Hedgehog signaling is important for embryonic development, patterning, and differentiation, which are required for the regulation of the self-renewal of normal mammary stem cells and CSCs [63]. Additionally, during development, Notch signaling controls cell fate, and aberrant activation contributes to tumorigenesis [64]. Niclosamide, a tineacide of the anthelmintic family, is an inhibitor of Wnt/β-catenin and Notch signaling [65], and it might be useful for inhibiting CSCs by blocking these signaling pathways. Mesenchymal stem cells in the tumor-associated stroma have been shown to affect cancer cell behavior. Mesenchymal stem cells influence the phenotypes of cancer cells, and prostaglandin E2 (PGE2) secreted by mesenchymal stem cells enables tumor progression via creating a CSC niche [66]. Targeting the tumor microenvironment may stimulate host antitumor responses by blocking tumor-promoting inflammation with PGE2 receptor antagonists, which is a promising treatment strategy. However, to effectively target CSCs, combination therapy may be needed.

The two major superfamilies of efflux transporters in CSCs are the ATP-binding cassette transporters and the solute carrier transporters. Interestingly, because of the ability of CSCs to extrude dye, targeting these efflux transporters as well as CSC-related surface markers in combination with conventional treatments can improve cancer treatment.

#### 5. Conclusions

Refractory cancer, which is associated with a long cellular life span, relative quiescence, and resistance to drugs, is derived from cellular heterogeneity produced by small populations of CSCs. Through their self-renewal and drug-resistant capacities, CSCs allow for proliferation and differentiation, even after treatment with anticancer therapy. No specific marker to identify all CSCs has been defined. Further, cancer cells gain plasticity during metastasis in colorectal cancer [38]. It is important to further understand the characteristics of CSCs and normal stem cells. If specific markers or tracing methods for these stem cells can be identified, we will be able to isolate, identify, and analyze these populations within heterogeneous tumors. Examining these characteristics will allow gene expression profiling, which provides crucial information that will allow prediction of patient prognosis [67]. Considering the tumor microenvironment and molecular subtypes, the elucidation of CSC properties could lead to the development of effective cancer treatments (Fig. 5).

Cancer stem cells are the cause of chemo- and radioresistance, which results in tumor recurrence. Cancer stem cells have the

capacity for self-renewal and multipotent differentiation, which is thought to lead to resistance to conventional treatments.

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#### References

- [1] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics, 2018. *CA A Cancer J Clin* 2018;68:394–424.
- [2] Colvin H, Mizushima T, Eguchi H, Takiguchi S, Doki Y, Mori M. Gastroenterological surgery in Japan: the past, the present and the future. *Ann Gastroenterol Surg* 2017;1:5–10.
- [3] Hamburger AW, Salmon SE. Primary bioassay of human tumor stem cells. *Science* 1977;197:461–3.
- [4] Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature* 1994;367:645–8.
- [5] Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 2001;414:105–11.
- [6] Pardal R, Clarke MF, Morrison SJ. Applying the principles of stem-cell biology to cancer. *Nat Rev Canc* 2003;3:895–902.
- [7] Miyoshi N, Mizushima T, Doki Y, Mori M. Cancer stem cells in relation to treatment. *Jpn J Clin Oncol* 2019;49:232–7.
- [8] Kondo T, Setoguchi T, Taga T. Persistence of a small subpopulation of cancer stem-like cells in the C6 glioma cell line. *Proc Natl Acad Sci U S A* 2004;101:781–6.
- [9] Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, et al. Identification of human brain tumour initiating cells. *Nature* 2004;432:396–401.
- [10] Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A* 2003;100:3983–8.
- [11] Kim CF, Jackson EL, Woolfenden AE, Lawrence S, Babar I, Vogel S, et al. Identification of bronchioalveolar stem cells in normal lung and lung cancer. *Cell* 2005;121:823–35.
- [12] Wang S, Garcia AJ, Wu M, Lawson DA, Witte ON, Wu H. Pten deletion leads to the expansion of a prostatic stem/progenitor cell subpopulation and tumor initiation. *Proc Natl Acad Sci U S A* 2006;103:1480–5.
- [13] Tsunedomi R, Yoshimura K, Suzuki N, Hazama S, Nagano H. Clinical implications of cancer stem cells in digestive cancers: acquisition of stemness and prognostic impact. *Surg Today* 2020;50:1560–77.
- [14] Clevers H. The cancer stem cell: premises, promises and challenges. *Nat Med* 2011;17:313–9.
- [15] Wagl N, Emery C, Berger MF, Davis MJ, Sawyer A, Pochanard P, et al. Dissecting therapeutic resistance to RAF inhibition in melanoma by tumor genomic profiling. *J Clin Oncol* 2011;29:3085–96.
- [16] Waclaw B, Bozic I, Pittman ME, Hruban RH, Vogelstein B, Nowak MA. A spatial model predicts that dispersal and cell turnover limit intratumour heterogeneity. *Nature* 2015;525:261–4. –11.
- [17] Fukusumi T, Ishii H, Konno M, Yasui T, Nakahara S, Takenaka Y, et al. CD10 as a novel marker of therapeutic resistance and cancer stem cells in head and neck squamous cell carcinoma. *Br J Canc* 2014;111:506–14.
- [18] Munakata K, Uemura M, Tanaka S, Kawai K, Kitahara T, Miyo M, et al. Cancer stem-like properties in colorectal cancer cells with low proteasome activity. *Clin Canc Res* 2016;22:5277–86.
- [19] Hayashi K, Tamari K, Ishii H, Konno M, Nishida N, Kawamoto K, et al. Visualization and characterization of cancer stem-like cells in cervical cancer. *Int J Oncol* 2014;45:2468–74.
- [20] Tamari K, Hayashi K, Ishii H, Kano Y, Konno M, Kawamoto K, et al. Identification of chemoradiation-resistant osteosarcoma stem cells using an imaging system for proteasome activity. *Int J Oncol* 2014;45:2349–54.
- [21] Merlos-Suárez A, Barriga FM, Jung P, Iglesias M, Céspedes MV, Rossell D, et al. The intestinal stem cell signature identifies colorectal cancer stem cells and predicts disease relapse. *Cell Stem Cell* 2011;8:511–24.
- [22] de Sousa E Melo F, Colak S, Buikhuisen J, Koster J, Cameron K, de Jong JH, et al. Methylation of cancer-stem-cell-associated Wnt target genes predicts poor prognosis in colorectal cancer patients. *Cell Stem Cell* 2011;9:476–85.
- [23] Takahashi H, Ishii H, Nishida N, Takemasa I, Mizushima T, Ikeda M, et al. Significance of Lgr5(+ve) cancer stem cells in the colon and rectum. *Ann Surg Oncol* 2011;18:1166–74.
- [24] Ishimoto T, Nagano O, Yae T, Tamada M, Motohara T, Oshima H, et al. CD44 variant regulates redox status in cancer cells by stabilizing the xCT subunit of system xc(-) and thereby promotes tumor growth. *Canc Cell* 2011;19:387–400.
- [25] Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, et al. Identification of a cancer stem cell in human brain tumors. *Canc Res* 2003;63:5821–8.

- [26] Beier D, Hau P, Proescholdt M, Lohmeier A, Wischhusen J, Oefner PJ, et al. CD133(+) and CD133(-) glioblastoma-derived cancer stem cells show differential growth characteristics and molecular profiles. *Canc Res* 2007;67:4010–5.
- [27] Yang ZF, Ho DW, Ng MN, Lau CK, Yu WC, Ngai P, et al. Significance of CD90+ cancer stem cells in human liver cancer. *Canc Cell* 2008;13:153–66.
- [28] Li C, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V, et al. Identification of pancreatic cancer stem cells. *Canc Res* 2007;67:1030–7.
- [29] Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, et al. Identification and expansion of human colon-cancer-initiating cells. *Nature* 2007;445:111–5.
- [30] O'Brien CA, Pollett A, Gallinger S, Dick JE. A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. *Nature* 2007;445:106–10.
- [31] Du L, Wang H, He L, Zhang J, Ni B, Wang X, et al. CD44 is of functional importance for colorectal cancer stem cells. *Clin Canc Res* 2008;14:6751–60.
- [32] Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC. Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. *J Exp Med* 1996;183:1797–806.
- [33] Goodell MA, Rosenzweig M, Kim H, Marks DF, DeMaria M, Paradis G, et al. Dye efflux studies suggest that hematopoietic stem cells expressing low or undetectable levels of CD34 antigen exist in multiple species. *Nat Med* 1997;3:1337–45.
- [34] Pan J, Zhang Q, Wang Y, You M. 26S proteasome activity is down-regulated in lung cancer stem-like cells propagated in vitro. *Plos One* 2010;5:e13298.
- [35] Adikrisna R, Tanaka S, Muramatsu S, Aihara A, Ban D, Ochiai T, et al. Identification of pancreatic cancer stem cells and selective toxicity of chemotherapeutic agents. *Gastroenterology* 2012;143:234–245.e7.
- [36] Vlashi E, Kim K, Lagadec C, Donna LD, McDonald JT, Eghbali M, et al. In vivo imaging, tracking, and targeting of cancer stem cells. *J Natl Cancer Inst* 2009;101:350–9.
- [37] Ayyaz A, Kumar S, Sangiorgi B, Ghoshal B, Gosio J, Ouladan S, et al. Single-cell transcriptomes of the regenerating intestine reveal a revival stem cell. *Nature* 2019;569:121–5.
- [38] Fumagalli A, Oost KC, Kester L, Morgner J, Bornes L, Bruens L, et al. Plasticity of Lgr5- negative cancer cells drives metastasis in colorectal cancer. *Cell Stem Cell* 2020;26:569–78.e7.
- [39] Morimoto Y, Mizushima T, Wu X, Okuzaki D, Yokoyama Y, Inoue A, et al. miR-4711-5p regulates cancer stemness and cell cycle progression via KLF5, MDM2 and TFDP1 in colon cancer cells. *Br J Canc* 2020;122:1037–49.
- [40] McConnell BB, Ghaleb AM, Nandan MO, Yang VW. The diverse functions of Krüppel-like factors 4 and 5 in epithelial biology and pathobiology. *Bioessays* 2007;29:549–57.
- [41] Ema M, Mori D, Niwa H, Hasegawa Y, Yamanaka Y, Hitoshi S, et al. Krüppel-like factor 5 is essential for blastocyst development and the normal self-renewal of mouse ESCs. *Cell Stem Cell* 2008;3:555–67.
- [42] Thomson JA, Itskovitz-Eldor J, Shapiro SS, Waknitz MA, Swiergiel JJ, Marshall VS, et al. Embryonic stem cell lines derived from human blastocysts. *Science* 1998;282:1145–7.
- [43] Hochedlinger K, Jaenisch R. Nuclear reprogramming and pluripotency. *Nature* 2006;441:1061–7.
- [44] Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 2006;126:663–76.
- [45] Takahashi K, Tanabe K, Ohnuki M, Narita M, Ichisaka T, Tomoda K, et al. Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 2007;131:861–72.
- [46] Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, et al. Induced pluripotent stem cell lines derived from human somatic cells. *Science* 2007;318:1917–20.
- [47] Yu J, Hu K, Smuga-Otto K, Tian S, Stewart R, Slukvin II, et al. Human induced pluripotent stem cells free of vector and transgene sequences. *Science* 2009;324:797–801.
- [48] Miyoshi N, Ishii H, Nagai K, Hoshino H, Mimori K, Tanaka F, et al. Defined factors induce reprogramming of gastrointestinal cancer cells. *Proc Natl Acad Sci U S A* 2010;107:40–5.
- [49] Dabelsteen S, Hercule P, Barron P, Rice M, Dorsainville G, Rheinwald JG. Epithelial cells derived from human embryonic stem cells display p16INK4A senescence, hypermotility, and differentiation properties shared by many P63+ somatic cell types. *Stem Cell* 2009;27:1388–99.
- [50] Hong H, Takahashi K, Ichisaka T, Aoi T, Kanagawa O, Nakagawa M, et al. Suppression of induced pluripotent stem cell generation by the p53-p21 pathway. *Nature* 2009;460:1132–5.
- [51] Li H, Collado M, Villasante A, Strati K, Ortega S, Cañamero M, et al. The Ink 4/Arf locus is a barrier for iPS cell reprogramming. *Nature* 2009;460:1136–9.
- [52] Bergin CJ, Zouggar A, Haebe JR, Masibag AN, Desrochers FM, Reiley SY, et al. G9a controls pluripotent-like identity and tumor-initiating function in human colorectal cancer. *Oncogene* 2020. <https://doi.org/10.1038/s41388-020-01591-7>.
- [53] Miyoshi N, Ishii H, Nagano H, Haraguchi N, Dewi DL, Kano Y, et al. Reprogramming of mouse and human cells to pluripotency using mature microRNAs. *Cell Stem Cell* 2011;8:633–8.
- [54] Houbaviy HB, Murray MF, Sharp PA. Embryonic stem cell-specific microRNAs. *Dev Cell* 2003;5:351–8.
- [55] Judson RL, Babiarz JE, Venere M, Blelloch R. Embryonic stem cell-specific microRNAs promote induced pluripotency. *Nat Biotechnol* 2009;27:459–61.
- [56] Suh MR, Lee Y, Kim JY, Kim SK, Moon SH, Lee JY, et al. Human embryonic stem cells express a unique set of microRNAs. *Dev Biol* 2004;270:488–98.
- [57] Okita K, Ichisaka T, Yamanaka S. Generation of germline-competent induced pluripotent stem cells. *Nature* 2007;448:313–7.
- [58] Ogawa H, Wu X, Kawamoto K, Nishida N, Konno M, Koseki J, et al. MicroRNAs induce epigenetic reprogramming and suppress malignant phenotypes of human colon cancer cells. *PloS One* 2015;10:e0127119.
- [59] Lopez-Bertoni H, Kotchetkov IS, Mihelson N, Lal B, Rui Y, Ames H, et al. A Sox 2:miR-486-5p axis regulates survival of GBM cells by inhibiting tumor suppressor networks. *Canc Res* 2020;80:1644–55.
- [60] Wu X, Yamamoto H, Nakanishi H, Yamamoto Y, Inoue A, Tei M, et al. Innovative delivery of siRNA to solid tumors by super carbonate apatite. *PloS One* 2015;10:e0116022.
- [61] Borovski T, De Sousa E Melo F, Vermeulen L, Medema JP. Cancer stem cell niche: the place to be. *Canc Res* 2011;71:634–9.
- [62] Bisson I, Prowse DM. WNT signaling regulates self-renewal and differentiation of prostate cancer cells with stem cell characteristics. *Cell Res* 2009;19:683–97.
- [63] Liu S, Dontu G, Mantle ID, Patel S, Ahn NS, Jackson KW, et al. Hedgehog signaling and Bmi-1 regulate self-renewal of normal and malignant human mammary stem cells. *Canc Res* 2006;66:6063–71.
- [64] Miele L. Notch signaling. *Clin Canc Res* 2006;12:1074–9.
- [65] Li Y, Li PK, Roberts MJ, Arend RC, Samant RS, Buchsbaum DJ. Multi-targeted therapy of cancer by niclosamide: a new application for an old drug. *Canc Lett* 2014;349:8–14.
- [66] Li HJ, Reinhardt F, Herschman HR, Weinberg RA. Cancer-stimulated mesenchymal stem cells create a carcinoma stem cell niche via prostaglandin E2 signaling. *Canc Discov* 2012;2:840–55.
- [67] Guinney J, Dienstmann R, Wang X, de Reynies A, Schlicker A, Soneson C, et al. The consensus molecular subtypes of colorectal cancer. *Nat Med* 2015;21:1350–6.