

Citation: Sakai Y, Takada T (2016) Pathogen Propagation Model with Superinfection in Vegetatively Propagated Plants on Lattice Space. PLoS ONE 11(5): e0154883. doi:10.1371/journal. pone.0154883

Editor: Vijai Gupta, National University of Ireland - Galway, IRELAND

Received: January 19, 2016

Accepted: April 20, 2016

Published: May 5, 2016

Copyright: © 2016 Sakai, Takada. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This research was funded in part by Grants-in-Aid from the Japanese Society for the Promotion of Science (JSPS) for Scientific Reserch: No. 15H04418 & 25340115 & 26291087. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

RESEARCH ARTICLE

Pathogen Propagation Model with Superinfection in Vegetatively Propagated Plants on Lattice Space

Yuma Sakai¹*, Takenori Takada²

1 Graduate School of Environmental Science, Hokkaido University, Sapporo, Hokkaido, Japan, 2 Graduate School of Environmental Earth Science, Hokkaido University, Sapporo, Hokkaido, Japan

* <u>sakai@ees.hokudai.ac.jp</u>

Abstract

Many clonal plants have two reproductive patterns, seed propagation and vegetative propagation. By vegetative propagation, plants reproduce the genetically identical offspring with a low mortality, because resources are supplied from the other individuals through interconnected ramets at vegetative-propagated offspring. However, the ramets transport not only resources but also systemic pathogen. Pathogens evolve to establish and spread widely within the plant population. The superinfection, which is defined as the ability that an established pathogen spreads widely by infecting to already-infected individuals with other strains of a pathogen, is important to the evolution of pathogens. We examine the dynamics of plant reproduction and pathogen propagation considering spatial structure and the effect of superinfection on genetic diversity of pathogen by analysis of several models, 1-strain and multiple-strain models, on two-dimensional square lattice. In the analysis of 1-strain model, we derive equilibrium value by mean-field approximation and pair approximation, and its local stability by Routh-Hurwitz stability criterion. In the multiple-strain models, we analyze the dynamics by numerical simulation of mean-field approximation, pair approximation and Monte Carlo simulation. Through the analyses, we show the effect of parameter values to dynamics of models, such as transition of dominant strain of pathogen, competition between plants and pathogens and density of individuals. As a result, (i) The strain with intermediate cost becomes dominant when both superinfection rate and growth rate are low. (ii) The competition between plants and pathogens occurs in the phase of coexistence of various strains by pair approximation and Monte Carlo simulation. (iii) Too high growth rate leads to the decrease of plant population in all models. (iv) Pathogens are easy to maintain their genetic diversity with low superinfection rate. However, if they do not superinfect, the maintenance becomes difficult. (v) When growth rate of plant is low, individuals are very influenced by distant individuals.

Introduction

Many clonal plants have two reproductive patterns, seed propagation and vegetative propagation (i.e. clonal growth). By seed propagation, plants reproduce genetically different offspring with a high mortality, because seed-propagated offspring are not supported by their parents (no physical connection). By contrast, in vegetative propagation, plants reproduce genetically identical offspring with a lower mortality, because resources are supplied from the other individuals through interconnected ramets at vegetative-propagated offspring [1, 2]. However, if some of systemic pathogens invade the population, the pathogens spread rapidly within the population, because ramets transport not only resources, but also pathogens [3].

According to Stuefer et al. (2004), pathogens have diverse negative effects on plants, such as lethality or severe damage. For example, they lead to deformation of leaves [4], reduction the growth rate [5-7], change of the growth form of plants [8, 9] and effectively block plant reproduction [10-12]. Thus, the possibility that the plant population suffers serious damage through effect of a pathogen is high, and plants have evolved to prevent the spread of infection, such as escaping by increasing their growth rate [8].

Contrastingly, pathogens have evolved to establish and spread widely within the plant population. A diversity of infections, e.g. superinfection, plays an important role in the evolution of pathogens [13-15]. Superinfection is defined as the ability of an established pathogen to spread widely by infecting already-infected individuals with other strains of a pathogen (secondary infection) [14]. Thus, competition among different strains of a pathogen occurs and leads to increased fitness of the pathogens relative to single infection [16].

There are several theoretical studies pertinent to evolution of pathogen virulence with superinfection [17-23]. They defined that the already-infected individual is supreinfected and taken over by other strain of pathogen with higher virulence, thus strains do not share in a host. Additionally, most of them assumed that there is trade-off between the infection rate and the virulence of the pathogen, and considered the evolution of virulence in a host population by the host-parasite model. If hosts are long-lived, pathogens should get a long-term benefit from the plants by a low infection rate. However, if hosts are short-lived, the infection rate is supposed to evolve to a higher value, because pathogens should propagate quickly in a new host before the death of the hosts from the high infection rate. In addition, previous studies analyzed the evolution of virulence in the two cases where a host is either infected by only one strain of a pathogen (single infection) or by several strains (superinfection) of a pathogen. As a result, in the model of single infection, the virulence compared with single infection. However, they did not consider the spatial structures, such as the configuration of ramets in plant population.

Thus, we focus on the spatial structures, which play an important role in evolution of both plants and pathogens. The interaction between a plant and a pathogen depends on the spatio-temporal dynamics of pathogen dispersal, the genetic diversity of the host plant population and the spatial positioning of ramets [24, 25]. According to Koubek and Herben (2008), there are several defense reactions against systemic pathogens in clonal plant, such as (i) increasing the growth speed of the plant [8, 9, 26–28], (ii) dispersing the risk of infection and spread of pathogen by splitting the physical connection of ramets [7, 29], and (iii) detaching the infected ramets or tissues deliberately [30]. Additionally it is expected that features of the host would assist local pathogen transmission and evolution of the pathogen to lower levels of virulence [21], because clonal growth increases the probability of finding susceptible hosts in the vicinity of the initially infected host. However, they did not consider the superinfection event.

To explain these dynamics, we constructed a model based on the contact process (CP) [31], especially the two stage contact process (TCP) [32, 33], in mathematical models. These are simple models that express the dynamics of the contact infection process. The CP assumes only two states, healthy and infected, and healthy individuals are infected by only neighboring infected individuals. By contrast, TCP assumes three states, such as empty, healthy and infected, and healthy individuals reproduce new offspring and infected individuals increase only by transmission to healthy ones. Therefore, we adopted TCP to express not only the infection process, but also the plant growth process. In addition, we analyzed this model on the lattice space for simplicity. However, an explicit solution was not obtained in TCP, because it is too difficult to solve analytically. Thus, we performed an approximation by applying the meanfield approximation (MA) and the pair approximation (PA) to discuss the behavior of the system analytically. MA is the simplest approximation method, which takes no account of the effect of other sites. PA assumes that the effects of distant sites will be less important than those of the nearest neighbor sites. Therefore, this approximation is very useful to analyze the effect of local connections. Several studies have applied PA to TCP [34-36]. Sato et al. (1994) analyzed TCP in detail using PA; however, they could not determine the stability of the epidemic equilibrium (coexistence of healthy and infected individuals) analytically. Satulovsky and Tomé (1994) studied a predator-prey system using an improved TCP that had similar transition rules to our model, and obtained the analytical result with respect to the coexistence equilibrium. However, we did not use their model to express the superinfection process directly because they assumed only one predator species (not many). Haraguchi and Sasaki (2000) considered mutants of pathogens, which have additional mortality, based on TCP and analyzed the ESS (Evolutionarily Stable Strategy) of the mortality and transmission rate by computer simulation. However they did not include the superinfection event in propagation process of pathogens. Thus, further modification of the models is needed to express the pathogen spread process, including superinfection.

In this paper, we explain the plant reproduction and pathogen propagation dynamics considering special structures (using lattice structured space) and the effect of superinfection events on genetic diversity of pathogens using several models, the 1-strain and multiple-strain models. In the analysis of the 1-strain model, we derived an equilibrium value by MA and PA, and its local stability by the Routh—Hurwitz stability criterion. In the multiple-strain models, we analyzed the dynamics by numerical simulation using MA, PA and Monte Carlo simulation (MCS). Through these analyses, we showed the effect of parameter values on the dynamics of the models, such as density of individuals, transition of a dominant strain of a pathogen, and competition between plants and pathogens. As a result, the superinfection event leads to the polymorphism of pathogen strains in the host plant population, the dominant strain changes depending on plant and pathogen ability; and the competition is observed in particular parameter range. Thus, the plant and pathogen affect in the course of evolution mutually. In addition, when the growth rate of a plant is low, our present model is largely affected by the spatial structure.

Analysis

We constructed a model of plant growth and pathogen propagation processes, including superinfection. In the model, we assume that there is a single species of plant and multiple strains of a pathogen, and that a healthy plant individual is infected by a strain of the pathogen and an already-infected plant is superinfected by other strains of the pathogen. The dynamics of our model is a continuous Markov process on a lattice space, and the states of each site, the transition rate and mortality of each state are represented by a vector $\Omega = (\sigma_0, \sigma_1, \sigma_2, \dots, \sigma_n)$, $\mathbf{B} = (\beta_{\sigma_0}, \beta_{\sigma_1}, \beta_{\sigma_2}, \dots, \beta_{\sigma_n})$ and $\mathbf{D} = (d_{\sigma_0}, d_{\sigma_1}, d_{\sigma_2}, \dots, d_{\sigma_n})$, respectively, where the total number of states is n+1. Let $\rho_{\sigma_i}(t)$ be the probability that a randomly chosen site has state σ_i at time t, in other words, $\rho_{\sigma_i}(t)$ means the global density of the site with state $\sigma_0 = "0"$, $\sigma_1 = "S"$, and $\sigma_{i+1} = "I_i"(i = 1, 2, \dots, n - 1)$, which mean the empty, susceptible (healthy) individual, and infected individual by a pathogen with *i*-strains, respectively. In addition, we assumed that the already-infected individuals with *i*-strain ("I_i") are superinfected (and taken over) by the more virulent *j*-strain than *i*-strain, because the strain with higher virulence often wins within-host competition [21, 37]. From the definition of d_0 and β_0 , $d_0 = 0$ and $\beta_0 = \frac{1}{n} \sum_{i=1}^n d_{\sigma_i}$ because the empty site does not die and transition to "0" means death of healthy and infected individuals.

We configured four demographic processes: (i) the plant growth process, (ii) the first infection process, (iii) the superinfection process and (iv) the death process. The growth process is represented by transition from state "0" to "S", which means that plants grow their ramets into an open area (i.e. an empty site is occupied by a healthy individual). The first infection process is represented by transition from state "S" to "I_i", which means healthy individuals are infected by pathogens of the *i*-strain. The superinfection process is represented by transition from state "I_i" to "I_j" (i > j). The death process is represented by transition from "S" or "I_i" to "0" which represent the death of healthy and infected individuals from natural causes and virulence of the pathogen, respectively. In addition, infected individuals are not able to recover to a healthy one. Thus, we describe these processes by the following notation, which is often used to explain the TCP.

(i)
$$0 \rightarrow S$$
 at rate $\frac{\beta_{S}n(S)}{z}$
(ii) $S \rightarrow I_{i}$ at rate $\frac{\beta_{I_{i}}n(I_{i})}{z}$
(iii) $I_{i} \rightarrow I_{j}$ at rate $s\frac{\beta_{I_{j}}n(I_{j})}{z}$
(iv) $S, I_{i} \rightarrow 0$ at rate $d_{S}, d_{I_{i}}$
(TP)

Parameter $n(\sigma_i)$ is the number of σ_i -sites in the nearest neighbor of focal sites, z is the number of nearest-neighbor sites (e.g. z = 4 for von Neumann neighborhood on the two-dimensional square lattice), and s is the superinfection rate. Thus $s\beta_{I_j}$ describes the proportion of superinfection to first infection [18]. The growth or infection event occurs at a rate proportional to the number of the healthy or infected state in nearest-neighbor sites, respectively.

Here, let $q_{\sigma_i/\sigma_i}(t)$ be the conditional probability that a randomly chosen nearest neighbor of a σ_i -site is a σ_j -site. In particular, q_{σ_i/σ_i} means the local density of σ_i -site. $P_{\sigma_i \sigma_j}(t)$ is the probability that a randomly chosen site has state σ_i and a randomly chosen nearest-neighbor site has state σ_j at time *t*. These variables have the following relationship [34, 38].

$$P_{\sigma_i \sigma_i} = \rho_{\sigma_i} q_{\sigma_i/\sigma_i}.$$
 (1)

Thus, we can describe the following set of master equations, which is referred to as the general model (GM) from the above dynamics.

$$\begin{split} \dot{P}_{00} &= 2 \left(\sum_{i=1}^{n-1} d_{i_{i}} P_{i_{10}} + d_{s} P_{s0} - \frac{\beta_{s}(z-1)q_{s/00}}{z} P_{00} \right), \\ \dot{P}_{s0} &= \frac{\beta_{s}(z-1)q_{s/00}}{z} P_{00} + d_{s} P_{ss} - \frac{\beta_{s}}{z} P_{s0} - \frac{\beta_{s}(z-1)q_{s/0s}}{z} P_{s0} \\ &- d_{s} P_{s0} + \sum_{i=1}^{n-1} \left[d_{i_{i}} P_{si_{i}} - \frac{\beta_{i_{i}}(z-1)q_{i_{i}/s0}}{z} P_{s0} \right], \\ \dot{P}_{ss} &= 2 \left(\frac{\beta_{s}}{z} P_{s0} + \frac{\beta_{s}(z-1)q_{s/0s}}{z} P_{s0} - \sum_{i=1}^{n-1} \frac{\beta_{i_{i}}(z-1)q_{i_{i}/s0}}{z} P_{ss} - d_{s} P_{ss} \right), \\ \dot{P}_{10} &= \frac{\beta_{1}(z-1)q_{1i/s0}}{z} P_{s0} + d_{s} P_{si_{i}} - \frac{\beta_{s}(z-1)q_{s/0i_{i}}}{z} P_{10} + \sum_{j=1}^{n-1} d_{i_{j}} P_{1,i_{j}} - d_{i_{i}} P_{i,0} \\ &+ s \left(\sum_{j=i+1}^{n-1} \frac{\beta_{i_{i}}(z-1)q_{i_{i}/s0}}{z} P_{s0} + d_{s} P_{si_{i}} - \frac{\beta_{s}(z-1)q_{s/0i_{i}}}{z} P_{si_{0}} - \sum_{j=1}^{n-1} \frac{\beta_{i_{j}}(z-1)q_{i_{j}/i_{0}}}{z} P_{i,0} \right), \\ \dot{P}_{si_{i}} &= \frac{\beta_{s}(z-1)q_{s/0i_{i}}}{z} P_{0i_{i}} + \frac{\beta_{i_{i}}(z-1)q_{i_{i}/si_{j}}}{z} P_{ss} - \frac{\beta_{i_{i}}}{z} P_{si_{i}} - \sum_{j=1}^{n-1} \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{i}}}{z} P_{si_{i}} \\ &- \left(d_{s} + d_{i_{i}} \right) P_{si_{i}} + s \left(\sum_{j=i+1}^{n-1} \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{j}}}{z} P_{si_{j}} - \sum_{j=1}^{n-1} \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{j}}}{z} P_{si_{i}} \right), \\ \dot{P}_{i,i_{j}} &= \frac{\beta_{i_{i}}(z-1)q_{i_{i}/si_{j}}}{z} P_{si_{j}} + \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{j}}}{z} P_{i,j_{i}} - \left(d_{i_{i}} + d_{i_{j}} \right) P_{i,i_{j}} \\ &+ s \left(\sum_{k=i+1}^{n-1} \frac{\beta_{i_{k}}(z-1)q_{i_{k}/si_{j}}}{z} P_{i,j_{i}} - \sum_{k=i}^{k-1} \frac{\beta_{i_{k}}(z-1)q_{i_{k}/si_{j}}}{z} P_{i,j_{i}} - \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{j}}}{z} P_{i,j_{i}} \right) \right) (i > j), \\ \dot{P}_{i,i_{i}} &= 2 \left[\frac{\beta_{i_{i}}}{z} P_{si_{i}} + \frac{\beta_{i_{i}}(z-1)q_{i_{i}/si_{j}}}{z} P_{si_{i}} - d_{i_{i}} P_{i,j_{i}} \right) - \sum_{j=i}^{i-1} \frac{\beta_{i_{j}}(z-1)q_{i_{j}/si_{j}}}{z} P_{i,j_{i}} \right) \right]. \end{split}$$

Here, for example, in the right side of the fifth equation in the set (i.e. differential equation of P_{SI_i}), the first term describes the transition from P_{0I_i} to P_{SI_i} , which means healthy individuals reproduce their offspring to empty sites (state $0 \rightarrow S$). In this term, the transition rate is determined by TP(i), n(S) is $(z - 1)q_{S/0I_i}$. Transition begins from P_{0I_i} ; therefore, one of the nearest-neighbor sites of the 0-state site is state I_i , and at least one of the other nearest-neighbor sites of the 0-state site (in (z-1) sites) should be S to transition

PLOS

from 0 to S. Thus, the probability is $q_{S/0I_i}$, and the expectation of n(S) is equal to $(z-1)q_{S/0I_i}$. In subsequent terms, the $n(\sigma)$ ($\sigma \in \Omega$) is obtained in a similar process, except the third term.

The second and third terms mean that healthy individuals are infected by pathogens with *i*-strain (state $S \rightarrow I_i$). These terms describe the transition from P_{SS} to P_{SI_i} and from P_{SI_i} to $P_{I_iI_i}$, respectively, and the transition rate of these terms is determined by TP(ii). In particular, the value of $n(I_i)$ in the third term is equal to 1. The transition begins from P_{SI_i} ; therefore, there is already a I_i -state site in nearest-neighbor sites of S-state site. Here, the case that the state of other sites in the nearest-neighbor sites is also I_i is included in the fourth term.

The fourth term means that healthy individuals are infected by pathogens with *j*-strain (state S \rightarrow I_{*j*}). This term describes the transition from P_{SI_i} to $P_{I_jI_i}$ ($i \in j$), and sums the transition rates in respect to all strains (from I₁ to I_{*n*-1}).

The fifth term means the death of healthy or infected individuals (state S, $I_i \rightarrow 0$). The term describes the transition from P_{SI_i} to P_{0I_i} or P_{S0} and the transition rate of each process is determined by TP(iv).

The last term means that the already-infected individuals are superinfected by other strains of the pathogen (e.g. state $I_2 \rightarrow I_1$). The first and second terms in parentheses describe the transition from P_{SI_j} to P_{SI_i} and from P_{SI_i} to P_{SI_i} ($j \neq i$), respectively, and the transition rate of these term is determined by TP(iii). The first term means that the infecting pathogens super-infect already-infected individuals with another strain, and the second term means that an already-infected individual is superinfeced by another strain of the pathogen. Thus, the range of summation in the first term is from i+1 to n-1 and in the second term is from 1 to i-1 from our assumption.

Results

In the subsequent sections, we introduce a new parameter m_i (we refer to the parameter as mortality cost in this paper) defined as β_{I_i}/d_{I_i} for n-1 strains ($m_i := \beta_{I_i}/d_{I_i}$, $m_1 < m_2 < \cdots < m_{n-1}$), and set $d_{I_i} = 1$ ($\forall I_i \in \Omega$) to standardize the parameter, for ease of analysis. The mortality cost means the expectation of the number of new infected-individual produced during a life-time of an infected-individual, thus, higher virulent strain has lower mortality cost. Consequently, already-infected individuals are superinfected by the strains with lower mortality costs. In addition, we set $d_S \approx 0$, because the plant mortality is generally smaller than plant growth rate in clonal plant (i.e. the plants are long lived).

1-strain model

Initially, we analyzed the simplest case (n = 2) in GM by MA and PA. The state of each site was denoted by $\sigma_i \in \mathbf{S} \equiv \{0, S, I\}(I: = I_1)$ from the assumption of only one strain of pathogen.

Therefore, we obtained the following set of master equations to rewrite Eq (2).

$$\begin{split} \dot{P}_{00} &= 2P_{10} - 2\frac{\beta_{\rm S}(z-1)q_{\rm S/00}}{z}P_{00}, \\ \dot{P}_{\rm S0} &= P_{\rm SI} + \frac{\beta_{\rm S}(z-1)q_{\rm S/00}}{z}P_{00} \\ &- \left[\frac{\beta_{\rm S}}{z} + \frac{\beta_{\rm S}(z-1)q_{\rm S/0S}}{z} + \frac{m_{\rm I}(z-1)q_{\rm I/S0}}{z}\right]P_{\rm S0}, \\ \dot{P}_{10} &= P_{\rm II} + \frac{m_{\rm I}(z-1)q_{\rm I/S0}}{z}P_{\rm S0} - \left[\frac{\beta_{\rm S}(z-1)q_{\rm S/0I}}{z} + 1\right]P_{\rm I0}, \\ \dot{P}_{\rm SS} &= 2\left[\frac{\beta_{\rm S}}{z} + \frac{\beta_{\rm S}(z-1)q_{\rm S/0S}}{z}\right]P_{\rm S0} - 2\left[\frac{m_{\rm I}(z-1)q_{\rm I/SS}}{z}\right]P_{\rm SS}, \\ \dot{P}_{\rm SI} &= \frac{\beta_{\rm S}(z-1)q_{\rm S/0I}}{z}P_{0\rm I} + \frac{m_{\rm I}(z-1)q_{\rm I/SI}}{z}P_{\rm SS} \\ &- \left[\frac{m_{\rm I}}{z} + \frac{m_{\rm I}(z-1)q_{\rm I/SI}}{z} + 1\right]P_{\rm SI}, \\ \dot{P}_{\rm II} &= 2\left[\frac{m_{\rm I}}{z} + \frac{m_{\rm I}(z-1)q_{\rm I/SI}}{z}\right]P_{\rm SI} - 2P_{\rm II}. \end{split}$$

In this model, superinfection does not occur because there is only one strain of the pathogen.

Mean-field Approximation. To close a set of Eq (3), we approximated several variables by MA, $q_{\sigma_i/\sigma_j} \approx \rho_{\sigma_i}$ (e.g. $P_{S0} = \rho_S q_{0/S} \approx \rho_S \rho_0$) and $q_{\sigma_i/\sigma_j \sigma k} \approx \rho_{\sigma_i}$. In addition, the equations were simplified by the definition of variables from Eq (S1.3) in S1 Appendix.

$$\dot{\rho_0} = 1 - \rho_0 - \rho_s (1 + \beta_s \rho_0),
\dot{\rho_s} = \rho_s (\beta_s \rho_0 - m_1 (1 - \rho_0 - \rho_s)).$$
(4)

The system has three equilibrium states.

$$\begin{split} \boldsymbol{E}_{\mathrm{M}} &\equiv \left(\tilde{\rho}_{0}^{*}, \, \tilde{\rho}_{\mathrm{S}}^{*}, \, \tilde{\rho}_{\mathrm{I}}^{*}\right) = (1, \, 0, \, 0), \\ \hat{\boldsymbol{E}}_{\mathrm{M}} &\equiv \left(\hat{\rho}_{0}^{*}, \, \hat{\rho}_{\mathrm{S}}^{*}, \, \hat{\rho}_{\mathrm{I}}^{*}\right) = (0, \, 1, \, 0), \\ \bar{\boldsymbol{E}}_{\mathrm{M}} &\equiv \left(\bar{\rho}_{0}^{*}, \, \bar{\rho}_{\mathrm{S}}^{*}, \, \bar{\rho}_{\mathrm{I}}^{*}\right) \\ &= \left(\frac{m_{\mathrm{I}} - 1}{\beta_{\mathrm{S}} + m_{\mathrm{I}}}, \, \frac{1}{m_{\mathrm{I}}}, \, \frac{\beta_{\mathrm{S}}(m_{\mathrm{I}} - 1)}{m_{\mathrm{I}}(\beta_{\mathrm{S}} + m_{\mathrm{I}})}\right). \end{split}$$

 \tilde{E}_{M} , \tilde{E}_{M} and \bar{E}_{M} mean the states of extinction, disease-free and epidemic, respectively. From local stability analysis of the each equilibrium (see S2 Appendix), \tilde{E}_{M} is always unstable, which means that plants do not become extinct at the positive parameter range in the system. When $m_{I} < 1$, \hat{E}_{M} is stable, which means that the pathogen is not able to survive when it has a low mortality cost. By contrast, when m_{I} exceeds 1, \hat{E}_{M} becomes unstable and \bar{E}_{M} becomes always stable, which means that the epidemic occurs because of the spread of the pathogens within the plant population. Thus, $m_{I} = 1$ is the threshold value of stability shifting. In conclusion, the

stability of the equilibrium states and the equilibrium density of healthy individuals (ρ_s^*) in the epidemic state depend only on m_I , regardless of β_s in MA.

Pair Approximation. To close a set of Eq (3) and consider the effect of local connections on the dynamics, we approximated several variables by PA, $q_{S/0\sigma} \approx q_{S/0}$ and $q_{I/S\sigma} \approx q_{I/S}$. In addition, the equations were simplified by definition of variables (see S1 Appendix), and we obtained following three equilibrium states.

$$\begin{split} \hat{\boldsymbol{E}}_{P} &\equiv (\tilde{\rho}_{0}^{*}, \tilde{\rho}_{S}^{*}, \tilde{\rho}_{1}^{*}, \tilde{q}_{0/0}^{*}, \tilde{q}_{S/0}^{*}, \tilde{q}_{1/0}^{*}) \\ &= (1, 0, 0, 1, 0, 0), \\ \hat{\boldsymbol{E}}_{P} &\equiv (\hat{\rho}_{0}^{*}, \hat{\rho}_{S}^{*}, \hat{\rho}_{1}^{*}, \hat{q}_{0/S}^{*}, \hat{q}_{S/S}^{*}, \hat{q}_{1/S}^{*}) \\ &= (0, 1, 0, 0, 1, 0), \\ \bar{\boldsymbol{E}}_{P} &\equiv (\bar{\rho}_{0}^{*}, \bar{\rho}_{S}^{*}, \bar{\rho}_{1}^{*}, \bar{q}_{0/0}^{*}, \bar{q}_{S/0}^{*}, \bar{q}_{1/0}^{*}, \bar{q}_{0/S}^{*}, \bar{q}_{S/S}^{*}, \bar{q}_{1/S}^{*}, \bar{q}_{0/1}^{*}, \bar{q}_{S/1}^{*}, \bar{q}_{1/1}^{*}) \\ &= (\text{see S3 Appendix}). \end{split}$$

 $E_{\rm p}$ means that the plants become extinct. Therefore, $q_{\sigma/S}$ and $q_{\sigma/I}$ are non-existent, because $\rho_{\rm S}$ and $\rho_{\rm I}$ are equal to 0. $\hat{E}_{\rm p}$ means the disease-free state; thus, $q_{\sigma/0}$ and $q_{\sigma/1}$ are nonexistent, because $\rho_{\rm S}$ and $\rho_{\rm I}$ are non-existent, because $\rho_{\rm S}$ and $\rho_{\rm I}$ are equal to 0. $\bar{E}_{\rm p}$ means the epidemic state, at which state all of the twelve variables exist and have positive values. The local stability of each equilibrium state was examined using the Routh—Hurwitz stability criterion (see <u>S3 Appendix</u>). From the stability analysis, we obtained the three stable-equilibrium phases, disease-free, epidemic, and periodic oscillation (Fig1). In particular, $\tilde{E}_{\rm p}$ is always unstable (i.e. plants do not become extinct), and we derived two thresholds, which are referred to as the epidemic and bifurcation thresholds. The stability of $\hat{E}_{\rm p}$ and $\bar{E}_{\rm p}$



Fig 1. Phase diagram of pair approximation showing the three regions of the equilibrium state. In the epidemic region, plants and pathogens coexist and the equilibrium is stable. In the oscillation region, plants and pathogens coexist, but the equilibrium is unstable. Therefore, Hopf bifurcation occurs and oscillation is observed. The solid line indicates the bifurcation threshold, and the dashed line the epidemic threshold, respectively. In the disease-free region, pathogens become extinct because of the too low mortality cost.

doi:10.1371/journal.pone.0154883.g001



Fig 2. Comparison of the simulation results from mean-field approximation (MA), pair approximation (PA) and Monte Carlo simulation (MCS), depending on mortality cost and growth rate; (a). $\beta_s = 10$, (b). $\beta_s = 30$, (c). $m_l = 15$. (i) equilibrium density of healthy individuals (ρ_s^*) (ii) equilibrium density of infected individuals (ρ_1^*). The dots, solid line and dashed line indicate the results of MA, PA and MCS, respectively. These results show that when m_l is low, both approximation methods present similar trend to MCS, although these methods overestimate the equilibrium value. (iii) The variance among 100 trials in MCS. A high variance means the oscillatory solution is observed.

depends on whether the parameter values exceed each threshold, especially the epidemic condition depends only on the mortality cost, irrespective of the growth rate because the epidemic threshold is $(m_I)_c = z/(z-1)$, which is similar to MA. That is, if m_I is low, pathogens become extinct (panels (a) and (b) in Fig 2), because the low m_I means a high virulence or low infection rate, thus pathogens die within the infected hosts before infecting other hosts. In addition, a large m_I leads to a decrease in both healthy and infected individuals. β_S affects the equilibrium value in the epidemic phase. For example, a large β_S leads to an increase in the equilibrium density of infected individuals (panel (c) in Fig 2), because pathogens can spread widely within the many hosts supplied by the fast growth rate. In the epidemic phase, when β_S is large, the equilibrium density of healthy individuals (ρ_S^*) decreases, thus a low growth rate has an advantage over a high growth rate for plants in this phase. In addition, Hopf bifurcation occurs when the parameter values exceed to the bifurcation threshold (panel (b) in Fig 2). In other words, the stability of \hat{E}_p shifts from stable to unstable, which is different from MA.

To check the validity of each approximation method, we compared equilibrium values among MA, PA and MCS. The MCS was conducted 100 times for each given parameter set; (a). $\beta_S = 10$ and $m_I = 0 \sim 30$, (b). $\beta_S = 30$ and $m_I = 0 \sim 30$, (c). $m_I = 15$ and $\beta_S = 0 \sim 25$ in Fig 2. We used a two-dimensional square lattice torus and calculated an average value of 100 trials at each parameter value (Fig 2). The result indicated that there are several discrepancies with respect to the equilibrium values and threshold values among them: MA and PA overestimate the equilibrium values and the periodic solution is observed in PA under higher parameter values compared with MCS. In Fig 2, the discrepancy of the threshold value between MCS and MA (PA) is large when the mortality cost is large ((a), (b)) or the growth rate is low ((c)). The use of these approximation methods, in which we neglected the effect from far sites, is probably the cause of the discrepancies. However, the periodic solution was observed in the result of PA and MCS [35], which was differently from MA. Thus, PA could explain the basic behavior of the system better than MA.

Multiple-strain models

We examined four models (n = 3, 4, 11, and 26) in GM (Eq (2)); however, the analysis was too complex to obtain the analytical result because of too many variables (e.g. MA and PA require at least n and $\sum_{i=1}^{n} i - 1$ variables, respectively). Therefore, we derived the equilibrium value by MA in the 2-strain and 3-strain models, and analyzed all the models through computer simulations (MCS and numerical simulation of MA and PA). We set $m_i = m_{i-1} + \Delta m_{i,i-1} (i = 2, 3, ..., n-1)$, s = 0, 0.5, 1.0, 1.5 and vary the β_S in the simulations. In this paper, we assumed that $\Delta m := \Delta m_{i,i-1}$ was constant just for the simplicity of the model. As a result of the numerical simulations of MA and PA, the healthy individuals did not become extinct in all models. However, the healthy and infected individuals did become extinct in MCS, especially when n is large and β_S is small. From the comparison among MA, PA and MCS, discrepancies became large as the number of strains increased.

The 2-strain model (*n* = 3). We obtained five equilibrium states by MA (Table 1 in detail): E_1 : extinction ($\rho_0^* = 1$), E_2 : disease-free ($\rho_{I_1}^* = 0$, $\rho_{I_2}^* = 0$), E_3 : occupation of a strain with high $\cos t (\rho_{I_1}^* = 0, \rho_{I_2}^* > 0)$, E_4 : occupation of a strain with low $\cos t (\rho_{I_1}^* > 0, \rho_{I_2}^* = 0)$, E_5 : coexistence ($\rho_{I_1}^* > 0, \rho_{I_2}^* > 0$). In particular, in the (equilibrium) phase of coexistence (i.e. E_5 is stable), the dominant strain changes depending on the plant growth rate.

When s > 0, Figs 3 and 4 show the equilibrium density of each state depending on the growth rate (β_S) for a given superinfection rate (s) and the difference in mortality cost (Δm), respectively, in the simulation of MA (panels (a)), PA (panels (b)) and MCS (panels (c)) and transition of equilibrium phase (panels (d)). As a result, when β_S is small, the strain with a high cost is dominant, then the dominant strain shifts to strain with a low cost as β_S increases and the strain occupies the pathogen population on a large β_S . The increase in s and decrease in Δm lead to lower threshold values of the phase transition (panels (d) in Figs 3 and 4); thus, when s is small or Δm is large, the parameter range of the phase of coexistence increases. Notably, when Δm is large (moderate competition), multiple strains are more likely to coexist, like 'limiting similarity of niche' proposed by competition theory [39, 40]. For the healthy individuals, they increase their density at the coexistence phase and decrease it at other phases in PA. Thus, if the growth rate is out of the range value of the coexistence phase, healthy individuals do not increase their population.

E 1	(1, 0, 0, 0)
E ₂	(0, 1, 0, 0)
E ₃	$igg(rac{m_2-1}{eta_5+m_2}, \ rac{1}{m_2}, \ 0, \ rac{eta_5(m_2-1)}{m_2(eta_5+m_2)}igg)$
E ₄	$\left(rac{m_1-1}{eta_{ m S}+m_1}, ~ rac{1}{m_1}, ~ rac{eta_{ m S}(m_1-1)}{m_1(eta_{ m S}+m_1)}, ~ 0 ight)$
E 5	$ \left(\frac{m_2 - m_1}{s\beta_5 m_1}, \ \frac{m_1 (s\beta_5 + 1) - m_2}{\beta_5 (m_2 + m_1 (s-1))}, \ \frac{s\beta_5 m_1 (m_2 - 1) - (m_2 - m_1) (\beta_5 + m_2)}{s\beta_5 m_1 (m_2 + m_1 (s-1))}, \ \frac{m_1 (m_2 - m_1) + \beta_5 (m_2 - m_1 - sm_1 (m_1 - 1))}{s\beta_5 m_1 (sm_1 + m_2 - m_1)} \right) $
$E \equiv (\rho_0^*, \ \rho_S^*, \ \rho_{I_1}^*, \ \rho_{I_2}^*)$).

doi:10.1371/journal.pone.0154883.t001



Fig 3. The equilibrium value of each state and transition of the equilibrium phase depending on the superinfection rate in the 2-strain model. We set $m_1 = 5$ and $\Delta m = 5$. The I, II and III differ in the value of s (=0.5, 1.0, 1.5). (a)–(c) show the variation of the equilibrium density of each state ("0", "S", "I₁" and "I₂". $\mathcal{E}_{\sigma}\rho_{\sigma} = 1$) with growth rate in each simulation: (a) MA, (b) PA, (c) MCS. (d) shows the transition of the equilibrium phase with $\beta_{\rm S}$ in MA, PA, and MCS.



Fig 4. The equilibrium value of each strain and transition of the equilibrium phase depending on the difference of mortality cost in the 2-strain model. We set $m_{l_1} = 5$ and $\Delta m = 5$ in all figures. The I, II and III differ in the value of s (=0.5, 1.0, 1.5). (a) MA, (b) PA, (c) MCS, (d) the transition of equilibrium phase.



Fig 5. The case of no superinfection (s = 0) in the 2-strain and 3-strain model by MA, PA and MCS. We plotted the global density of each strain of the pathogen at the equilibrium state depending on β_s . The black circles and triangles indicate I_1 and I_2 , respectively, and squares indicate I_3 in (b). The parameter values are (a) n = 3, $m_{I_1} = 5$, $\Delta m = 10$, (b) n = 4, $m_{I_1} = 5$, $\Delta m = 5$.

When s = 0 (no superinfection), the shift of the equilibrium phase along the gradient of β_S is different from the case of s > 0 in MCS. For example, when β_S is large, the strain with the highest mortality cost occupies and coexists with healthy plants (i.e. strains with lower cost become extinct). In addition, the range of the coexistence phase decreases compared with the case of when s > 0 (Fig 5(i)). In MA and PA, the strain with a high cost always occupies regardless of other parameter values (panels (a) and (b) in Fig 5(i)). Thus, when s = 0 and β_S is large, the pathogen population is occupied by the strain with highest cost, contrary to the case of s > 0.

From the comparison of numerical simulations (MA and PA) with MCS, when β_S is small, the result is at extreme variance with MCS. Thus, in the parameter range, the approximation method cannot apply. When β_S is large enough, the discrepancy between them increases with the decrease in *s* (Fig 3) and Δm (Fig 4). However, MA and PA can explain the transition process of the equilibrium phase (panels (d) in Figs 3 and 4). In addition, the oscillatory solution, which means the solution is oscillating for a long time although unproven, is not observed in



Fig 6. Time development in the 2-strain model. We plotted the equilibrium value of the global density of each state. In all figures, we set s = 1.0 and $\Delta m = 15$. I. $m_{l_1} = 5$, II. $m_{l_2} = 30$ and (a) $\beta_S = 10$, (b) $\beta_S = 15$, (c) $\beta_S = 25$.

the coexistence phase ($\underline{Fig 6}$). However, in the occupation phase of a strain, the oscillatory solution is observed because the behavior of the model follows the 1-strain model in that phase.

The 3-strain model (*n* = 4). We obtained nine equilibrium states by MA (<u>Table 2</u> in detail): *E*₁: extinction, *E*₂: disease-free, *E*₃₋₅: occupation of a strain ($\rho_{l_i}^* > 0$, $\rho_{l_j}^* = 0$, $\rho_{l_k}^* = 0$), *E*₆₋₈: coexistence of two strains ($\rho_{l_i}^* > 0$, $\rho_{l_j}^* > 0$, $\rho_{l_k}^* = 0$), *E*₉: coexistence of all strains ($\rho_{l_i}^* > 0$, $\rho_{l_j}^* >$

When s > 0, Figs 7 and 8 show the equilibrium density of each state depending on β_s in a given *s* and the difference in mortality costs (Δm), respectively, by the numerical simulation of MA (panels (a)), PA (panels (b)) and MCS (panels (c)). Among them, the occupation phase of a strain and the coexistence phase of two strains showed a generally similar response to the parameter values in the 1-strain and 2-strain models. The effect of *s* and Δm on equilibrium values, threshold values and discrepancies among simulation results were similar to the

Table 2. Equilibria of the 3-strain model in MA.

E ₁	(1, 0, 0, 0, 0)
E ₂	(0, 1, 0, 0, 0)
E ₃	$\left(rac{m_1-1}{eta_5+m_1}, \ rac{1}{m_1}, \ rac{eta_5(m_1-1)}{m_1(eta_5+m_1)}, \ 0, \ 0 ight)$
E ₄	$\left(rac{m_2-1}{eta_5+m_2}, \ rac{1}{m_2}, \ 0, \ rac{eta_5(m_2-1)}{m_2(eta_5+m_2)}, \ 0 ight)$
E ₅	$\left(rac{m_3-1}{eta_5+m_3}, \ rac{1}{m_3}, \ 0, \ 0, \ rac{eta_s(m_3-1)}{m_3(eta_5+m_3)} ight)$
E ₆	$ \left(\frac{m_2 - m_1}{s\beta_5 m_1}, \ \frac{m_1(s\beta_5 + 1) - m_2}{\beta_5(m_2 + m_1(s-1))}, \ \frac{s\beta_5 m_1(m_2 - 1) - (m_2 - m_1)(\beta_5 + m_2)}{s\beta_5 m_1(m_2 + m_1(s-1))}, \ \frac{m_1(m_2 - m_1) + \beta_5(m_2 - m_1 - sm_1(m_1 - 1))}{s\beta_5 m_1(sm_1 + m_2 - m_1)}, \ 0 \right) $
E ₇	$\left(\frac{m_3 - m_1}{\mathfrak{s}\beta_8 m_1} , \ \frac{m_1(\mathfrak{s}\beta_8 + 1) - m_3}{\beta_8(m_3 + m_1(\mathbf{s}-1))} , \ \frac{\mathfrak{s}\beta_8 m_1(m_3 - 1) - (m_3 - m_1)(\beta_8 + m_3)}{\mathfrak{s}\beta_8 m_1(m_3 + m_1(\mathbf{s}-1))} , \ 0 , \ \frac{m_1(m_3 - m_1) + \beta_8(m_3 - m_1 - \mathfrak{s}m_1(m_1 - 1))}{\mathfrak{s}\beta_8 m_1(\mathfrak{s}m_1 + m_3 - m_1)} \right)$
E ₈	$ \begin{pmatrix} \underline{m_3 - m_2} \\ \underline{s\beta_5 m_2} \\ \\ n_5(\underline{m_3 + m_2(s-1))} \end{pmatrix}, \ 0, \ \frac{\underline{s\beta_5 m_2(\underline{m_3 - 1}) - (\underline{m_3 - m_2})(\underline{\beta_5 + m_3})}{\underline{s\beta_5 m_2(\underline{m_3 + m_2(s-1))}} \\ \\ \\ n_2(\underline{s-1}) \\ n_2(s$
E 9	$\left(\frac{m_1m_3-m_2}{\beta_Sm_2+m_1m_3}, \ \frac{m_2}{m_1m_3}, \ \frac{\beta_Sm_2(s(m_1m_3-m_2)-m_3+m_2)-m_1m_3(m_3-m_2)}{sm_1m_3(\beta_Sm_2+m_1m_3)}, \right)$
	$ \left. \frac{m_2(m_3-m_1)-sm_1(m_2+m_1m_3)}{sm_1m_3(\beta_Sm_2+m_1m_3)}, \ \frac{m_1m_3(s\beta_Sm_1-m_2+m_1)-\beta_Sm_2(sm_1+m_2-m_1)}{sm_1m_3(\beta_Sm_2+m_1m_3)} \right) $

 $E \equiv (\rho_0^*, \ \rho_{S}^*, \ \rho_{I_1}^*, \ \rho_{I_2}^*, \ \rho_{I_3}^*).$

doi:10.1371/journal.pone.0154883.t002



Fig 7. The equilibrium value of each strain depending on the superinfection rate in the 3-strain model. We set $m_{I_1} = 5$ and $\Delta m = 5$ in all figures. The I, II and III differ in the value of s (=0.5, 1.0, 1.5). (a) MA, (b) PA, (c) MCS.



Fig 8. The equilibrium value of each strain depending on the difference in the mortality cost in the 3-strain model. We set $m_{l_1} = 5$ and s = 1.5 in all figures. The I, II and III differ in the value of Δm (=5, 10, 15). (a) MA, (b) PA, (c) MCS.

2-strain model. Thus, the increase or decrease of the density of healthy individuals depends on β_S in PA, which is different from MA (i.e. the growth rate negatively affects the density of healthy individuals in MA). However, when *s* and Δm are both large, the transition of the equilibrium phase in MA (PA) was different from the MCS, because the strain with highest cost



Fig 9. Time development of global density of each state in the 3-strain model. We plotted the equilibrium value of the global density of each state. We set the parameter values; I. s = 1.0, $m_{l_1} = 10$, $\Delta m = 15$, II. s = 1.5, $m_{l_1} = 10$, $\Delta m = 20$ and (a) $\beta_S = 3$, (b) $\beta_S = 5$, (c) $\beta_S = 7$. In the 3-strain model, the oscillatory solution was observed in a particular parameter range at the coexistence phase (panel(b)).

 (I_3) became extinct in the MCS. In addition, the oscillatory solution was observed in a parameter range (Fig 9) and the strain with the middle cost (I_2) was dominant, similar to the results of the previous studies [17–21](Figs 7 and 8). In addition, when s = 0 (Fig 5(ii)), the response to β_S was the same as in the 2-strain model (i.e. the range of the coexistence phase decreased and pathogen population was occupied by strain with the highest cost at a large β_S).

Multiple-strain models. In the multiple-strain models, there are many equilibrium states: extinction, disease-free, occupation of a strain, coexistence of various strains, and coexistence of all strains.

We plotted the results of the 10-strain (Fig 10) and 25-strain models (Fig 11) at $\Delta m = 5$ with varying β_S and *s*. As a result, a smaller value of β_S or *s* led to the dominance of the strain with higher cost (Figs 10 and 11) and the cost of the dominant strain shifted to a lower value as these parameter values increased, similar to the 2-strain and 3-strain model. In addition, when the β_S was small, the oscillatory solution was observed (e.g. Fig 12), and when the *s* was also small, there was a possibility of extinction in MCS when the *n* was too large (Fig 11).

Discussion

The physical connection through ramets, especially through the vascular system of plants, is important to spread pathogens within a clonal plant population [25]. Viruses are able to spread through the vascular system [41], and fungal pathogens are able to grow their hyphae through the intercellular spaces of vascular vessels [42]. By contrast, plants have evolved to increase the clonal growth rate to escape from disease when (systemic) pathogens invade the population [8, 9, 26–28]. Thus, clonal plants and pathogens have been mutually affected in the course of evolution.

Several studies have analyzed the pathogen spread process by an approximation method on the lattice space [34-36] and the superinfection process using mathematical models without spatial structures [18]. Sato et al. (1994) studied the case of $d_s = 1$ in our 1-strain model. Their model had three equilibrium states, (disease-free, endemic and epidemic), and they derived equilibrium values at two states (disease-free and endemic) and their local stability condition, explicitly. However, they did not obtain the epidemic equilibrium value and its local stability. Haraguchi and Sasaki (2000) considered that there is no trade-off between the infection rate



Fig 10. The equilibrium density distribution of strains in 10-strain model. The left, center and right panels show the result in simulation by MA, PA and MCS, respectively. We set $m_{l_1} = 5$, $\Delta m = 5$ and other parameter values are: I. s = 0.5, $\beta_S = 25$, II. s = 1.0, $\beta_S = 25$, III. s = 1.5, $\beta_S = 25$, IV. s = 1.5, $\beta_S = 5$.

and the virulence of a pathogen and assumed that multiple pathogens have different virulences. They examined the ESS of the infection rate by numerical simulation and discussed the evolution of the infection rate of the pathogen. Their simulation suggested that pathogens evolve to an intermediate infection rate. Satulovsky and Tomé (1994) considered a transition rule similar to our model and assumed the correlation between the transition rates of each state (i.e. $\beta_{\rm S} + m_{\rm I} + d_{\rm I} = 1$). According to their results of PA and MCS, there are four equilibrium phases (three stationary and one oscillation phase), and if the transition rate to state 0 ($d_{\rm I}$) is small, the Hopf bifurcation occurs. Nowak and May (1994) examined the superinfection events using ODE (Ordinary Differential Equation). As a result, superinfection leads to maintenance of the polymorphism of a parasite strain, and the oscillatory solution (i.e. competition among plants and pathogens) is observed when there is more than one strains. However, they did not consider the spatial structure and host reproductive dynamics (they assumed that the host increases constantly).

To consider the effect of spatial structures, we analyzed plant reproduction and pathogen propagation dynamics with superinfection in a clonal plant population on the lattice space. We analyzed five models (the 1-strain model and four multiple-strain models), including interaction among plant and several strains of pathogen, and adopted the MA and PA to analyze the dynamics of the models. In addition, we checked the validity of the approximation methods in comparison with MCS.

PLOS



Fig 11. The equilibrium density distribution of strains in the 25-strain model. The left, center and right panels show the result in simulation by MA, PA and MCS, respectively. We set $m_{l_1} = 5$, $\Delta m = 5$ and other parameter values are: I. s = 0.5, $\beta_S = 5$, II. s = 0.5, $\beta_S = 25$, III. s = 1.0, $\beta_S = 25$, IV. s = 1.5, $\beta_S = 25$.

Using the 1-strain model (n = 2), the value of the epidemic threshold depended only on the mortality cost, which means that to establish a pathogen within a plant population depends only on the ability of the pathogens, regardless of plants. In addition, the density of healthy individuals decreases with increasing growth rate and mortality cost (panels (i) in Fig 2). By contrast, the density of infected individuals increases with the growth rate and decreases with the increase in mortality cost (panels (ii) in Fig 2). Therefore, plants should not increase their growth rate to maintain a large population size when infected by a systemic pathogen, and the





doi:10.1371/journal.pone.0154883.g012

pathogen should evolve a low mortality cost that is higher than the epidemic threshold to maintain their population and that of their hosts.

In addition, if the parameter values exceed the bifurcation threshold, the Hopf bifurcation occurs and the periodic solution, which means plants and pathogens continue to compete forever, is observed in PA and MCS, which is different from MA. Thus, considering the effect of local interaction is important to express the dynamics of the pathogen propagation process. However, MA (which neglect the local interaction) is useful to analyze the dynamics in a measure such as the number of equilibrium states in the model.

In the multiple-strain models (n > 2), we assumed that multiple strain of a pathogen had different mortality costs, and that the already-infected individuals are superinfected by strains with lower mortality costs. The analytical result of MA showed that there are a lot of equilibrium states: extinction, disease-free, occupation of a strain, coexistence of various strains, and coexistence of all strains (Tables 1 and 2). From the results of the simulation of MA, PA and MCS, the equilibrium phase and the dominance of a strain in the coexistence phases depend on parameter values and the oscillatory solution is observed in the coexistence phases, except for the 2-strain model in PA and MCS (Figs 6 and 9).

In addition, the genetic diversity of a pathogen is maintained by a decrease in superinfection events. In fact, the parameter range of the coexistence phase increases with the decrease in superinfection rates, even when the difference in mortality cost is small. This is because a superinfection event is conducive to the strong competition among strains from a decrease in the difference of spread-speed among them by additional transmission routes. But, if superinfection does not occur (s = 0), the range of the coexistence phase decreases (Fig 5), thus super-infection is important to maintain genetic diversity. However, when the plant growth rate increases, the pathogen population is occupied by a strain eventually, regardless of the superinfection rate. (Figs 3, 5 and 7). That is, the increase in the growth rate provides them with no benefit. Healthy individuals can increase their abundance via the growth rate in the coexistence phase of several strains; however, if the growth rate is too high, an equilibrium phase shifts to the phase of occupation of one strain, and the density of healthy individuals then decreases with increasing growth rate. The dynamics of the model follows the 1-strain model in this phase.

In summary, pathogens maintain their genetic diversity through superinfection events and a moderate mortality cost relative to growth rate; thus they evolve their mortality cost and superinfection rate depending on plant growth ability to maintain their population. By contrast, the number of healthy individuals (plants) increases in (all and several strains) coexistence phases with the growth rate. Thus, when systemic pathogens invade the plant population, plants evolve the growth rate to be slightly lower than a threshold value at which the equilibrium phase shifts to the phase of occupation of a strain to increase their population.

In this paper, we constructed simple models to express the relationship between clonal growth and pathogen spread through superinfection, considering the spatial structure. We showed that: (i) The strain with an intermediate cost becomes dominant, similar to the previous studies [17-21], when both the superinfection rate and the growth rate are low. However, a high superinfection rate or growth rate leads to dominance of the strain with lowest cost in our model. Actually, pathogen gets more benefit due to low mortality cost when hosts grow rapidly [3]. (ii) The competition among strains occurs in the coexistence phase of various strains by PA and MCS in the model of n > 3. (iii) Too high a growth rate leads to occupation of the strain with lowest cost. Thus, the competition between the strain and the hosts occurs, thereby the host population decreases in all models. (iv) Pathogens easily maintain their genetic diversity with a low superinfection rate. However, if they do not superinfect, such maintenance

becomes difficult. (v) When the growth rate of a plant is low, an individual at a local site is strongly interconnected by distant individuals because MA and PA do not apply in this case. In conclusion, the superinfection is one of the important factors to evolution of virulence by maintenance of genetic diversity, and the spatial structure plays a more important role in a slowly growing plant relative to the speed of spread of the pathogen. In addition, too high a growth rate disadvantages clonal plants, because the rapid growth of the plant assists the spread of the pathogen due to the increase in susceptible individuals, and leads to lower mortality costs of the dominant strain. However, from a biological requirement, we will have to construct and analyze more complicated models to express the dynamics of real plants. In the future, more details will have to be analyzed, such as the bifurcation condition, the values of the equilibrium, and stability of the equilibrium state. In addition, to analyze real pathogens, it is necessary to change the assumption for the mortality cost, such as a more complex relationship between infection rate and virulence, and we will estimate the parameter value, notably difference in morality costs among strains (Δm), by comparison between our result and available quantitative data.

Supporting Information

S1 Appendix. Simplification of master equation. (PDF)

S2 Appendix. Local stability analysis in mean-field approximation. (PDF)

S3 Appendix. Local stability analysis in pair approximation. (PDF)

Acknowledgments

We thank Ryo Oizumi, Kazunori Sato and our laboratory member for supporting our study.

Author Contributions

Conceived and designed the experiments: YS. Performed the experiments: YS. Analyzed the data: YS. Contributed reagents/materials/analysis tools: YS. Wrote the paper: YS TT.

References

- 1. Slade AJ, Hutchings MJ. Clonal integration and plasticity in foraging behaviour in Glechoma hederacea. The Journal of Ecology. 1987: 1023–1036. doi: 10.2307/2260311
- Marshall C. Sectoriality and physiological organisation in herbaceous plants: an overview. Vegetatio. 1996; 127(1): 9–16. doi: <u>10.1007/BF00054842</u>
- Stuefer JF, Gómez S, Van Mölken T. Clonal integration beyond resource sharing: implications for defence signalling and disease transmission in clonal plant networks. Evolutionary Ecology. 2004; 18 (5–6): 647–667. doi: <u>10.1007/s10682-004-5148-2</u>
- 4. Gibbs A, Harrison B. Effects of viruses on plants. Edward Arnold, London. 1976.
- 5. Jones R. Further studies on losses in productivity caused by infection of annual pasture legumes with three viruses. Crop and Pasture Science. 1992; 43(5): 1229–1241. doi: 10.1071/AR9921229
- 6. Potter L. The effects of white clover mosaic virus on vegetative growth and yield of clones of S. 100 white clover. Plant pathology. 1993; 42(5): 797–805. doi: 10.1111/j.1365-3059.1993.tb01567.x
- Piqueras J, Klimeš L. Demography and modelling of clonal fragments in the pseudoannual plant shape Trientalis europaea L. Plant ecology. 1998; 136(2): 213–227. doi: <u>10.1023/A:1009753515163</u>
- 8. Wennstrom A, Ericson L. Environmental heterogeneity and disease transmission within clones of Lactuca sibirica. Journal of ecology. 1992: 71–77. doi: <u>10.2307/2261064</u>

- Piqueras J. Infection of Trientalis europaea by the systemic smut fungus Urocystis trientalis: disease incidence, transmission and effects on performanc of host ramets. Journal of Ecology. 1999; 87(6): 995–1004. doi: 10.1046/j.1365-2745.1999.00409.x
- Garcia-Guzman G, Burdon J. Impact of the ower smut Ustilago cynodontis (Ustilaginaceae) on the performance of the clonal grass Cynodon dactylon (Gramineae). American journal of botany. 1997; 84 (11): 1565–1565. doi: 10.2307/2446618 PMID: 21708559
- Groppe K, Steinger T, Sanders I, Schmid B, Wiemken A, Boller T. Interaction between the endophytic fungus Epichloë bromicola and the grass Bromus erectus: effects of endophyte infection, fungal concentration and environment on grass growth and owering. Molecular ecology. 1999; 8(11): 1827–1835. doi: <u>10.1046/j.1365-294x.1999.00772.x</u> PMID: <u>10620227</u>
- 12. Pan JJ, Clay K. Infection by the systemic fungus Epichloë glyceriae and clonal growth of its host grass Glyceria striata. Oikos. 2002; 98(1): 37–46. doi: 10.1034/j.1600-0706.2002.980104.x
- González-Jara P, Fraile A, Canto T, García-Arenal F. The multiplicity of infection of a plant virus varies during colonization of its eukaryotic host. Journal of virology. 2009; 83(15): 7487–7494. doi: <u>10.1128/</u> JVI.00636-09 PMID: 19474097
- Folimonova SY, Robertson CJ, Shilts T, Folimonov AS, Hilf ME, Garnsey SM, et al. Infection with strains of Citrus tristeza virus does not exclude superinfection by other strains of the virus. Journal of virology. 2010; 84(3): 1314–1325. doi: <u>10.1128/JVI.02075-09</u> PMID: <u>19923189</u>
- Folimonova SY. Superinfection exclusion is an active virus-controlled function that requires a specific viral protein. Journal of virology. 2012; 86(10): 5554–5561. doi: <u>10.1128/JVI.00310-12</u> PMID: 22398285
- 16. Frank SA. Multiplicity of infection and the evolution of hybrid incompatibility in segmented viruses. Heredity. 2001; 87(5): 522–529. doi: 10.1046/j.1365-2540.2001.00911.x PMID: 11869342
- Levin S, Pimentel D. Selection of intermediate rates of increase in parasite-host systems. American Naturalist. 1981: 308–315. doi: <u>10.1086/283708</u>
- Nowak MA, May RM. Superinfection and the evolution of parasite virulence. Proceedings of the Royal Society of London Series B: Biological Sciences. 1994; 255(1342): 81–89. doi: <u>10.1098/rspb.1994.</u> 0012 PMID: <u>8153140</u>
- May RM, Nowak MA. Coinfection and the evolution of parasite virulence. Proceedings of the Royal Society of London Series B: Biological Sciences. 1995; 261(1361): 209–215. doi: <u>10.1098/rspb.1995.</u> 0138 PMID: 7568274
- Frank SA. MODELS OF PARASITE VIRULENGE. Quarterly Review of Biology. 1996; 71(1): 37–78. doi: 10.1086/419267 PMID: 8919665
- Gandon S, Jansen VA, Van Baalen M. Host life history and the evolution of parasite virulence. Evolution. 2001; 55(5): 1056–1062. doi: 10.1111/j.0014-3820.2001.tb00622.x PMID: 11430642
- Day T. Virulence evolution and the timing of disease life-history events. Trends in Ecology & Evolution. 2003; 18(3): 113–118. doi: <u>10.1016/S0169-5347(02)00049-6</u>
- Day T, Proulx SR. A general theory for the evolutionary dynamics of virulence. The American Naturalist. 2004; 163(4): E40–E63. doi: 10.1086/382548 PMID: 15122509
- van Mölken T, Stuefer JF. Virulence in clonal plants: conflicting selection pressures at work? Evolutionary Ecology. 2008; 22(3): 467–470.
- Koubek T, Herben T. Effect of systemic diseases on clonal integration: modelling approach. Evolutionary Ecology. 2008; 22(3): 449–460. doi: <u>10.1007/s10682-007-9219-z</u>
- Frantzen J. The role of clonal growth in the pathosystem Cirsium arvense-Puccinia punctiformis. Canadian Journal of Botany. 1994; 72(6): 832–836. doi: 10.1139/b94-107
- 27. van Groenendael JM, Kroon HD. Clonal growth in plants: regulation and function. 1990.
- Van Groenendael J, Mony C, Garbey M, Campillo F, El Hamidi A. Introduction to the special issue of Ecological Modelling: "Modelling clonal plant growth: From ecological concepts to mathematics". Ecological Modelling. 2012; 234: 1–2. doi: 10.1016/j.ecolmodel.2012.03.013
- Eriksson O. Dynamics of genets in clonal plants. Trends in Ecology & Evolution. 1993; 8(9): 313–316. doi: 10.1016/0169-5347(93)90237-J
- McCrea KD, Abrahamson WG. Evolutionary impacts of the goldenrod ball gallmaker on Solidago altissima clones. Oecologia. 1985; 68(1): 20–22. doi: <u>10.1007/BF00379467</u>
- Harris TE. Contact interactions on a lattice. The Annals of Probability. 1974; 2(6): 969–988. doi: <u>10.</u> <u>1214/aop/1176996493</u>
- 32. Krone SM. The two-stage contact process. Annals of Applied Probability. 1999: 331–351.
- Li Y, Han D. A two-stage contact process on scale-free networks. Journal of Statistical Physics. 2013; 153(2): 312–324. doi: 10.1007/s10955-013-0832-7

- Satō K, Matsuda H, Sasaki A. Pathogen invasion and host extinction in lattice structured populations. Journal of mathematical biology. 1994; 32(3): 251–268. doi: <u>10.1007/BF00163881</u> PMID: <u>8182357</u>
- Satulovsky JE, Tomé T. Stochastic lattice gas model for a predator-prey system. Physical Review E. 1994; 49(6): 50–73. doi: <u>10.1103/PhysRevE.49.5073</u>
- Haraguchi Y, Sasaki A. The evolution of parasite virulence and transmission rate in a spatially structured population. Journal of Theoretical Biology. 2000; 203(2): 85–96. doi: <u>10.1006/jtbi.1999.1065</u> PMID: <u>10704294</u>
- Ebert D, Mangin KL. The influence of host demography on the evolution of virulence of a microsporidian gut parasite. Evolution. 1997: 1828–1837. doi: 10.2307/2411005
- Matsuda H, Ogita N, Sasaki A, Satō K. Statistical Mechanics of Population The Lattice Lotka-Volterra Model. Progress of theoretical Physics. 1992; 88(6): 1035–1049. doi: <u>10.1143/ptp/88.6.1035</u>
- MacArthur R, Levins R. The limiting similarity, convergence, and divergence of coexisting species. American naturalist. 1967: 377–385. doi: <u>10.1086/282505</u>
- Hutchinson GE. Concluding remarks. In: Cold Spring Harbor symposia on quantitative biology. vol. 22. Cold Spring Harbor Laboratory Press; 1957: 415–427.
- Cheng NH, Su CL, Carter SA, Nelson RS. Vascular invasion routes and systemic accumulation patterns of tobacco mosaic virus in Nicotiana benthamiana. The Plant Journal. 2000; 23(3): 349–362. doi: 10.1046/j.1365-313x.2000.00788.x PMID: 10929128
- Mendgen K, Hahn M. Plant infection and the establishment of fungal biotrophy. Trends in plant science. 2002; 7(8): 352–356. doi: <u>10.1016/S1360-1385(02)02297-5</u> PMID: <u>12167330</u>