

Research Paper: Intra-cerebroventricular Administration of Crocin Attenuates Sleep Deprivation-induced Hyperalgesia in Rats



Faezeh Rezaei¹, Mohammad Reza Saebipour^{1*}, Kazem Ghaemi², Mohammad Mehdi Hassanzadeh-Taheri³, Mohsen Foadoddini⁴, Mehran Hosseini¹

1. Department of Anatomy, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran.

2. Department of Neurosurgery, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran.

3. Department of Anatomy, Cellular and Molecular Research Center, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran.

4. Department of Physiology, Cardiovascular Diseases Research Center, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran.



Citation: Rezaei, F., Saebipour, M. R., Ghaemi, K., Hassanzadeh-Taheri, M. M., Foadoddini, M., & Hosseini, M. (2020). Intra-cerebroventricular Administration of Crocin Attenuates Sleep Deprivation-induced Hyperalgesia in Rats. *Basic and Clinical Neuroscience*, 11(3), 261-268. <http://dx.doi.org/10.32598/bcn.11.2.144.3>

doi <http://dx.doi.org/10.32598/bcn.11.2.144.3>



Article info:

Received: 27 May 2018

First Revision: 20 Jun 2018

Accepted: 06 May 2019

Available Online: 01 May 2020

Keywords:

Secondary insomnia, Pain, Crocin, Analgesic

ABSTRACT

Introduction: Sleep deprivation can cause hyperalgesia and interfere with analgesic treatments. The aim of the present study was to establish an obligatory sleep-abstinence model and also evaluate the effects of Intracerebroventricular (ICV) injection of crocin on pain perception in Wistar rats.

Methods: In this experimental study, 35 adult male Wistar rats were randomly divided into 5 groups (n=7). The intra-ventricular cannulation was done for all rats before sleep deprivation. Sleep deprivation was performed by placing animals on a chamber equipped with an automatic animated conveyor (5 s with an interval of 3 min) for 72 h. Subsequently, the sleep-deprived animals received ICV injection of saline (MOD), Morphine 10 µg (MOR), Crocin 10 µg (Cr10), and Crocin 40 µg (Cr40) using a microsyringe. Besides, a non-sleep-deprived group was allocated as a Control Group (NC) and only received an ICV injection of saline. Fifteen minutes after the ICV injections, pain perception was evaluated by the hot plate test (54±0.4°C).

Results: Compared with the NC group, latency significantly decreased in the MOD group (6.28±0.48 vs. 4.28±0.48, P<0.0001). In comparison with the MOD group, both morphine (8.42±1.53) and crocin (7.60±1.45 for Cr10 and 8.14±0.89 for Cr40) could significantly increase latency in the sleep-deprived animals (P<0.0001). There was no statistically significant difference between the Cr10 and Cr40 (P=0.42), Cr10, and MOR (P=0.059) and Cr40 with MOR (P=0.86) groups.

Conclusion: Our results indicated that crocin could attenuate hyperalgesia induced by sleep deprivation in rats.

* Corresponding Author:

Mohammad Reza Saebipour, MD.

Address: Department of Anatomy, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran.

Tel: +98 (915) 9611261

E-mail: saebipour@gmail.com

Highlights

- Central analgesic effect of Crocin is comparable with morphine.
- Sleep deprivation for 72 hours in a customized less harmful deprivation device decreases pain threshold.
- Social behavior of a group of sleep-deprived rats can be studied using our sleep deprivation device.

Plain Language Summary

Enough sleep is vital for human health and productivity. Chronic pain is a very common condition, which not only is annoying during wake time but also affects the quality and duration of sleep. Treating pain-related sleep problems is a challenging issue for physicians due to the side effects of hypnotics. A pain killer, which concomitantly can support sound sleep, will be very helpful in chronic pain. Crocin is one of the ingredients of saffron with the considerable analgesic property. In this study, we simulated chronic sleep loss in a group of sleep-deprived animals in a specific conveyor belt equipped with a chamber for 72 h. Then, we tested their pain threshold both with and without crocin injection into their brain ventricles. We also compared the effect of crocin with morphine, which is well known for its potent painkilling properties. Our results indicated that saffron may be the main source for developing drugs addressing pain-related sleep problems.

1. Introduction

Pain is a distressing feeling often caused by intense or damaging stimuli to escaping from threats, and an alert indicating there is a minor problem somewhere in the body. Meanwhile, this life-saving alarm is annoying when becomes chronic. Chronic pain with a prevalence of about 46.5% not only decreases the quality of life by depressing mood directly (Elliott, Smith, Penny, Smith, & Chambers, 1999) but also its role in sleep disruption imposes more emotional, physical, and economic burden (Magni, Marchetti, Moreschi, Merskey, & Luchini, 1993; Turk & Rudy, 1988). Chronic pain is considered as an important issue in health care systems and is the most common cause of seeking medical service (Black, 1975).

On the other hand, a great body of evidence shows that insomnia, loss of sleep, or Sleep Deprivation (SD) is associated with several neurological disorders, like memory deficits, Alzheimer disease, Parkinson disease, pain perception, etc. (Atkinson, Ancoli-Israel, Slater, Garfin, & Gillin, 1988). Primary insomnia prevalence is estimated at about 15–30% in different societies (Campbell et al., 2015; Cheatle, Foster, Pinkett, Lesneski, Qu, & Dhingra 2016; Modalen et al., 2016). Comorbid insomnia prevalence in patients suffering from chronic pain is around 50%. On the other hand, depression is concomitant with sleep disturbances in a bidirectional relationship (Augustinavicius, Zanjani, Zakzanis, & Shapiro, 2014; Murphy & Peterson, 2015). These facts highlight the importance of a medical approach, by which

these three common interrelated pathologies can be addressed. Common hypnotics, such as benzodiazepines increase total sleep time; however, they are unable to provide a refreshing sleep and also may increase pain sensitivity. Accordingly, physicians may choose opioids as an alternative (Lamberg, 1999). On the other hand, the chronic use of opioids is a medical challenge. Trazodone, an antidepressant medication, decreases sleep-onset latency, and increases sleep efficiency (O'malley et al., 2000). In spite of these effects of Trazodone, it can lead to muscle contraction and hypersensitivity to touch (Arnold, Keck, & Welge, 2000).

Although research on herbal remedies is still in its early stages, many herbs are thought to provide pain management and improve sleep quality (Uritu et al., 2018). *Crocus sativus* (Sarris, Panossian, Schweitzer, Stough, & Scholey, 2011), commonly known as saffron, is an appropriate candidate, which may target all these three conditions "pain, insomnia, and depression" at the same time. Traditionally, saffron has been used for depression, fear, sleep disorders, pain relief, etc. (Safakhah et al., 2016) which has been approved in different experimental studies (Vahdati Hassani, Naseri, Razavi, Mehri, Abnous, & Hosseinzadeh, 2014; Schmidt, Betti, & Hensel, 2007; Shafiee, Arekhi, Omranzadeh, & Sahebkar, 2018). The analgesic effect of saffron has also been indicated (Ahmad Dar, Brahaman, Tiwari, & Pitre, 2012). Crocin, picrocrocin, and safranal are three main biologically active ingredients of saffron. Crocin has reported responsible for the analgesic activity of saffron. The pain-relieving effect of crocin in orofacial pain has been documented by

injecting crocin into the cerebral fourth ventricle (Tamaddonfard, Tamaddonfard, & Pourbaba, 2015).

However, to the best of our knowledge, there is no evidence addressing the analgesic effect of crocin on sleep deprivation-induced hyperalgesia. The aim of the present study was to investigate the analgesic effect of crocin in a sleep deprivation condition.

2. Methods

2.1. Animals

Healthy male Wistar rats (60 days old) were purchased from the laboratory animal facility in Birjand University of Medical Sciences, Birjand, Iran. The rats were housed in a temperature-controlled room ($22\pm 2^{\circ}\text{C}$) with a 12 h light/dark cycle and had free access to standard laboratory animal diet (Behparvar, Iran) and tap water.

Experimental design and sleep deprivation

The animals were divided randomly into five equal groups ($n=7$) as follows:

- Control (CON): Intra-cerebroventricular (ICV) injection of saline without Sleep Deprivation (SD).
- Model (MOD): ICV injection of saline after SD.
- Morphine (MOR): ICV injection of morphine (40 $\mu\text{g}/\text{rat}$) after SD.
- Crocin 10 $\mu\text{g}/\text{rat}$ (Cr10): ICV injection of crocin (Sigma, USA) (dissolved in saline) at a dose of 10 $\mu\text{g}/\text{rat}$ after SD.
- Crocin 40 $\mu\text{g}/\text{rat}$ (Cr40): ICV injection of crocin (dissolved in saline) at a dose of 40 $\mu\text{g}/\text{rat}$ after SD.

2.2. ICV injection

To deliver the chemical agents into the brain, a permanent guide cannula was implanted in each lateral ventricle of the brain. In brief, each rat was anesthetized with an intraperitoneal injection of a mixture of ketamine (80 mg/kg) and xylazine (10 mg/kg), and a 3-mm stainless-steel guide cannula (25 gauge) was stereotaxically (Stoelting Stereotaxic Apparatus, Wood Dale, IL, USA) placed in the lateral ventricle of the brain.

The stereotaxic coordinates, according to the Paxinos and Watson (1997) were: -1.08 mm posterior to the bregma, 2 mm lateral to the midline, and 4 mm below the top of the skull. The guide cannula was anchored with two screws and dental acrylic. To verify the correctness of the cannula implantation procedure, three rats were subjected to an ICV injection of 5 μl Hematoxylin into each ventricle. Animals were then euthanized by the administration of a high dose of ether, and their brains were removed and placed in a formalin solution (10%). After 24 h, the brains were sectioned coronally (50-100 μm) and viewed under a loop to observe the distribution of Hematoxylin in the lateral ventricles according to the atlas of Paxinos and Watson (1997).

Intraventricular injections of the drugs were performed using a 13-mm length injection needle connected via a 20-cm polyethylene catheter to a 5- μl Hamilton syringe. The intra-cerebral injection was performed over 5 min with a total volume of 5 μl per ventricle. After injections, the injection needle was left in place for a further 30 s to facilitate diffusion of the drug solution. Crocin and morphine were injected 15 min before the Hot Plate (HP) test.

2.3. Sleep deprivation setup

In this study, we designed a Programmable Sleep Deprivation Device (PSDD). The PSDD consisted of a chamber (130 \times 45 cm) equipped with an automatic animated



Figure 1. The Programmable Sleep Deprivation Device (PSDD)

A. PSDD consisted of a water chamber; and B. An automatic animated conveyor belt

conveyor belt. In one side of the cage, a water chamber (25 cm in depth) was placed (Figure 1). When animals did not walk contrary to the conveyor direction, they fell into the water chamber. PSDD was set up on 3 min immobility followed by a 5 s transition time consequently. Accordingly, when rats were immobile more than a certain time (3 min), the conveyor belt moved them, therefore, they had to walk or fall into the water.

2.4. HP Test

Nociceptive sensitivity was assessed using the HP test. The HP surface heat was set at 55°C and a plexiglass cage was fixed over the HP. All rats were placed in the plate 15 min after ICV infusion. The start time was determined, and as soon as the hands or feet were started licking, animal tolerance was recorded. The cutoff point time was considered as 40 s. The apparatus was thoroughly cleaned between trials.

2.5. Statistical analysis

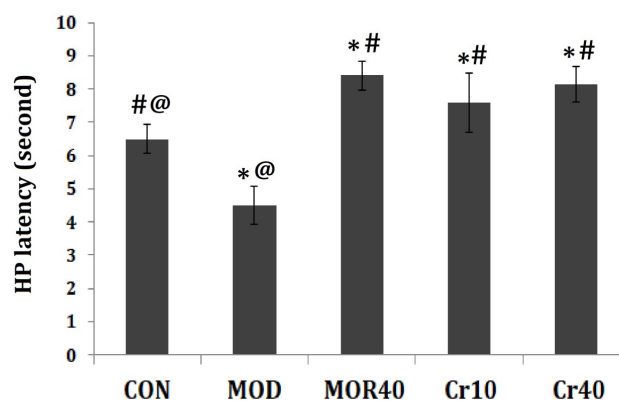
Statistical comparisons were performed using Graph-Pad Prism software v. 5 (GraphPad Software, San Diego, CA, USA). The differences among experimental groups were compared using the One-way ANOVA followed by Tukey's test. Values were expressed as Mean±SD. $P < 0.05$ was considered statistically significant.

3. Results

No death was observed during the research procedure. The Shapiro-Wilks test indicated the normal distribution of data ($P = 0.062$); therefore, the One-way ANOVA was used. The results of the HP test are depicted in Figure 2. Accordingly, SD caused a significant decrease in the latency of rats compared with non-SD animals (4.2 ± 0.48 vs. 6.2 ± 0.47 , $P < 0.0001$). As expected, the latency of SD rats receiving ICV injection of morphine (40 µg/rat) significantly improved even more than the control non-SD group (8.4 ± 0.53 , $p < 0.0001$). Likewise morphine, crocin at both doses (10 and 40 µg/rat) significantly ($P < 0.0001$) increased latency of the SD rats (7.6 ± 0.45 and 8.14 ± 0.89 respectively). There was no significant difference between crocin doses regarding latency in the HP test ($P = 0.42$). Compared with the morphine-treated animal, there was no statistical difference in the latency of rats received Cr10 ($P = 0.059$) and Cr40 ($P = 0.86$).

4. Discussion

In this study, we measured the pain threshold in sleep-deprived rats in sham or ICV crocin-injected groups. The aim of sleep deprivation in this study was to produce a natural sleep deprivation model, not a total sleep deprivation. Fortunately, no rats died due to exposure to the PSDD. Sleep-deprived rats showed decreased latency to noxious thermal stimulus compared with the control group. All rats chose the farthest locations from the water chamber, indicating their awareness about the rules of



NEURSCIENCE

Figure 2. Effects of Intracerebroventricular (ICV) injection of crocin on sleep deprivation-induced hyperalgesia in rats

Values are presented as Mean±Standard Deviation.

CON: Control rats with no intervention; MOD: Sleep-deprived model group; MOR40: Sleep-deprived rats that received an ICV injection of morphine (40 µg/rat); Cr10 and Cr40: Sleep-deprived rats that received ICV injection of crocin at the doses of 10 µg/rat and 40 µg/rat, respectively

* $P < 0.001$ versus CON; # $P < 0.001$ versus MOD; @ $P < 0.05$ versus MOR40

the PSDD. PSDD can be used to measure pain threshold for many rats at the same time, which reduces the rats' unwanted anxiety due to animal isolation.

Consistent with the results of crocin injection to the third ventricle of the rats (Erfanparast, Tamaddonfard, Taati, & Dabbaghi, 2015), crocin injection to the lateral ventricles also showed a central analgesic effect. It has shown that the analgesic mechanism of crocin probably is not mediated by opioid receptors, because its effect is not affected by naloxone (Tamaddonfard & Hamzeh-Gooshchi, 2010). Crocin plays an antagonistic role against the long-term potentiation inhibitory effect of ethanol on the N-methyl-D-aspartate (NMDA) receptor (Abe & Saito, 2000; Abe, Sugiura, Shoyama, & Saito, 1998). Such an effect has been reported about memory impairment induced by scopolamine, as well (Ghadami & Pourmotabbed, 2009). Oral administration of aqueous extract of saffron has increased dopamine and glutamate neurotransmission in the brain (Ettehad et al., 2013), whereas crocetin has found with an anti-glutamatergic effect (Berger, Hensel, & Nieber, 2011). ICV infusion of crocin exerts antiepileptic effect when it is injected in combination with diazepam, which may be due to its effects on the GABAA receptors (Tamaddonfard et al., 2012). The molecular mechanism of action of crocin in the brain seems more complicated considering that microglial cells are also targeted by active components of saffron (Nam et al., 2010; Xie, Huo, & Tang, 2009).

In a triple-blinded clinical trial, oral administration of capsules containing 250 mg of saffron has shown a mild analgesic effect for labor pain (%11.8 reduction in pain perception) (Azhari, Ahmadi, Rakhshandeh, Jafarzadeh, & Mazlom, 2014). In a rat model for chronic constriction injury, 30 µg/kg of oral crocin but not 15 mg/kg decreased the neuropathic pain (Safakhah et al., 2016). In a rat model of neuropathic pain, 50 and 100 mg/kg of intraperitoneal injection of saffron extract in combination with 3 mg/kg amitriptyline, showed a synergistic analgesic effect (Amin, Hosseini, & Hosseinzadeh, 2017). Several studies have focused on analgesic properties of saffron extract in rat models of neuropathic pain (Amin & Hosseinzadeh, 2012), however, no comparison has been made with morphine.

Crocetin, even at high doses, cannot cross the Blood-Brain Barrier (BBB), but crocin is converted to crocetin in the gastrointestinal system after oral administration and crocetin can cross the BBB (Hosseini, Razavi, & Hosseinzadeh, 2018; Lautenschläger et al., 2015; Xi, Qian, Du, & Fu, 2007; Zhang et al., 2017). Therefore, the potential central analgesic properties of crocetin, as a

metabolite of crocin, should be investigated. Interestingly, according to the results of a pharmacokinetic study, crocetin at an oral dosage of 22.5 mg/kg showed a rapid absorption and a mean half-life of about 7 h with no serious side effects (Umigai et al., 2011). However, it is still unknown that crocetin at which dose can reach the brain.

In conclusion, the analgesic effect following an ICV infusion of crocin can be a promising approach for more sophisticated research, which hopefully may provide a translational therapeutic strategy for chronic pain and concomitant comorbidities.

Ethical Considerations

Compliance with ethical guidelines

All animal procedures were conducted in accordance with the guide for the laboratory animals' care and usage of Birjand University of Medical Sciences, Birjand, Iran (Ethics Code: IR.BUMS.REC.1396.47).

Funding

This article is based on an MSc. the thesis of Faezeh Rezaei in the Department of Anatomy, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, also this article was supported by Birjand University of Medical Sciences (Grant No.455233).

Authors' contributions

Performed experiments: Faezeh Rezaei; Designed the study: Mohammad Reza Saebipour and Kazem Ghaemi, Developed sleep deprivation machine and provide animal facilities: Mohsen Foadoddini; Analyzed the data and co-wrote the first manuscript draft: Mohammad Mehdi Hassanzadeh-Taheri and Mehran Hosseini; Read and approved the final version of the manuscript: All authors.

Conflict of interest

The authors declared that there are no conflicts of interest.

Acknowledgments

The authors thank Professor Mahmoud Hosseini and Psychiatry and Behavioral Research Center, Mashhad University of Medical Sciences for his kindly technical assistance.

Reference

- Abe, K., & Saito, H. (2000). Effects of saffron extract and its constituent crocin on learning behaviour and long-term potentiation. *Phytotherapy Research*, 14(3), 149-52. [DOI:10.1002/(SICI)1099-1573(200005)14:3<149::AID-PTR665>3.0.CO;2-5]
- Abe, K., Sugiura, M., Shoyama, Y., & Saito, H. (1998). Crocin antagonizes ethanol inhibition of NMDA receptor-mediated responses in rat hippocampal neurons. *Brain Research*, 787(1), 132-8. [DOI:10.1016/S0006-8993(97)01505-9]
- Ahmad Dar, R., Brahaman, P. K., Tiwari, S., & Pitre, K. S. (2012). Indirect electrochemical analysis of crocin in phytochemical sample. *Journal of Chemistry*, 9(2), 918-25. [DOI:10.1155/2012/967079]
- Amin, B., & Hosseinzadeh, H. (2012). Evaluation of aqueous and ethanolic extracts of saffron, *Crocus sativus* L., and its constituents, safranal and crocin in allodynia and hyperalgesia induced by chronic constriction injury model of neuropathic pain in rats. *Fitoterapia*, 83(5), 888-95. [DOI:10.1016/j.fitote.2012.03.022] [PMID]
- Amin, B., Hosseini, S., & Hosseinzadeh, H. (2017). Enhancement of antinociceptive effect by co-administration of amitriptyline and *Crocus sativus* in a rat model of neuropathic pain. *Iranian Journal of Pharmaceutical Research*, 16(1), 187-200. [DOI:10.22037/IJPR.2017.2004] [PMID] [PMCID]
- Arnold, L. M., Keck, P. E., & Welge, J. A. (2000). Antidepressant treatment of fibromyalgia: A meta-analysis and review. *Psychosomatics*, 41(2), 104-13. [DOI:10.1176/appi.psy.41.2.104] [PMID]
- Atkinson, J., Ancoli-Israel, S., Slater, M. A., Garfin, S. R., & Gillin, C. (1988). Subjective sleep disturbance in chronic back pain. *The Clinical Journal of Pain*, 4(4), 225-32. [DOI:10.1097/00002508-198812000-00007]
- Augustinavicius, J. L. S., Zanjani, A., Zakzani, K. K., & Shapiro, C. M. (2014). Polysomnographic features of early-onset depression: A meta-analysis. *Journal of Affective Disorders*, 158, 11-8. [DOI:10.1016/j.jad.2013.12.009] [PMID]
- Azhari, S., Ahmadi, S., Rakhshandeh, H., Jafarzadeh, H., & Mazlom, S. R. (2014). [Evaluation of the effect of oral saffron capsules on pain intensity during the active phase of labor (Persian)]. *The Iranian Journal of Obstetrics, Gynecology and Infertility*, 17(115), 1-10. <http://eprints.mums.ac.ir/4201/>
- Berger, F., Hensel, A., & Nieber, K. (2011). Saffron extract and trans-crocin inhibit glutamatergic synaptic transmission in rat cortical brain slices. *Neuroscience*, 180, 238-47. [DOI:10.1016/j.neuroscience.2011.02.037] [PMID]
- Black, R. G. (1975). The chronic pain syndrome. *Surgical Clinics of North America*, 55(4), 999-1011. [DOI:10.1016/S0039-6109(16)40697-3]
- Campbell, C. M., Buenaver, L. F., Finan, P., Bounds, S. C., Redding, M., & McCauley, L., et al. (2015). Sleep, pain catastrophizing, and central sensitization in knee osteoarthritis patients with and without insomnia. *Arthritis Care & Research*, 67(10), 1387-96. [DOI:10.1002/acr.22609] [PMID] [PMCID]
- Cheatle, M. D., Foster, S., Pinkett, A., Lesneski, M., Qu, D., & Dhingra, L. (2016). Assessing and managing sleep disturbance in patients with chronic pain. *Anesthesiology Clinics*, 34(2), 379-93. [DOI:10.1016/j.anclin.2016.01.007] [PMID]
- Elliott, A. M., Smith, B. H., Penny, K. I., Smith, W. C., & Chambers, W. A. (1999). The epidemiology of chronic pain in the community. *The Lancet*, 354(9186), 1248-52. [DOI:10.1016/S0140-6736(99)03057-3]
- Erfanparast, A., Tamaddonfard, E., Taati, M., & Dabbaghi, M. (2015). Effects of crocin and safranal, saffron constituents, on the formalin-induced orofacial pain in rats. *Avicenna Journal of Phytomedicine*, 5(5), 392-402. [PMID] [PMCID]
- Ettehad, H., Mojabi, S. N., Ranjbaran, M., Shams, J., Sahraei, H., & Hedayati, M., et al. (2013). Aqueous extract of saffron (*Crocus sativus*) increases brain dopamine and glutamate concentrations in rats. *Journal of Behavioral and Brain Science*, 3(3), 315-9. [DOI:10.4236/jbbs.2013.33031]
- Ghadami, M. R., & Pourmotabbed, A. (2009). [The effect of crocin on scopolamine induced spatial learning and memory deficits in rats (Persian)]. *Physiology and Pharmacology*, 12(4), 287-95. <http://ppj.phypha.ir/article-1-462-en.pdf>
- Hosseini, A., Razavi, B. M., & Hosseinzadeh, H. (2018). Pharmacokinetic properties of saffron and its active components. *European Journal of Drug Metabolism and Pharmacokinetics*, 43(4), 383-90. [DOI:10.1007/s13318-017-0449-3] [PMID]
- Lamberg, L. (1999). Chronic pain linked with poor sleep; exploration of causes and treatment. *JAMA*, 281(8), 691-2. [DOI:10.1001/jama.281.8.691-JMN0224-2-1] [PMID]
- Lautenschläger, M., Sendker, J., Hüwel, S., Galla, H., Brandt, S., & Düfer, M., et al. (2015). Intestinal formation of trans-crocin from saffron extract (*Crocus sativus* L.) and in vitro permeation through intestinal and blood brain barrier. *Phytomedicine*, 22(1), 36-44. [DOI:10.1016/j.phymed.2014.10.009] [PMID]
- Magni, G., Marchetti, M., Moreschi, C., Merskey, H., & Luchini, S. R. (1993). Chronic musculoskeletal pain and depressive symptoms in the National Health and Nutrition Examination. I. Epidemiologic follow-up study. *Pain*, 53(2), 163-8. [DOI:10.1016/0304-3959(93)90076-2] [PMID]
- Modalen, E., Hand, M., Remeniuk, B., Perry, S., Swedberg, L., & Pressman, A., et al. (2016). (190) Effects of sleep continuity disruption on cold pain tolerance and accompanying pain catastrophizing in healthy, good sleepers. *The Journal of Pain*, 17(4 Suppl), S23. [DOI:10.1016/j.jpain.2016.01.093]
- Murphy, M. J., & Peterson, M. J. (2015). Sleep disturbances in depression. *Sleep Medicine Clinics*, 10(1), 17-23. [DOI:10.1016/j.jsmc.2014.11.009] [PMID] [PMCID]
- Nam, K. N., Park, Y. M., Jung, H. J., Lee, J. Y., Min, B. D., & Park, S. U., et al. (2010). Anti-inflammatory effects of crocin and crocetin in rat brain microglial cells. *European Journal of Pharmacology*, 648(1-3), 110-6. [DOI:10.1016/j.ejphar.2010.09.003] [PMID]
- O'malley, P. G., Balden, E., Tomkins, G., Santoro, J., Kroenke, K., & Jackson, J. L. (2000). Treatment of fibromyalgia with antidepressants. *Journal of General Internal Medicine*, 15(9), 659-66. [DOI:10.1046/j.1525-1497.2000.06279.x] [PMID] [PMCID]
- Paxinos, G., & Watson, C. (1997). *Atlas of anatomy of rat brain. The Rat Brain in Stereotaxic Coordinates*, 3rd ed. San Diego, Calif: Academic Press Inc.
- Safakhah, H. A., Taghavi, T., Rashidy-Pour, A., Vafaei, A. A., Sokhanvar, M., & Mohebbi, N., et al. (2016). Effects of saffron (*Crocus sativus* L.) stigma extract and its active constituent crocin on neuropathic pain responses in a rat model of chronic constrict-

- tion injury. *Iranian Journal of Pharmaceutical Research*, 15(1), 253-61. [PMID] [PMCID]
- Sarris, J., Panossian, A., Schweitzer, I., Stough, C., & Scholey, A. (2011). Herbal medicine for depression, anxiety and insomnia: A review of psychopharmacology and clinical evidence. *European Neuropsychopharmacology*, 21(12), 841-60. [DOI:10.1016/j.euroneuro.2011.04.002] [PMID]
- Schmidt, M., Betti, G., & Hensel, A. (2007). Saffron in phytotherapy: Pharmacology and clinical uses. *Wiener Medizinische Wochenschrift*, 157(13-14), 315-9. [DOI:10.1007/s10354-007-0428-4] [PMID]
- Shafiee, M., Arekhi, S., Omranzadeh, A. R., & Sahebkar, A. H. (2018). Saffron in the treatment of depression, anxiety and other mental disorders: Current evidence and potential mechanisms of action. *Journal of Affective Disorders*, 227, 330-7. [DOI:10.1016/j.jad.2017.11.020] [PMID]
- Tamaddonfard, E., & Hamzeh Gooshchi, N. (2010). Effects of intraperitoneal and intracerebroventricular injection of crocin on acute corneal pain in rats. *Phytotherapy Research*, 24(10), 1463-7. [DOI:10.1002/ptr.3169] [PMID]
- Tamaddonfard, E., Hamzeh Gooshchi, N., & Seidnejad-Yamchi, S. (2012). Central effect of crocin on penicillin-induced epileptiform activity in rats. *Pharmacological Reports*, 64(1), 94-101. [DOI:10.1016/S1734-1140(12)70735-1]
- Tamaddonfard, E., Tamaddonfard, S., & Pourbaba, S. (2015). Effects of intra-fourth ventricle injection of crocin on capsaicin-induced orofacial pain in rats. *Avicenna Journal of Phytomedicine*, 5(5), 450-7. [PMID] [PMCID]
- Turk, D. C., & Rudy, T. E. (1988). Toward an empirically derived taxonomy of chronic pain patients: Integration of psychological assessment data. *Journal of Consulting and Clinical Psychology*, 56(2), 233-8. [DOI:10.1037/0022-006X.56.2.233] [PMID]
- Umigai, N., Murakami, K., Ulit, M., Antonio, L., Shirotori, M., & Morikawa, H., et al. (2011). The pharmacokinetic profile of crocetin in healthy adult human volunteers after a single oral administration. *Phytomedicine*, 18(7), 575-8. [DOI:10.1016/j.phymed.2010.10.019] [PMID]
- Uritu, C. M., Mihai, C. T., Stanciu, G. D., Dodi, G., Alexa-Stratulat, T., & Luca, A., et al. (2018). Medicinal plants of the family Lamiaceae in pain therapy: A review. *Pain Research & Management*, 2018, 7801543. [DOI:10.1155/2018/7801543] [PMID] [PMCID]
- Vahdati Hassani, F., Naseri, V., Razavi, B. M., Mehri, S., Abnous, Kh., & Hosseinzadeh, H. (2014). Antidepressant effects of crocin and its effects on transcript and protein levels of CREB, BDNF, and VGF in rat hippocampus. *Daru*, 22(1), 16. [DOI:10.1186/2008-2231-22-16] [PMID] [PMCID]
- Xi, L., Qian, Z., Du, P., & Fu, J. (2007). Pharmacokinetic properties of crocin (crocetin digentiobiose ester) following oral administration in rats. *Phytomedicine*, 14(9), 633-6. [DOI:10.1016/j.phymed.2006.11.028] [PMID]
- Xie, Y. F., Huo, F. Q., & Tang, J. S. (2009). Cerebral cortex modulation of pain. *Acta Pharmacologica Sinica*, 30(1), 31-41. [DOI:10.1038/aps.2008.14] [PMID] [PMCID]
- Zhang, Y., Fei, F., Zhen, L., Zhu, X., Wang, J., & Li, S., et al. (2017). Sensitive analysis and simultaneous assessment of pharmacokinetic properties of crocin and crocetin after oral administration in rats. *Journal of Chromatography B*, 1044-1045, 1-7. [DOI:10.1016/j.jchromb.2016.12.003] [PMID]

